

March 2011



The International Treaty

ON PLANT GENETIC RESOURCES FOR FOOD AND AGRICULTURE

**E****Item 15 of the Provisional Agenda****FOURTH SESSION OF THE GOVERNING BODY****Bali, Indonesia, 14 – 18 March 2011****DRAFT REVISED GENE BANK STANDARDS***Note from the Secretary*

The Governing Body, at its Third Session, welcomed the revision of *Genebank Standards* and invited the Commission on Genetic Resources for Food and Agriculture (Commission) to commence and coordinate the process. It requested its Bureau to coordinate with the Bureau of the Commission the agendas of the respective bodies for the purpose of discussing modalities for such revision and the ways and means for input of the Governing Body in the process. The Second Joint Bureau meeting held in November 2010 requested that the draft revised *Genebank Standards* for orthodox seeds be made available to the Fourth Session of the Governing Body as an information document. The draft revised Genebank Standards are presented in this document.

TABLE OF CONTENTS

	<i>Pages</i>
I. Introduction	3
II. Preparation of the draft revised Genebank Standards	3
III. Key features of the draft revised Genebank Standards	4
<i>Appendix:</i> Draft Revised Genebank Standards for Orthodox Seeds	5- 37

I. INTRODUCTION

1. The *Genebank Standards* were published in 1994, under the aegis of the then Commission on Plant Genetic Resources (now the Commission on Genetic Resources for Food and Agriculture (Commission)).¹ The Commission at its Twelfth Regular Session, agreed on the need for revising the *Genebank Standards* in order to ensure that plant genetic resources are conserved under conditions that meet recognized and appropriate standards, based on current and available technological and scientific knowledge. It requested FAO, in cooperation with the International Treaty on Plant Genetic Resources for Food and Agriculture (International Treaty), the Consultative Group on International Agricultural Research (CGIAR) and other relevant international institutions, to undertake this review for consideration by the Intergovernmental Technical Working Group on Plant Genetic Resources (Working Group), at its Fifth Session.²
2. The Governing Body of the International Treaty, at its Third Session, invited the Commission to commence and coordinate the process for the revision of the *Genebank Standards* and requested its Bureau to coordinate with the Bureau of the Commission the agendas of the respective bodies for the purpose of discussing modalities for such revision and the ways and means for input of the Governing Body in the process.³
3. The status of revision of the *Genebank Standards* was reviewed at the Second Joint Bureau Meeting of the Commission and the International Treaty held on 13 November 2010. The Bureaus agreed with the approach taken to limit the review, in a first step, to the original scope of the standards (i.e. orthodox seeds) and to consult the Commission on possible further steps to elaborate additional standards for plant species that cannot be maintained under relatively uniform conditions. The Joint Bureau also requested that the draft revised *Genebank Standards* for orthodox seeds be made available to the Fourth Session of the Governing Body as an information document.

II. PREPARATION OF THE DRAFT REVISED GENE BANK STANDARDS

4. As requested, FAO revised the *Genebank Standards* in cooperation with the International Treaty, Bioversity International, other CGIAR centres, GCDT and other relevant international institutions. In addition, through technical consultations, FAO sought and received inputs from various experts from national, regional and international genebanks.
5. A first draft of the revised *Genebank Standards* was shared, through a Circulate State Letter (AGP/GPA 11) with all National Focal Points for plant genetic resources, including the National Focal Points for the International Treaty, for comments and inputs and further distribution to relevant stakeholders.⁴
6. The consultations provided valuable inputs so that current scientific knowledge and changes in the conditions for *ex situ* conservation of orthodox seeds could be reflected in the revised version. The importance of active management of collections to optimize resource availability with the demands of germplasm maintenance was highlighted. The need for consideration of standards for non-orthodox seeds and associated conservation practices was also identified.

¹ CPGR/93/5 Annex.

² CGRFA-12/09/Report, paragraph 28.

³ IT/GB-3/09/Report, *Appendix A*, paragraph 20-21.

⁴ <http://www.fao.org/agriculture/crops/core-themes/theme/seeds-pgr/itwg/5th/en/>

III. KEY FEATURES OF THE DRAFT REVISED GENE BANK STANDARDS

7. The draft revised *Genebank Standards*, like the previous edition, is composed of separate sections. However their structure and presentation have been improved to provide more specificity. The Standards contain four main sections: Introduction, Underlying Principles, Standards and Appendices. The 'Introduction' covers the context of the revision and scope of the Standards. It is followed by the 'Underlying Principles', which provide a framework for setting the Standards and serve as the overarching principles of genebank management. The section on 'Standards' provides the specificity to adhere to the underlying principles. These Standards are presented upfront in a straightforward manner, followed by a narrative on technical aspects, contingencies and selected references. As in the earlier edition, these standards are essentially targets to aim for and remain nonbinding and voluntary in nature.
8. The draft revised *Genebank Standards* solely concern the conservation of seeds of orthodox species, including wild species, i.e. those species whose seed can survive considerable desiccation, and for which longevity can be improved by reducing seed storage moisture content and/or temperature. These standards should not be used for non-orthodox seeds or clonally-propagated material.⁵
9. The draft revised *Genebank Standards* presents one standard⁶, in contrast to the two levels, 'preferred' and 'acceptable' standards, used in the previous edition, mainly to avoid ambiguity or unnecessary duplications and optimize the use of limited resources. The revised standards take into account the changes in *ex situ* conservation conditions, diversity in storage requirements, purpose and period of germplasm conservation, ranging from temperate to tropical provenances. The revised standards are provided for nine areas that cover acquisition, seed drying and storage, viability monitoring, regeneration, characterization, documentation, distribution, safety duplication and security/personnel.
10. The draft revised *Genebank Standards* can be aimed at by all genebanks for conserving orthodox seed collections; however, in light of the continuous technological advances in conservation methods, much of it species-specific, they should be used in conjunction with species-specific information.
11. The draft revised *Genebank Standards* are provided as an Appendix to this document.

⁵ Orthodox seeds include many food crops, annual, biennial and horticultural species including grains, pulses, vegetables and temperate fruit and forest trees. Non-orthodox seeds also known as recalcitrants include aquatic habitats species, tropical fruit species, underutilized plants and temperate woody perennials.

⁶ A standard defines that lowest level of performance of a routine genebank operation below which there is a very high risk of losing genetic integrity (e.g. a probability of five percent or more of losing an allele in an accession over the storage period).

*Appendix***Draft Revised Genebank Standards for the Conservation of Orthodox Seeds****March 2011**

TABLE OF CONTENTS

1. Introduction	6
2. Underlying principles	7
3. Standards	10
3.1. Standards for acquisition	11
3.2. Standards for drying and storage	12
3.3. Standards for seed viability monitoring	18
3.4. Standards for regeneration	22
3.5. Standards for characterization	25
3.6. Standards for documentation	27
3.7. Standards for distribution	29
3.8. Standards for safety duplication	32
3.9. Standards for security/personnel	35
4. Appendices	
List of Acronymns and Abbreviations	37

1. Introduction

Genebanks around the world hold collections of a broad range of plant genetic resources, with the overall aim of long-term conservation and accessibility of the plant germplasm to plant breeders, researchers and other users. A sustainable conservation of these plant genetic resources depends on effective and efficient management of genebanks through the application of standards and procedures that ensure the survival and availability of plant genetic resources at present and in the future. For any conservation effort to be sustainable and successful it should also be cost effective and well managed.

The draft revised *Genebank Standards* arise from the revision of the FAO/IPGRI *Genebank Standards*, published in 1994. The revision was undertaken at the request from the Commission on Genetic Resources for Food and Agriculture (CGRFA) in light of the changes in the global policy landscape and advances in field of science and technology. The main policy developments that impact the conservation of plant genetic resources in genebanks lie within the context of availability and distribution of germplasm arising from the adoption of international instruments such as Convention on Biological Diversity (CBD), the International Treaty on Plant Genetic Resources (ITPGRFA) and the International Plant Protection Convention (IPPC). In 2010, the CBD adopted the Nagoya Protocol on Access to Genetic Resources and Equitable Sharing of Benefits Arising from their Utilization – that could have an impact on germplasm exchange. On the scientific front, advances in seed storage technology, biotechnology, and information and communication technology (ICT), have added new dimensions to plant germplasm conservation.

The draft revised *Genebank Standards* are concerned solely with the conservation of seeds of orthodox species, including wild species, i.e. those species whose seed can survive considerable desiccation, and in which longevity can be improved by reducing seed storage moisture content and/or temperature. The standards are underpinned by a set of broad underlying principles that provide the overarching framework for an effective and efficient management of genebanks. These key principles at the core of a genebank operation are the preservation of germplasm identity, maintenance of viability and genetic integrity, promoting access including the associated information to facilitate use of the stored plant material in accordance with relevant national and international regulatory instruments. The standards provide the specificity to ensure that a genebank can adhere to these underlying principles.

It is noted that these standards are voluntary and nonbinding and have not been developed through a formal standard-setting procedure. They should be viewed as targets for developing an efficient, effective, rational and transparent global system of *ex situ* conservation that provide optimal maintenance of seed viability and genetic integrity in genebanks, thereby ensuring access to and use of high quality seeds of conserved plant genetic resources.

These standards do not cover *ex situ* conservation of non- orthodox seeds or clonally propagated crops. Appropriate standards for such collections will be developed in due course.

The draft revised *Genebank Standards* can be aimed at by all genebanks for conserving orthodox seed collections, but they should not be used uncritically because there are continuous technological advances in conservation methods, much of it species-specific, as well as in the context of the purpose and period of germplasm conservation and use. It is therefore recommended that the draft revised *Genebank Standards* be used in conjunction with other reference sources, particularly references to species-specific information.

This document is divided into three parts, namely the Underlying Principles, Standards and the Appendices. The standards are detailed in nine sections and a selective list of references is provided for all the standards.

2. Underlying principles

Genebanks globally share many of the same basic goals, but their missions, resources, and the systems they operate within often differ. As a result, curators have to optimize actively their own overall genebank system and this objective requires management solutions which may differ substantially across institutions, but that would still lead to the same goals. Underlying principles explain why and for what purpose plant genetic resources are being conserved. These principles provide the basis for establishing the norms and standards essential for the smooth operation of a genebank. The major underlying principles for conservation are described in the section below.

Identity of accessions

Care should be taken to ensure that the identity of seed sample accessions conserved in genebanks is maintained throughout the various processes, beginning with acquisition through to storage and distribution. Proper identification of seed samples conserved in genebanks is closely related to careful documentation of data and information about the material. This will begin with recording passport data including collecting information and, if applicable, donor information; information should also be recorded for older collections in the genebanks for which passport data was not recorded earlier or is incomplete. Often herbarium voucher specimen and seed reference collections may play an important role in the correct identification of seed samples. Modern techniques such as bar-coding accessions can greatly facilitate the management of the germplasm without incurring errors and thus ensuring the identity of the accessions in question.

Maintenance of viability

Maintaining viability and genetic integrity of seed samples and making them available for use is the ultimate aim behind genebank management. It is, therefore, critically important that all processes adhere to the standards necessary to ensure that acceptable levels of viability are maintained. A high initial viability can ensure the attainment of a maximum period of conservation under long-term conditions, thus decreasing the frequency of regeneration to avoid loss of alleles. To meet these aims, particular attention needs to be paid to standards on germplasm acquisition, processing and storage. In general, seed samples accepted into the genebank at the point of acquisition should have high viability and as far as possible meet the standards for acquisition of germplasm. Collecting the seeds as close as possible to the time of natural dispersal can ensure the highest physiological seed quality. A monitoring system should be in place to check viability status of stored samples at appropriate intervals depending on expected seed longevity. Costly regeneration can be avoided or at least delayed if correct attention is paid to post-harvest handling, drying and storage.

Maintenance of genetic integrity

The need to maintain genetic integrity is closely related to maintenance of viability and diversity of the original collected sample. All genebank processes, starting from collecting and acquisition through to storage, regeneration and distribution, are important for the maintenance of genetic integrity. Adequately representative seed samples of good quality and sufficient quantity should be obtained during acquisition as far as possible. Ensuring that viability is maintained according to the standards contributes to the maintenance of genetic integrity. To minimize genetic erosion it is important to follow recommended protocols for regenerating seed accessions, with as few regeneration cycles as possible, sufficiently large effective population sizes, as well as pollination control. A special mention is made here on the importance of safety duplication to respond to risks that can occur in genebank facilities.

Maintenance of seed health

Genebanks should strive to ensure that the seeds they are conserving and distributing are free from quarantine seed-borne diseases and pests to the extent possible. Often genebanks do not have the capacities or the resources necessary to test themselves whether samples collected or acquired

and samples harvested from regeneration/multiplication plots are free quarantine seed-borne diseases and pests. This is particularly the case with germplasm received from third parties. So it is important that relevant import and phytosanitary certificates accompany seeds materials when exchange of germplasm takes place to ensure the health status of samples received. Some infected/infested samples may be easily cleaned, while others may required more elaborate methods for cleaning.

Physical Security of collections

An underlying principle of germplasm conservation is that the physical structures of the genebank facilities in which germplasm are conserved are of adequate standard to secure the materials from any external factors including natural disasters, thefts and riots. Adequate security systems are also required to ensure that genebank cooling equipment is in good running conditions and monitoring devices are available for track the essential parameters against time. Another important security issue for genebanks is to ensure materials are safely duplicated in one or more locations such that if the collection is destroyed for some reasons, it can be restored from the duplicated sets.

Availability and use of germplasm

The conserved material must be available for current and future use. It is, therefore, important that all processes in genebank operations and management contribute to this goal. There will be a need to maintain sufficient quantities of seed and related information on the accessions. If information on the conserved germplasm is made easily available and accessible it will enhance its use. Further this will help the genebank curators to better plan their multiplication and regeneration activities in order to keep adequate stocks of their accessions.

Availability of information

In order to ensure communication of information and accountability, essential, detailed, accurate, and up-to-date information at all stages should also be recorded, including historical as well as current information, especially in relation to the management of individual accessions, subsequent to their acquisition. Access, availability and sharing of this information should be treated with high priority, as it leads to better and more rational conservation. Search-query interactive databases containing phenotypic evaluation data can assist germplasm clients in the targeting of germplasm requests, and in turn feedback of further evaluation data adds to the value and utility of the collection.

Proactive management of genebanks

Sustainable and effective conservation of genetic resources depends on active management of the conserved germplasm material. Proactive management is critical for ensuring that germplasm is efficiently conserved and made timely and in adequate quantity available for further use by plant breeders, farmers, researchers and other users. It emphasizes the importance of securing and sharing material as well as the related information, and sets in place a functional strategy for management of human and financial resources for a rational system. It includes a risk management strategy and encourages a participatory role of genebanks in the efforts to conserve biodiversity. Adherence to the legal and regulatory frameworks at national and international levels, in particular as they relate to access, availability and distribution of materials and plant and seed health is necessary. A Standard Material Transfer Agreement (SMTA) should be used in all cases under the Multilateral System of the ITPGRFA. The IPPC regulations provide the framework for quarantine and health regulations to prevent the introduction and spread of plant pests and diseases. Above all, there is a need for long-term and continuous commitment of the institutions holding genebanks with regards to the availability of human and financial resources.

Furthermore, proactive management would encourage application of practical experiences and knowledge to new germplasm in a genebank and seek to apply the genebank standards to the extent possible under the locally prevailing conditions. This could sometimes mean that although

a particular standard is not entirely met but precautionary measures are taken to uphold the underlying principles of genebank management.

3. Standards – structure and definitions

The Standards as described in this document, define the lowest level of performance of a routine genebank operation below which there is a high risk of losing genetic integrity (e.g. a probability of five percent or more of losing an allele in an accession over the storage period⁷).

Each section is divided into

- A. Standards
- B. Context
- C. Technical aspects
- D. Contingencies
- E. Selected references

The standards are detailed in nine sections: acquisition, seed drying and storage, viability monitoring, regeneration, characterization, documentation, distribution, safety duplication and security/personnel.

The **Context** provides the basic necessary information in which the standards apply. It provides a brief description of the routine genebank operation for which the standards are defined and the underlying principles for them.

The **Technical Aspects** explain technical and scientific principles important to understand and underpin the standards.

The **Contingencies** provide recommendations in the case that standards cannot be applied to a given species, for example exceptions, alternative routes, and risk management options.

Selected sources of information and references are provided in all sections.

⁷ The storage period will depend on the frequency of regeneration and use and may vary for short-term, medium term and long-term storage.

3.1. Standards for acquisition

A. Standards

3.1.1. All seed samples added to the genebank collection have been acquired legally with relevant technical documentation.

3.1.2. Seed collecting is made as close as possible to the time of natural seed dispersal to ensure maximum seed quality.

3.1.3. To maximize seed quality, the period between seed collecting and transfer to a controlled drying environment is as short as possible.

3.1.4. All seed samples are accompanied by a minimum of associated data as detailed in the FAO/IPGRI multi-crop passport descriptors.

3.1.5. The minimum size of a seed sample must capture 95 percent of alleles in the sampled population.

B. Context

Acquisition is the process of collecting or requesting seeds for inclusion in the genebank, together with related information. The material should be legally acquired, be of high seed quality and properly documented.

Acquisition is made in accordance with relevant international and national regulations such as phytosanitary/quarantine laws, ITPGRFA or CBD access regulations, and national laws for genetic resources access. Adherence to Standard 3.1.1 will allow the export of seeds from the origin/donor country and the import into the country of the genebank, and determine the management and distribution regime (for example SMTA or bilateral Material Transfer Agreements (MTA)).

There is a need to ensure maximum seed quality and avoid conservation of immature seeds and seeds that have been exposed for too long to the elements. The way that seeds are handled after collection and before they are transferred to controlled conditions is critical for seed quality. High temperatures and humidity during the post-collecting period and during transport to the genebank could cause rapid loss in viability and reduce longevity during storage. The same applies to post-harvest handling within the genebank. The seed quality and longevity is affected by the conditions experienced prior to storage within the genebank.

During the acquisition phase, it is important to ensure that passport data for each accession is as complete as possible and fully documented. Passport data are crucial in identifying and classifying the accession and will function as an entry point in selecting and using the accession.

C. Technical aspects

For material collected outside the genebank country, there must be a Material Acquisition Agreement (MAA) or Access and Benefit Sharing Agreement (ABSA) drafted and signed by the authorized person in the country of collecting, and according to the national laws for genetic resources access for the country where the collecting will take place (ENSCONET, 2009). Phytosanitary regulations and any other import requirements must be sought from the relevant national authority of the receiving country. For material donated both from within and external to the genebank country, the provisions for the donation, if any, should be made explicit i.e. as SMTA or other type of MTA.

Seeds that are freshly harvested from the field may have high water content and need to be ventilated to prevent fermentation. They should be placed into suitable containers that allow for good air circulation, and that ensure the contents do not become moist through inadequate air exchange and are neither mixed nor damaged during collecting and transport. Monitoring the temperature and relative humidity (RH) to ensure that seeds are not exposed to conditions above 30 °C or 85 percent RH after collecting and transport, as well as during post-harvest processing will help to maintain seed quality. If fully mature seeds need to be processed and dried in the field, technical recommendations for the particular or similar species should be applied to reduce the risk of deterioration.

During collecting appropriate collecting forms should be used. These forms should include information such as the initial taxonomic classification of the sample, the global positioning system coordinates of the collecting site, a description of the habitat of the collected plants, the number of plants sampled and other relevant data that are important for proper conservation. If possible, the FAO/IPGRI multi-crop passport descriptors should be used (FAO/IPGRI, 2001). Very useful additional information, such as cultural practices, previous generations of seed history and origin, uses etc, can be obtained with farmer interviews when seed is collected from farmer fields/stores.

In case of donation of the seeds (from a seed company, research programme or genebank), the taxonomic classification, donor, identification number of the donor, and names in addition to the available passport data should be provided. Adequate information about how the germplasm received was maintained should be sought from the donor. Seeds should be assigned a unique identification number (either temporary or permanent, according to the practice used in the genebank) that accompanies the seeds at all times, and that will link the seeds to the passport data and any other collected information, and guarantee the authenticity of the seed sample. Whenever possible a herbarium voucher specimen collected from the same population as the seed samples should be taken, and a record should be made of the method and reason for acquisition.

D. Contingencies

Collecting should not take place without meeting the legal requirements especially if the germplasm is taken out of the country of collection afterwards.

Seeds collected in the field are rarely in such condition (physiological and phytosanitary status) that long-term conservation is automatically guaranteed. In this case multiplication in controlled conditions for the specific purpose of long-term conservation is recommended.

When collections contain a significant proportion (>10 percent) of immature seeds or fruits, measures should be taken to encourage post-harvest ripening. This can usually be achieved by holding material in well ventilated, ambient conditions protected from rainfall. Visual improvements in maturity should be monitored and the material should be transferred to controlled drying conditions as soon as the collected seeds are deemed more mature.

Allowances will have to be made for rare species where seeds might not be available in optimal conditions or quantity.

E. Selected references

¹ SGRP. Crop Genebank Knowledge Base (<http://cropgenebank.sgrp.cgiar.org>)

² Guarino, L., Ramanatha Rao, V. & Reid, R. eds. 1995 *Collecting Plant Genetic Diversity: Technical Guidelines*, Wallingford: CAB International on behalf of IPGRI. in association with FAO, IUCN and UNEP, 748 pp.

³ **Engels, J.M.M. & Visser L.** eds. *A guide to effective management of germplasm collections*. IPGRI Handbooks for Genebanks, No. 6. IPGRI, Rome, Italy, 2003.

⁴ **ENSCONET** *Seed Collecting Manual for Wild Species*, ENSCONET. 2009. ISBN: 978-84-692-3926-1 (www.ensconet.eu).

⁵ **Eymann, J., Degreef, J., Häuser, C., Monje, J.C., Samyn, Y. & VandenSpiegel, D.** eds. 2010. *Manual on Field Recording Techniques and Protocols for All Taxa Biodiversity Inventories and Monitoring, Vol. 8*. Chapters can be downloaded from: <http://www.abctaxa.be/volumes/volume-8-manual-atbi>

⁶ **FAO/IPGRI.** 2001. *Multi-Crop Passport Descriptors*. FAO, Rome, 4 pp. Available online from: [http://www.biodiversityinternational.org/index.php?id=19&user_biodiversitypublications_pi1\[showU id\]=2192](http://www.biodiversityinternational.org/index.php?id=19&user_biodiversitypublications_pi1[showU id]=2192)

⁷ **Genebank Standards 1994 FAO/IPGRI, Rome**
<ftp://ftp.fao.org/docrep/fao/meeting/015/aj680e.pdf>

⁸ **Guerrant, E.O., Havens, K. & Maunder, M.** eds. 2004. *Ex Situ Plant Conservation: supporting species survival in the wild*. Island Press, Washington D.C. USA.

⁹ **Lockwood, D.R., Richards, C.M. & Volk, G.M.** 2007. *Probabilistic models for collecting genetic diversity: comparisons, caveats and limitations*. *Crop Science* 47: 859-866.

¹⁰ Model MAA and source of authorized persons (CBD, Treaty focal points)

¹¹ **Probert, R.J.** 2003. Seed viability under ambient conditions and the importance of drying, pp 337-365 In: R.D. Smith, J.D. Dickie, S.H. Linington, H.W. Pritchard, R.J. Probert eds. *Seed Conservation: turning science into practice*: Royal Botanic Gardens, Kew, UK.

¹² **Probert, R., Adams, J., Coneybeer, J., Crawford, A. & Hay, F.** 2007. Seed quality for conservation is critically affected by pre-storage factors. *Australian Journal of Botany* 55, 326-335.

¹³ **RBG, Kew,** Millennium Seed Bank Technical information sheet 04: post-harvest handling of seed collections: <http://www.kew.org/msbp/scitech/publications/04-Post%20harvest%20handling.pdf>

¹⁴ **Smith, R.D., Dickie, J.D., Linington, S.L., Pritchard, H.W. & Probert, R.J.** 2003. *Seed Conservation: turning science into practice*: Royal Botanic Gardens, Kew. Chapters can be downloaded from: <http://www.kew.org/msbp/scitech/publications/sctsip.htm>

¹⁵ **Upadhyaya H. D. & Gowda C.L.L.** 2009. *Managing and enhancing the use of germplasm – Strategies and methodologies*. Technical Manual no. 10. International Crops Research Institute for the Semi-Arid Tropics. 236 pp. Patancheru 502 324, Andhra Pradesh, India.

3.2. Standards for drying and storage

A. Standards

3.2.1. All seed samples are dried to equilibrium in a controlled environment of 5-20 °C and 15-25 percent of relative humidity.

3.2.2. After drying, all seed samples need to be sealed in a suitable air-tight container prior to storage at the chosen temperature and relative humidity of 15 percent \pm 3 percent.

3.2.3. Most-original-samples and safety duplicate samples are stored under long-term conditions at a temperature of -18 ± 3 °C.

3.2.4. For medium-term conditions samples are stored under refrigeration at 5-10 C.

B. Context

Maintaining seed viability is a critical genebank function that ensures germplasm is available to users and is genetically representative of the population from which it was acquired (i.e. the most-original-sample). A critical objective of seed drying and storage standards is to reduce the frequency of regeneration of the most-original-sample by maximizing seed longevity, thereby reducing the cost of genebanking and the risks of genetic erosion. For this purpose, long-term storage is required for all most-original samples and for safety duplication of the collection (see Standards for safety duplication). In addition storage standards are also required for circumstances where the objective is to store seeds over the medium- or short-term to keep them alive long enough for distribution to users and evaluation of germplasm. In such cases the standard need not be as stringent as in the case of long-term conservation.

Prior to storage, seed samples need to be dried to appropriate moisture content. A variety of methods can be used for seed drying, the most common being the use of a desiccant or using a dehumidified drying chamber. The methods chosen will depend on the available equipment, number and size of the samples to be dried, local climatic conditions and cost considerations. However, there is a limit to which drying can increase longevity. At a critical moisture level, maximum longevity for the storage temperature is attained and drying below this level does not increase seed longevity further. To realize the full benefit of refrigerated or freezer storage, it is recommended that genebanks dry seeds to the critical moisture level (see Technical aspects). Various RH-temperature combinations can be used during drying, with faster drying possible at higher temperatures but the potential for physiological aging reduced by lower drying temperatures.

Long-term storage conditions as recommended above are expected to provide high seed quality for about 100 years for seed of most agricultural species; medium-term storage conditions are adequate for 30 years and will generally require refrigerated storage. Short-term storage is expected to provide high quality seed for at least eight years and may be accomplished at ambient temperatures (under as cool and stable temperatures as possible but not more than 25 °C) for some longer-lived species if relative humidity is controlled according to Standard 3.2.2. It should be pointed out that the longevity of mature, high quality seeds may vary among species and even among seed lots of the same species (Probert *et al.* 2009; Nagel and Börner 2009; Crawford *et al.* 2007; Walters *et al.* 2005). The variation among species and among seed lots of the same species, particularly if seeds are harvested with variable maturity, requires the genebank curator's vigilance to monitor viability (see Standards for viability monitoring).

As seed equilibrium moisture content varies depending on oil content, the best measurement for the drying standard is equilibrium relative humidity (eRH) which is constant depending on the relative humidity and temperature of the drying environment. However, it should be noted that in sealed containers during storage, seed eRH will fall or increase if the storage temperature is lower or higher than the drying temperature.

C. Technical aspects

Seed longevity is determined by interactions of biological factors intrinsic to the seed and the quality and consistency of the storage environment, namely the storage temperature and the control of seed moisture content (equilibrium relative humidity). It is well known that seed longevity increases as the seed moisture content and storage temperature decreases, within limits (Ellis and Roberts, 1980; Harrington, 1972). Studies have demonstrated that drying seed beyond a certain critical seed moisture content provides little or no additional benefit to longevity (Ellis *et al.* 1995; Ellis and Hong, 2006) and may even accelerate seed-aging rates (Vertucci and Roos 1990; Walters, 1998). The storage standards as presented are intended to ensure that seeds are stored at this optimum moisture content. However, it has been shown that lowering the storage temperature increases the optimum seed moisture content level (Walters and Engels, 1998; Ellis and Hong, 2006), which suggests there might be danger of over-drying seeds. Conversely, there are reports of successful long-term storage of seeds under 'ultra-dry' conditions (Pérez-García *et al.* 2009). However, there is still uncertainty and requires further research (Ellis and Hong, 2006; Vertucci and Roos 1990; Walters, 1998).

Drying conditions that achieve the critical moisture level at the storage temperature should be determined using water sorption isotherms which show the relationship between the amount of water in the seeds, usually expressed as a percentage of the total seed weight, and their RH. There could be different combinations of relative humidity and drying temperature for given species. Isotherm relationships, predicted based on seed oil content, are available online at the Kew Seed Information Database (SID) website (see references). Genebank operators should clearly understand the relationship between relative humidity and storage temperature to be able to decide about the best combination for their seed drying environment.

As soon the seeds have reached the desired moisture content they should be packaged and stored. After drying, seed moisture should be maintained using moisture-proof containers. Either glass containers that are sufficiently thick to avoid breakage or laminate packaging with a metal foil layer of at least 20 μm thickness will maintain desired moisture levels for up to 40 years, depending on the ambient relative humidity at the genebank's location and the quality of the seal. Seed moisture content or eRH should be measured periodically to confirm that storage moisture is adequately maintained.

The storage temperature defines the maximum longevity possible for a seed sample and a stable storage environment is critical to maintaining seed viability. However, there is limited data from long-term storage at a range of low temperatures. Storage at $-18\text{ }^{\circ}\text{C}$ has been recommended in the past for long-term storage as it is the lowest temperature that can be achieved with a single stage standard deep freezer compressor. For long-term stored seeds, all attempts should be made to maintain storage temperatures within $\pm 3\text{ }^{\circ}\text{C}$ of the set temperature and to limit the total duration of fluctuations outside this range to less than one week per year. Genebanks should maintain records of storage temperature deviations and periods when seed accessions are removed from the storage environment. For short-term storage, the seeds should be dried at the same temperature as they are stored, e.g. if ambient condition is $20\text{ }^{\circ}\text{C}$, seeds should then be dried at that same temperature.

D. Contingencies

Seeds in long-term storage should be removed rarely and only when samples in medium-term storage are exhausted. Desired storage conditions are not achieved when mechanical environmental controls fail or when seeds are repeatedly removed from controlled storage environment. Back-up generators with an adequate fuel supply should be available on-site.

All containers leak and seed moisture will eventually equilibrate to environmental conditions within the storage vault. This occurs faster in containers for which thermal plastics are used as the moisture barrier or if glass or foil laminate containers have faulty seals or imperfections. Seeds may need to be re-dried occasionally and containers or gaskets replaced within 20-40 years.

If clear (for example, glass) containers are used, perforated transparent plastic sachets containing self-indicating silica gel, equilibrated to the drying environment, can be used to monitor container performance during long-term storage. A change in colour of the silica gel inside the sachet (stored alongside the seeds) will indicate moisture ingress if the container seal fails.

Orthodox seeds with short life spans or seeds with low initial quality may deteriorate more rapidly in storage and not meet long-term storage standards unless cryogenic conditions are used.

E. Selected references

- ¹ Crawford, A.D., Steadman, K.J., Plummer, J.A., Cochrane, A. & Probert, R.J. 2007. Analysis of Seed Bank Data Confirms Suitability of International Seed Storage Standards for the Australian Flora. *Australian Journal of Botany*, 55: 18-29.
- ² Dickie J.B., Ellis, R.H., Kraak, H.L., Ryder, K. & Tompsett, P.B. 1990. Temperature and seed storage longevity. *Annals of Botany*, 65: 197-204.
- ³ Ellis, R.H. & Roberts, E.H. 1980. Improved equations for the prediction of seed longevity. *Annals of Botany*, 45: 13-30.
- ⁴ Ellis, R.H. & Hong, T.D. 2006. Temperature sensitivity of the low-moisture-content limit to negative seed longevity-moisture content relationships in hermetic storage. *Annals of Botany*, 97: 785-91.
- ⁵ Engels, J.M.M. & Visser, L. *A guide to effective management of germplasm collections*. IPGRI Handbooks for Genebanks No. 6. IPGRI, Rome, Italy.
- ⁶ Gomez-Campo, C. 2006. Erosion of genetic resources within seedbanks: the role of seed containers. *Seed Science Research* 16:291-294
- ⁷ Harrington, J.F. 1972. Seed storage longevity. In: T.T. Kozlowski, ed. *Seed biology*, Vol. III. pp. 145-245 Academic Press, New York, USA.
- ⁸ **Kew Seed Information Database: predict seed viability module**
(<http://data.kew.org/sid/viability/percent1.jsp> ; Convert RH to water content
(<http://data.kew.org/sid/viability/mc1.jsp>) and Convert water content to RH
(<http://data.kew.org/sid/viability/rh.jsp>)
- ⁹ Nagel, M. & Börner A. 2009. The longevity of crop seeds stored under ambient conditions. *Seed Science Research*, 20: 1-12.
- ¹⁰ Pérez-García, F., Gómez-Campo, C. & Ellis, R.H. 2009. Successful long-term ultra dry storage of seed of 15 species of *Brassicaceae* in a genebank: variation in ability to germinate over 40 years and dormancy. *Seed Science and Technology*, 37(3): 640-649.
- ¹¹ Probert, R.J., Daws, M.I. & Hay, F.R. 2009. Ecological Correlates of *Ex Situ* Seed Longevity: a Comparative Study on 195 Species. *Annals of Botany*, 104 (1): 57-69.
- ¹² Smith, R.D., Dickie, J.D., Linington, S.L., Pritchard, H.W. & Probert, R.J. 2003. Seed Conservation: turning science into practice: Royal Botanic Gardens, Kew. Chapters can be downloaded from: <http://www.kew.org/msbp/scitech/publications/sctsip.htm> (see chapters 17 and 24).
- ¹³ Vertucci, C.W. & Roos, E.E. 1990. Theoretical Basis of Protocols for Seed Storage. *Plant Physiology*, 94:1019-1023.
- ¹⁴ Walters, C. 1998. Understanding the mechanisms and kinetics of seed aging. *Seed Science Research*, 8:223-244.
- ¹⁵ Walters, C. 2007. Materials used for Seed Storage Containers. *Seed Science Research*, 17: 233-242.

-
- ¹⁶ **Walters, C., Wheeler, L.J. & Stanwood, P.C.** 2004. Longevity of cryogenically-stored seeds. *Cryobiology*, 48: 229-244.
- ¹⁷ **Walters, C. & Engels, J.** 1998. The effect of storing seeds under extremely dry conditions. *Seed Science Research*, 8. Supplement 1, pp 3-8.
- ¹⁸ **Walters, C., Wheeler, L.J. & Grotenhuis, J.** 2005. Longevity of seeds stored in a genebank: species characteristics. *Seed Science Research* 15:1-20.
- ¹⁹ **Gomez-Campo, C.** 2006. Erosion of genetic resources within seedbanks: the role of seed containers. *Seed Science Research* 16:291-294

3.3. Standards for seed viability monitoring

A. Standards

3.3.1. The initial seed viability test is conducted after cleaning and drying the accession or at the latest within 12 months after receipt of the sample at the genebank.

3.3.2. The initial germination value should exceed 85 percent for most seeds e.g. cereals, and 75 percent for some vegetables and lower for some wild or forest species, which do not normally reach high levels of germination

3.3.3. Viability monitoring test intervals to be set at one-third of the time predicted for viability to fall to 85 percent⁸ of initial viability. If this deterioration period cannot be estimated and accessions are being held in long-term storage at -18 °C in hermetically closed containers, the interval should be ten years for species expected to be long lived and five years or less for species expected to be short lived.

3.3.4. The viability threshold for regeneration or other management decision such as re-collection is 85 percent of initial viability.

B. Context

Good seed storage conditions maintain germplasm viability, but even under excellent conditions viability declines with period of storage. It is therefore necessary to assess viability periodically. The initial viability test should be conducted as early as possible before the seeds are packaged and enter the storage, and subsequent tests are conducted at intervals during storage. If for practical reasons of workflow and efficiency the initial viability test cannot be made prior to storage, it should be made as soon as possible and not later than 12 months after storing. This can be the case of multi-species genebanks, where a wide range of germination regimes is required and samples of the same species are tested all together once a year.

The purpose of viability monitoring is to detect loss in viability during long-term storage before viability has fallen below the threshold for regeneration. The important guiding principle is one of active management of the collection. Too frequent monitoring will result in unnecessary waste of seeds and resources. On the other hand, significant viability decline may not be detected if monitoring is delayed or infrequent; advanced aging of the sample may result in genetic changes (random or directed selection), unrepaired mutations fixed in the sample, or ultimate loss of the accession.

When it is predicted that viability will fall to 85 percent before the next scheduled retest, the time of the retest should be anticipated or the accession directly scheduled for regeneration.

Risk of genetic erosion during storage is lower for homogeneous samples and germination decline to less than 85 percent is allowable as long as plant establishment during regeneration remains adequate. For heterogeneous samples such as wild species and landraces, the 85 percent standard should be adhered. For some wild species such as cereal and legume wild relatives, a viability of 85% in newly replenished seed s rarely achievable. In these situations, the curator can set the viability standard trigger for selected species to a lower threshold, such as 70 percent.

Models to predict seed longevity from ambient to freezer conditions are available for diverse agricultural species. Genebank staff should use available predictive tools documented for particular species and storage conditions to anticipate duration that seeds will maintain high viability and to guide other genebank operations such as viability monitoring and regeneration

⁸ The time for seed viability to fall can be predicted for a range of crop species using an online application based on the Ellis/Roberts viability equations (see <http://data.kew.org/sid/viability/>)

frequencies (see Standards for viability monitoring and regeneration). Longevity predictions based on general species characteristics should be considered as estimates with large confidence intervals. Genebanks are encouraged to develop and report new information that describes and updates species responses to storage conditions.

C. Technical aspects

Viability monitoring intervals should be adjusted according to the data received from germination tests. As soon as a significant decline is detected, monitoring intervals should be reduced in order to 'fine tune' the prediction of time to reach the viability standard.

Accessions with very high initial viability (> 98 percent) may show a statistically significant decline in viability long before the predicted time for viability to fall to 85 percent, when germination is still well above 90 percent. Regeneration or recollection at this point is probably too soon and unnecessary. However, future retest intervals should be brought forward (for example from ten years to five years) in order to track the decline more accurately.

For accessions of lower quality, the accession might be dangerously close to the tipping point if viability declines comparatively rapidly. Such accessions should be managed carefully and the first viability monitoring tests should be after 3-5 years of storage intervals at first. Infrequent (for example ten-year) monitoring might fail to detect rapid deterioration and the viability threshold of 85 percent could be missed with negative consequences to the genetic integrity of the collection

Sample sizes for viability monitoring will inevitably be dependent upon the size of the accession but should be maximized to achieve statistical certainty.

It is difficult to establish a strict standard for the number of seeds for germination tests in genebanks. As a general guideline 200 seeds are recommended to be used for initial germination tests (ISTA, 2008) followed by sequential testing, if the initial germination is less than 90 percent (Ellis *et al.* 1985) during storage. But in practice the actual sample size for germination will depend on the size of the accession, which in general is very limited (the recommended minimum size for self pollinated is 1500 and for cross pollinated species 3000 seeds) in genebanks. It is important to minimize the use of valuable seeds required for germination tests. For small accession sizes (as is often the case for wild species) sample sizes of 50 seeds or less could be acceptable. However it must be realized then that there may be a higher chance of germination being below the threshold. The genebank curator should assess the risk of this occurrence.

The germination test should always be used in preference to alternatives such as the tetrazolium test. However, in circumstances where it is not possible to remove seed dormancy, alternative tests may be carried out. It is recommended that germination often be measured at two different times so as to have an idea of fast and slow germinating seeds. Records of the number of abnormally germinating seeds should also be kept. Slower germination and increasing abnormals are often early indicators that deterioration is occurring.

Every effort should be made to germinate all viable seeds in a collection using optimum conditions and appropriate dormancy-breaking treatments where needed. Non-germinated seeds remaining at the end of a germination test should be cut-tested to assess whether they are dead or dormant. Seeds with firm, fresh tissue are likely to be dormant and should be counted as viable seeds.

All data and information generated during viability monitoring should be recorded and entered into the documentation system.

D. Contingencies

It is recognized that viability monitoring is an expensive activity and that genebanks would wish to seek cost-cutting procedures. One such procedure may entail measuring seed quality in a subsample of accessions of the same species grown in the same harvest year. This practice may reveal overall trends on the effect of harvest year on seed quality, but will not take genotype x harvest year interactions into consideration that are known to be important for seed quality. In the event that subsampling is unavoidable, it should be undertaken with sufficient statistical rigor to ensure usefulness of the data in future analyses. For example, performing germination tests on less than ten accessions may not provide sufficient statistical power to compare accessions harvested in different years. Hence, should a subsampling strategy be used, at least 10 percent of same-species accessions harvested in the same year should be evaluated with a minimum of ten accessions evaluated.

Where different harvest conditions occur over a wide range of maturities across accessions, then a sampling strategy can be from separate sub groups harvested. An additional strategy would be to focus retesting on the accessions that gave the lowest viability result in the initial tests. Retest data from these accessions should provide an early warning on the performance of the batch as a whole.

The initial germination test at harvest for known hard seeded species and accessions frequently found in some forage legume species and WCR's can be as low as 45%, and increases after 10-15 years to 95% or more and remains so for long periods of time. If the initial germination is less than 90 percent, then regenerate/recollect at first detectable significant decline established by an appropriate statistical test.

However it is recognized that intraspecific variation among accessions has been observed for a wide range of accessions, thus there are risks associated with the above strategies, which should be considered. Viability monitoring of accessions of wild species is generally more problematic compared with crop species. Seed dormancy is likely to be much more prevalent and small accession sizes often mean that smaller minimum sample sizes have to be adopted for germination tests, as this will inevitably affect the ability to detect the onset of seed deterioration.

With reference to the initial seed viability testing it is also possible that genebanks receive small quantities of seeds. In that case it is not necessary to carry out initial seed viability testing since the samples is sent for regeneration. However the regenerated seeds must then be tested for viability prior to storage.

The initial germination test at harvest for known hard seeded species and accessions frequently found in some forage legume species and WCR's. can be as low as 45%, and increases after 10-15 years to 95% or more and remains so for long periods of time. If the initial germination is less than 90 percent, then regenerate/recollect at first detectable significant decline established by an appropriate statistical test.

The range of inherent longevity is also wider in wild species with some species from Mediterranean and tropical dryland habitats expected to be extremely long lived and conversely some species from cold, temperate regions expected to be short lived. For the latter, retesting intervals of as few as three years should be considered as well as duplication into cryo-storage as a precautionary measure. In the event that storage conditions are not met (as will occur if there is a prolonged power cut when seeds are stored in refrigeration units), viability will be affected negatively depending on the species, length of disruption and conditions during the disruption. In such an event a disaster management plan should be activated. For example some representative samples may need to be tested immediately following resumption of adequate storage conditions.

E. Selected references

- ¹ **Association of Official Seed Analysts (AOSA)** 2005. Page 113 in: Capashew, ed. *Rules for Testing Seeds*, 4-0, 4-11. Las Cruces, New Mexico, USA.
- ² **Dickie, J.B., Ellis, R.H., Kraak, H.L., Ryder, K. & Tompsett, P.B.** 1990. Temperature and seed storage longevity. *Annals of Botany*, 65:197-204.
- ³ **Ellis, R.H. & Roberts, E.H.** 1980 Improved equations for the prediction of seed longevity. *Annals of Botany*, 45, 13-30.
- ⁴ **Ellis, R.H., Hong, T.D. & Roberts, E.H.** 1985. Sequential germination test plans and summary of preferred germination test procedures. *Handbook of seed technology for genebanks: Vol I .Principles and methodology*, Chapter 15, pp 179-206. International Board for Plant Genetic Resources. Rome, Italy.
- ⁵ **Engels, J.M.M. & Visser, L.** eds. 2003 *A guide to effective management of germplasm collections*. IPGRI Handbooks for Genebanks No. 6. IPGRI, Rome, Italy.
- ⁶ **ENSCONET** manual: http://www.ensconet.eu/PDF/Curation_protocol_English
- ⁷ **Harrington, J.F.** 1972. Seed storage longevity. In: T.T. Kozlowski, ed. *Seed biology, Vol III*, pp.145-245, Academic Press, New York, USA.
- ⁸ **International Seed Testing Association (ISTA)**. 2008. *International Rules for Seed Testing*. Bassersdorf, Switzerland.
- ⁹ **Nagel, M. and Börner, A.** 2010: The longevity of crop seeds stored under ambient conditions. *Seed Science Research* 20, 1-12
- ¹⁰ **Nagel, M., Rehman Arif, M.A., Rosenhauer, M. and Börner, A.** 2010: Longevity of seeds - intraspecific differences in the Gatersleben genebank collections. Tagungsband der 60. Jahrestagung der Vereinigung der Pflanzenzüchter und Saatgutkaufleute Österreichs 2009, 179-181.
- ¹¹ **Royal Botanical Gardens, Kew Seed Information Database (SID)**:at <http://data.kew.org/sid/>
- ¹² **Smith, R.D., Dickie, J.D., Linington, S.L., Pritchard, H.W. & Probert, R.J.** 2003. *Seed Conservation: turning science into practice*. Royal Botanic Gardens, Kew. Chapters can be downloaded from: <http://www.kew.org/msbp/scitech/publications/sctsip.htm> (see chapters 17 and 24).

3.4. Standards for regeneration

A. Standards

3.4.1. Regeneration should be carried when the viability drops below 85 percent of the initial viability. The most-original-sample should be used to regenerate those accessions.

3.4.2. The sample size of the accession to-be-regenerated contains a minimum number of plants which capture at least 95 percent of alleles with a minimum frequency of 0.05 .

3.4.3. The regeneration has to be carried out in such a manner that the genetic integrity of a given accession is maintained; for example, regenerated material should contain less than 1 percent of contamination arising from geneflow of pollen that originated from other accessions of the same species or from other species.

3.4.4. At least 50 seeds of the original and the subsequent most original samples are archived in long-term storage for reference purposes.

B. Context

Regeneration is a key operation and an integral responsibility of any genebank that maintains orthodox seeds. It is a process that leads to an increase of the stored seeds (also called “multiplication”) in the genebank and/or to an increase of the viability of the seeds equal to or above an agreed minimum level, which is referred to as the regeneration threshold. An accession will be regenerated when it does not have sufficient seeds for long-term storage (i.e. 1 500 seeds for a self-pollinating species and 3 000 for an out-crossing species) or when its viability has dropped below an established minimum threshold (i.e. below 85 percent of initial germinability of the stored seeds).

As regeneration is an activity that could easily affect the genetic composition of an accession (and thus its genetic integrity) utmost care is required. Consequently, genebank operators will have to strike a delicate balance between avoiding regeneration as much as possible versus the potential loss of viability and thus, the risk of affecting the genetic integrity of an accession. Active management of the collections will greatly help to decide on the best moment to regenerate.

Regeneration should be undertaken with the least possible change to the genetic integrity of the accession in question. This means that in addition to sampling considerations (see paragraph below) of the accession in question we need to pay due attention to the environment in which the activity will be undertaken, as such environment might cause severe selection pressure on the accession. It has been suggested that the regeneration environment should be as similar as possible to that at the collecting site, in particular when a population collected in the wild is being regenerated, in order to minimize genetic drift and shift as well as to produce the best possible quality of seeds. It can often be difficult to harvest sufficient quantity of seed from wild relatives due to lower seed/plant numbers compared to other species, or plant dispersal mechanisms such as seed shattering. It is therefore necessary to ensure that appropriate technical practices are used to capture as much seed as possible (i.e. nets to capture dropped seeds). Repeat regeneration cycles may also be required to ensure that sufficient seed is conserved.

To preserve the genetic integrity of genebank collections during seed regeneration, it is important that sampling of accessions be carried out effectively. The number of seeds to be used for the regeneration process must be of sufficient size to be representative of the genetic diversity in an accession and to capture one or more rare alleles with a certain probability.

The methodology to be used for regeneration might vary from species to species and depends, among others on the population size, breeding system and pollination efficacy. Therefore, it is of significant importance to collate as much as possible of the relevant biological information related

to the species in question. In addition, when possible and meaningful, it is recommended that the regeneration event be used also for the characterization of regenerated accessions (see Characterization Standards).

C. Technical aspects

In order to maintain the genetic integrity of accessions it is recommended to use seeds from the most-original-sample for regeneration. For multiplication it is recommended to use seeds from the working collection for up to five cycles of multiplication without returning to the most original sample (IPGRI, 2003).

It should be noted that in cases where the original collection or donation is a small sample, it is necessary to regenerate immediately following receipt of the material in order to obtain an adequate quantity of seeds for long-term storage. It is important to record the number of the regeneration cycle and enter the information into the documentation system. It is recommended that the receiving genebank always keep some seeds from the initial seed sample for future reference purposes. Even if these original seeds lose their viability, they can be useful in confirming morphology or genotype of later generations of the respective accession.

The size of the seed sample to be used in the regeneration activity has to reflect the genetic composition of the accession, i.e. the reproductive biology of the species in question as well as the degree of homogeneity/heterogeneity of the accession. For this purpose the effective population size (N_e) is a key parameter that will have a bearing on the degree of genetic drift that is associated with the regeneration of the accession. This minimal size of N_e to minimize loss of alleles can be estimated for individual accessions based on the pollination biology, growing conditions and harvest techniques. .

To avoid geneflow/contamination it is critically important to use proper isolation methods between plots of accessions of cross-pollinated species being regenerated. This also applies to self-pollinated species, depending on the regeneration environment. The principle of disjunctive cultivation, that is planting accessions of different taxa in adjacent plots, has been recommended (Lehmann and Mansfeld 1957). For species that depend on specific pollinators, isolation cages and the corresponding pollinators should be used (Dulloo, M.E. *et al.* 2008). Contamination and genetic drift/shift can be assessed with morphological, enzymatic or other distinctive traits that can be used as markers (e.g. flower colour; seed colour, etc.), or with molecular markers.

Reference collections (herbarium specimen, photographs and/or descriptions of the original accessions) are essential for conducting the true-to-type verification (Lehmann and Mansfeld 1957). Close inspections of obtained seeds and during the first regeneration of a new genebank accession are required to collect important reference information.

In order to avoid differences in seed maturity in a seed sample, multiple harvests should be carried out during the fruiting season.

D. Contingencies

The management of a genebank and of a germplasm collection is a multifaceted task in which scientific considerations have to be combined with economical, infrastructural, personnel and other aspects and where an optimum balance must be aspired. However, as already indicated, the underlying principles such as genetic integrity and identity have to be given the highest attention while regenerating accessions. Nevertheless, there will always be a risk management dimension to the curatorship role. Solid biological knowledge of the species in question is a key factor in making the best possible decisions under constrained conditions. Aspects such as sample size, distance between individual accessions and other forms of isolating accessions, respecting

established thresholds for viability loss, growing conditions and others, all need to be given due attention when planning the regeneration activity.

In view of this complexity it is not meaningful to look for possible contingencies. In case of emergency it would be advisable to seek advice from experts and/or collaboration with other genebanks that could provide assistance.

E. Selected references

- ¹ **Breese, E.L.** 1989. *Regeneration and multiplication of germplasm resources in seed genebanks: the scientific background*. Available online at:
http://www2.bioversityinternational.org/publications/Web_version/209/
- ² **SGRP** Crop genebank knowledge base <http://cropgenebank.sgrp.cgiar.org>
- ³ **Crossa, J.** 1995. Sample size and effective population size in seed regeneration of monocious species. In: J.M.M. Engels, R. Ramantha Rao, eds. *Regeneration of seed crops and their wild relatives. Proceedings of a consultation meeting, 4-7 December 1995*. ICRISAT, Hyderabad, India. International Plant Genetic Resources Institute, Rome, Italy. pp.140–143.
- ⁴ **Dulloo, M.E., Hanson, J., Jorge, M.A. & Thormann, I.** 2008. Regeneration guidelines: general guiding principles. In: M.E. Dulloo, I. Thormann, M.A. Jorge & J.Hanson, eds. *Crop specific regeneration guidelines* [CD-ROM]. CGIAR System-wide Genetic Resource Programme (SGRP), Rome, Italy. 6 pp.
- ⁵ **Engels, J.M.M. Ramantha Rao, R.** editors. 1995. *Regeneration of seed crops and their wild relatives. Proceedings of a consultation meeting, 4-7 December 1995*. ICRISAT, Hyderabad, India. International Plant Genetic Resources Institute, Rome, Italy. pp.140–143.
- ⁶ **Engels, J.M.M. & Visser, L.** 2003. *A guide to effective management of germplasm collections*. IPGRI Handbooks for Genebanks No. 6. IPGRI, Rome, Italy.
- ⁷ **Lawrence, L.** 2002. *A comprehensive collection and regeneration strategy for ex situ conservation. Genetic resources and crop evolution* 49 (2): 199-209.
- ⁸ **Lehmann C.O. & Mansfeld R.** 1957. Zur Technik der Sortimentserhaltung. Kulturpflanze 5: 108-138.
- ⁹ **Rao, N.K., Hanson, J., Dulloo, M.E., Ghosh, K., Nowell, D. & Larinde, M.** 2006. *Manual of seed handling in genebanks. Handbooks for Genebanks* No. 8. Bioversity International, Rome, Italy.
- ¹⁰ **Sackville Hamilton, N.R. & Chorlton, K.H.** 1997. *Regeneration of accessions in seed collections: a decision guide*. J. Engels, ed. Handbook for Genebanks No. 5. International Plant Genetic Resources Institute, Rome, Italy.

3.5. Standards for characterization

A. Standards

3.5.1. Around 95 percent of accessions are characterized within five years of acquisition or the first regeneration cycle.

3.5.2. Characterization is based on standardized and calibrated measuring formats and characterization data follow internationally agreed descriptor lists and are made publicly available.

B. Context

Characterization is the description of plant germplasm. It determines the expression of highly heritable characters ranging from morphological, physiological or agronomical features to seed proteins and oil or molecular markers.

Characterization can be carried out at any stage of the conservation process, as long as there are sufficient numbers of seeds to sample. It is essential that the germplasm being conserved is known and described to the maximum extent possible to assure their maximum use by plant breeders. Therefore, characterization should be carried out as soon as possible to add value to the collection. The use of a minimum set of phenotypic physiological and seed qualitative traits and morphological descriptors and information on the breeding system, such as those published by Bioversity is helpful for characterisation. Useful descriptors can also be found in the publications of the International Union for the Protection of New Varieties of Plants. Use of internationally agreed standards for characterization data increases the usefulness of the published data.

With the advances in biotechnology, molecular marker technologies , genomics are increasingly used for characterization (de Vicente, *et al.* 2004). Characterization will allow for detecting intra-accessions diversity. Means such as splitting samples may be necessary for ensuring the preservation of rare alleles or for improving access to defined alleles. Documentation of observations and measures taken is extremely important.

C. Technical aspects

Characterization is time consuming and expensive. Effort can be made to combine characterization with multiplication or regeneration to the extent possible. Curators should make all possible efforts to record characterization data.

Characteristics and traits for crops are defined by crop experts and/or curators in consultation with genebank managers. A wide range of crop descriptor lists has been developed for example by Bioversity International and also minimum sets of key descriptors for utilization have been established for several of these. Furthermore there are regional and national descriptor lists available. Data recording needs to be carried out by trained staff using calibrated and standardized measuring formats as indicated in the internationally agreed and published crop descriptor lists. The data need to be validated by curator and documentation officers before being uploaded into the genebank database and made publicly available. It is also recognized that reference collections (herbarium specimens, seed herbarium, photographs) play an essential role for true-to-type identification.

D. Contingencies

Reliability of data might vary among data collectors if they are not well trained and experienced. Therefore trained technical staff in the field of plant genetic resources should be available during the entire growth cycle to record and document characterization data. Access to expertise in taxonomy, seed biology and plant pathology (in-house or from collaborating institutes) during the process of characterization is desirable.

Characterization is very labor-intensive and requires sufficient funding to allow for good quality data. Carrying out full characterization of accessions during regeneration cycles may reduce the number of accessions which can be regenerated per cycle.

The incidence of pests and diseases can limit the collection of quality data. The determination of some traits like oil or protein content requires laboratory assays which are not always available or could be costly.

E. Selected references

¹ **Bioversity** Crop Descriptor Lists available online at:

http://www.bioversityinternational.org/research/conservation/sharing_information/descriptor_lists.html and from the SGRP Crop Genebank Knowledge Base Bioversity

² **Bioversity International**. 2007. Developing crop descriptor lists, Guidelines for developers.

Bioversity Technical Bulletin No. 13. Bioversity International, Rome, Italy. 71p. Available online at:

[http://www.bioversityinternational.org/index.php?id=19&user_bioversitypublications_pi1\[showU id\]=3070](http://www.bioversityinternational.org/index.php?id=19&user_bioversitypublications_pi1[showU id]=3070)

³ **de Vicente, M.C., Metz, T. & Alercia, A.** 2004. *Descriptors for Genetic Marker Technologies*.

International Plant Genetic Resources Institute, Rome, Italy. 30p. Available online at:

[http://www.bioversityinternational.org/index.php?id=19&user_bioversitypublications_pi1\[showU id\]=2789](http://www.bioversityinternational.org/index.php?id=19&user_bioversitypublications_pi1[showU id]=2789)

⁴ **FAO/IPGRI**. 2001. *Multi-Crop Passport Descriptors*. FAO, Rome, 4 pp. Available online from:

[http://www.bioversityinternational.org/index.php?id=19&user_bioversitypublications_pi1\[showU id\]=2192](http://www.bioversityinternational.org/index.php?id=19&user_bioversitypublications_pi1[showU id]=2192)

⁵ **Lehmann C.O. & Mansfeld R.** 1957. Zur Technik der Sortimentserhaltung. Kulturpflanze 5: 108-138.

UPOV : (http://www.upov.int/en/publications/tg_rom/tg_index.html)

3.6. Standards for documentation

A. Standards

3.6.1. Passport data of 100 percent of the accessions are documented using FAO/IPGRI multi-crop passport descriptors.

3.6.2. All data and information generated in the genebank relating to all aspects of conservation and use of the material are recorded in a suitably designed database.

3.6.3. A duplicate of all genebank data and information is established outside the genebank for safety reasons.

B. Context

Information about accessions is essential for the genebank to manage and maintain their collection; it is also important to share this information and make it available publicly for potential germplasm users, and should be attached to any distributed material. Passport data are the minimum data that should be available for each accession to guarantee proper management, and international standards such as the FAO/IPGRI multi-crop passport descriptors (FAO/IPGRI 2001) should be used to record passport data. The use of internationally agreed standards will very much facilitate data exchange.

Major advances in information technology and bioinformatics have taken place over the last decade or so and much of it is available online. A majority of genebanks also have access to computers and the internet. This new technology makes it possible to record and exchange data and information efficiently. Ultimately conservation and usability of conserved germplasm are promoted through good data and information management. All data and information generated throughout the process of acquisition, registration, storage, monitoring, regeneration, characterization, evaluation, and distribution should be recorded in a suitably-designed database and employed to improve conservation and use of the germplasm. Such data and information ranges from details of the genetic characteristics of individual accessions and populations to distribution networks and clients.

Documentation of characterization and evaluation data is particularly important to enhance the use of the respective collection and help identification of distinct accessions.

With advances in biotechnology, there is a need to complement phenotypic trait data with molecular data. Efforts must be made to record the molecular data being generated through genomics, proteomics and bio informatics.

C. Technical aspects

Computer-based systems for storing data and information allow for more comprehensive storage of all information associated with genebank management. The adoption of data standards which today exist for most aspects of genebank data management helps to make the information management easier and to improve use and exchange of data. For example, the FAO/IPGRI List of Multi-crop Passport Descriptors should be used for documenting passport data as it is instrumental for data exchange among different genebanks and countries.

Germplasm information management systems exist, such as GRIN-Global, which have specifically been developed for genebanks and their documentation and information management needs. Another germplasm information management system is the International Crop Information System (ICIS) platform in which germplasm data from 1 or more genebanks can be stored, and published online with a search-query capacity to allow users to set criteria for selection of germplasm by single or by multiple trait criteria, as well as bounded by GPS coordinates for a region and/or overlaid with climatic and soil maps, for targeted selection of germplasm.

Evaluation data are often produced by the users to which seeds have been distributed. The genebank should solicit the user to share the evaluation data, which should then be included in the genebank's documentation system. Such information could address resistances to biotic and abiotic stresses, growth and development features of the accession, quality characteristics of yield etc. Adding this type of information allows more focused identification of germplasm to meet prospective client needs.

However, it is recognized that using information generated by users may not be so simple and may involve copy right and institutional issues.

D. Contingencies

Lack of documentation or loss of it compromises the optimal use of the seeds or can even lead to their loss, if it impedes planning regeneration properly.

E. Selected references

¹ **de Vicente, C., Alercia, A. & Metz, T.** 2004. *Descriptors for Genetic Marker Technologies*. International Plant Genetic Resources Institute, Rome, Italy. 30p. Available online at: [http://www.biodiversityinternational.org/index.php?id=19&user_biodiversitypublications_pi1\[showU id\]=2789](http://www.biodiversityinternational.org/index.php?id=19&user_biodiversitypublications_pi1[showU id]=2789).

² **FAO/IPGRI.** 2001. *Multi-Crop Passport Descriptors*. FAO, Rome, 4 pp. Available online at: [http://www.biodiversityinternational.org/index.php?id=19&user_biodiversitypublications_pi1\[showU id\]=2192](http://www.biodiversityinternational.org/index.php?id=19&user_biodiversitypublications_pi1[showU id]=2192)

³ **ICIS** International Crop Information System. <http://irri.org/knowledge/tools/international-crop-information-system>

3.7. Standards for distribution

A. Standards

3.7.1. Seeds are distributed in compliance with national laws and relevant international treaties and conventions.

3.7.2. Seed samples are provided with all relevant documents required by recipient country.

3.7.3. At least 95 percent of the accessions and associated data are readily available for distribution and the remaining after multiplication/regeneration.

3.7.4. The time span between receipt of a request for seeds and the dispatch of the seeds is kept to a minimum.

3.7.5. A sample of a minimum of 30-50 viable seeds is supplied for accessions with sufficient seeds. For accessions with too little seed at the time of request and in the absence of a suitable alternative accession, samples are supplied after regeneration/multiplication, based on a renewed request.

B. Context

Conservation should be linked to utilization. Germplasm distribution is the supply of a representative sample of seed accessions from a genebank in response to requests from plant germplasm users. The CBD and ITPGRFA emphasize this continuum between conservation and sustainable utilization, along with facilitated access and equitable sharing of benefits arising from use.

There is a continuous increase in demand for genetic resources to meet the challenges posed by climate change, by changes in virulence spectra of major pests and diseases and by invasive alien species. This demand has led to wider recognition of the importance of using germplasm from genebanks - which ultimately determines the germplasm distribution. The time between receipt of a request for seeds from a user and the following response and dispatch of seeds (along with relevant information) should be kept as short as possible.

The diversity of the legal systems with respect to their procedural rules governing access to courts and to arbitration, and the obligations arising from international and regional conventions applicable to these procedural rules is recognized.

The ITPGRFA within the framework of its Multilateral System both to facilitate access to Plant Genetic Resources for Food and Agriculture and to share, in a fair and equitable way, the benefits arising from the utilization of these resources, on a complementary and mutually reinforcing basis, has developed the SMTA for Annex1 crops. While other distribution models also exist, the SMTA can also be used for non-Annex1 crops.

Genebanks that hold working collections should promote the availability of genetic resources for uses including research, breeding, education, farming and repatriation. Internationally, genebanks can be a source of land race germplasm re-supply to countries which are initiating their own genebank, or which suffered a disaster such as fire, flood or civil strife.

The exchange of seeds is not accompanied by the risk of spreading some quarantine diseases, insects or invasive and exotic weed species that could seriously affect national production.

C. Technical aspects

Germplasm should be distributed in a way that ensures the germplasm reaches its destination in good condition. Environmental conditions can be harmful to the quality of seed during transport therefore seeds should be carefully packed and sealed in airtight envelopes for protection during transit.

Samples to be distributed should comply with the requirements of the quality standards as defined in this document and the requirements of seed health as requested by the recipient country. The distribution should also comply with national regulation laws.

Easy and speedy clearance of shipments from customs offices and plant protection departments will most often necessitate the availability of documents required by the recipient country and the requestor.

Phytosanitary certificate, certificate of donation, certificate of no commercial value and import permit are among the documents required by the recipient country. It is therefore important to maintain and update the list of documents requested by different countries.

The list of the material and associated information (passport data as a minimum) should be provided to the recipient together with any legal agreement related to access and use of genetic resources provided.

It is highly recommended to reduce as much as possible the time between the dispatch and the delivery of the shipment. When seeds are not available responses include a detailed description of the reason, an estimated date when the accession will be available, and alternative accessions that may suit the requestor's needs.

Genebank clients are encouraged to do their own seed bulking for their trials needs and experiments. This is particularly relevant for wild species for which seed stock are often low, and for replicated field trials where supply of the required seed quantity cannot be considered.

The distributing genebank should insist on the flowback of information about the usefulness of the supplied germplasm.

D. Contingencies

Political decisions or crisis situations or bureaucratic delays might extend the time span between receipt of a seed request and the distribution of the material. Limitations related to regeneration and/ or multiplication of the accessions may also affect and delay the distribution process.

E. Selected references

¹ **Convention on Biological Diversity (CBD)**. 1992.

<http://www.cbd.int/convention/convention.shtml>

² **Engels, J.M.M. & Visser, L.** 2003. *A guide to effective management of germplasm collections*. IPGRI Handbooks for Genebanks No. 6. IPGRI, Rome, Italy.

³ **FAO/IPGRI**. 1994. Genebank Standards.

⁴ **International Treaty on Plant Genetic Resources for Food and Agriculture (ITPGRFA)**:

<http://www.itpgrfa.net/International/>

⁵ **Rao, N.K., Hanson, J., Dulloo, M.E., Ghosh, K., Nowell, D. & Larinde, M.** 2006. *Manual of seed handling in genebanks*. Handbooks for Genebanks No. 8. Bioversity International, Rome, Italy.

⁶ **SGRP**. Crop Genebank Knowledge Base: <http://croptgenebank.sgrp.cgiar.org>

⁷ **Standard Material Transfer Agreement (SMTA)**: <http://www.itpgrfa.net/International/>

3.8. Standards for safety duplication

A. Standards

3.8.1. A safety duplicate sample for every original accession is stored in a geographically distant area, under the same or better conditions than those in the original genebank.

3.8.2. Each safety duplicate sample is accompanied by relevant associated information

B. Context

Safety duplication is that of a genetically identical subsample of the accession to mitigate the risk of its partial or total loss caused by natural or human-caused catastrophes. The safety duplicates are genetically identical to the long-term collection and are referred to as the secondary most original sample (Engels and Visser, 2003). Safety duplication includes both the duplication of material and its related information and these are deposited in long-term storage at a different location, often outside the country. The location is chosen to minimize possible risks and provides the best possible storage facilities.

Safety duplication is generally made under a ‘black-box’ approach. This means that the repository genebank has no entitlement to the use and distribution of the germplasm. It is the depositor’s responsibility to ensure that the deposited material is of high quality, to monitor seed viability over time and to use their own base collection to regenerate the collections when they begin to lose viability. The germplasm is not touched without permission from the depositor and is only returned on request when the original collection is lost or destroyed. Recall of the deposit is also possible when it is replaced with newly regenerated germplasm. It is recognized however that the black-box is not the only approach. There may be cases where the safety collection is also taken care of by the recipient genebank.

Safety duplication should be made for all original seeds collected by the genebank or when only held by the genebank. Seeds which are duplicates from other collections can usually be retrieved from those collections and do not require safety duplication unless there is doubt about their security in the other collection.

Any safety duplication arrangement requires a clearly signed legal agreement between the depositor and the recipient of the safety duplicate that sets out the responsibilities of the parties and terms and conditions under which the material is maintained.

This safety duplication is now available at the Svalbard Global Seed Vault on Spitsbergen island, Norway. Countries which have joined the ITPGRFA are eligible to deposit, and retain ownership and management of their own duplicate collection. Sealed “Black Box” arrangements for quarantine clearance can be made with the relevant quarantine authority for countries requiring a generation in quarantine for restricted plant types. The prearranged quarantine clearance makes the germplasm available for immediate use upon return from Svalbard.

C. Technical aspects

When selecting the location for safety duplication, primary consideration is given to the geographic location and environmental conditions of the location. Facilities must ensure low radiation (radioactivity) and stability (low probability of earthquakes). The facility must be situated at an elevation that guarantees proper drainage during seasonal rains and eliminates the risk of flooding in the event of rising sea levels due to global warming. Equally important is economic stability and socio-political certainty. Koo *et al.* (2004) suggest that safety duplicate

samples should be located away from the risk of political embargo, military action or terrorism that could disrupt international access.

Samples are prepared for safety duplication in the same way as for the base collection. Conditions should be at least as stringent as those for long-term storage of germplasm in a genebank and the quality of seed preparation (i.e., drying) is important.

In some cases it is helpful to sort material according to short, medium and long living seed groups before sending for safety duplication.

Sample size should be sufficient to conduct at least three regenerations. A safety duplicate of an accession in a seed genebank should contain at least 500 viable seeds for outbreeders and heterogeneous accessions with high diversity and a minimum of 300 seeds for genetically uniform accessions. For accessions with seeds of low viability more seeds are necessary. Storage temperatures should be -18°C to -20°C .

The packaging material for safety duplication should be of trilaminate material of which the middle metal foil layer should be at least $20\ \mu\text{m}$ in thickness. It should be formed into a pouch seamed on all four sides with no gusset. This would provide an adequate water barrier for transport and storage at -18°C for at least 30 years.

An outer and inner label should be placed on each packet of seeds to ensure that the germplasm is properly identified.

As the storage conditions for the safety duplicate should be the same or better than that of the base collection, seed viability can be monitored on seed lots of the same accession maintained in long-term storage in the genebank and extrapolated to the safety duplicate if basic standards for storage conditions are met and the same containers are used. In some cases, samples for germination testing may be sent in a separate box with the safety duplicate and monitored for germination by agreement with the depository.

Strong cold-resistant boxes (thick carton or polypropylene boxes) are the best options for transporting and storing seeds. Boxes should be sealed properly. Shipment should consider the fastest means of transport available either by air freight, courier or by land to avoid deterioration of seed quality during transit.

Samples should be renewed from the sender when the viability of the samples in similar storage conditions in the base collection of the sender starts to decline. The duplicate samples can be either destroyed or returned to the sender and replaced with a new batch.

D. Contingencies

When extrapolating the viability of the safety duplicate from viability monitoring results of the sample in the base collection, some caution should however be taken. Seeds may age at different rates if there is a difference in ambient RH at the two sites and/or differences in extent or frequency of temperature fluctuations, though the average storage temperature is the same.

Issues of liability may occur related to sending samples in sealed black-box conditions. One issue is on liability for contents of the sealed box and handling by customs officers and other authorities for entry into a country. In some cases boxes are opened and special seals are applied by the authorities to confirm that the samples are not medicinal or other prohibited plants. Another issue is that on liability of the recipient institution should material be damaged or lose viability earlier than expected as a result of stress during transit, faulty seal of containers, or temperatures that fluctuate from specified standards. Under the conditions described here, the safety duplicate repository should only be "liable" if the temperature becomes uncontrollable; this should be reported immediately to the primary institution so that they can decide on what action to take. The primary institution should bear full responsibility for transport disasters or uncontrolled moisture.

The standards and technical aspects may be difficult to implement for some species due to the inherent biology of the samples, e.g. short-lived seeds, large-seeded species where space and cost may be limiting.

E. Selected references

¹ **Engels, J.M.M. & Visser L.** 2003. *A guide to effective management of germplasm collections*. IPGRI Handbooks for Genebanks No. 6. IPGRI, Rome, Italy. Available in [English](#) (1.4 MB) and [Spanish](#) (1.5 MB).

² **SGRP.** Crop Genebank Knowledge Base. The page on safety duplication, available on line at http://croptgenebank.sgrp.cgiar.org/index.php?option=com_content&view=article&id=58&Itemid=207&lang=english contains detailed background documents, a list of references and a standard safety deposit agreement template.

3.9. Standards for security and personnel

A. Standards

3.9.1. A genebank should have a risk management strategy in place which includes inter alia measures against power cut, fire, flooding and earthquakes.

3.9.2 A genebank should follow the local Occupational Safety and Health (OSH) requirements and protocols.

3.9.3 A genebank employs the requisite staff to fulfill all the routine responsibilities to ensure that the genebank can acquire, conserve and distribute germplasm according to the standards.

B. Context

Achieving a genebank's goal of acquisition, conservation and distribution of germplasm not only require adequate procedures and equipment for germplasm handling be in place, but that properly trained staff be employed to carry out the required work and to guarantee the security of the genebank.

Active genebank management requires well-trained staff, and it is crucial to allocate responsibilities to suitably competent employees. A genebank should therefore have a plan or strategy in place for personnel, and a corresponding budget so as to guarantee that a minimum of properly trained personnel is available to fulfill the responsibilities of ensuring that the genebank can acquire, conserve and distribute germplasm. Access to specialists in a range of subject areas is desirable, depending on the mandate and objectives of each individual genebank. However, staff complements and training will depend on specific circumstances. The health and usefulness of the seeds stored in the genebank depend also on issues related to safety and security of the genebank. Arrangements need to be in place for electricity back-up; fire extinction equipment has to be in place and regularly checked genebank buildings need to be earthquake-proof if situated in a seismic-prone area, to mention some. A genebank should therefore implement and promote systematic risk management that addresses the physical and biological risks in the every-day environment to which the collections and related information are exposed.

C. Technical aspects

Staff should have adequate training acquired through certified training and/or on-the-job training and training needs should be analyzed.

Genebank personnel should be aware of and trained in safety procedures to minimize risks to the germplasm.

The genebank facilities should be constructed so as to withstand natural disasters, such as hurricanes, cyclones, earthquakes, or floods that are known to occur in the location where the genebank has been built.

Storage facilities should be protected with standard security facilities such as fences, alarm systems, security doors and any other system that helps to shield the genebank from burglars and other intruders. Security of the seed collections in the genebank will be enhanced by allowing entry strictly to authorized personnel into the actual storage facilities.

Protective clothing should be provided and used in the storage area. Adequate precautions should be taken and safety equipment, including alarms and devices to open doors from inside drying rooms and refrigerated rooms, should be installed.

Refrigeration will almost certainly be reliant on electrical power and it is therefore necessary that the power supply is adequate and reliable. Failure in power supply can result in complete loss of genebank accessions. Consideration should be given to the provision of a back-up generator that automatically cuts in when the main power supply fails. This will require stockpiling adequate amounts of fuel to run the generator during power cuts.

Monitoring devices for temperature should be available in the drying and storage room to track the actual parameters against time.

It should be considered whether it is better to store seed without refrigeration if refrigeration is inherently unreliable. If refrigeration is to be used to conserve germplasm, it must meet necessary standards as unreliable refrigeration can be far more damaging than non-refrigerated storage.

If refrigeration and/or electric power are unreliable, a facility can be built in the soil at a depth of 10-20 m, where temperature can be averaged at 10 °C. This could be attractive in several tropical regions under no risk of flooding. Drying should be well carried out however, and seeds should be kept in properly-sealed vials.

Fire alarm and fire-fighting equipment is required in the genebank. Most fires begin from faulty electrical circuits and therefore periodic checks should be made on the electrical circuitry to ensure compliance with safety standards. Firefighting equipment will include extinguishers and fire blankets. For areas affected by thunderstorms, a lightning rod should be fitted to the genebank.

D. Contingencies

When suitably trained staff is not available, or when there are time or other constraints, it might be a solution to outsource some of the genebank work or to approach other genebanks for assistance. The international community of genebanks should be informed, if the functions of the genebank are endangered.

Unauthorized entry to genebank facilities can result in direct loss of material, but can also jeopardize the collections through inadvertent introduction of pests and diseases and interference in management systems.

E. Selected references

¹ **Engels J.M.M. & Visser, L.** 2003. *A guide to effective management of germplasm collections*. IPGRI Handbooks for Genebanks No. 6. IPGRI, Rome, Italy. Available in English (1.4 MB) and Spanish (1.5 MB).

² **SGRP.** Crop Genebank Knowledge Base, Section on risk management:
http://croptgenebank.sgrp.cgiar.org/index.php?option=com_content&view=article&id=135&Itemid=236&lang=english

Appendices

List of acronyms and abbreviations

ABSA	Access and Benefit Sharing Agreement
CBD	the Convention on Biological Diversity
CGIAR	the Consultative Group on International Agricultural Research
CGRFA	Commission on Genetic Resources for Food and Agriculture
FAO	Food and Agriculture Organisation
GPS	Global Positioning System
GRIN	Genet Resources Information Network
ICT	Information and Communication Technologies
ICIS	International Crop of Information Systems
IPPC	International Plant Protetcion Convention
ITPGRFA	International Treaty on Plant Genetic Resources for Food and Agriculture
ISTA	Intenational Seed Testing Association
MAA	Material Acquisition Agreement
MTA	Material Transfer Agreement
PGRFA	Plant genetic resources for food and agriculture
RH	Relative Humidity
SID	Seed Information Database
SMTA	Standard Material Transfer Agreement