



manual

PREPARING FOR
HIGHLY PATHOGENIC
AVIAN INFLUENZA



PREPARING FOR HIGHLY PATHOGENIC AVIAN INFLUENZA

V. Martin, A. Forman, J. Lubroth
Animal Production and Health Division
FAO, Rome, Italy

Acknowledgements

The manual "Preparing for highly pathogenic avian influenza" was prepared by the Animal Production and Health Division of FAO.

FAO would like to thank the World Organisation for Animal Health (OIE) for its contribution in reviewing and approving the document.

Special thanks to Phil Harris and Cecilia Murguia for their help with editing this manual.

The designations employed and the presentation of material in this information product do not imply the expression of any opinion whatsoever on the part of the Food and Agriculture Organization of the United Nations concerning the legal or development status of any country, territory, city or area or of its authorities, or concerning the delimitation of its frontiers or boundaries.

ISBN 92-5-105548-3

All rights reserved. Reproduction and dissemination of material in this information product for educational or other non-commercial purposes are authorized without any prior written permission from the copyright holders provided the source is fully acknowledged. Reproduction of material in this information product for resale or other commercial purposes is prohibited without written permission of the copyright holders. Applications for such permission should be addressed to the Chief, Electronic Publishing Policy and Support Branch, Information Division, FAO, Viale delle Terme di Caracalla, 00100 Rome, Italy or by e-mail to copyright@fao.org

Contents

1. Introduction	1
2. Avian influenza and the virus that causes it	3
2.1. Clinical signs	3
2.2 Gross pathology	5
2.3 Differential diagnosis	6
3. The risk of introduction and dissemination of avian influenza	7
3.1 Risk of introduction by migrating birds	7
3.2 Risk of importation	8
3.3 Risk of spread from infected poultry	8
3.4 Virus survival in the environment	9
4. Preparing for an outbreak	11
4.1 Early detection	11
4.1.1 Wild bird surveillance	11
4.1.2 Domestic poultry surveillance	11
4.2. Rapid response	13
4.2.1 Disease surveillance	13
4.2.2 Planning for avian influenza control	14
4.3 Avian influenza control strategies	15
4.3.1 Culling	15
4.3.2 Financial support	24
4.3.3 Vaccination	25
4.4 Management of disease control	27
4.5 Laboratory diagnosis	29
4.5.1 Samples	29
4.5.2 Laboratory tests	30
4.5.3 Communication and public awareness	31
4.5.4 Human health and safety considerations	31
5. Prevention and biosecurity	33
5.1 Restricted access means keeping the disease out	33
5.2 Clean areas mean healthy chickens, geese and ducks	33
5.3 Buy healthy: keep healthy	34
5.4 Use of clean equipment – keeping disease out	34
5.5 Report early signs of a problem that could be devastating	34

5.6 A period of rest	35
5.7 Vaccination against avian influenza or other diseases	35
5.8 Compartmentalisation	35
Annex 1: Selected references for further information	39
Annex 2: Sample - Tender document for inactivated avian influenza vaccines	41
Annex 3: OIE/FAO reference laboratories and experts for avian influenza	43
Annex 4: Information for shipping international diagnostic specimens	45
Annex 5: Criteria for defining infected areas and disease control zones	51
Annex 6: Leaflet example	53
Annex 7: Avian influenza vaccine producers and suppliers	55

1. Introduction

Highly pathogenic avian influenza (HPAI) has seriously affected poultry farmers whenever and wherever it has appeared. Historically, outbreaks of HPAI have occurred in all continents. The current avian influenza epidemic, caused principally by the H5N1 strain, has been continuing since it was first recognised in the Republic of Korea in December 2003. Despite concerted attempts at control, Thailand, Viet Nam, Indonesia and China are still recording outbreaks and there are major control campaigns being implemented in Viet Nam and Indonesia. Some outbreaks are still being recorded in Cambodia. Lao People's Democratic Republic, where a few outbreaks occurred, is now apparently free at the time of writing.

However, two circumstances have increased international concern about the behaviour and spread of this disease. The first is that, at the time of publication of this manual, more than 230 cases of transmission of the virus to humans have been recorded, with an approximately 50 percent fatality rate. There is increasing concern that in the future the virus will adapt to enable human-to-human transmission with ease and result in a global human influenza pandemic if not contained in time. Secondly, between August and December 2005, the disease has spread over wide geographical area and was reported in the Russia Federation, Turkey, Croatia, Romania and Ukraine. In February 2006, the disease was reported on the African continent with the first notification of the HPAI H5N1 strain in Nigeria. The occurrence of the disease in Africa is of major concern, putting at immediate risk the livelihood of millions of people relying on poultry production for income generation and sources of protein¹. If this situation runs out of control, it will have a devastating impact on the poultry population in the region and will increase the exposure of humans to the virus.

It is difficult to predict the severity of either of these threats. The virus has been present in China since at least 1996 and probably disseminated to Southeast Asian countries at least some months before it developed into the epidemic beginning in 2003. There has been enormous opportunity for the virus to infect humans, which has probably occurred much more than has been identified, and yet adaptation for human-to-human transmission has not yet occurred. However, this does not imply that it will not occur and the greater the shedding of virus from infected poultry the greater the risk of adaptation leading to a human pandemic. Similarly, despite opportunities for the virus to spread in wild birds, to date it has caused minimal disease in poultry outside Southeast and East Asia. Again, whether this will occur in the future is difficult to predict.

¹ In this document, poultry is referred as 'all birds reared or kept in captivity for the production of meat or eggs for consumption, for the production of other commercial products, for restocking supplies of game, or for breeding these categories of birds'. This definition has been recently adopted by OIE in the 2005 edition of the *Terrestrial Animal Health Code*, Chapter on Avian Influenza. (OIE, 2005a)

Countries may be under threat of introduction of avian influenza through exposure of poultry to wild birds, especially waterfowl. They may also be at risk from introduction of infected or contaminated poultry, poultry products or fomites. This represents a threat to poultry industries around the world, to people's livelihoods, and to a source of high quality and inexpensive protein complement to diets. Human populations are also at risk if an influenza pandemic occurs.

This manual is intended to assist national animal health authorities and other stakeholders consider the needs for preparing for a possible incursion of HPAI, to detect disease at the earliest opportunity and to respond as rapidly as possible to contain the disease.

The international community has a vested interest in minimising the spread of this disease. FAO, together with World Organization for Animal Health (OIE) and World Health Organization (WHO), are the key agencies for coordinating an international response to the threat. This manual also assists countries in determining means of obtaining outside assistance to improve their preparedness for highly pathogenic avian influenza and its detection.

2. Avian influenza and the virus that causes it

Avian influenza is caused by influenza viruses which are common in wild birds and occasionally infect poultry. When poultry are infected, they may have no disease, mild disease or very severe disease. Chickens, quail and turkeys are especially susceptible while ducks more commonly show no disease, but act as a reservoir for the virus. Other poultry species, including guinea fowl and pheasants, and also ostriches, can become affected. While generally wild birds are not clinically affected by the AI viruses that they carry, occasionally they can suffer disease. This has been observed in Asia and parts of Europe as a result of infection with the H5N1 virus and may be because they have become infected with the virus from domestic birds.

Influenza viruses have two main surface antigens, haemagglutinin (H) and neuraminidase (N). There are many H and N subtypes, but historically highly pathogenic avian influenza viruses have been either H5 or H7, and to a lesser degree H9. The avian influenza virus currently causing the major epidemic in Asia is H5N1, with some occurrences of H5N2 being reported as well. The virus causing disease in Pakistan in recent years is H7N3 and H9N2. AI viruses are also classified by pathotype – highly pathogenic (HPAI) and low pathogenic (LPAI) – a biological characteristic of the virus' virulence in chickens. Currently, the pathotype definition has been expanded to include the genetic sequence coding for basic amino acids in the cleavage site of the H protein. All AI viruses that have these sequences at the critical site are considered notifiable and the viruses are denoted as HPNAI (highly pathogenic notifiable avian influenza) and LPNAI.

2.1. CLINICAL SIGNS

The clinical signs of AI infection are variable and influenced greatly by the virulence of the viruses involved, the species infected, age, concurrent viral or bacterial disease and the environment. The virulence exhibited in chickens can vary during an outbreak.

Infection with non-pathogenic viruses

- No clinical signs in infected birds, with seroconversion.
- Some of these viruses have the potential to become virulent through genetic mutation.

Infection with low or mild pathogenic viruses

- Clinical signs in chickens and turkeys range from inapparent to mild or severe respiratory disease and can be confused with infectious laryngotracheitis and other respiratory tract infections.
- Mortality ranges from 3 percent in caged hens (layers) to 15 percent in meat chickens (broilers).

- Egg production in layers can drop sometimes to 45 percent of the expected egg yield of a large flock, returning to normal levels of production in 2–4 weeks.
- Mutation to virulence has been demonstrated in outbreaks.

Infection with highly pathogenic viruses

- In peracute cases involving sudden death, as in the 2004-5 outbreak in Viet Nam, clinical signs may not be seen and mortalities occur within hours after onset of depression. Overall mortality rates for peracute/acute cases nearing 100 percent have been reported.
- In acute cases, mortalities occur as early as 24 hours after the first signs of the disease, and frequently within 48 hours. In other cases, more diverse visible signs are seen and mortalities can be delayed for as long as a week.
- Clinical signs in chickens and turkeys include severe respiratory distress with excessively watery eyes and sinusitis, cyanosis of the combs, wattle and shanks, oedema of the head and eyelids, ruffled feathers, diarrhoea and nervous signs.
- Eggs laid after the onset of illness frequently have no shells.
- Some severely affected hens may recover, but rarely come back into lay.

The disease in turkeys is similar to that in chickens, but is often complicated by secondary bacterial infections such as those due to fowl cholera (*Pasteurella multocida*), turkey coryza (*Hemophilus gallinarum*), or colibacillosis (*Escherichia coli*).



CREDIT: USDA

Oedematous cyanotic comb and wattle of a chicken with Highly Pathogenic Avian Influenza



CREDIT: USDA

Oedematous wattles

2.2 GROSS PATHOLOGY

In many cases, poultry dying from the peracute form of the disease lack visible gross pathological lesions. With acute infections in chickens, there is severe lung congestion, haemorrhage and oedema in dead chickens; other organs and tissues appear normal. More varied visible lesions are seen in chickens surviving 3 to 5 days, including congestion and/or cyanosis of the comb and wattles and swollen heads. The changes in the combs and wattles progress to depressed areas of dark red to blue areas of ischaemic necrosis. Internally, the characteristics of acute infections with viruses causing HPAI are haemorrhagic, necrotic, congestive and transudative changes. The oviducts and intestines often have severe haemorrhagic changes.

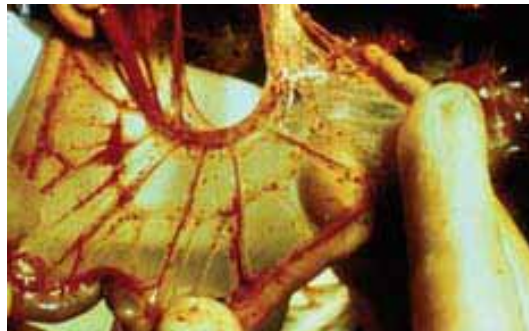
As the disease progresses, the pancreas, liver, spleen, kidney and lungs can display yellowish necrotic foci. Haemorrhages (petechial and ecchymotic) cover the abdominal fat, serosal surfaces and peritoneum. The peritoneal cavity is frequently filled with yolk from ruptured ova, associated with severe inflammation of the airsacs and peritoneum in birds that survive 7–10 days. Haemorrhages may be present in the proventriculus, particularly at the junction with the ventriculus (gizzard).

In cases due to mild pathogenic avian influenza viruses, lesions may be seen in the sinuses characterised by catarrhal, serofibrinous, mucopurulent or caseous inflammation. The tracheal mucosa may be oedematous with exudates varying from serous to caseous. The airsacs may be thickened and have fibrinous to caseous exudates. Catarrhal to fibri-

Oedematous wattles dissected



Haemorrhage in the mesentery of the small intestine



Large haemorrhages in the fat on the serosal surfaces of the abdominal organs

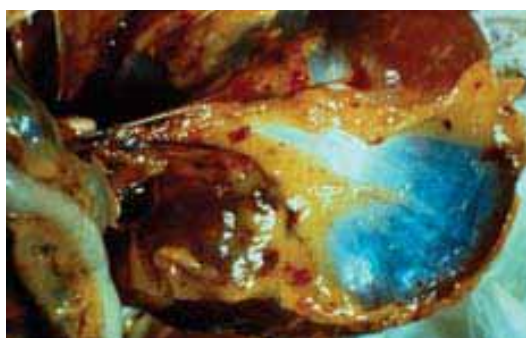


Haemorrhage in the muscle and the fat around the heart



CREDIT: USDA

Ecchymotic haemorrhages in the proventriculus.



CREDIT: USDA

Haemorrhage in the muscle and fat around the gizzard.

nous peritonitis and egg yolk peritonitis may be seen. Catarrhal to fibrinous enteritis may be seen in the caeca and/or intestine, particularly in turkeys. Exudates may be seen in the oviducts of laying birds (Easterday *et al.* 1997). Histopathological lesions seen in the gross changes described above are not definitive for HPAI, although vasculitis in the brain and other organs may be highly suggestive of the disease.

2.3 DIFFERENTIAL DIAGNOSIS

The following diseases must be considered in the differential diagnosis of virulent AI:

- Other diseases causing sudden high mortality
 - Newcastle disease
 - Infectious laryngotracheitis
 - Duck plague
 - Acute poisonings
- Other diseases causing swelling of the combs and wattles
 - Acute fowl cholera and other septicaemic diseases
 - Bacterial cellulitis of the comb and wattles

Less severe forms of the disease may be confused with, or complicated by, many other diseases with respiratory or enteric signs. HPAI should be suspected in any disease outbreak in poultry that persists despite the application of preventive and therapeutic measures for other diseases or when the epidemiological context is highly suggestive of the introduction of the infection.

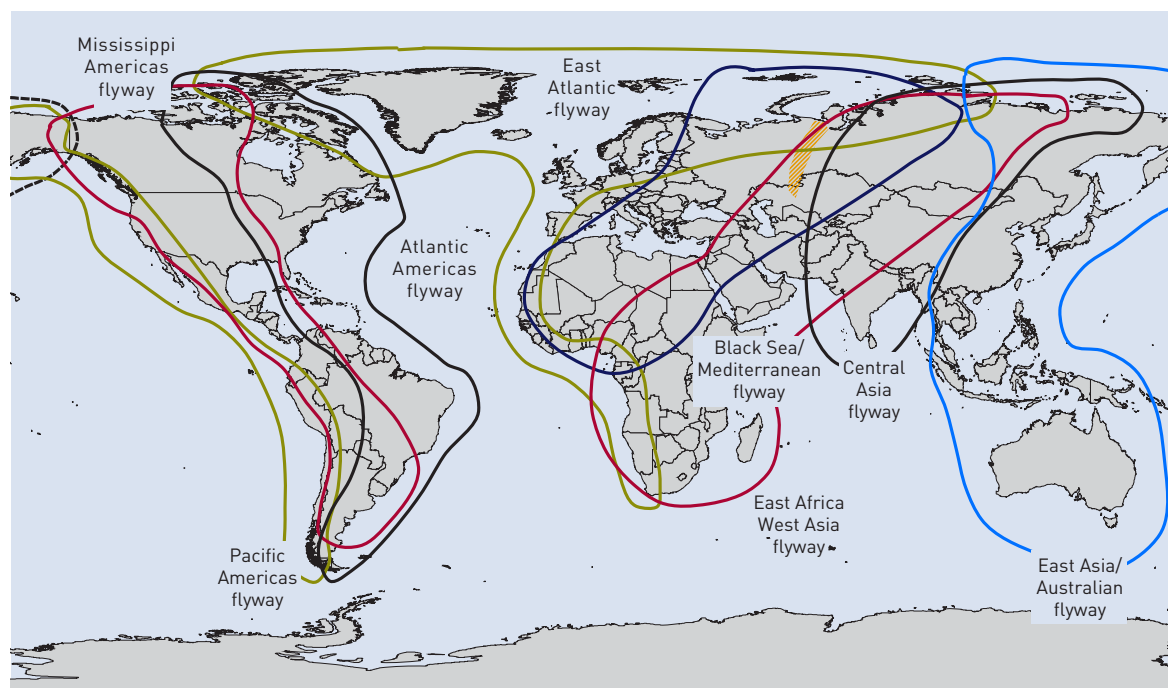
3. The risk of introduction and dissemination of avian influenza

3.1 RISK OF INTRODUCTION BY MIGRATING BIRDS

Migration of water birds represents the main risk of carrying AI viruses over long distances and provides a complex network because different bird flyways overlap geographically.

Birds infected with AI virus can shed virus for up to one month. Birds from different regions intermingle with each other in areas where large water bodies attract them and transmission of viruses can occur between them. The outcome is that potentially viruses can be transmitted from infected countries in Southeast and East Asia to Central Asia, Eastern Europe, the Middle East and Africa, North and South America. In the course of the current epidemic, a large number of wild bird species have been found dead, with AI virus type

FIGURE 1
Major flyways of migratory birds (wader species)



FAO 2005.
All right reserved. Compiled by FAO AGAH, EMPRES Programme.
Flyways: Wetlands International

H5N1 being isolated. Recent findings show that the virus can be isolated from other bird species without signs of disease. However, it is not yet fully determined which species are implicated in the long distance introduction of the virus and its transmission to poultry.

If infection occurs in domestic poultry, it is likely to be in areas where wild waterfowl congregate and where poultry are not in bird-proof sheds. Transmission of virus can occur from contaminated water as well as from direct contact of wild birds with poultry. Good biosecurity therefore requires physical barriers erected between poultry and wild birds and also the provision of clean or treated water before being provided to poultry.

An important component of preparedness in the context of the current avian influenza epidemic is to identify wild bird migratory patterns, timing and destination sites and to assess the risk of close contact with domestic poultry providing an entry point for establishment of avian influenza.

3.2 RISK OF IMPORTATION

Many countries currently impose bans on the importation of poultry and products from infected countries with notifiable AI. Given the potential for transboundary spread of the disease, it would be wise to take great care with all poultry products, especially those that can carry the virus. Live birds represent by far the greatest risk but dressed carcasses of infected birds, eggs from infected hens, poultry waste and fomites contaminated with faeces can all be a source of infection. A detailed risk assessment for each poultry product has been carried out by the European Food Safety Agency (EFSA) and is available on their web site (see Annex 1).

It should be recognised that illegal movement of live birds also represents a risk that will not be mitigated by imposing bans on legal importation.

3.3 RISK OF SPREAD FROM INFECTED POULTRY

To prevent further spread of H5N1, surveillance in poultry as well as in wild birds should be strengthened in countries at immediate risk, especially where birds come to rest along their migrating routes. Resources should be focused on the reduction of close contacts between humans, poultry and wildlife through better management practices and improved biosecurity practices in poultry production enterprises, especially those that are small and 'open-air'- where poultry and waterfowl mingle with wild birds. The influenza viruses are easily spread by fomites and generally survive well in water. Furthermore, certain species of ducks are able to carry influenza viruses without exhibiting any clinical signs of disease. Juvenile ducks have the highest rates of infection and shedding. High titres of virus occur in late summer, when birds leave their northern breeding areas, although these titres decrease as birds continue southwards.

Once a highly pathogenic virus (HPAI) has been recognised in the marketing environment or country, all persons working with poultry should greatly increase the level of hygienic practices to avoid bringing virus into an operation (bio-exclusion) and to prevent virus exiting (bio-containment) if it has already entered a flock, village or region. The main ways in which the virus passes from one region to another area are: sale of infected birds to markets, departure of wild waterfowl which have visited infected backyard poultry units, the wearing of contaminated footwear or clothing by people working or selling poultry, or the

transfer of contaminated cages and egg crates to markets or poultry farms. Poultry keepers and communities must therefore take practical measures to avoid introducing the virus, and to reduce the risk of spread when disease has been detected.

3.4 VIRUS SURVIVAL IN THE ENVIRONMENT

Survival of influenza viruses is prolonged by low relative humidity and low temperature in aerosols, whereas low temperature and high moisture levels prolong survival in faeces. Most studies on viral environmental persistence have been carried out in cool northern climates with following findings:

- AI virus can survive in faeces for at least 35 days at 4°C. AI virus can survive within the poultry house environment for up to 5 weeks (Webster *et al.* 1978).
- Virus may remain infective in lake water for up to 4 days at 22°C and over 30 days at 0°C (Webster *et al.* 1978).
- As an enveloped virus, influenza virus is susceptible to several disinfectants, including detergents.
- The virus is stable over a pH range of 5.5-8.
- AI virus can be isolated from lake water where waterfowl are present (Hinshaw *et al.* 1979). Acidification of potentially contaminated drinking water to pH 2.5 or chlorination should minimise spread of infection.

4. Preparing for an outbreak

Good preparedness planning can be of enormous benefit in successfully managing an outbreak and minimising its impact. This planning should involve a consideration of how best to rapidly detect an outbreak, confirm the diagnosis and implement a rapid and effective control programme. It requires an assessment of veterinary service capabilities and capacity and the legal framework in which these services operate. The structure of national poultry industries should be examined to determine the potential for entry of virus. Industry support should be sought in the planning process. Provision for good public awareness programmes should be made as it is critical to have public support for disease control activities and good public knowledge to minimise the risk of human infection

4.1 EARLY DETECTION

4.1.1 Wild bird surveillance

Where the risk is from migrating birds, it is essential to identify the migratory habits of different species, their origins, destinations and timing of migration. While disease in wild birds would not be expected, it is of value to alert wildlife personnel to report unusual deaths in wild birds. Active surveillance can be practised by catching wildfowl species and sampling (generally by taking cloacal swabs) to test for the presence of AI viruses.

4.1.2 Domestic poultry surveillance

The identification of poultry at risk should involve poultry flocks located in high-risk agro-ecological systems where migrating birds congregate as well as in high-risk farming systems and practices such as free-grazing ducks associated with rice production systems.



CREDIT: WILLIAM KARESH, WILDLIFE CONSERVATION SOCIETY

Team collecting samples along lake shore (Mongolia).

At times of particularly high perceived risk, such as the arrival of migrating birds, it might be considered useful to undertake active serological and virological surveillance of sentinel birds within flocks, especially domestic ducks which are most likely to be exposed. It is therefore advisable that countries perform risk assessment studies of the introduction of AI in order to define the most appropriate surveillance strategy.

In any event, each country will have particular priorities and surveillance systems which should be refined to reflect these priorities. For example, HPAI-free countries or those with a lower risk of becoming infected will seek access to detailed, updated information on risks and will focus on the detection of incursions — making early warning and surveillance their priority. For infected countries or those at high risk of infection introduction, surveillance priorities will include the collection of detailed, current information on human health risks, ecological zones and production systems representing the highest risk for introduction and maintenance of HPAI infection.

The FAO guidelines for HPAI surveillance are particularly relevant to countries exposed to the risk of HPAI introduction and should be read in conjunction with this document (see Annex 1).

The objectives of HPAI surveillance and monitoring include:

- To detect clinical disease and infection.
- To understand the epidemiology and ecology of AI, as well as its socioeconomic impact, to help to design effective and implementable control programmes for poultry production systems.

Minimum requirements for effective surveillance (taken from “FAO Guiding Principles for HPAI Surveillance”)

The following minimum requirements apply to all countries and compartments:

- HPAI is a notifiable disease (i.e. there should be a legal requirement for suspected cases of disease to be reported to the official veterinary services).
- The official veterinary services must have a formal system for detecting and investigating outbreaks of disease and for reporting confirmed cases internationally, in accordance with OIE guidelines.
- The country and/or region must have the technical capability to diagnose HPNAI and LPNAI (see Annex 2).
- The country and/or region must have a system for recording, managing and analysing diagnostic and surveillance data.
- The country should participate in the regional surveillance and diagnostic network, including the public health sector, to enable sharing of information to characterize risk, prevent disease spread, and enhance control efforts.
- The frequency of surveillance could be a minimum of every six months within a country or could be less than this if selected ‘pilot’ areas are targeted for more frequent surveillance.

- To assess temporal and spatial patterns and thereby improve the effectiveness of control efforts.
- To understand the evolution in Asia of AI virus variants.
- To help define and control risks to public health.
- To monitor for antigenic drift in AI viruses through frequent analysis at competent laboratories.
- To maintain livelihoods and assist in ensuring food security through the implementation of appropriate control measures.
- To demonstrate freedom from clinical disease and absence of infection in a country or compartment and thereby facilitate trade.
- To assess the efficacy of vaccination when used as part of a comprehensive disease control programme.

In addition to formal surveillance and reporting procedures, raising awareness of the risk of avian influenza is an important means of increasing the likelihood that an outbreak of disease in poultry will be reported rapidly. However, it must also be kept in mind that such public awareness raising should not be undertaken in such a manner that it causes undue concern within the community. Also, veterinary disease control authorities will need to be prepared for an increased work load, since if formal and informal reporting mechanisms work, there will probably be a significant number of investigations that must be made following increased publicity of the risk of disease.

4.2 RAPID RESPONSE

4.2.1 Disease surveillance

Active surveillance should be initiated as soon as a country is considered at high risk of HPAI incursion. In cases of suspected disease, a sample of all domestic species of bird that die in the restricted area should be investigated and specimens submitted to approved laboratories for virus analysis and characterisation. Field surveillance examinations should seek to detect changes in flock health. Trained personnel should be aware of the potential risk to human health and wear protective gear (goggles or face shield, mask, gloves, and disposable gowns or coveralls, and rubber boots that can be disinfected before leaving the investigation site).

Surveillance needs to include:

- integrated commercial level poultry producers carrying out their own surveillance and timely reporting;
- local disease control centre officers carrying out regular telephone surveillance of independent premises; and
- targeted surveillance of premises in the restricted area and control area, particularly focusing on:
 - infected places, suspect places and dangerous contact places; and
 - premises with unusual sickness and/or mortality.

All reports of a decline in the health status of birds or egg production should be investigated and samples taken.

Although surveillance will begin immediately around the infected place or flock, it will have to be extended very quickly to all other sites where birds, products and contaminated materials might have been moved from the infected area. Surveillance of wild birds to determine their potential involvement in the dissemination of the disease may also be attempted, but this is likely to have a limited impact on controlling spread of the disease if biosecurity mechanisms in poultry farms are high.

4.2.2 Planning for avian influenza control

It is difficult to put in place a rapid and effective response to an outbreak of a new disease if the process has not been planned and the required resources made available. FAO has published guidelines in *“Manual on the preparation of national animal emergency preparedness plans”*, which recommends the development of four sets of complementary technical contingency plans:

1. Specific disease contingency plans that document the strategies to be followed in order to detect, contain and eliminate the disease.
2. Standard operating procedures that may be common to several or all emergency disease campaigns.
3. Enterprise manuals that set out zoosanitary guidelines for enterprises that may be involved in and emergency animal disease outbreak.
4. Simple job description cards for all individual officers.

Each national veterinary authority needs to consider its particular needs and be careful not to embark on a programme of emergency manual development that is not sustainable with its resources. A critical element of emergency plans is that they must specifically consider the situation and needs of a particular country (e.g. structure of the poultry sector, organisation of veterinary services). In addition, the implementation of contingency plans through simulation exercises is key to defining gaps or overlap in responsibilities or resources during an outbreak.

The Australian veterinary contingency plan, AUSVETPLAN, is a set of such plans, including a Disease Strategy for Highly Pathogenic Avian Influenza. It can be downloaded from the internet (Annex 1). Some of the most important considerations for the contingency planning process are:

1. Consideration of the disease control strategies that are available, the implications of applying them and the most appropriate strategy in different circumstances in a particular country.
2. Financial planning to determine where the necessary funds will come from and ensure that there is a mechanism and commitment to provide them immediately. A particular issue that needs to be considered is whether adequate compensation will be provided to poultry owners whose birds are destroyed. In the current context of the global avian influenza threat, countries might consider negotiating commitments from international donors to assist in control, should a disease incursion occur.
3. Resource planning, which include the needs for personnel, equipment and other physical resources. Plans will indicate resource requirements at the time of an outbreak. However, they should also provide a means to determine requirements in

- advance and either obtain them or make provision to obtain them rapidly when they are needed.
4. The need for appropriate legislation must be considered since this requires long-term planning in most administrative systems. Laws, regulations and proclamations are required to give authorised people the power to:
 - proclaim a notifiable disease
 - enter a poultry enterprise to inspect birds or collect specimens
 - define infected areas and disease control zones
 - institute quarantine of affected or suspect premises
 - place movement controls on poultry, poultry products and potentially contaminated materials
 - destroy and dispose of infected or potentially infected birds and contaminated materials
 - undertake other disease control operations, such as compulsory vaccination
 - place controls on the operation of enterprises, such as poultry processing plants.
 5. Obtaining consensus and commitment from all regulatory authorities and industry, as appropriate.
 6. Undertaking training of personnel so that the appropriate skills are available.
 7. Conducting simulation exercises to identify deficiencies and undertaking periodical review of the contingency plan.

4.3 AVIAN INFLUENZA CONTROL STRATEGIES

The primary aim of an early and rapid response to any occurrence of HPAI is to contain the disease before it spreads and eliminate it by stamping out affected premises and establishing proper disposal. Only if this fails should other measures be considered, including vaccination using effective and quality controlled vaccines. This approach is endorsed by OIE, FAO and WHO. Where it proves impossible to eradicate the disease in the short- to medium-term, it may be possible to aim for compartment freedom (e.g., freedom within the commercial sector in which birds are housed and protected from infection), or zone freedom (freedom in defined geographic areas). In the case of compartmentalisation, the poultry industry must take responsibility for its biosecurity (bioexclusion) with the necessary veterinary regulatory oversight in place; and in the latter, commercial compliance must follow national restrictions to ensure infection does not enter the free zone, and regulators prove to be trading with partners of the zone's clean status at any given time.

There is no pre-described strategy to control avian influenza outbreaks. In order to effectively control the disease, countries should have a complete plan of action and the financial and human resources to implement it under the particular conditions prevailing in the country. A regional approach is also necessary.

4.3.1 Culling

The basis of HPAI eradication by stamping out is to:

- immediately impose quarantine of the affected area (premises or village).
- slaughter all infected and potentially infected birds and dispose of the carcasses.



CREDIT: V. MARTIN

Restriction zone. Outbreak in Anhui province, China (July 2004)

- decontaminate sheds and other poultry housing areas.
- carry out rapid surveillance of surrounding areas to determine the extent of possible spread.
- close and disinfect markets.
- keep sick and dead birds out of the human food chain, and not sell for feed to other animals (i.e. zoos).

Quarantine and movement controls

AI is readily transmitted via contaminated objects, so strict control of movement of anything that may have become contaminated with virus and immediate imposition of tightly controlled quarantine on all places suspected of being infected are essential to a successful eradication programme. Ideally, quarantine should be imposed on all farms/villages on which infection is either known or suspected and should be strictly policed to ensure that no one, including the residents, owners, staff and other visitors, leaves without changing clothes and footwear.

Particular attention needs to be paid to workers on poultry farms who keep backyard poultry at home.

Strict on-farm biosecurity and hygiene is needed to control spread of the disease from wild birds. Access of wild birds to commercial poultry sheds and flocks should also be considered during depopulation operations. In areas where poultry are raised in a village environment, particular consideration needs to be given as to how effective quarantine disposal and decontamination can be imposed.

Effective quarantine of an area requires around-the-clock security to ensure that only authorised personnel in protective clothing are allowed to enter. It will be necessary to

supervise the movements of residents onto and off the property and to ensure that all pets are confined. It is also strongly recommended to ban cockfighting, pigeon racing and other avian concentrations in the outbreak area.

Slaughter of infected and potentially infected poultry

All susceptible poultry species in infected and dangerous contact premises, or in a large area if this is deemed necessary, must be slaughtered whether they are obviously diseased or apparently healthy.

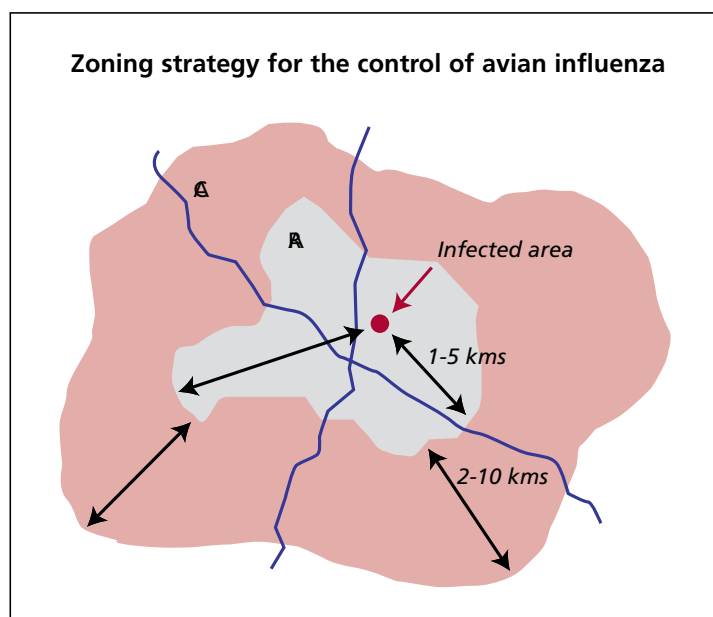
Although it not possible to provide specific and universally applied standards for controlling the disease through zoning for all potential outbreak occurrences, the following definitions and distances should be considered as a guide to rapidly contain HPAI. It must be noted that distances are indicative and subject to changes according to epidemiological characteristics, physical and geographical barriers, poultry density and farming systems (more details are provided in Annex 5).

Infected area

An area classified as an infected area (IA) will be a defined area (village, farm) in which HPAI has been detected. Infected premises (IPs) will be subject to quarantine and all susceptible animals will be destroyed in this area.

Restricted area

A restricted area (RA) will be a relatively small declared area (compared to the control area - see below) around infected places which is subject to intense surveillance and movement controls. Movement out of the RA will, in general, be prohibited, while movement into the area would only be through regulatory approval. Multiple RAs may exist within one control area (CA). The RA does not need to be circular but can have an irregular perimeter depending on known physical and geographical barriers, markets, poultry density and



farming systems. This distance will vary according to the size and nature of the potential source of virus, but will be approximately 1–5 km around the IP, depending on the density of poultry premises. The boundary could be the perimeter fence of the IP if the IP is in an isolated location. The boundary in a densely populated area will take into account the distribution of susceptible birds and traffic patterns to markets, service areas, abattoirs and areas that constitute natural barriers to movement. If possible hatcheries should be kept out of the RA.

Control area

The CA will be a larger declared geographical area around one or several RAs (possibly as large as a province initially) where restrictions will reduce the risk of disease spreading from the RAs. The boundary of the CA will be adjusted as confidence about the extent of the outbreak becomes clearer but must remain consistent with the OIE Code chapters on surveillance and zoning (Chapters 1.3.4 and 1.3.5; see Appendix 3). In general, surveillance and movement controls will be less intense and animals and products may be permitted to move under permit from the area.

The declaration of a CA also helps to control the spread of the outbreak from within the RA. The perimeter of the CA is a buffer zone between the RA and the rest of the country. The boundary does not have to be circular or parallel to that of the RA but should be 2–10 km from the boundary of the RA. In general, the movement of possibly contaminated articles and materials within the CA is allowed but movement out of the CA is prohibited without approval from the Chief Veterinary Officer. This type of control area allows reasonable and safe commercial activities to continue.

When declaring RAs and CAs, the areas must not be larger than necessary, thus restricting the number of properties to be quarantined to only those deemed prudent. If flocks in a quarantine area are not depopulated, then the cost of keeping the birds beyond their normal market age could be substantial.

Birds should be slaughtered by methods that take account of animal welfare concerns and the safety of operations, preferably without moving them from the site.

- For *small numbers of birds*, the preferred method is dislocation of the neck (using burdizzos, bone cutters, secateurs or bare hands). Burdizzos are particularly useful when large numbers of poultry with strong necks (geese, ducks, etc.) are to be destroyed.
- For *large numbers of birds* in commercial poultry units the preferred method is gassing with carbon dioxide. This method involves lining large garbage waste bins (skips) with plastic sheeting that also forms a canopy over the top of the bin. Birds can be caught using teams of labourers. Experienced catching teams may be available. Chicks are easily caught under heaters and are transferred to skips in plastic garbage bins. Broilers on the ground are driven, using a movable hessian wall, to the catching area where they are caught and placed directly into skips.
- *Caged birds* are more difficult and progress is slower. Each catcher removes 3 or 4 birds from cages and carries them by the legs to skips. Layers on perches are best caught at night or during low light when they are quiet.

Carbon dioxide (CO₂) is transferred to the bottom of the skips through 2.5 cm gar-



CREDIT: V. MARTIN

Burial site in Ahnui Province, China (July 2004).

den hose fitted to the top of the cylinders. The carbon dioxide should be decanted in 30–45 second bursts. It is essential not to decant too quickly or the bottles will freeze when about half empty. The concentration of CO₂ must be in the range of 60–70 percent in the skip, with the lid tightly closed for a 1–2 minute period to properly stun and kill the birds. On average, half a 45 kg cylinder of carbon dioxide is needed for the three cubic metre skips and three or more cylinders for the 20 cubic metre skips. Carbon dioxide should be added at a sufficient rate to ensure birds succumb before other birds are placed on top of them. Skips should be three quarters (75 percent) filled with birds, sealed and transported to the disposal site. Care must be taken to ensure no bird is still alive when dropped into the burial pit. Should this happen birds must be immediately caught and humanely killed.

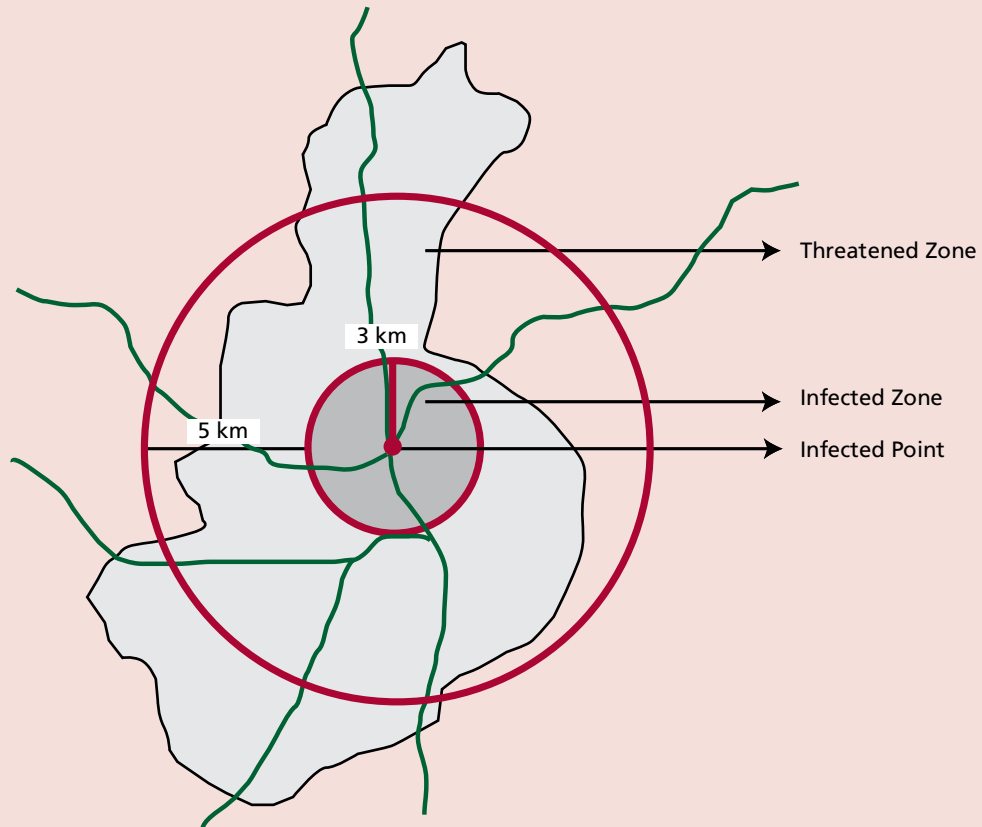
Safe disposal of carcasses

Disposal of dead birds, poultry litter and other contaminated waste is best done by burial. Sometimes this is not practicable or desirable because the required equipment is not available or it would have adverse environmental effects, such as potential contamination of ground water. In these circumstances, the best alternative might be composting.

Burial - Burial is best undertaken at the infected site. It is best to minimise the distance that infected material needs to be transported. A burial place outside infected premises may be the best option in situations where a number of infected foci would have to be depopulated and decontaminated in a given area and where a common burial site would be more efficient. The preferred equipment for digging burial pits is an excavator. This equipment is the most efficient available for the construction of long, deep, vertically sided pits. Other advantages include the ability to easily store topsoil separate to subsoil and the equipment can be used if required to fill the pit with carcasses or other materials and close the pit without disturbance of the carcasses. Loaders, bulldozers, road graders and backhoes (for

Example of zoning: the case of China

Premises with infected poultry or relevant slaughtering houses and other departments are considered as infected points; areas within the 3 km radius are considered as infected zones; areas within 5 km around the infected zones are considered as threatened zones.



Stamping out. All poultry within infected zones should be stamped out.

Emergency vaccination. All susceptible poultry in the threatened zones are vaccinated compulsorily with the vaccines approved by Ministry of Agriculture. Only healthy birds should receive vaccine.

Disposal. All poultry carcasses and poultry products in infected points, and excretion material, contaminated feed, litter and sewerage from the infected points shall be subject to bio-treatment or disposal.

Cleaning and disinfection. All contaminated items within the infected zones, transportation vehicles, utensils, poultry counters and ground must be cleaned and disinfected.

Movement control. Warning signs widely posted around the infected zone, disinfection stations set up in the transportation entrance of infected zones to disinfect vehicles and items entering and exiting zones; movement of all susceptible live birds and their products controlled.

Closing the market. All poultry and poultry product markets in infected zones and live birds markets within a 10 km radius of infected zones must be closed.

Tracing. If poultry and their products are sold out during the incubation and clinical manifestation period or moved out, tracing should be conducted on the suspect contaminated items to prevent these items from spreading disease.

Financial support. Financial support systems are established for all poultry destroyed because of HPAI.

Public health, surveillance of staff of poultry rearing, trade and transportation and processing units, especially of staff in the infected zones, should be intensified, and epidemiological investigation should be conducted. Stringent protective measures must be implemented by staff participating in the destruction of infected birds and cleaning of contaminated premises.

Lifting quarantine. The conditions for lifting a quarantine are stipulated as follows: 21 days for infected point and infected zone after strict treatment according to “*National Contingency Plan for Highly Pathogenic Avian Influenza*” and standard technical requirements for treatment of HPAI; over 14 days for the threatened zone where all the susceptible birds are vaccinated with nationally approved vaccines and after no new cases occur, it is inspected and the findings accepted by the relevant authorities. When quarantine is lifted, live bird trade markets 10 km around the infection zone may be reopened. The infected points are re-stocked 6 months after strict disposal.

small jobs) may be used if excavators are unavailable. With the exception of backhoes, all other equipment requires the continual movement of the machine over the site while digging the pit. Excavators and backhoes essentially remain in a fixed position while digging; hence they move soil faster, with less cost and less damage to the site surrounding the pit. Most excavators have an attachable hammer for rock work if necessary. The dimensions of the burial pit will depend on the equipment used, site considerations and the volume of material to be buried. The preferred dimensions are for pits to be as deep as practically possible (reach of machinery, soil type and water-table level being the usual constraints), with vertical sides.

Gas production from decomposition within unopened carcasses may result in considerable expansion in the volume of the buried material to the extent that the surface of the closed pit may rise and carcasses may be expelled from the pit. Lime may be added to pits to prevent earthworms bringing contaminated material to the surface after pit closure. Covering the carcasses with 40 cm of soil is suggested, and add an unbroken layer of slaked lime [Ca(OH)₂] before filling is completed. Lime should not be placed directly on carcasses because it slows, and may prevent, decomposition.

Inspection of the burial site after closure is recommended so that appropriate action can be taken in the event of seepage or other problems.

Composting – Biological decomposition, or composting, is an effective way of dealing with manure and litter waste and can be carried out within sheds or otherwise on site, thus

overcoming the risks of disseminating the virus during transport. Composting should be done in a secure area not accessible to susceptible birds. The procedure requires the piling of carcasses with other bulky contaminated or non-contaminated material, such as wood chips or straw bedding, to allow for proper aeration and covered with a biological filter (i.e. not whole plastic). The pile should not be pressed or otherwise compacted. The site selection is important – away from any residences, with at least 1 m of ground between the pile and any known water source, with any run-off water from the decomposing material collected and treated. Care must be taken to ensure that susceptible animals or pets (dogs) do not have access to the compost pile. Temperatures for proper composition should reach 55-60°C within 10 days and the material kept in place for several weeks; subsequently mix but never press the material within the pile. Properly decomposing material at this time should be dark in colour with minimal foul odour.

Burning/Incineration - A burning area outside an infected place may be the best option in situations where a number of infected foci would have to be depopulated and decontaminated and where a common burning site would be more practical. The principle is to place carcasses on top of sufficient combustible material, ensuring the arrangement of fuel and carcasses allows adequate air flow to enter the pyre from below, thus achieving the hottest fire and the most complete combustion in the shortest time.

When loading of the carcasses is complete and weather conditions suitable, saturate the fire-bed and carcasses with diesel or heating oil (NOT PETROL) and prepare ignition points about every 10 metres along the length of the fire-bed. These can be made of rags soaked in kerosene. Move all vehicles, personnel and other equipment well away from the fire-bed. Start the fire by walking into the wind and lighting the ignition points along the way. The fire must be attended at all times and re-fuelled as necessary; use a tractor with a front-mounted blade or a front-loader. Ensure any carcasses or parts thereof that fall off the fire are replaced on the fire. A well-constructed fire will burn all carcasses within 48 hours. The ashes should be buried and the site restored as well as possible.

Rendering - Rendering is a closed system for mechanical and thermal treatment of waste, and a good way for carcass disposal if the plant has sufficient capacity and if it is possible to effectively decontaminate the rendering plant afterwards. A medium-sized facility could render some 12 tonnes per hour of operation. However, private rendering plants may not be willing to handle infected birds and eggs unless an emergency order is imposed. A disadvantage is that infected material would need to be transported from infected sites to the plant.

Where burial, cremation or rendering are not considered practical or are difficult to carry out in the infected place, permission should be sought to transfer carcasses and/or infectious material to another site for disposal by burial, cremation or rendering. Transport should be in a leak-proof container, such as a large skip, covered with tough polyethylene covers and sealed at the top. It should not be overloaded – half a metre or more (depending on distance to be travelled and temperature) should be left clear for expansion of carcasses. Vehicles should travel slowly to avoid splashing of contaminated material and should be accompanied by a police vehicle to minimise the chances of accidents and to

TABLE 1
Selection and application of decontamination procedures

Item to be disinfected	Disinfectant/chemical/procedures
• Live birds	• Euthanase (carbon dioxide gas; dislocation of neck)
• Carcasses	• Bury or burn
• Animal housing/equipment	• 1, 2, 3 (See key below)
• Humans	• 1
• Electrical equipment	• 5
• Water	• Drain to pasture where possible
• Feed	• Bury
• Effluent, manure	• Bury or burn; 4, 3
• Human housing	• 1, 2
• Machinery, vehicles	• 1, 3
• Clothing	• 1, 2, 3

Key

1. Soaps and detergents: leave in contact for 10 minutes
2. Oxidising agents:
 - a. sodium hypochlorite: liquid, dilute to final 2-3 percent available chlorine, not good for organic materials. 10-30 minute contact time.
 - b. calcium hypochlorite: Solid or powder, dilute 2-3 percent available chlorine (20 g/litre powder, 30 g/litre solid), not good for organic materials. 10-30 minute contact time.
 - c. Virkon®: 2 percent (20 g/litre). 10 minutes contact time.
 - d. Virocid®: 0.25 percent (1:400). 10 minutes contact time on non porous surfaces.
3. Alkalis: (do not use with aluminium and similar alloys)
 - sodium hydroxide (NaOH): 2 percent (20 g/litre). 10 minute contact time.
 - sodium carbonate anhydrous (Na₂CO₃ .10H₂O): 4 percent (40 g/litre from powder, 100g/litre from crystals), recommended for use in presence of organic materials as above. 10-30 minute contact time.
4. Acids:
 - hydrochloric acid (HCl): 2 percent (20 ml/litre), corrosive, use only when other chemicals are not available
 - citric acid: 0.2 percent (2 g/litre), safe for clothes and body decontamination. 30 minute contact time.
5. Formaldehyde gas: Toxic, only if others cannot be used. 15-24 h exposure time.

prevent breaches of biosecurity. The escorting officer must carry a supply of an approved disinfectant and basic equipment to deal with minor spills en route. All vehicles must be cleaned and disinfected before leaving the infected place and after unloading.

Decontamination

Soapy water and detergents are the first choice for decontamination. The avian influenza virus is more easily destroyed than many other viruses since it is very sensitive to detergents which destroy the outer lipid envelope of the virus. Therefore washing of contaminated surfaces should always be with detergents (soapy water) or specific disinfectants. The most difficult material to decontaminate are bird droppings since the virus can survive in moist environments with high organic content; it is essential to thoroughly clean and disinfect items that have been in contact with bird droppings – cages, shoes, clothes – before working with poultry or entering a place where poultry are kept. Simple hygienic measures can reduce risk - but national authorities are encouraged to prepare and communicate specific guidance for each type of poultry enterprise. More guidance for veterinary services on selection and application of decontamination procedures is given in table 1 (Ausvetplan Manual). Adaptation to specific country circumstances will be needed.

Destocking period

After slaughter, disposal and decontamination procedures have been completed the premises must be left without susceptible species (destocked) for a period of time, determined by the estimating survival time of the pathogen in the particular environment. **Restocking** should not take place until at least 21 days after satisfactory cleaning and disinfection has been completed and the outbreak has been brought under control in the area. Restocking should be undertaken by introducing a small number of poultry first, and these **monitored** daily for signs of disease. Should this occur, notification to the authorities must be immediate and sampling of the sick or dead birds done to determine the cause. If the poultry remain healthy, full repopulation can be carried out. Of course, improvement of biosecurity should be instituted at all stages of production to decrease the likelihood of AI or other diseases entering the recovered premises. After repopulation, **monitoring** should be continuous through the sampling of dead birds to determine whether re-infection has occurred.

4.3.2 Financial Support

The issue of compensation for slaughtered birds, property damaged during decontamination and/or loss of income needs to be carefully considered. In principle, offering compensation encourages owners to report disease. However, it can become a very expensive undertaking and guidelines are usually developed that strictly limit the categories for compensation. It is important to consider what the cost of compensation might be and how it would be funded should a major outbreak of disease occur.

If compensation is paid, it can be controlled as follows:

- Have a registration process
- Only pay for animals slaughtered, not those that have died.
- Pay promptly and at a level that is close to market value (some countries have used innovative strategies such as paying slightly more than market value for healthy in-contact birds, and less for sick birds – which allowed for quick reporting to authorities).
- Do not compensate losses other than livestock (birds).
- Ensure that people with very small flocks are also compensated

There are alternatives to payment of compensation in cash and experience has sometimes shown them to be acceptable.

- Rather than paying cash, provide replacement birds (can be difficult sometimes; governments have to think in advance about logistical questions of where to obtain replacements, their transport and the required destocking times).
- Provide credit for owners to re-establish their poultry production, including village birds or facilitate entry into alternative livelihoods.
- Provide area assistance to enable market conditions to become re-established without undue delay.
- Provide farmers with free technical and veterinary services in re-establishing production schemes.

4.3.3 Vaccination

Vaccination as a support strategy may be considered when the disease has spread to such an extent that it has overwhelmed the resources of disease control authorities or the economic cost of a widespread slaughter campaign cannot be borne. It can also be considered at an earlier stage when veterinary service infrastructures and capacities prove to be very weak and insufficient to curb the spread of the disease. FAO and OIE have made recommendations for the use of OIE-approved AI vaccines, and several such vaccines are commercially available. If used in accordance with FAO/OIE recommendations (*FAO Position Paper, September 2004*) and the *OIE Manual for Diagnostic Tests and Vaccines for Terrestrial Animals*, these vaccines provide excellent protection against clinical disease in chickens by reducing mortality and production losses. Vaccination of poultry also reduces the viral load in the environment, thus decreasing the risk of transmission to poultry and humans. According to current OIE recommendations, HPAI-vaccinated poultry are not excluded from international trade, although specific technical guidelines must be followed to ensure that the vaccine is being applied properly and monitored effectively.

TABLE 2
Vaccine properties

Vaccine type	Advantages	Disadvantages
INACTIVATED HOMOLOGOUS VACCINE The same H and N antigens as the strain isolated in the outbreak	<ul style="list-style-type: none"> • Readily available • Rapid onset of immunity with adjuvants • Inexpensive • Safe 	<ul style="list-style-type: none"> • Impossibility of differentiating vaccinated from infected birds serologically. • Monitoring by using sentinel unvaccinated birds (identification, bleeding and swabbing) is time-consuming, requires planning and monitoring • Requires boosters in long-lived species • Requires percutaneous injection
INACTIVATED HETEROLOGOUS VACCINE (DIVA Strategy: Differentiation of Infected from Vaccinated Animals) The same HA subtype and a different NA subtype compared to the virus isolated in the outbreak	NA: marker of field infection. Serology can determine whether birds in a vaccinated flock have also been infected.	<ul style="list-style-type: none"> • Laboratory capacity to perform the discriminatory test based on the N antigen • Serology is expensive, requires additional reagents and requires a complete knowledge of circulating N antigen sub-types • Requires boosters in long-lived species • Requires percutaneous injection
RECOMBINANT FOWLPOX VIRUS	<ul style="list-style-type: none"> • Enables the differentiation between infected and vaccinated birds by serologic tests • Specificity of the immune response directed exclusively against HA components • Vaccination is rapid and only one dose is required • Inexpensive 	<ul style="list-style-type: none"> • Can only be used to vaccinate chickens without previous fowlpox exposure. Therefore, usually applied only to day-old chicks • Cannot be used in ducks/geese • Requires percutaneous injection

Vaccination, when it is applied, must be done in combination with other disease control measures, including the slaughter of affected flocks. Efforts to control the disease by vaccination alone, without slaughtering affected birds to reduce the virus load in the environment, will probably not be successful. Depending on the incidence and distribution of outbreaks, vaccination may be undertaken around outbreaks (ring vaccination) or throughout the poultry population (mass vaccination).

Sourcing vaccine

There are a number of different avian influenza vaccines available. Conventional vaccine is prepared from the allantoic fluid of infected eggs, which is inactivated and emulsified with an adjuvant. Attenuated live influenza virus vaccines are not recommended because of the risk that the vaccine virus could either mutate or reassort with other influenza viruses to become virulent. However, recombinant vaccines have been produced, including fowl-pox virus with the influenza haemagglutinin gene inserted.

Generally, conventional inactivated vaccines are used. The main immunogenic component is the haemagglutinin protein. It must be of the same subtype as the outbreak virus (for the current widespread Asian epidemic, this is H5). The neuraminidase antigen can be the same as the outbreak strain. However, if differential serology is to be undertaken for monitoring vaccine response or virus activity (the DIVA method), then a different neuraminidase should be used in the vaccine (e.g., H5N2 or H5N9).

It is also possible to leave a small number of identified sentinel birds unvaccinated which will aid monitoring for flock infection. If sentinel birds show disease symptoms or die, virus isolation and serological test have to be done to confirm flock infection.

The DIVA strategy requires testing of serum samples for antibody to the neuraminidase, to differentiate that of field strain(s) from that of the vaccine strain(s). It assumes that the heterologous N antigen is not circulating in the field; and therefore that knowledge of circulating AI viruses (virulent or not) is known. In circumstances, such as currently prevail in Asia, where there may be several strains of avian influenza virus circulating, and where in some countries there are vaccines in use with several different neuraminidase antigens, it may be difficult to apply the strategy. Also, the differentiating assay involves the use of additional reagents, which may be beyond the scope of most national laboratories because it represents an additional expense. Therefore, a decision to apply the DIVA strategy needs to be carefully considered against these constraints; it may be more appropriate towards the end of a successful control campaign.

For inactivated vaccines, two doses of the vaccine must be given, approximately 30 days apart, to achieve adequate protection. Vaccinated birds are generally not fully protected from infection but have increased resistance to infection, suffer less clinical disease and shed substantially less virus in the event that they become infected. Longer lived species (ducks, geese, yellow chickens) require booster injections of vaccine to maintain protection.

Recombinant fowl pox vaccine can be used for vaccinating day-old chicks. Since it is a live pox virus, it can be applied by stab inoculation into the wing web, which can be performed quickly with minimal training. This vaccine cannot be used in older birds, since they

are likely to have already become exposed to fowl pox and will not respond to vaccination. A disadvantage of the fowl pox-vectored AI vaccine is that it is ineffectual in ducks. Specifications for purchasing vaccine and a list of potential suppliers is provided in Annex 2.

4.4 MANAGEMENT OF DISEASE CONTROL

The *FAO Manual on the Preparation of National Animal Disease Emergency Preparedness Plans* makes recommendations in regard to management of disease control operations. The following recommendations should be considered:

1. To manage disease control on a national basis, there needs to be a suitable command structure for veterinary services. It is essential that information can flow quickly and efficiently from the field to national headquarters and that conversely, control mechanisms are continuous from headquarters to the field. In recent years, government veterinary services in many countries have been restructured, including regionalisation, rationalisation and downsizing, privatisation, separation of policy and operational functions, and separation of authority for field and laboratory operations. Countries may need to consider whether they need to make structural changes or alternative arrangements to adequately deal with animal health emergencies.
2. It is often advisable to have a consultative committee which can meet during the period of an animal disease emergency to provide the best technical advice to outbreak management personnel. The committee might comprise the CVO, national directors of field and laboratory services, head of the epidemiology unit, AI expert, directors of state, regional or provincial veterinary services, representatives of private industry and other key groups, and other technical experts as required. Gaining the cooperation of police, military and public works offices would be beneficial for the success of any plan.
3. At the time of an emergency, it can be a great advantage to have made arrangements in advance for a national animal disease control centre and local animal disease control centres. The control of diseases in difficult or marginalised areas of some countries may require special consideration.
4. Arrangements need to be made in advance of an outbreak. They should include negotiation with all government authorities and others who will be involved in assisting in disease control. Because certain strains of avian influenza viruses can infect humans, the respective roles of human health and veterinary services need to be considered in advance.

Field services capacity

Veterinary services must have the capacity to undertake disease surveillance, investigate and respond to disease outbreaks and report to various levels of the official veterinary service structure. In designing a proposed strategy to control avian influenza, careful consideration needs to be given to the capacity of the field services to assume disease management activities in accordance with legislation that enables and empowers decision makers. Very few national veterinary administrations have the capacity within their own resources to

Lessons learned from the 2004 epidemic in Asia:

As observed in South East Asia during the 2004 epidemic, many countries did not have a plan of action supported by national legislation at the time of HPAI detection.

The following points were particularly lacking in several affected countries:

- A structured surveillance program including surveillance protocols in suspected or at-risk farms.
- Protective equipment not available for workers and animal health personnel.
- Investigation procedures: standardised protocols and sets of epidemiological questions for outbreak investigation and mapping. These protocols should also include information on collection of a standard set of samples for disease investigation from infected farms, neighbouring farms and uninfected farms outside of the control areas to allow for more detailed pathological examination of dead birds.
- Restocking programmes were often incomplete with no detailed method to control the flock after restocking.
- Vaccination plans were often incomplete and did not have a coherent strategy for post-vaccination surveillance activities. Vaccines used were often of unknown quality and of dubious efficacy.
- Declaration of disease freedom from areas or zones were made without proper scientific data supporting such claims and were subsequently proven to be false creating additional uncertainty in the population and loss of consumer confidence.
- Absence of adequate legislation or difficulties of enforcement.

undertake all the necessary measures required to combat a major transboundary disease incursion. Assistance is often sought from police, military, national disaster and other emergency organisations late in the process. The immediate and early needs from other public offices and cooperation from private industry must be anticipated to assist field veterinary services in performing their duties. Access to extra budgetary resources should be part of the contingency planning process and appropriate arrangements for collaboration negotiated in advance.

An electronic disease information database can be a great asset in tracking the huge amount of detail that can be accumulated in the course of a disease epidemic. There are many systems available, including the FAO-designed and supported system, TADinfo. FAO, OIE and WHO have collaborated in establishing a Global Early Warning and Response System (GLEWS) to enable international animal and human health authorities to receive early notice of outbreaks, or the likelihood of certain diseases occurring based on disease intelligence and forecasting risks. Data is analysed and information made available to international and national disease control authorities.

4.5 LABORATORY DIAGNOSIS

Laboratory services must be able to:

- undertake testing of samples collected for surveillance activities.
- test samples collected for diagnosis.
- test samples collected for monitoring response to vaccination.

Consideration needs to be given to the capacity of veterinary services to collect and deliver specimens to a competent national laboratory. There are benefits in having regional laboratories that might have better access to the field. However, in many countries expert personnel are lacking and it may be more effective to have just one central laboratory that is well resourced and properly staffed.

4.5.1 Samples

Samples taken from dead birds should include intestinal contents (faeces) or cloacal and oropharyngeal swabs. Samples from trachea, lungs, air sacs, spleen, kidney, brain, liver and heart may also be collected and processed either separately or as a pool. Intestines (loops) should always be collected last and packaged separately to avoid bacterial contamination. Samples from live birds should include both tracheal and cloacal swabs, although swabs of the latter are the most likely to yield virus. Since small delicate birds may be harmed by swabbing, the collection of fresh faeces may serve as an adequate alternative. To optimize the chances of virus isolation, it is recommended that at least one gramme of faeces be processed either as faeces or coating of the swab. Should the investigator not be sufficiently well trained to perform a necropsy (autopsy), whole birds should be bagged twice (one bag inside another), maintained refrigerated at all times and submitted to a laboratory where a proper necropsy and sample collection can be completed.

The samples should be placed in isotonic phosphate buffered saline (PBS), pH 7.0–7.4, containing antibiotics. The antibiotics can be varied according to local conditions, but could be, for example, penicillin (2000 units/ml), streptomycin (2 mg/ml), gentamycin (50 µg/ml) and mycostatin (1000 units/ml) for tissues and tracheal swabs, but at five-fold higher concentrations for faeces and cloacal swabs. It is important to adjust the pH of the PBS solution to pH 7.0–7.4 following the addition of the antibiotics. Faeces and finely minced tissues should be prepared as 10–20 percent (w/v) suspensions in the antibiotic solution. Suspensions should be processed as soon as possible after incubation for 1–2 hours at room temperature (22–25°C). When immediate processing is impracticable, samples may be stored at 4°C for up to 4 days. For prolonged storage, diagnostic samples and isolates should be kept at –80°C without PBS. Never use alcohol to preserve samples.

Submission of samples to any laboratory outside the country of origin should always be subject to prior agreement with the recipient laboratory (see Annex 4 *Information for shipping International Diagnostic Specimens*) and be transported in containers meeting IATA (International Air Transport Association) regulations. Infectious substances which cause disease only in animals are categorized as UN 2900. Infectious substances which cause disease in humans (or both in humans and animals) must be assigned to UN 2814. All H5 and H7 samples must be assigned to UN 2814. All materials should be in leak-proof containers.

4.5.2 Laboratory tests

National laboratory services should be able to carry out the following tests.

As a minimum requirement:

1. Virus isolation in eggs (SPF or SAN), identification of isolates as “A” influenza virus, haemagglutinin and neuraminidase typing.
2. Serology – including ELISA (for antibody to matrix protein), haemagglutination-inhibition testing.
3. Antigen detection – ELISA or haemagglutinin testing.

In addition, the capacity to perform the following tests is highly desirable:

4. Neuraminidase antigen identification.
5. Pathogenicity testing of virus isolates by chicken inoculation.
6. Polymerase chain reaction (PCR) technology for rapid detection of virus genome.

Methods are described in the *OIE Manual*, Chapter 2.1.14. Consideration needs to be given not just to having the appropriate technology but to having the capacity to handle a large number of specimens in the event of a disease control emergency.

Countries should also have access to international expertise to assist in confirming the identity and characteristics of isolates and undertaking further key studies that are of particular importance at the international level. It is important to monitor field viruses for early detection of changes and this can be done by systematic sampling of birds during an epizootic and in inter-epizootic periods and sending suspect samples to a reference laboratory for confirmation and further characterisation.

A list of the OIE/FAO reference laboratories is in Annex 3.

In April 2005, OIE and FAO launched a joint scientific worldwide network to support the veterinary services in the control of avian influenza – OFFLU. The objectives of the new network are:

- to collaborate with the WHO human influenza network on issues relating to the animal-human interface, including early preparation of vaccines for possible human use.
- to promote research on avian influenza.
- to offer veterinary expertise and new skills to countries to assist in the control and eradication of HPAI.

Through active and permanent scientific cooperation, the network will develop collaborative research proposals, provide multidisciplinary teams to countries requiring assistance, and act as a link between OIE/FAO reference laboratories, and regional and national laboratories. Sharing permanently updated scientific information and expertise on efficient control methods will provide a pro-active approach in helping infected countries to progressively control and eradicate the disease and for disease-free countries to better protect themselves.

For more detailed information, see the OFFLU website www.offlu.net

Field diagnosis

Often decisions need to be made immediately in the field during an emergency. Waiting for a laboratory diagnosis before taking action to quarantine an area can lead to critical

delays. Clinical and pathological findings in the field can often be sufficient to make a presumptive diagnosis of HPAI and actions taken accordingly. Though some rapid on-site diagnostic assays are available, to date these have been of poor sensitivity and expensive. Several institutes are actively researching the development of improved sensitive, specific and cost-effective rapid on-site assays, but these remain in the development stage and require validation in the field before their recommendation.

4.5.3 Communication and Public Awareness

There are several objectives which should be considered in public awareness campaigns:

- Inform farmers and consumers on the infection channels and risks related to AI.
- Communicate information (e.g. time and venues for vaccination, procedures for obtaining compensation).
- Promote better farming practices and improved hygiene.
- Raise awareness of the risk of poultry infection (see Annex 6: poster example).
- Recognise that in many village situations it may be unrealistic to expect to be able to improve biosecurity.
- Seek assistance from the community by having poultry owners report unusual sickness and mortalities in their flocks.
- Cooperate during disease control activities, should an emergency occur.
- In conjunction with human health authorities, raise awareness of the risk of transmission from poultry to human and about disease in humans.

4.5.4 Human health and safety considerations

Certain avian influenza viruses can infect humans, occasionally causing severe disease and sometimes death. The WHO website (www.who.int) provides updates and recommendations with regard to symptoms, patient care and management, and recommendations to reduce occupational risks (i.e., personal protective equipment). The following should be considered:

General considerations for the human population

- vaccination with seasonal human influenza vaccine (which contain H1, H3 and B virus antigens) to minimise the risk of multiple influenza infections causing a new virus to emerge.
- avoid unnecessary contact with infected or exposed poultry, poultry products and poultry waste.
- in particular, people in poor health and children should avoid all contact with affected birds and their environment.
- do not purchase or consume poultry that are not healthy or wholesome.

Specific considerations for all personnel involved in disease control

- selection of workers should avoid those in high risk categories.
- workers should wear protective overalls and, where gross contamination is likely, a waterproof apron.
- disposable or rubber work gloves should be worn.

- a disposable P2 or N97 respirator should be worn.
- goggles or a visor should be worn to protect against eye splash.
- disposable footwear or rubber boots should be worn.
- protective clothing should be disinfected after use.
- hands should be washed after protective clothing is removed.
- use of antiviral drugs is recommended, during and for 7 days after exposure.
- workers should monitor their health, watching for signs of fever, respiratory symptoms (e.g., cough) and conjunctivitis (eye inflammation).
- illness in workers or their close contacts should be reported to public health authorities immediately.

5. Prevention and biosecurity

Areas that have not been affected by an HPAI outbreak or those that have undergone culling, disinfection and even vaccination should improve their biosecurity. Improved biosecurity at whatever level is cost-effective in comparison to the losses from disease, depopulation and further anguish, be it at the village level or commercial farm.

The most difficult environment in which to improve biosecurity and disease prevention is likely to be at village level, where poultry and other animals are allowed to move without any restrictions and there are no costs to animal care (feeding), but their losses due to disease or scavenging animals (dogs, cats, wildlife) are high. Under these circumstances, the role of rural developing agencies can be beneficial in promoting the advantages of keeping animals in a fenced enclosure where environmental stresses are minimised, theft less likely, animals are safer from scavenging animals, and the loss of valued animals through being run over by motorcycles, cars or lorries is eliminated.

5.1 RESTRICTED ACCESS MEANS KEEPING THE DISEASE OUT

Restricting access to a property or farm through the use of fences and enclosures creates a barrier between clean areas where the poultry are kept and the outside environment. Access to where poultry are kept should be restricted to people known by the owner, people who do not have poultry of their own, and to people who do not participate in events where birds congregate, such as cockfights. Particular attention needs to be paid to workers on poultry farms who keep backyard poultry at home – best practices would stipulate that no workers should have poultry of their own, since this is a high risk avenue for disease introduction. Wild birds – resident fowl or migratory birds – should have no contact with the flock through the use of screens or overlying nets. Visitors wishing to see poultry should wash their hands and change their shoes and use footwear provided by the owner (e.g. rubber boots that are kept for such visitors). If visitors have birds of their own, they should not be allowed near the birds.

Ducks kept in ponds or paddy fields with other ducks of a different owner represent a high risk, unless all duck owners agree on the measures that can be taken collectively. For instance, erecting poles with netting that separate one owner's flock from another and taking turns in scaring away wild birds from landing or feeding within production flocks.

5.2 CLEAN AREAS MEAN HEALTHY CHICKENS, GEESE AND DUCKS

Keep the area of the flock clean from garbage (food waste, plastic bottles, glass bottles, tins or drums). When the owner or care person needs to attend to chickens or other poultry (e.g., collecting eggs, feeding or watering chores, change the bedding or the repair of fencing material), a change of clothing and boots should be required. These clothes and boots can be cleaned and disinfected upon exiting the enclosure and be ready for the next use. Dirty clothes should be washed with detergent and hung out to dry in the sun; boots

should be washed in chlorinated water, or with soapy water. Washing hands with soap before entering the caged area should be practised always. Tools (feeding scoops, shovels, brooms) and feeding pans used in the caged areas should be kept clean daily. All manure should be removed and disposed of properly (i.e. compost pile). Keeping a wide pale with chlorinated or soapy water for frequent use before entering or exiting the enclosure is a good reminder to follow biosecurity.

Keeping the cages clean prevents pathogens from accumulating and causing health problems. Clean cages keep the birds and eggs clean as well - which translates into better market prices.

Sick or dead chickens must be removed quickly and community animal health workers or the local veterinarian informed of such illness or death.

5.3 BUY HEALTHY: KEEP HEALTHY

Transportation of birds to the farm can represent a considerable risk – not only should the owner be aware of the “good” price obtained, but also that vehicles, (trucks, motorcycles, bicycles), cages, equipment and feed may be contaminated when returning or entering the farmer’s property. Newly-purchased equipment should be thoroughly washed with soapy water or otherwise disinfected before use. Newly-purchased birds should be housed in a separate enclosure for at least two weeks before allowing them to mix with birds already on the farm. Owners are advised that it is important to keep species separate, and not mix ducks with chickens, chickens with pigs, or ducks with pigs. It is also good practice not to mix animals of different ages.

5.4 USE OF CLEAN EQUIPMENT – KEEPING DISEASE OUT

Poultry equipment, such as cages, egg crates, shovels or rakes, should not be shared between family or neighbours. Wooden pallets, wooden handles, or egg crates can be porous, and even though they can be treated with disinfectant, it is difficult to ensure that they are completely disinfected. Metal cages can be cleaned and disinfected; if these are borrowed because of necessity, they must be cleaned and disinfected by the owner of the birds before they are reused.

5.5 REPORT EARLY SIGNS OF A PROBLEM THAT COULD BE DEVASTATING

Many bird diseases look similar. Early detection and prompt reporting will probably help stop the spread of disease. Owners must know who and where to report abnormalities on the farm when they begin not when they end. Signs to be reported include: sudden death, depression and decreased appetite, diarrhoea, breathing difficulties such as coughing, sneezing and gasping, nervous twitching or dropped wings or paralysis, and swelling of the head with darkened combs, wattles, or legs.

Owners must be assured that early reporting of a problem will benefit them, their families and their village in the long run. In this regard, the government - in conjunction with the poultry industry - should be prepared to react and provide proper compensation (see section 4.3.3 above). **Failure to provide an incentive for compensation for disease reporting will undoubtedly lead to disease spread.**

5.6 A PERIOD OF REST

One prevention measure that can be instituted but requires planning and several enclosures is the practice of “all-in all-out”. This method is used in many countries and envisages a complete growth cycle of chickens (or other species) from the moment of introduction – as in day-old-chicks – all the way to marketing age. At no time are other birds introduced into the enclosure. Once sent to market, the floor is scraped clean of faecal and feather debris, and bedding and feed removed; cages and other equipment are cleaned and prepared for the introduction of young healthy birds. It would be wise to keep the enclosures free of birds and other animals for a rest period of say, seven days, before bringing in the next batch of poultry.

With duck operations, the “all-in all-out” operation may be more difficult, unless there is planning in the growth cycle between birds using the same pond and double netting used between age groups. Double netting – 2-3 metres apart – implies additional cost, but decreases the opportunity of pathogens from contacting susceptible ducklings. Although ducks share the same water ponds in which avian influenza viruses may survive, it still decreases the likelihood of disease transmission.

5.7 VACCINATION AGAINST AVIAN INFLUENZA OR OTHER DISEASES

Vaccination, in general, increases the resistance of poultry to disease but does not eliminate the possibility that infection may occur in a flock. Prevention of disease and infection can only be accomplished with other aspects of prevention and improved biosecurity.

When poultry are to be vaccinated, it would be wise for the owner ensure that the vaccination team changes clothes and cleans and disinfects boots, gloves and equipment before entering poultry enclosures. Should the vaccination team resist such instructions, the owner should report the non-observance of biosecurity measures to the appropriate veterinary authorities.

5.8 COMPARTMENTALISATION

In the *OIE Terrestrial Animal Health Code*, compartmentalisation refers to one or more establishments under a common biosecurity management system containing an animal sub-population with a distinct health status with respect to a specific disease or specific diseases for which required surveillance, control and biosecurity measures have been applied for the purpose of international trade (more information available is in chapter 1.3.5 on *Zoning and compartmentalisation*).

In countries where the disease may be present in some areas or confined to some production systems, this concept can be applied to poultry operations that will adopt strict biosecurity measures to prevent the introduction of the disease all along the production process.

In poultry operations that are tightly controlled by producers, a strict method of operation must be assured to prevent disease from entering the operation. Besides the measures mentioned in the above sections of this manual, operators need to constantly monitor areas or risk and practise “all-in all out” measures. For example: the origin of fertilised eggs, certified biosecured and reliable hatcheries and their incubators, certified feed sources and

transport companies must be registered, dated and documented. The poultry operation must register a complete account of their activities and sources, which include:

- Census of production – stages and location
- Protocols for training of operators
- Instructions to operators within the farm (clothing, cleaning, vaccination, feeding, reporting, etc.)
- Protocols for cleaning and disinfection
- Purchases and location of suppliers
- Vermin and insect control measures
- Egg crate circulation, management and acquisition
- Employee profiles and responsibilities
- Transport control on and off the premises
- Employee and employee-family awareness
- Registries open to frequent regulatory inspection

One area of concern for regulatory authorities is the reality that many highly developed production poultry operations have their own diagnostic laboratories that may carry out diagnostic assays without reporting to the authorities. It is indispensable that such commercially associated laboratories and their managers be made aware of the importance of reporting disease occurrence and consequences to international trade for the country and their enterprise.

Commercial operators should be linked with prevention, contingency and emergency plans for national and regional success and health.

Annexes

Annex 1

Selected references for further information

1. *Guiding principles for highly pathogenic avian influenza surveillance and diagnostic networks in Asia*. <http://www.fao.org/ag/againfo/subjects/en/health/diseases-cards/Guiding%20principles.pdf>
2. *FAO position paper on AI control strategy*.
3. *Global strategy for the progressive control of highly pathogenic avian influenza (HPAI)*. <http://www.fao.org/ag/againfo/subjects/documents/ai/HPAIGlobalStrategy31Oct05.pdf>
4. Ausvetplan. *Disease Strategy – Avian influenza; Operational Procedures Manual – Destruction; Operational Procedures Manual – Disposal*. www.animalhealthaustralia.com.au/aahc/index.cfm
5. *FAO EMPRES Good Emergency Management Practice*. <http://www.fao.org/ag/aga/agah/empres/gemp/intro/2300-gemp.html>
6. Geering WA, Forman AJ and Nunn MJ (1995). *Exotic Diseases of Animals: A field guide for Australian Veterinarians*. Bureau of Resource Sciences, Australian Government Publishing Service, Canberra.
7. Geering WA, Roeder PL and Obi TU (1999). *Manual on the Preparation of National Animal Disease Emergency Preparedness Plans*. FAO Animal Health Manual No. 6. FAO. Rome, 1999.
8. *OIE Manual for Diagnostic Tests and Vaccines for Terrestrial Animals*. http://www.oie.int/eng/normes/mmanual/A_summry.htm
9. *OIE Guidelines for HPAI surveillance*.
10. *OIE Terrestrial Animal Health Code*.

Annex 2

Sample - Tender document for inactivated avian influenza vaccines

To be used for the purchase of inactivated avian influenza vaccine by governments or donor organizations for control of disease in infected countries.

INTRODUCTION

These are specifications for the purchase of inactivated avian influenza vaccine for use in poultry, including chickens and ducks. Vaccination is used to provide protection in the face of possible exposure or to reduce the viral load in an infected environment. Vaccinated birds are generally not fully protected from infection but have increased resistance to infection, suffer less clinical disease and shed substantially less virus.

There are several options for vaccine. Conventional vaccine, for which this specification is written, is prepared from the allantoic fluid of infected eggs, which is inactivated and emulsified with adjuvant. Attenuated live influenza virus vaccines are not recommended because of the risk that the vaccine virus could either mutate or re-assort with other influenza viruses to become pathogenic. However, recombinant vaccines have been produced, including fowl-pox virus, with the influenza haemagglutinin gene inserted and haemagglutinin produced in a baculovirus expression system. This specification does not cover the requirements for recombinant vaccines.

The virus type used for vaccine production must be of the same haemagglutinin type as the outbreak virus. For maximum potency, it is preferable for the vaccine virus to be closely related to the outbreak strain. If post-vaccination monitoring depends on serology to determine whether antibody-positive birds have been infected or vaccinated (the DIVA test), the neuraminidase type should be different to that of the outbreak strain.

TENDER SPECIFICATION

General requirements

1. Vaccine manufacture must be undertaken in accordance with OIE Guidelines – Chapters 1.1.7 and 2.7.12 of the *Manual of Diagnostic Tests and Vaccines for Terrestrial Animals*, 5th edition 2004. It must be produced under Good Manufacturing Practice (GMP) and under acceptable third-party audited quality assurance.
2. In assessing the acceptability of a vaccine, [FAO or other purchaser] may require documentation to be furnished to validate GMP and quality assurance practices and the production details for a specific vaccine batch. [The purchaser] may also seek to undertake an audit of the manufacturing plant(s).

3. The vaccine must be registered or otherwise acceptable for use, by the government of *[insert country]*.

Specific requirements

1. The requirement is for *[insert number]* of doses of vaccine for use in *[insert species]*.
2. The vaccine must contain haemagglutinin antigen of H *[insert type – for current SE Asia epidemic, H5]* type. Evidence (challenge or VN test) should be provided that the vaccine protects against the virus strains currently circulating in *[insert country/region]*.
3. Evidence should be provided that vaccine produced by the same means (i.e. not an individual batch requirement) in the same manufacturing plant significantly reduces virus transmission from vaccinated birds when subsequently infected.
4. The vaccine virus must be derived from an LPAI virus strain.
5. The virus should be grown in specific antibody negative or specific pathogen free eggs.
6. The virus is to be inactivated with formalin or beta-propiolactone.
7. The vaccine should be emulsified with a mineral oil adjuvant or with an alternative adjuvant with similar immuno-stimulating efficacy.
8. The vaccine must have undergone appropriate sterility, safety and potency tests in accordance with international standards.
9. The vaccine must have a minimum of one microgramme per dose of haemagglutinin protein. Vaccine of a higher haemagglutinin concentration will be considered favourably. Alternatively, the potency of the batch may be demonstrated by live bird challenge with virulent virus or by a minimum HI antibody response of 1:32 in vaccinated birds.
10. Packaging of the vaccine should be in containers of *[insert number of doses]*.
11. Labelling in *[insert language/s]* must indicate manufacturer, type of vaccine, batch identification, volume of contents, storage recommendations and expiry date. Package insert in *[insert language/s]* to include instructions for vaccinating poultry, recommended species to which the vaccine applies, vaccination regime and dose.
12. Vaccine to have a minimum of six months period prior to expiry, on delivery.
13. Vaccine must be delivered to cold storage in *[insert place or country]*. Verification will be required of continuity of appropriate storage of the vaccine from production to delivery.

Annex 3

OIE/FAO reference laboratories and experts for avian influenza

Dr Ilaria Capua

Istituto Zooprofilattico Sperimentale delle Venezie, Laboratorio Virologia

Via Romea 14/A, 35020 Legnaro, Padova

ITALY

Tel: (39.049) 808.43.69 Fax: (39.049) 808.43.60

Email: icapua@izsvenezie.it

Dr Ian Brown

VLA Weybridge

New Haw, Addlestone, Surrey KT15 3NB

UNITED KINGDOM

Tel: (44.1932) 34.11.11 Fax: (44.1932) 34.70.46

Email: i.h.brown@vla.defra.gsi.gov.uk

Dr Paul W. Selleck

CSIRO, Australian Animal Health Laboratory (AAHL)

5 Portarlington Road, Private Bag 24, Geelong 3220, Victoria

AUSTRALIA

Tel: (61.3) 52.27.50.00 Fax: (61.3) 52.27.55.55

Email: paul.selleck@csiro.au

Dr B. Panigrahy

National Veterinary Services Laboratories

P.O. Box 844, Ames, IA 50010

UNITED STATES OF AMERICA

Tel: (1.515) 663.75.51 Fax: (1.515) 663.73.48

Email: brundaban.panigrahy@aphis.usda.gov

Dr Ortrud Werner

National Reference Laboratory for Highly Pathogenic Avian Influenza and Newcastle Disease,

Institute of Diagnostic Virology, Federal Research Centre for Virus Diseases of Animals (BFAV)

Insel Riems, Boddenblick 5a, 17493 Greifswald - Insel Riems

GERMANY

Tel: (41) 383.517.152 Fax: (41) 383.517.151

Email: ortrud.werner@rie.bfav.de

Dr Hiroshi Kida

Graduate School of Veterinary Medicine, Hokkaido University, Department of Disease Control

Kita-18, Nishi-9, Kita-ku, Sapporo 060-0818

JAPAN

Tel: (81.11) 706.52.07 Fax: (81.11) 706.52.73

Email: kida@vetmed.hokudai.ac.jp

For more information, visit the OFFLU website www.offlu.net

Annex 4

Information for shipping international diagnostic specimens

**OIE/FAO AND NATIONAL REFERENCE LABORATORY FOR NEWCASTLE DISEASE AND AVIAN INFLUENZA VIROLOGY DEPARTMENT.
ISTITUTO ZOOPROFILATTICO SPERIMENTALE (IZS) DELLE VENEZIE (ITALY)
(As of November 2005)**

Important: Contact the IZSVe in order to discuss testing and testing materials before shipping. Provide the name and details of a contact person.

Types of specimen

Specimens submitted may be virus isolates made in the submitting country or clinical specimens, such as tissues or swabs, collected from diseased birds.

Packaging requirements

All materials should be in leak-proof containers. Packaging should be composed of (1) a primary receptacle, (2) a secondary packaging and (3) a rigid outer packaging. Packaging of "diagnostic samples" should be coded UN3373 and comply with IATA PI650 standard. "Virus isolates" should be coded UN2814 for AIV and UN2900 for NDV complying with IATA PI602 standards.

Contact couriers to confirm the provision of boxes complying with these requirements.

Documents to be accompanied for clearing

Import permissions from the Italian Ministry of Health (formerly provided by the IZSVe) and a *signed proforma invoice* (The template will be formerly provided by the IZSVe) should be attached firmly to the box.

Shipping modality

Air freight or couriers to Milan Malpensa Airport (recommended), Rome Fiumicino Airport (courier only) or Venice Marco Polo Airport (no virus isolates). Informing the IZSVe 1 week in advance is recommended. *Arrange for shipments to arrive in Italian airports from Monday to Thursday only.*

Shipping Address

Instituto Zooprofilattico Sperimentale delle Venezie
Virology Department
Viale dell'Università 10, 35020 Legnaro
Padova, Italy

Notification of shipment

Before shipping, please supply the following information to the IZS contact person

- Embarkation date
- Airline name and flight number
- Name of the destination airport
- Date of arrival in Italy
- Airway bill number (the air bill should be faxed as soon as possible to: Fax + 39 049 8084360 or sent via e-mail to the contact person)
- Contact person to whom the results should be sent (name, fax number, e-mail address)

Contact people at the IZS

To ship diagnostic samples and virus isolates:

Elena Ormelli

E-mail: eormelli@izsvenezie.it

Micaela Mandelli

E-mail: mmandelli@izsvenezie.it

For reagents:

William Dundon

E-mail: wdundon@izsvenezie.it

Micaela Mandelli

E-mail: mmandelli@izsvenezie.it

Other contact persons:

Giovanni Cattoli

E-mail: gcattoli@izsvenezie.it

Alessandro Cristalli

E-mail: acristalli@izsvenezie.it

Paola De Benedictis

E-mail: pdebenedictis@izsvenezie.it

**AVIAN VIROLOGY LABORATORY, VETERINARY LABORATORIES AGENCY,
WEYBRIDGE, UK. FROM OUTSIDE THE EU
(As of February 2006)**

Packaging requirements

All materials should be in leak-proof containers. At least two layers of packaging should be used and the inner layer treated lightly with disinfectant.

The outer packaging must be marked as follows:

ANIMAL PATHOGEN – PACKAGE ONLY TO BE OPENED AT THE AVIAN VIROLOGY SECTION, VETERINARY LABORATORIES AGENCY, WEYBRIDGE, SURREY

and with one of the following IMPORT LICENCE NUMBERS:

For Newcastle disease:

AHZ/2232/2002/5

For Avian Influenza, other viruses, avian tissue, serum, faeces and eggs:

AHZ/2074C/2004/3

Shipping Address

Ruth Manvell

Avian Virology, VLA Weybridge
New Haw, Addlestone, Surrey KT15 3NB,
United Kingdom

A letter should accompany the parcel with as much history about the isolates as possible, to include species and age, area/country of isolation, any clinical history, etc. If sending by **AIR FREIGHT** it is essential that the **AIRWAY BILL NUMBER** is given to VLA-Weybridge by FAX, telephone or e-mail before the arrival of the materials in order to facilitate an early delivery.

Packages sent by air freight should be clearly marked: CARE OF TRANSGLOBAL to ensure rapid processing at the airport.

Notification of shipment

Please notify the VLA-Weybridge, Avian Virology Laboratory of the shipment details and the contact person for information on the result (name, fax number, e-mail address) before dispatch.

Direct FAX: +44 (0)1932 357856

Direct Tel: +44 (0)1932 357 736

E-mail: r.manvell@vla.defra.gsi.gov.uk

Contact

If you wish to discuss a submission and options for support from the International Reference Laboratory for Avian Influenza and Newcastle Disease, please contact:

Dr. I.H.Brown

Direct Tel: +44 (0)1932 357 339

Direct FAX: +44 (0)1932 357 239

E-mail: i.h.brown@vla.defra.gsi.gov.uk

AUSTRALIAN ANIMAL HEALTH LABORATORY (AAHL) (As of November 2005)

Types of specimen

Specimens submitted to AAHL for disease diagnosis may be either virus isolates made in the submitting country or clinical specimens, such as tissues or swabs, collected from diseased birds.

Import permit and packing

Copies of Australian import permits are available from AAHL by contacting aahl-accessions@csiro.au. All specimens must be packed in leak-proof containers in accordance with the appropriate IATA regulation and appropriately labelled. Suitable transport containers and packing instructions are also available from AAHL by contacting aahl-accessions@csiro.au. Copies of the import permit and other consignment details should be attached to the outside of the package to expedite clearance through Australian customs.

Notification of shipment

If submitting specimens, please notify the accessions clerk on accessions@csiro.au, the Duty Veterinarian on dutyvet@csiro.au or Dr. Peter Daniels on + 61 3 5227 5000 of the consignment details so that specimens can be collected upon arrival in Australia. Alternatively send the information by fax to +61 3 5227 5555. Consignment details include the consignment note/air weigh bill number, courier/airline and expected arrival date.

Shipping address

The Director

Australian Animal Health Laboratory
5 Portarlington Road, Geelong, 3220
Australia
Telephone: +61 3 5227 5000
Facsimile: +61 3 5227 5555
<http://www.csiro.au/aahl>

Contact for Avian Influenza

You may also wish to discuss the testing required with Peter Daniels (peter.daniels@csiro.au) or Paul Selleck (paul.selleck@csiro.au) on +61 3 5227 5000 prior to submitting the specimens.

**NATIONAL VETERINARY SERVICES LABORATORIES (NVSL),
AMES, IOWA, USA
(As of November 2005)**

Import permit

Packages containing diagnostic specimens or organisms (infectious materials) imported from foreign locations into the United States must be accompanied by a permit issued by the U.S. Department of Agriculture. The importation permit, with proper packaging and labelling, will expedite clearance of the package through U.S. customs. One copy of the permit should be attached to the outside of the shipping container and a second copy placed just inside the lid of the outer shipping container. The importation permit can be obtained from the laboratory (NVSL, Ames, Iowa).

Packaging requirements

All materials should be in leak-proof containers and packaged to withstand breakage. All materials should be properly labelled.

Shipping address

Director,

National Veterinary Services Laboratories
Diagnostic Virology Laboratory
1800 Dayton Avenue, Ames
Iowa, USA 50010

Notification shipment

Please notify the Diagnostic Laboratory with shipping information (date of arrival, airline/courier, weigh bill number, etc.) as soon as it is available. Fax information to +1 (515) 663-7348 or telephone +1 (515) 663-7551.

Contact for Avian Influenza

Dr. Beverly J. Schmitt

Direct Tel: +1 515/663-7532

Direct Fax: +1 515/663-7348

E-mail: beverly.j.schmitt@usda.gov

Annex 5

Criteria for defining infected areas and disease control zones

INFECTED PLACES

A place classified as an *infected place* (IP) will be a defined area (village, farm) in which *highly pathogenic avian influenza* (HPAI) disease or a highly virulent strain of AI virus exists, or is believed to exist. An IP will be subject to quarantine. A mildly or lowly virulent AI virus may be declared an agent for eradication if it has the potential to mutate into virulent AI virus.

DANGEROUS CONTACT PLACES

Places classified as *dangerous contact places* (DCPs) will be those that contain birds, poultry products or poultry waste that have recently been introduced from an IP (usually up to 21 days before the premises were declared infected) and are likely to be infected or contaminated or any of these items that may have been in substantial contact with people, vehicles and equipment that have been associated with an infected premises within three days of visiting the DCPs.

SUSPECT PLACES

Places classified as *suspect places* (SPs) will be those that contain birds that have possibly been exposed to an AI virus, such that quarantine and surveillance, but not pre-emptive slaughter, are warranted; or birds not known to have been exposed to an AI virus but showing clinical signs requiring differential diagnosis.

The classification SPs is a temporary classification and should be treated as infected until determined otherwise. High priority should be given to clarifying the status of the suspect birds so that the SPs can be reclassified as either an infected premise (IP) and appropriate quarantine and movement controls implemented, or as free from disease, in which case no further disease control measures are required.

RESTRICTED AREA

A *restricted area* (RA) will be a relatively small declared area compared to a *control area* (CA) around IP that are subject to intense surveillance and movement controls. Movement out of the area will, in general, be prohibited, while movement into the area would only be by allowed. Multiple RAs may exist within one CA.

The RA does not need to be circular but can have an irregular perimeter provided the boundary is initially an appropriate distance from the nearest IP, DCP or SP. This distance will vary with the size and nature of the potential source of virus, but will be approximately

1-5 km around the IP, depending on the density of poultry premises. The boundary could be the perimeter fence of the IP if the IP is in an isolated location. The boundary in a densely populated area will take into account the distribution of susceptible birds and traffic patterns to markets, service areas, abattoirs and areas that constitute natural barriers to movement. If possible, hatcheries should be kept out of the RA.

CONTROL AREA

The *control area* (CA) will be a larger declared area around the RA(s) and, initially, possibly as large as a province where restrictions will reduce the risk of disease spreading from the RA(s). The boundary of the CA will be adjusted as confidence about the extent of the outbreak becomes clearer but must remain consistent with the *OIE Code* chapters on surveillance and zoning (Chapters 1.3.4 and 1.3.5; see Appendix 3). In general, surveillance and movement controls will be less intense and animals and products may be permitted to move under permit from the area.

The declaration of a CA also helps to control the spread of the outbreak from within the RA. The CA is a buffer zone between the RA and the rest of the country. The boundary does not have to be circular or parallel to that of the RA but should be 2–10 km from the boundary of the RA. In general, the movement of possibly contaminated things and materials within the CA is allowed but movement out of the CA is prohibited without CVO approval. This type of control area allows reasonable commercial activities to continue.

NB: When declaring RAs and CAs, the areas must not be larger than necessary, thus restricting the number of properties to be quarantined to only those deemed prudent. If flocks in a quarantine area are not depopulated, then the cost of keeping the birds beyond their normal market age could be substantial.

INTERNATIONAL CONSIDERATIONS

Under *OIE Code* definitions, an *infected zone* means a clearly defined territory in which a disease (listed in the *Code*) has been diagnosed. This area must be clearly defined and decreed by the veterinary authorities in accordance with the environment, the different ecological and geographical factors as well as all the epidemiological factors and the type of husbandry being practised. The territory in question should have a radius from the centre or centres of the disease of at least 10 km in areas with intensive livestock raising, and 50 km in areas where extensive livestock raising is practised.

In June 1993, the European Union published a decision laying down the criteria for classifying 'third countries' with regard to avian influenza and Newcastle disease. Annex C point 4 of this decision states:

Around confirmed outbreaks of disease a protection zone with a minimum radius of 3 km and a surveillance zone with a minimum radius of 10 km shall be implemented. In these zones stand still measures and controlled movements of poultry shall be in force until at least 21 days after the end of disinfection operations on the infected holding. Before lifting the measures in these zones the authorities shall carry out the necessary inquiries and sampling of the poultry holdings to confirm that disease is no longer present in the region concerned. The practicality of declaring a zone, the intensity of the industry and the transmissibility of virus causing an outbreak might mean a decision is taken to declare larger areas than those used by the EU.

Annex 6

Leaflet example

BIRD FLU (HIGHLY PATHOGENIC AVIAN INFLUENZA)

The purpose of this leaflet is to inform the public, especially poultry farmers and those responsible for meat markets with basic information on Avian Influenza, commonly known as Bird Flu. This leaflet also informs them about the rights and responsibilities in case of eventual disease outbreaks.

WHAT IS BIRD FLU

Bird Flu (avian influenza) is a highly contagious viral disease of poultry and other birds. According to the virulence (ability and degree to cause disease), the bird flu virus is often characterized as:

- Highly pathogenic – causing illness with a high death rate (over 75%)
- Low pathogenic – causing mild signs of disease, but in case of secondary infections can cause serious problems with death rates of up to 50%.

Some low pathogenic types can change over time to become highly pathogenic.

A current strain of avian flu virus, known as H5N1, is circulating in many countries in Southeast Asia, and has recently been confirmed also in Russia, Romania, Croatia and Turkey. The H5N1 can cause disease in humans, too.

WHO CAN BE INFECTED BY BIRD FLU?

Domestic poultry – chickens and turkeys - are most often affected, while ducks and geese often develop only mild signs of the disease. Wild birds, especially wild ducks may be naturally resistant to the infection and do not show clinical signs of the disease.

Birds that do not show signs of the disease can still carry the virus and represent a danger for the introduction of the virus in poultry operations.

HOW IS THE DISEASE SPREADING?

The main sources of infection in poultry are the following:

- Live infected birds even if they don't appear sick
- Faecal droppings and discharges from sick birds
- Dead Birds
- Contaminated objects, including equipment (egg crates, cages), shoes or clothing, and contaminated ground

It is believed that the disease can be spread over long distances by migrating birds (especially wild ducks and geese, but possibly other water or shore birds). If infected wild birds have contact with domestic poultry during their resting times along their migrating routes, transmission of the virus could occur.

Within the country the disease is most commonly spread in poultry through the movement of people, birds and goods in an infected area and marketing practices used when infected poultry come into contact with healthy birds. Humans play a very important role in spreading the disease since the virus can easily be carried on dirty clothes, shoes, contaminated equipment, vehicles and in the transportation of sick poultry.

Though rare, the disease can be introduced by importing healthy live poultry or poultry products (meat, unprocessed feathers, laying eggs etc.). The virus can also be introduced to an unaffected area or country through illegal trade, especially live birds.



HOW TO RECOGNIZE THE DISEASE IN THE FLOCK

Bird flu spreads very quickly within the flock, so almost all units will be infected in a very short time and birds may die within days. The spread of the disease is likely to be slower in layers - chickens that are used to produce eggs – since they are often in cages and do not mix with each other.

Signs of disease are:

- Depression and lack of appetite
- Drastic drop in egg production
- Swollen head and neck
- Dark and swollen wattles and combs
- Bleeding under the skin
- Sudden death which can reach 100% of the flock

If any of your birds show these signs you should immediately report to the nearest veterinary station or to the veterinary inspector.

Failure to notify the authorities could put more animals -and even humans - at risk of becoming infected.



HOW TO PREVENT BIRD FLU

Basic prevention measures include:

- Decrease the opportunity for wild birds coming into contact with domestic poultry through the use of protective nets, or keeping the poultry in enclosed and protected buildings.
- Apply bio-safety measures:
 - Fence the farms to keep unwanted animals and visitors out. Lock doors!
 - Disinfect vehicles before they enter the farm.
 - Prohibit the entrance of unauthorized people to the farm.
 - Establish disinfection areas (foot-baths) before entering the farm or in each of the poultry houses.
 - Use boots and outer clothing that can be cleaned or changed between houses or farms. Use rubber gloves as well.
 - Clean and disinfect all surfaces regularly (cages, walls, poultry eating and watering areas).
 - Do not borrow equipment from other farms, as these may be contaminated.
 - Disinfect with detergents or hypochlorite solutions
- Replace animals from within the flock or from controlled and healthy flocks.
- Apply the principle of "all in – all out". This means that all animals in a poultry house are taken off the farm at the same time (for the market), the ground and house must be cleaned and disinfected, and only then can young stock be introduced into the cleaned house. Lock doors!
- Avoid keeping ducks, chickens and turkeys in the same yard.
- In case of an outbreak on your farm, you should immediately report to the veterinary authorities. Do not sell your animals. Do not eat or feed sick or dead birds to other animals.



CSIRO

PLANS FOR ERADICATING BIRD FLU

If the disease appears, the following measures would be undertaken:

- killing of all poultry in affected yards/farms and settlement areas
- safe disposal of poultry carcasses
- disinfection of contaminated yards/farms
- prohibition for movement of all kinds of live poultry, meat and other poultry products from contaminated settlement areas
- increased control for movement of people, mechanical equipment, vehicles, etc. from contaminated yards, farms and settlement areas
- other measures ordered by the veterinary inspectors.

Owners will be compensated for slaughtered birds as part of the measures for preventing the spread of the disease. Compensation will also be paid for destroyed poultry products and equipment according to their market value.

IS BIRD FLU A THREAT FOR HUMAN HEALTH?

The bird flu virus rarely infects people. If it does, the disease is usually not serious and is often characterized as reddening of the eye (conjunctivitis) with mild respiratory symptoms.

However, the highly pathogenic avian influenza H5N1 strain can be a serious health problem, and can kill people that are infected. Avoid contact with birds that are thought to be affected.

HOW DO PEOPLE BECOME INFECTED?

Avian influenza is not a food-borne disease. The bird flu virus is killed by the heat of normal cooking. There is no risk of getting avian influenza from properly cooked poultry meat and eggs. However, sick chickens should not be eaten, as a sick bird often releases toxins and has other microorganisms that may pose a danger to you and your family.

WHICH GROUPS OF PEOPLE ARE AT RISK?

People that are at higher risk are those who work with poultry (breeders, buyers, transporters, slaughterhouse workers and also veterinarians), who are in contact with infected poultry material or are employed in disease control activities (veterinarians employed in laboratories). Hunters and ornithologists are only at risk if they are handling sick or animals that have died due to avian influenza virus.

HOW PEOPLE CAN BE PROTECTED?

Currently, there is no medicine that could fully protect people from bird flu. Present seasonal vaccines against human flu are only effective against known circulating human influenza. Should you or anyone in your family have a fever with flu-like symptoms, it is advised to see medical attention immediately.

WHO DO I CONTACT?

This area to be used for information for local or regional veterinary authority, diagnostic laboratory, help desk, or call-free number.

NB: *this poster was designed by authorities of Macedonia*

Annex 7

Avian influenza vaccine producers and suppliers

Compiled by FAO's Emergency Prevention System for Transboundary Animal and Plant Pests and Diseases (EMPRES). The manufacturers/suppliers and their vaccines are not necessarily endorsed by FAO and it is the [importing] country's responsibility to establish independent quality assurance/quality control for safety, purity, potency and efficacy parameters.

China

Laboratory	Affiliation	Location	Strain	Type	Antibody levels	Established Production	Website	Comments
Harbin Veterinary Research Institute		Harbin, Heilongjiang province	A/Turkey/England/N-28/73 subtype H5N2, Low pathogenicity	Inactivated monovalent	8log2 reached by the fifth week after vaccination and maintained for 4 weeks, protective antibody titers maintained for 23 weeks	Dec. 2003	www.hvri.ac.cn	
Harbin Veterinary Research Institute		Harbin, Heilongjiang province	A/Goose/Guangdong/1996, Subtype H5N1	Inactivated monovalent				Used for ducks and other waterfowl in Vietnam
Harbin Veterinary Research Institute		Harbin, Heilongjiang province	A recombinant virus from A/Goose/Guangdong/1996 and human influenza vaccine virus H5N1	Inactivated recombinant	9log2 reached by the third week after vaccination and maintained for 4 weeks, protective antibody titers maintained for 25 weeks	Jan. 2005	www.hvri.ac.cn	High specificity, antibody level and long maintain time, vaccinated water fowl do not shed virus and are resistant against infection proved by the laboratory results
Harbin Veterinary Research Institute		Harbin, Heilongjiang province	A recombinant avian pox virus expressed H5 from A/Goose/Guangdong/1996	Live recombinant avian pox virus vectored H5	7log2 reached by the second week after vaccination protective antibody titers maintained for 26 weeks	Jan. 2005	www.hvri.ac.cn	Induced only antibody against specific protein, helpful in differentiation of the immune and the infection. The immune efficiency is less than the other two vaccines, and price is higher. Only for chicken
Zhengzhou Bio-pharm Co. Ltd	China Animal Husbandry Group	Zhengzhou City, Shandong province	A/Turkey/England/N-28/73 subtype H5N2	Inactivated monovalent	8log2 reached by the fifth week after vaccination and maintained for 4 weeks, protective antibody titers maintained for 23 weeks	Dec. 2003		
Qingdao Yebio Bioengineering Co. Ltd	National Animal Quarantine Institute of the Ministry of Agriculture	Qingdao City, Shandong province	A/Turkey/England/N-28/73 subtype H5N2	Inactivated monovalent	8log2 reached by the fifth week after vaccination and maintained for 4 weeks, protective antibody titers maintained for 23 weeks	Dec. 2003	www.yebio.com.cn	
Qingdao Yebio Bioengineering Co. Ltd	National Animal Quarantine Institute of the Ministry of Agriculture	Qingdao City, Shandong province	H9	Inactivated			www.yebio.com.cn	
Qingdao Yebio Bioengineering Co. Ltd	National Animal Quarantine Institute of the Ministry of Agriculture	Qingdao City, Shandong province	H5N2/H9	Inactivated bivalent			www.yebio.com.cn	
Guangdong Yongshun Bio-pharm Co. Ltd		Guangdong province	A/Turkey/England/N-28/73 subtype H5N2	Inactivated monovalent	8log2 reached by the fifth week after vaccination and maintained for 4 weeks, protective antibody titers maintained for 23 weeks	Dec. 2003		
Zhaoqing Dahua agriculture Bio-pharm Co. Ltd	Veterinary College of Southern China Agriculture University	Zhaoqing City, Guangdong	A/Turkey/England/N-28/73 subtype H5N2	Inactivated monovalent	8log2 reached by the fifth week after vaccination and maintained for 4 weeks, protective antibody titers maintained for 23 weeks	Dec. 2003	http://www.un-pur.org/gongyingshangmulu/yiyao/zhaqing/zhaqing.htm	
Liaoning Yikang Bioengineering Co. Ltd		Liaoyang City, Liaoning Province	A/Turkey/England/N-28/73 subtype H5N2	Inactivated monovalent	8log2 reached by the fifth week after vaccination and maintained for 4 weeks, protective antibody titers maintained for 23 weeks	Dec. 2003		
Nanjing Merial Animal Products Co., Ltd	China Animal Husbandry Group	Nanjing City, Jiangsu province	A/Turkey/England/N-28/73 subtype H5N2	Inactivated monovalent	8log2 reached by the fifth week after vaccination and maintained for 4 weeks, protective antibody titers maintained for 23 weeks	Dec. 2003		
Qilu Animal Health Products Factory		Ji'nan City, Shandong province	A/Turkey/England/N-28/73 subtype H5N2	Inactivated monovalent	8log2 reached by the fifth week after vaccination and maintained for 4 weeks, protective antibody titers maintained for 23 weeks	Dec. 2003	www.qiludb.com	
Chengdu Jianghua Bioproducts Co. Ltd	Jianghua Group	Ziyang City, Sichuan province	A/Turkey/England/N-28/73 subtype H5N2	Inactivated monovalent	8log2 reached by the fifth week after vaccination and maintained for 4 weeks, protective antibody titers maintained for 23 weeks	Dec. 2003	http://www.jinghuagroup.net/main.asp	
Merial International Trading Company	Merial (France)		H5N2	Inactivated monovalent				Commercial name: FLU H5N2 Formalin inactivated. Mineral oil adjuvant

France

Laboratory	Affiliation	Location	Strain	Type	Antibody levels	Established Production	Website	Comments
Meriel			H9N2 or H7N1 (Some H7N3) strains as separate vaccines	Inactivated monovalent				
Meriel			A/chicken/Italy/1067/99 (H7N1) and A/chicken/Italy/22A/98 (H5N9)	Inactivated bivalent			www.merial.com	Commercial name: BioFlu H7N1 and H5N9. For chicken
Meriel			H5 from A/Turkey/Ireland/83 recombinant Fowlpox vector	Recombinant live			www.merial.com	Commercial name: Trovac AIV H5. Protection after day old vaccination birds up until 20 weeks of age. The vaccine is produced in USA
Laprovét S.A.S		Tours, Cedex 2, France	H5N2	Inactivated monovalent				Commercial name: ITA-FLU. Formalin inactivated. Mineral oil adjuvant. The vaccine is produced in Mexico.
Ceva Santé Animale SA		Libourne, France	A/Chicken/Mexico/232/94/CPA. Oil emulsified. H5N2	Inactivated	8Log 2.Four weeks after vaccination at 10 days/Subcutaneous	Approved 2004	www.ceva.com	Commercial name: FLU-KEM. Binary ethyleneimine (BEI) inactivated. Mineral oil adjuvant. The vaccine is produced in Mexico through CEVA de MEXICO

Germany

Laboratory	Affiliation	Location	Strain	Type	Antibody levels	Established Production	Website	Comments
Boehringer Ingelheim Vetmedica, GmbH		Ingelheim am Rhein, Germany	H5N2. A/Chicken/Mexico/232/94/CPA. Oil emulsified	Inactivated monovalent	8Log 2.Four weeks after vaccination at 10 days/Subcutaneous.			Mineral oil adjuvant. *The vaccine is produced in Mexico by Boehringer Ingelheim Vetmedica S.A. de C.V.

Italy

Laboratory	Affiliation	Location	Strain	Type	Antibody levels	Established Production	Website	Comments
Meriel Italy Spa	Meriel (France)	Italy	H5N9	Inactivated monovalent	Minimum HI antibody response of 1:32 in vaccinated birds			Commercial name: Gallimune Flu H5N9. Beta-propiolactone inactivated. Mineral oil adjuvant.

Mexico

Laboratory	Affiliation	Location	Strain	Type	Antibody levels	Established Production	Website	Comments
Intervet		Mexico City	A/Chicken/Mexico/232/94/CPA. Oil emulsified.	Inactivated monovalent	8Log 2. Four weeks after vaccination at 10 days of age/Subcutaneous	Approved 2004	www.intervet.com.mx	
Intervet Mexico	Intervet (Netherlands)	Huixquilucan	H5N2 or H5N6	Inactivated monovalent				Commercial name: Nobilis influenza H5. Mineral oil adjuvant. The same vaccine is supplied by Intervet international b.v. in Netherlands
Boehringer Ingelheim Vetmedica, S.A. de C.V.		Guadalajara, Jalisco	A/Chicken/Mexico/232/94/CPA. Oil emulsified H5N2	Inactivated monovalent	8Log 2. Four weeks after vaccination at 10 days/Subcutaneous.	Approved 2004	www.lineavolvac.com	Mineral oil adjuvant. This same vaccine produced in Mexico is supplied in Germany through Boehringer Ongelheim Vetmedica GmbH
Avimex laboratories		Mexico City	A/Chicken/Mexico/232/94/CPA. Oil emulsified. H5N2	Inactivated monovalent	8Log 2. Four weeks after vaccination at 10 days/Subcutaneous.	Approved 2004	www.avimex.com.mx	
Investigación Aplicada S.S. (IASA)		Tehuacan, Puebla	A/Chicken/Mexico/232/94/CPA. Oil emulsified. H5N2	Inactivated monovalent	8Log 2. Four weeks after vaccination at 10 days/Subcutaneous.	Approved 2004	www.iasa.com.mx	
Ceva de Mexico	Ceva Santé Animale SA (France)	Mexico City	A/Chicken/Mexico/232/94/CPA. Oil emulsified. H5N2	Inactivated monovalent	8Log 2. Four weeks after vaccination at 10 days/Subcutaneous.	Approved 2004	www.ceva.com	Commercial name: FLU-KEM. Mineral oil adjuvant. This vaccine is supplied in France through CEVA Santé Animale
Ceva de Mexico	Ceva Santé Animale SA (France)	Mexico City	A/Chicken/Mexico/232/94 (H5N2+LaSota NDV)					Commercial name: NEW-FLU-KEM AI and Newcastle disease

Netherlands

Laboratory	Affiliation	Location	Strain	Type	Antibody levels	Established Production	Website	Comments
Intervet International b.v.		Boxmeer, Netherlands	A/Chicken/Mexico/232/94/CPA H5N2 or H5N6	Inactivated			www.intervet.com	Commercial name: Nobilis Influenza H5N2 or Nobilis Influenza H5N6. Formalin or beta-propiolactone inactivated. Mineral oil adjuvant. The vaccine is produced in Mexico and Spain.
Intervet			A/CK/Italy/473/99 (H7N1)	Inactivated			www.intervet.com	Commercial name: Nobilis Influenza H7N1. Oil adjuvant.
Intervet			A/duck/Potsdam/15/80 (H7N7)	Inactivated			www.intervet.com	Commercial name: Nobilis Influenza H7N7.

Pakistan

Laboratory	Affiliation	Location	Strain	Type	Antibody levels	Established Production	Website	Comments
Sindh Vaccine Production Centre		Karachi, Pakistan	H7N3 and H9N2 strains, separate or combined products					
Biolab (pvt) Ltd		Rawalpindi, Pakistan	H7N3 and H9N2 strains for both separate and combined products					
Avicina Laboratories		Lahore, Pakistan	H7N3 and H9N2 strains for both separate and combined products					
Otoman Pharma		Lahore, Pakistan	H7N3 and H9N2 strains for both separate and combined products					

USA

Laboratory	Affiliation	Location	Strain	Type	Antibody levels	Established Production	Website	Comments
Biomune vaccines	Ceva Santé Animale SA	Lenexa - Kansas, USA	A/Turkey/Wisconsin/68 (H5N9)	Inactivated				Commercial name: Layermune AIV H5N9. Beta-propiolactone inactivated. Mineral oil adjuvant.
Biomune vaccines	Ceva Santé Animale SA		A/Chicken/New York/273874/03 (H7N2)	Inactivated				Commercial name: Layermune AIV H7N2
Biomune vaccines	Ceva Santé Animale SA		A/Turkey/Utah/24721-10/95 (H7N3)	Inactivated				Commercial name: Layermune AIV H7N3
Fort Dodge Animal Health		Overland Park, USA	H5N3	Inactivated				Commercial name: Poulvac Flu Fend I H5N3 RG. Formalin inactivated. Water in oil adjuvant.
Fort Dodge Animal Health		Overland Park, USA	H5N3	Inactivated				Commercial name: Inactivated AIV type A, H5N2, H5N9, H7N2, H7N3. Formalin inactivated. Water in oil adjuvant.

FAO ANIMAL PRODUCTION AND HEALTH MANUALS

1. Small-scale poultry production, 2004 (E, F)
2. Good practices for the meat industry, 2004 (E)
3. Preparing for highly pathogenic avian influenza, 2006 (E)

Availability: June 2006

Ar	-	Arabic	Multil	-	Multilingual
C	-	Chinese	*		Out of print
E	-	English	**		In preparation
F	-	French			
P	-	Portuguese			
R	-	Russian			
S	-	Spanish			

The FAO Animal Production and Health Manuals are available through the authorized FAO Sales Agents or directly from Sales and Marketing Group, FAO, Viale delle Terme di Caracalla, 00100 Rome, Italy.

FAO ANIMAL HEALTH MANUALS

1. Manual on the diagnosis of rinderpest, 1996 (E)
2. Manual on bovine spongiform encephalopathy, 1998 (E)
3. Epidemiology, diagnosis and control of helminth parasites of swine, 1998
4. Epidemiology, diagnosis and control of poultry parasites, 1998
5. Recognizing peste des petits ruminant - A field manual, 1999 (E, F, A)
6. Manual on the preparation of national animal disease emergency preparedness plans, 1999 (E)
7. Manual on the preparation of rinderpest contingency plans, 1999 (E)
8. Manual on livestock disease surveillance and information systems, 1999 (E)
9. Recognizing African swine fever. A field manual, 2000 (E, F)
10. Manual on Participatory Epidemiology - Method for the Collection of Action-Oriented Epidemiological Intelligence, 2000 (E)
11. Manual on the preparation of african swine fever contingency plans, 2001 (E)
12. Manual on procedures for disease eradication by stamping out, 2001 (E)
13. Recognizing contagious bovine pleuropneumonia, 2001 (E, F)
14. Preparation of contagious bovine pleuropneumonia contingency plans, 2002 (E, F)
15. Preparation of Rift Valley fever contingency plans, 2002 (E, F)
16. Preparation of foot-and-mouth disease contingency plans, 2002 (E)
17. Recognizing Rift Valley fever, 2003 (E)

Highly pathogenic avian influenza (HPAI) represents a threat to poultry industries worldwide and to people's livelihoods, and a potential threat to human health. The international community has a vested interest in minimizing the spread of this disease. Countries may be under threat of introduction of avian influenza through exposure of poultry to wild birds, especially waterfowl. They may also be at risk from introduction of infected or contaminated poultry, poultry products or fomites.

The Food and Agriculture Organization of the United Nations and the World Organisation for Animal Health have prepared this manual to help national animal health authorities and other stakeholders prepare for a possible incursion of HPAI, detect disease as soon as possible and respond as rapidly as possible to contain the disease.

The manual offers practical advice on disease identification, pathology and diagnosis; detection, response and control strategies; and biosecurity measures to prevent outbreaks. It is an invaluable source of useful information for anyone involved in poultry-keeping and animal health practices.

ISBN 92-5-105548-3 ISSN 1810-1119



9 789251 055489

TC/M/A0632E/1/07.06/5000