

Food and Agriculture Organization of the United Nations



EGGS AND LARVAE OF COMMON MARINE FISH SPECIES OF NORTHWEST AFRICA



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EGGS AND LARVAE OF COMMON MARINE FISH SPECIES OF NORTHWEST AFRICA

by

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PREPARATION OF THIS DOCUMENT

This identification guide was conceived and supported by the EAF-Nansen Programme *"Supporting the Application of the Ecosystem Approach to Fisheries Management considering Climate and Pollution Impacts"* of the Fisheries and Aquaculture Division (NFI), Food and Agriculture Organization of the United Nations (FAO).

The EAF-Nansen Programme supports fisheries research and management institutions in partner countries in their efforts to sustainably manage their fishery resources. Fisheries and environmental surveys are conducted with its research vessel, the *Dr. Fridtjof Nansen* to strengthen the knowledge base and develop capacity at the institutional and human resources levels. The Programme's science plan guides the research work, and improving knowledge on the ecology of the early life stages of fish is the goal of its first thematic area. Information on early life history stages of fish is important for fisheries management. For example, parameters related to the dynamics of fish populations, such as spatio-temporal ichthyoplankton distribution patterns, larval pool composition and seasonal recruitment could help to improve our understanding of the factors affecting the fishable stock and their potential resilience to fishing pressure. However, fish eggs and larvae identification, which is a crucial point for any biological and ecological study, is challenging even for the experienced scientists. In developing countries, larval fish identification remains an even more significant challenge, mainly due to the dearth of both specialists and identification guides.

Noting that the northwestern coast of Africa is lacking specific tools for the identification of ichthyoplankton, the Programme decided to support the preparation of this identification guide, building on the work previously completed by the author for the western Mediterranean Sea¹.

The work started in early 2021 and was completed at the end of 2022. Before finalizing the guide, a selection of species sheets were tested at the "*EAF-Nansen Programme Regional Training Workshop on the Identification of Ichthyoplankton*" that was held at the University of Cape Coast, Ghana from 20 to 24 June 2022.

This guide is designed to assist researchers in the identification of the early life stages of the fish species that are most likely to be present in the plankton samples collected in the Canary Current Large Marine Ecosystem region waters.

EAF-Nansen Programme coordinator: Merete Tandstad (FAO, Rome)

Technical editing, scientific revision, and formatting: Edoardo Mostarda (FAO, Rome).

¹ Rodriguez, J.M., Alemany, F. & Garcia A. 2017. A guide to the eggs and larvae of 100 common Western Mediterranean Sea bony fish species. FAO, Rome, Italy, 256 pp.

ABSTRACT

This guide presents the egg and larval descriptions of 150 species of fishes belonging to 57 families, which are most likely to be present in plankton samples collected in the continental shelf and oceanic waters off northwest Africa. The guide is structured in two parts. The first introductory part describes the different applications of ichthyoplankton studies in fisheries research and management, and fish population ecology, the main sampling strategies, methods and gears, and the problems related to sample representativeness. It also describes the early life history of fishes, and how to identify them. A brief description of the hydrography of the study area is also presented. The second part of the guide features the species identification sheets. Each species sheet includes the following information: an illustration of the adult fish and information on its distribution, habitat, spawning season, and meristic characters; a description of the main features useful towards identifying the egg, yolk-sac and larval stages; and illustrations and photos of different larval stages. Finally, the guide provides a comprehensive list of references.

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1. INTRODUCTION

What is ichthyoplankton?

The early life stages (eggs and larvae) of marine fishes are referred to as ichthyoplankton (Figure 1). Some species of marine fish produce demersal eggs that lay on the sea bottom, or are even deposited in nests. Demersal eggs of a number of species are under parental care, extended in some exceptional cases, such as in sea horses, to the larval stage. However, most marine fish species spawn pelagic eggs. Larvae hatched from pelagic and from most demersal eggs are pelagic.



Due to their null (eggs) or limited (larvae) swimming Figure 1. Ichthyoplankton sample abilities, they are both integrated into the plankton

community as part of the meroplankton (organisms that only spend part of their life cycle within the plankton community). Ichthyoplankton is largely found in the upper layer of the water column (from 200 m depth to the surface), where it is subject to spatial dispersion by marine currents mainly. The planktonic stage of fishes usually lasts from a couple of weeks to a few months (Victor, 1986; Brothers et al. 1983).

During the larval stage, larvae develop specializations for the planktonic life and other important changes occur. For example, most fishes increase their weight by five orders of magnitude throughout their life and three of them occur during the planktonic stage (Werner and Gilliam, 1984; Houde, 1987; Miller et al., 1988).

The end of the larval stage is marked by a metamorphosis or transformation process, more or less abrupt depending on the species, during which the larva becomes a juvenile, which is morphologically similar to the adult and possesses the meristic characters of the species.

Juveniles of pelagic species become part of the nekton community and those of most demersal species migrate directly to the bottom, after the metamorphosis. However, in several demersal species, where the transformation phase is prolonged, there is an intermediate stage (called the pre-juvenile stage) where the fish develops specializations that are distinct from both larvae and juveniles. These pelagic pre-juveniles eventually transform into demersal juveniles (Kendall et al., 1984). Pelagic pre-juveniles of demersal species and the early juveniles of the pelagic ones maintain an intense relationship, mainly trophic, with the plankton community, but because of their behaviour and swimming abilities, they cannot be considered ichthyoplankton, they become part of the micronekton.

Why study ichthyoplankton?

There are several reasons for studying ichthyoplankton. For example, information regarding the distribution and abundance of fish eggs and larvae can provide clues to spawning locations and environmental requirements of important (from a fisheries standpoint) fish species. Moreover, the knowledge of ichthyoplankton is necessary because, as one of the components of the pelagic food web (Raymont, 1983), it can represent an important link between smaller planktonic and larger nektonic organisms. Finally, the survival of fish larvae may directly influence the future abundance of adult fish stocks. The latter has been and still is the most important reason for studying ichthyoplankton, as most processes determining the recruitment strength and the spatial distribution of fish populations occur during the planktonic stage of fishes, resulting in important interannual fluctuations in fish stock biomasses. Such fluctuations have been known for centuries, but they started to worry scientists and fishery managers only at the end of the nineteenth century (Petersen, 1894; Garstang, 1900).

Initially, it was thought that fishing pressure itself or fish migrations were responsible for such fluctuations, but at the beginning of the twentieth century, Hjort (1914, 1926), after analysing the causes of the successful herring year-class of 1904, suggested that the variability in "year-class success" was determined during the early life stages of fishes. He proposed two hypotheses to explain the fish stock fluctuations resulting from interannual recruitment variability, the *Critical Period Hypothesis* and the *Aberrant Drift Hypothesis*.

According to the Critical Period Hypothesis, the strength of a year-class is determined shortly after yolk-sac absorption, at the beginning of exogenous feeding, when larvae must find suitable prey and in sufficient amounts (Figure 2). Failing to find adequate feeding conditions would lead to massive larval mortality, in a short period of time. Hjort's hypotheses laid the groundwork for future research on recruitment variability, mainly focused on the Critical Period Hypothesis (Cowan and Shaw. 2002; Houde, 2008), fostering the



Figure 2. Illustration showing the Hjort's Critical Period Hypothesis (1914, 1926)

Source: modified after Houde, E.D. 2008. Emerging from Hjort's Shadow. *Journal of Northwest Atlantic Fisheries Science*, 41: 53-70

development of "recruitment fisheries oceanography" as a multidisciplinary science to investigate the spectrum of oceanographic processes controlling recruitment (Kendall and Duker, 1998).

In the *Aberrant Drift Hypothesis*, larvae (due to interannual variability in ocean circulation) may be transported to unfavourable areas, failing to recruit to the exploited stock.

Cushing (1974, 1990) merged both Hjort's hypotheses into the *Match/Mismatch Hypothesis*. He hypothesized that a fixed time of spawning coupled with a variable time of plankton blooms generate variability in larval fish survival and, hence, variable recruitment. In this hypothesis, food limitation at any time of the larval period, rather than mortality being concentrated at a specific larval stage, could be a major contributor to the recruitment variability. In addition, abiotic factors that regulate water-column mixing, and the timing and intensity of seasonal production cycles may be involved.

Lasker (1981), in the *Stable Ocean Hypothesis*, proposed that the occurrence and frequency of calm periods in upwelling ecosystems (named Lasker events) lead to stratification of the water column and, hence, the aggregation of fish larvae and their prey, supporting high larval feeding, survival and recruitment.

A further extension of Lasker's *Stable Ocean Hypothesis* was the *Optimal Environmental Window* (OEW) *Hypothesis* by Cury and Roy (1989). These authors hypothesized that, in upwelling ecosystems, the relationship between the annual recruitment of small pelagic fish species (sardines and anchovies) and the upwelling intensity is dome-shaped. This confirms the existence of an optimal environmental window for recruitment, with recruitment most successful under moderate (roughly 5-6 m*s⁻¹) wind stress that controls both ichthyoplankton advective losses and food availability for fish larvae. On the contrary, weak winds, if prolonged, disrupt the upwelling process and the renewal of nutrients in the surface layer, inducing nutrient limitation, which reduces primary productivity (Huntsman and Barber, 1977) and food availability for fish larvae. Finally, strong winds generate strong turbulence that also limits primary productivity (Huntsman and Barber, 1977) and increases the offshore transport of fish eggs and larvae (Bakun and Parrish, 1982).

Based on the Hjort's *Aberrant Drift Hypothesis*, Iles and Sinclair (1982) and Sinclair (1988) proposed the *Larval Retention* or *Member/Vagrant Hypothesis*. In this hypothesis, larval retention, not the levels of available prey, is critical in the recruitment process.

For tropical reef fish species, Sale (1978, 1991) proposed the *Lottery Hypothesis*, which states that recruitment in tropical reef fishes is strongly dependent on the potential to deliver settlers to the reef, and that post-settlement processes could control recruitment levels. However, nowadays, both pre- and post-settlements processes are acknowledged to influence recruitment (Jones, 1991; Doherty, 2002).

The above hypotheses consider starvation and physical processes as the main drivers of recruitment variability. However, there is strong evidence that predation is the main source of ichthyoplankton mortality and, consequently, the main factor influencing recruitment (Hunter, 1981; Bailey and Houde, 1989).

Mortality from nutritional deficiencies and predation may not act independently. Slow growth, caused by a lack of food, can result in fish larvae remaining in the plankton for longer periods and thus being more vulnerable to predation than fast-growing larvae. This is the foundation of the *Stage-Duration Hypothesis*, which implies that large size ("bigger is better") and fast growth improve the survival potential (Houde, 1987; Anderson, 1988; Houde, 2008).

In summary, although Hjort focused on starvation of first-feeding larvae as the main driver of larval mortality, he already envisaged that recruitment variability was the result of complex, interacting factors, stating that "the simultaneous investigation of meteorology, hydrography and biology seems the only way to a deeper understanding of the conditions in which the destiny of the spawned ova is being decided" (Hjort, 1926).

In this line, nowadays it is widely agreed that recruitment variability is the outcome of complex trophodynamic and physical processes, operating on different temporal and spatial scales and throughout the prerecruit life stages of fishes (Figure 3). That is to say, recruitment success is not determined during a particular ontogenetic stage and it depends on the populations species, and environmental conditions (Houde, 2008). In this way, the knowledge of the ecology of the early life stages of fishes is crucial to understanding the



Figure 3. Schematic diagram of different processes acting on the early stages of a marine fish (eggs, yolk-sac larvae, larvae and juveniles) Source: modified after Houde, E.D. 1987. Fish early life dynamics and recruitment variability. *American Fisheries Society Symposium*, 2: 17-29

population dynamics of fish stocks, but also, the functioning of marine ecosystems.

New discoveries, technological advances and new analytical techniques, developed in the last decades, have allowed for significant advances in the knowledge of the causes of mortality in the early life fish stages. One of the major findings was the discovery by Pannella (1971) that growth rings in fish otoliths are deposited daily. Brothers *et al.* (1976) confirmed the presence of daily growth rings in otoliths of fish larvae (Figure 4). This finding allowed for the estimation of growth and mortality rates during the larval stage, and for the cohort contributions to recruitment (Methot, 1983; Miller *et al.*, 1988; Houde, 1997). The analysis of the otolith's microstructure also allows for assessing the environmental influence on larval survival, dating, with high precision, early life events and relating them to environmental conditions.

Other indicators useful for determining the nutritional status and recent growth rate in fish larvae are the RNA/DNA ratio (Buckley, 1979, 1980, 1984) and the nitrogen (N) and carbon (C) stable isotope analysis (SIA). The latter is used to assess the trophic position and C flow to consumers

in food webs (Minagawa and Wada, 1984, Peterson and Fry 1987, Post, 2002). Specifically, 15N provides an estimate of the trophic level, and 13C can be used to assess the sources of C for an organism when the isotopic nature of the sources is different, as C isotope ratios undergo small changes within the food web (Peterson and Fry, 1987, France and Peters, 1997). The SIA analysis has also been used recently to assess the trophic ecology of fish larvae (e.g. Laiz-Carrión et al., 2013, Laiz-Carrión et al., 2015, Laiz-Carrión et al., 2022). Finally, advances in telemetry have allowed synoptic sampling of environmental variables over large areas and advances in computer power the development of biophysical models, simulating larval drift and survival under real or hypothetical environmental scenarios.



Figure 4. Otolith of a sardine (Sardina pilchardus) larva

Applications of ichthyoplankton studies in fisheries research

One of the main objectives of ichthyoplankton field studies is the direct assessment of the number or biomass in exploitable populations (Heath, 1992). This is based on the fact that for some fish populations there may be a relationship between the number of eggs and/or larvae in a given area and the number or biomass of spawning adult fish, and this can be used to estimate the size of the adult population (Heath, 1992). There are six direct methods that use ichthyoplankton production for estimating the abundance of fish stocks, three based on eggs and three on larvae, with the latter used for demersal spawners (Stratoudakis *et al.*, 2006). The underlying principle of the ichthyoplankton-based methods is that the abundance of an early life stage of a fish species can be used to estimate the reproductive outcome of the population over a time period (Stratoudakis *et al.*, 2006).

Of the three methods based on eggs production, the *Daily Egg Production Method* (DEPM), the most widely used, was developed for small pelagic fishes (such as anchovies and sardines) that exhibit indeterminate fecundity (Lasker, 1985; Parker, 1980). Fecundity is indeterminate when the potential annual fecundity of a female is not fixed prior to the onset of spawning and unyolked oocytes continue to be matured and spawned during the spawning season (Hunter *et al.*, 1992). The advantage of the DEPM is that it only requires a single survey to estimate the daily egg production at sea and the daily specific fecundity at the peak of the spawning period (Parker, 1980). In addition, the application of this method provides information about the reproductive biology and reproductive behaviour of a population and about the distribution, mortality and development of the early life fish stages, which are of particular importance for studying the underlying mechanisms of recruitment (Murua *et al.*, 2010).

The other two methods based on eggs, the *Annual Egg Production Method* (AEPM) (Lockwood *et al.*, 1981) and the *Daily Fecundity Reduction Method* (DFRM) (Lo *et al.*, 1992) were developed for species with determinate fecundity, for which the potential annual fecundity becomes fixed prior to the onset of spawning (Hunter *et al.*, 1992).

Where egg production methods are impractical, a larval census has been employed to obtain a relative index of the spawning biomass (Heat, 1992).

Larval survey data can be used to obtain estimates of recruitment. The estimation or forecasting of recruitment strength, under variable environmental scenarios, is essential for the proper management of fish stocks, particularly in a context of relatively rapid climate change. Moreover, the implementation of the *Ecosystem Approach to Fisheries* (EAF), where fisheries management recognizes the full range of interactions within a marine ecosystem (Katsanevakisa *et al.*, 2011), has led to an increase in studies aimed at understanding recruitment and its underlying processes.

In addition, a holistic understanding of ecosystems, of the environmental influence on fish populations, especially on their highly vulnerable early life stages, is required to implement the EAF. This knowledge is also essential for implementing the *Ecosystem-Based Fisheries Management* (EBFM), in which the order of management priorities is inverse to the customary norm, addressing the status of ecosystems, rather than that of the fishery resources (Pikitch, *et al.*, 2004; Cogan *et al.* 2009).

Ichthyoplankton surveys can also be used for the prospection of new fish resources, determining the timing and location of spawning areas and their variations or to estimate, as said above, the relative abundance of different stocks and monitoring its abundance trends over time.

Other applications of ichthyoplankton studies

Ichthyoplankton studies can provide a lot of information on the ecology and structure of fish populations, in a cheaper and simpler way than information obtained by studying juvenile or adult populations. Indeed, a single collection of plankton hauls can give information about most fish species spawning in a given area, both pelagic and demersal, whereas sampling of adults would require larger vessels and a variety of sampling gears and methodologies. According to Koslow and Wright (2016) "ichthyoplankton surveys provide a relatively low-cost, efficient means to monitor marine fish populations and communities".

On the other hand, because of their null (eggs) or limited (larvae) swimming abilities, fish larvae are displaced by surface marine currents and other water movements, such as those related to e.g. upwelling, upwelling filaments, and eddies. Therefore, fish larvae have been used as tracers of hydrographic processes in several regions of the world oceans, e.g. the Northeast Atlantic Ocean, off the Iberian Peninsula (Rodríguez, 2008; Rodríguez *et al.*, 2015), the Northwest African upwelling region (Rodríguez *et al.*, 1999; Rodriguez *et al.*, 2004), the Taiwan Strait (Hsieh *et al.*, 2012), Australian waters (Smith and Suthers, 1999; Smith *et al.*, 1999), and the Sicilian channel (García la Fuente *et al.*, 2002).

Moreover, in the current context of global warming, ichthyoplankton studies could be used to monitor and evaluate changes in the composition and structure of fish populations and communities of an area, as a consequence of the water temperature increase. Fish species move poleward with water warming, to remain within suitable "climate envelopes" (Walther, 2002). Thus, "each 1 °C of temperature change moves ecological zones on earth by about 160 km" (Thuiller, 2007). In the marine environment, shifts in the distribution to the north have been observed for several fish species in the Northeast Atlantic (Stebbing *et al.*, 2002; Beare *et al.*, 2004). However, for these species to be able to establish in a new region, they must reproduce successfully, and their offspring survive in the new area (Sabatés *et al.*, 2006). Other species can change their spawning areas (Ibaibarriaga *et al.*, 2007). In both cases, changes in fish species composition or in their spawning areas would be reflected in the composition and structure of the ichthyoplankton assemblage, and these changes could be evaluated through ichthyoplankton studies.

Ichthyoplankton sampling strategies, methods and gears

Any study on ichthyoplankton communities is based on two pillars: an adequate sampling strategy to achieve the specific objectives of the study, and a correct taxonomic identification of the target taxa. There is a great variety of sampling strategies, from large-scale surveys to intensive sampling of a single patch of larvae tracked by means of a Lagrangian buoy (a free-floating buoy that moves with a parcel of water). The sampling methodology to be followed and the sampling gear to be used will depend on the objectives of the study, on the target species and/or the early life stages to be sampled.

It is out of the scope of this guide to explain in detail all possible sampling strategies or describe the different types of ichthyoplankton sampling devices and sampling methods. However, it is worth illustrating those that are most frequently used and the potential sources of bias affecting sample representativeness.

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As already explained, many ichthyoplankton studies are aimed at estimating the biomass of adult fish stocks, either by using eggs or larvae. When using eggs, for e.g. applying the Daily Egg Production Method (DEPM), the gear employed is the CalVET net (Figure 5) in vertical hauls, and the sampling strategy consists of sampling stations arranged in a regular grid, with a short distance between stations, covering the whole spawning area of the fish stock to be studied.

When the objective is to estimate the biomass of an adult fish stock using larvae, to study the horizontal distribution of fish eggs and larvae of specific species or, in general, the ichthyoplankton community of an area, the most adequate sampling gear to be used is the Bongo net (Figure 5) with a 60 cm diameter mouth opening. Hauls are oblique and the sampling strategy consists of sampling a regular grid of stations. Ichthyoplankton hauls must cover the depth distribution range of the target species or, if the goal is to sample the entire ichthyoplankton community, at least the upper 200 m of the water column, where most fish eggs and larvae are concentrated.

When the aim of ichthyoplankton studies is to investigate the vertical distribution of fish eggs and larvae and/or the vertical (diel and/or ontogenetic) migrations of fish larvae, a multiple net system, capable of sampling successive water layers during the same tow, is required (Figure 6). Hauls are also oblique.



Figure 5. Left, a CalVET net and right, a Bongo net



Figure 6. Multiple net systems. Left, a Hydrobios MultiNet and right, a MOCNESS (Multiple Opening-Closing Net and Environmental Sampling System) net

Neuston nets are used to sample the very most surface water layer. Large micronekton nets, fitted with meshes of the size of 1 mm or larger, may also be necessary for effectively sampling late larval fish stages (Figure 7). Light traps are used for capturing advanced post-larvae in the presettlement stages.

Ichthyoplankton samples can also be obtained with continuous sampling systems, such as the Continuous Underway Fish Egg Sampler (CUFES), capable of collecting plankton as the research vessel is sailing. It consists essentially of a pump that draws seawater at a given depth and sends it through a plankton collector, where plankton is retained (Figure 8). The collector, with its load of plankton, is changed after a fixed period of time. The plankton samples are preserved in a solution of formalin and seawater. After a certain period of time (about half an hour), the preservation liquid is eliminated, and the fish eggs of the target species are identified and counted under a stereomicroscope. The purpose of keeping plankton samples for a while in formalin is to kill eggs and make them opaque, as live eggs are transparent and invisible under the microscope. While CUFES is running, a data logger records the date, time, and position of each sample, as well as other environmental data from the ship's sensors.

In the last decades, new and more sophisticated sampling systems have been implemented. In situ ichthyoplankton imaging systems or plankton

Figure 7. Neuston net





Source: Marine Life Research Group, Scripps Institution of Oceanography, Continuous Underway Fish Eggs Sampler website, http://cufes.ucsd.edu/text/descr.htm, accessed on 9/9/23

acoustics, despite not providing biological samples for other types of analyses, can produce impressive amounts of data on larval abundance or biomass, over large areas in relatively shorts periods of time (Cowen and Guigand, 2008; Garcia-Seoane *et al.*, 2016).

Any sampling aimed at obtaining quantitative data on ichthyoplankton requires knowing the exact volume of water filtered by the net during each haul. This is done by using flowmeters attached to the mouth of the net. Flowmeters measure the length of the haul path, which, multiplied by the surface of the mouth of the net, gives the volume of water filtered by the net during the haul. This allows standardization of fish eggs and larvae captured in each haul to the number of individuals per cubic meter, or densities. Ichthyoplankton counts can also be standardised to the number of individuals beneath a unit of sea surface (square meter), by multiplying densities by the maximum depth of tow in meters (Smith and Richardson, 1977). Haul depth can be determined by using any



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device equipped with pressure sensors, such as a conductivity-temperature depth instrument (CTD) of small size or simple depth gauges, such as those used for scuba diving, attached to the sampling gear structure. If flowmeters or depth gauges are not available, the volume of water filtered can be estimated from the duration of the tow and the vessel's speed, which allows for estimating the distance covered by the net's path. Haul depth can be estimated by multiplying the maximum length of wire led out by the cosine of the wire angle at the moment when the maximum amount of wire has been let out. Finally, to get a reliable estimation of ichthyoplankton densities or abundances, it is necessary to homogeneously sample the water column over the whole haul depth range.

Once the haul is finished, plankton must be concentrated into the cod ends (Figure 5), by gently washing the nets. After that, plankton is transferred into jars and fixed for long-term preservation. The most commonly used preservatives are a 5 percent solution of buffered formalin and seawater, or ethanol at different concentrations. Formalin is more adequate for preserving samples to be used in taxonomic studies, since the egg and larval morphology is better preserved, but presents the problem that formalin is carcinogenic. Ethanol dehydrates both eggs and larvae. Eggs lose their shape, and larvae tend to curl making them difficult to measure. However, it has the advantage that specimens can be used for genetic and for larval growth studies. Other fixatives are liquid nitrogen, used to preserve larvae for biochemical and daily growth analyses, and RNAlater, when it is necessary to stabilize and protect cellular RNA. For some specific studies, larvae must be quickly sorted and identified on board, prior to fixation.

Sample representativeness

There are several papers dealing with problems associated to ichthyoplankton sampling (Hempel, 1974; Smith and Richardson, 1977; Lasker, 1981; Heath, 1992). Specifically, difficulties in obtaining representative samples of ichthyoplankton communities derive from the spatial and temporal heterogeneity in ichthyoplankton distribution and from the interaction of fish eggs and larvae with sampling gears (Ahlstrom *et al.*, 1973).

Fish eggs and larvae are not distributed randomly, but they tend to be concentrated in patches that become part of the plankton during specific periods of the year. This is the result of the distribution of adult fishes, their spawning strategies, hydrodynamic processes, egg and larval density, and larval behaviour, which varies throughout ontogeny. The only way to deal with the non-random distribution of ichthyoplankton is through an adequate sampling design.

The main sources of sampling bias, related to the direct interactions between fish eggs and larvae with the plankton nets, are escapement and avoidance. The escapement is the passage of smaller ichthyoplankton organisms through the meshes of the net. It may be active or passive and it is a function of the size, shape and behaviour of the organisms in relation to mesh size (Vannucci, 1968).

The active escapement is the process by which fish larvae caught in the net may squeeze through the meshes. This involves behavioural patterns that vary with species and with the developmental stage (Vannucci, 1968).

Passive escapement or extrusion is the forced passage of eggs or larvae through the net's meshes. When the organisms are larger than the meshes, extrusion is aided by the compressibility of the organisms and the flexibility of the meshes (Saville, 1958). This source of sampling bias is easily overcome by using net meshes of an adequate size to retain smaller fish eggs and larvae. Considering the fact that most fish eggs have a diameter of over 0.5 mm and that most newly hatched larvae are more than 2.0 mm long, the adequate mesh size for sampling ichthyoplankton is between 300 and 500 microns. However, mesh size should not be too small for avoiding, or at least reducing, mesh clogging produced by plankton. Net clogging reduces the filtration efficiency of the net.

Avoidance is the process by which larger fish larvae avoid capture, swimming out of the path of the approaching net or migrating below the net's maximum sampling depth (Morse, 1989). This is

probably the most important source of sampling bias (Clutter and Ankaru, 1968). Evidence of this phenomenon is the observed difference in larval fish abundances between night and day samples (Russell, 1976; Bridger, 1956; Ahlstrom, 1954, 1959). However, it seems that avoidance occurs both day and night (Murphy and Clutter, 1972). Larvae are not only able to see the approaching nets (visual avoidance), but they are capable of detecting the water vibrations produced by the towing wires and also the wave pressure produced by the net ahead of it (Smith and Clutter, 1965; Mahnken and Joss, 1967; Tranter and Smith, 1968). Thus, to reduce or minimize net avoidance, it is useful to use nets without bridles in front of the net mouth, such as Bongo-type nets (McGowan and Brown, 1966).

Ichthyoplankton sorting

For the taxonomic identification of ichthyoplankton or for other analyses, fish eggs and larvae are usually sorted from the plankton samples. The first step in ichthyoplankton sorting is to eliminate formalin from plankton samples. To do this, samples must be sieved through meshes (of smaller sizes than those used for sampling at sea), washed with seawater and placed into jars with seawater. This task must be carried out under a fume hood to avoid breathing the carcinogenic formalin vapours. Then, the sample can be placed in Petri dishes and analysed under a stereoscopic microscope, at low magnification (10x is adequate), to detect and sort fish eggs and larvae.

Considering the low abundances of larvae of some species, it is recommendable, for studies on ichthyoplankton communities, to analyse the whole sample. However, for specific studies focusing on eggs or larvae of more abundant species, it is acceptable to analyse 50 or 25 percent of the sample or even less, depending on the abundance of the early life stages of the target species. Fish eggs and larvae must be handled very carefully, with soft tweezers, thin brushes or pipettes, to separate and keep them in vials with the appropriate preservative medium.

Taxonomic identification of fish eggs and larvae

The taxonomic identification of fish eggs and larvae is not an easy task. It is more difficult than identifying the juvenile and adult stages of fishes. This is due to several reasons. First, because of the small size of fish eggs and larvae, characters useful for their identification can only be visualized under a stereoscopic microscope. Then, in the case of fish larvae, the main problem is that they undergo continuous and, in some cases, dramatic morphological, meristic, morphometric and pigmentary changes throughout their development. In addition, other characters, such as pigmentation patterns, can present an important geographical and even individual variability. This has generally prevented the building of dichotomous keys, such as those existing for adult fishes, although partial dichotomous keys were developed for the western Mediterranean Sea by Abossouan (1964) and Marinaro (1971). However, these keys only include some developmental stages of a relatively small number of the species present in the area. Therefore, they easily lead to identification dead ends or to misidentifications. To overcome this problem, the use of intelligent software, capable of managing extensive databases and integrating all the available information on larval stages of different fish species, including ecological information, such as the distribution of adults, reproduction season, etc. has been proposed (Froese and Papasissi, 1989).

However, the design and implementation of such information systems are not easy, due to the heterogeneity of the available information, or even the lack of information on the early life stages of many marine fish species. Indeed, even in well-studied areas, eggs and larvae of many species are still unknown and, in many cases, some larval stages remain undescribed.

There are two basic ways to describe the early life stages of fishes. One consists of rearing eggs and larvae in captivity, and the other to use ichthyoplankton collections to construct these series, working backwards from juveniles to larvae and, in some cases, to eggs (Moser and Ahlstrom, 1981). More recently, molecular techniques are being used to identify fish eggs and larvae, or for validating previous descriptions of these, but they cannot be routinely applied.

Because of all the above difficulties, the "look-alike" method remains the method//approach usually used to identify fish eggs and larvae. It consists of comparing the individuals with descriptions made by other authors. This elimination process ultimately results in "assigning" an individual to a species. This is also the method followed in this guide.

However, to properly use the "look-alike" method, it is first necessary to compile and analyse all the available information about the ichthyofauna of the study area. That is to say, faunistic lists and all the available information on the spawning strategies, spawning seasons and sites of the fish species inhabiting the study area. Another prerequisite for using this method is being able to accurately determine the developmental stage of the individual under analysis and compare it with the corresponding developmental stage described in the literature.

The nomenclature of the different early developmental stages of fishes varies by author. In this guide, we follow the nomenclature suggested by Kendall *et al.* (1984) (Figure 9), one of the most widely accepted. They divided the early life-history of fishes into three stages: **egg**, from spawning to hatching; **larva**, from hatching to the attainment of complete fin-ray counts and beginning of squamation (juvenile); and **juvenile**, young fish, fundamentally like the adult in the meristic characters (excluding squamation) but smaller and reproductively inactive. Kendall *et al.* (1984)



Figure 9. Early life history stages of Trachurus symmetricus

Source: redrawn by L. Rodriguez after Kendall *et al.* 1984. Early life history of fishes and their characters. *In* G. Moser, W.J. Richards, D.M. Cohen, M.P. Fahay, A.W. Kendall Jr. & S.L. Richardson, eds. *Ontogeny and systematics of fishes.* Spec. Publ. No. 1, pp. 11-22. American Society of Ichthyologists and Herpetologists, La Jolla, California

also divided the larval stage into four sub-stages: **yolk-sac larva**, from hatching to exhausting of yolk reserves; **preflexion larva**, from yolk exhausting to the beginning of upward flexion of the notochord; **flexion larva**, that ends when the urostyle is in its final position, at approximately 45° from the notochord axis and **post-flexion larva**, that ends when metamorphosis begins. The transformation, or metamorphosis is a transitional stage between larva and juvenile during which the young fish loses larval characters and acquires those of the adult (Kendall *et al.*, 1984; Moser, 1996).

Most marine fish eggs encountered in plankton samples are spherical and transparent, with a diameter of about 1.0 mm. Eggs are enclosed by a thin membrane or chorion. There is a space between the chorion and the yolk sac, the perivitelline space. Most fish eggs have oil globules (Figure 10).

The main anatomical, morphological and morphometric features used in fish egg identification are: egg shape (spherical or elliptic); egg size (ranges from 0.5 to more than 5.5 mm in diameter); type of surface membrane (smooth, sculptured, with a single protuberance or filaments); the presence of a second internal membrane; type of yolk (homogeneous or segmented); the size of the perivitelline space; absence/presence, number, position and colour (for live individuals) of oil globules. When the







embryo is well developed, embryonic characters, such as morphological features, pigment patterns and special structures are also used to identify fish eggs (Rass, 1946; Russell, 1976; Matarese and Sandknop, 1984; Ahlstrom and Moser, 1980).

Scanning electron microscopy has proved to be a good tool for the taxonomic identification of fish eggs, but cannot be used in routine analysis (Boehlert, 1984).

The identification of fish eggs is usually a more difficult task than larval fish identification. This is due to the low number of fish species with the egg stage described. In the Mediterranean Sea, it is worth mentioning the pioneer identification guide to fish eggs developed by Marinaro (1971), which has been recently revised and extended by Crec'hriou *et al.* (2015).

In most fish species with pelagic eggs, the newly hatched larvae are in general less than 4 mm long (Russell, 1976). The size and state of development at hatching are generally related to yolk-sac size. Typically, the body length at hatching is 2.5 to 3.0 times the diameter of the yolk sac (Moser, 1996). In general, yolk-sac larvae hatched from eggs less than 1.5 mm in diameter have

an unformed mouth, unpigmented eyes and pectoral-fin buds, while yolk-sac larvae hatched from larger eggs are comparatively well developed, with the mouth formed, eyes pigmented and pectoral fins developed (Moser, 1996). In both cases, the locomotion is aided by a prominent fin-fold that extends from the top of the head, around the caudal region, and ventrally forward to the posterior margin of the yolk sac (Figure 11).



Figure 11. Main anatomic features of a yolk sac larva

Source: modified after Russell, F.S. 1976. *The eggs and planktonic stages of British marine fishes*. Academic Press, London. 524 pp.

The identification of yolk-sac larvae is very difficult because some structures, such as fins and most of the specialized larval characters, resulting from the evolutionary adaptation to the plankton realm, are not yet well developed. Consequently, yolk-sac larvae of different species may be very similar. The yolk-sac stage is characterized by the migration, coalescence and rearrangement of pigment cells or melanophores (Moser, 1996). Melanophores are amoeboid and capable of migrating from their point of origin in the neural crest to various sites in the larva, to establish the species-specific larval pigmentation pattern, at the end of the yolk-sac stage. Useful characters for the identification of yolk-sac larvae are the shape and relative size of the yolk sac, the presence and the relative position of oil globules in the yolk sac, the position of the anus in relation to the yolk sac and, in some, the species-specific pigmentation patterns.

The complete utilization of the yolk marks the end of the yolk-sac stage. By this time, most of the organs and the sensory system required to capture prey are functional. The mouth and gut are formed, the anus is open at the margin of the ventral fin-fold, the eyes are pigmented, and the primordial and pectoral fins are present. It is now an early larva and during the larval development, the fish gradually acquires the characters of the adult, thus facilitating its identification (Moser, 1996; Russell, 1976). At first, the body is still surrounded by the primordial fin, the urostyle is straight and rudiments of the hypural elements are visible in its ventral side. As the larva grows, the urostyle bends upwards, the hypural elements become defined, caudal-fin rays develop, and the

first signs of the formation of the dorsal and anal fins appear as interspinous areas. At this stage, both the meristic characters and the pigmentation patterns that are characteristic of the adult of the species, have usually appeared (Russell, 1976). The main anatomical features of a larva are shown in figure 12.

The main characters used in larval fish identification are the body form, the





pigmentation pattern and the meristic and morphometric characters. The body form allows for separating larvae into several major groups (Russell, 1976). For example, larvae with narrow, elongated bodies (e.g. Families Clupeidae, Engraulidae, Stomiidae, etc.); larvae with laterally compressed bodies (includes all flatfishes, e.g. Families Bothidae, Pleuronectidae, Soleidae, etc.); bodies with the typical fish shape (includes larvae of most fish species, e.g. Families Gadidae, Triglidae, Gobiidae); bodies with aberrant shapes (e.g. Family Belonidae) or showing specialized larval characters for the plankton life, such as cranial armatures (e.g. Family Scorpaenidae), elongate fin rays (e.g. Families Carapidae, Lophiidae), stalked eyes (e.g. some Myctophidae species) or early developed and large fins (e.g. Family Trachinidae).

Meristic characters are countable structures appearing in series, such as the number of myomeres, vertebrae or fin rays. They have a high diagnostic value (at least the combination of several counts), because they are species-specific. However, they have the disadvantage that some of them, such as fin rays, are completely formed only in older larvae, which are scarce in plankton samples. Others, such as myomeres or vertebrae, are difficult to visualize, even using staining methods, or other techniques, such as X-rays (Pothoff, 1984; Tucker and Laroche, 1984).

Morphometric characters include the different measurements of a larva. The main measurements of a larva, shown in figure 13, are the total length (TL), or the distance from the tip of the snout to the caudal-fin end; the standard length (SL), or the distance between the tip of the snout and



Figure 13. Body regions and the most important measurements of a fish larva Source: author's own elaboration

the urostyle end; the preanus length, or the distance between the tip of the snout and the anus; the head length, or the distance from the tip of the snout to the border of the cleitrum, and the eye diameter, or the maximum diameter of the eye. When measuring the different body regions for identification purposes, the stage of development of the larvae must be taken into account, since larval growth is allometric. It must be noted that formalin-preserved larvae suffer shrinkage that increases with the time of preservation and decreases with larval growth (Theilacker, 1978).

Fish larvae show a variety of pigmentary cells or chromatophores. Those containing black or brown pigments are known as melanophores; those with yellow pigments, xanthophores and those with red pigments, erythrophores (Russell, 1976). However, in formalin-preserved specimens, only black pigmented cells or melanophores remain. For this reason, the latter are usually the only ones used in ichthyoplankton identification. Melanophores are situated in different parts of the body, defining species-specific pigmentation patterns. These patterns are one of the chief diagnostic characters used for the identification of the larval stage of fish species. Often, the identification of species with similar larvae is made possible thanks to the presence or absence of a single melanophore or by its position (Russell, 1976). The general appearance of preserved specimens will differ very much according to the degree of expansion or contraction of the melanophores. Moreover, in specimens preserved for a long time, the pigmentation will fade, especially if kept in the light (Russell, 1976). Besides, there may be an intraspecific and geographical variability in pigmentation patterns.

The importance of proper taxonomic identification of ichthyoplankton

The first step in any ichthyoplankton study is the proper taxonomic classification of eggs and/or larvae since, as Powles and Markle (1984) pointed out, small errors in their identification can result in important misinterpretations about the biology and ecology of fish species. Moreover, there are several examples in the literature showing that wrong identifications of fish eggs or larvae have led to biased stock evaluations and, subsequently, inadequate management measures (Daniel and Graves 1994; Armstrong *et al.* 2001; Fox *et al.* 2005). Unfortunately, these identification errors are probably more frequent and important than desirable. For example, a recent study focusing on the ability of researchers from five different laboratories in Taiwan to identify fish larvae determined that the average accuracy of identification was 80.1, 41.1 and 13.5 percent at the family, genus and species levels, respectively (Ko *et al.* 2013). Recently, Puncher *et al.* (2015a) revealed that the Atlantic bluefin tuna (*Thunnus thynnus*) larvae have been misidentified in the Mediterranean Sea. These authors demonstrated, through genetic analysis, that more than half of the larvae, submitted by three Mediterranean institutions to a bluefin tuna research project, funded by the International Commission for the Conservation of Atlantic Tunas (ICCAT), were misidentified.

Some of the errors in identifying ichthyoplankton are caused by the persistence in the literature of wrong descriptions. However, it can also happen that in species whose larvae are accurately

described, the inexperience or lack of training of the people in charge of the taxonomic identification of the ichthyoplankton can lead to massive misidentifications (Puncher *et al.*, 2015b). Because of this, it is of paramount importance to produce identification guides with accurate descriptions, taking advantage of molecular genetic techniques, to validate doubtful identifications, as well as to organize courses given by experienced ichthyoplanktologists, and aimed at properly training new generations of technicians and researchers in charge of the taxonomic classification of fish eggs and larvae.

The study area

The geographical area covered by this guide (Figure 14) includes one of the most important coastal upwelling areas of the world: the Canary Current upwelling system that extends from about 10°N to the Strait of Gibraltar.



Figure 14. Geographic area covered in this guide Source: authors' own elaboration, conforms to UN Map of the World, 2022

The upwelling between Gibraltar and Cape Blanc is produced by favourable northeasterly winds throughout the year, although winds and upwelling are more intense during the summer months. Between Cape Blanc and Cape Vert, the upwelling has a marked seasonal periodicity, reaching its peak of intensity during winter (Arístegui *et al.*, 2004, Mittlelstaedt, 1991).

In coastal upwelling areas, the wind blows parallel to the coastal line or at a slight angle to it. As the wind begins to blow across the surface of the ocean, it transmits its force by friction to the surface layer of the sea, and a thin surface slab of water (25 - 50 m thick) is set in motion (Figure 15). As a result of the Coriolis force, the wind-driven layer (named Ekman layer) has a net movement 90° to the right (left) of the wind in the Northern (Southern) Hemisphere. To replace the surface water mass that moves offshore, subsurface, cool and rich nutrient waters, at a depth of 200 m at most, flow inshore and up to the surface layer, and then offshore in the surface divergence layer (Barber and Smith, 1988; Mann and Lazier, 2006).

The Ekman layer transports offshore the cool, nutrient-rich upwelled water with its load of phytoplankton, zooplankton and ichthyoplankton. The upwelling has a limit, between 50-100 Kms offshore, where the interface between the upwelled water and the offshore waters is located (Man and Lazier, 2006). This interface constitutes the upwelling front (Barber and Smith, 1988).



Figure 15. Schematic representation of the coastal upwelling phenomenon

Source: NOAA. What is upwelling?. National Ocean Service website, https://oceanservice.noaa.gov/facts/upwelling.html, accessed on 9/9/23

Upwelling fronts are recognized as an important part of the coastal upwelling processes (Brink, 1987) that have an enormous biological importance. These are zones of high primary production, where the zooplankton and the ichthyoplankton accumulate (e.g. Le Febre, 1986; Owen, 1981; Sabatés and Masó, 1990), and are even used by some fish species as spawning and nursery areas (Olivar, 1990). Moreover, they work as a barrier preventing fish larvae from being transported into the open sea, where they would die of starvation.

In upwelling regions, fish larvae of inshore-shelf spawning species may follow different strategies to remain within the appropriate areas for the development and avoid the offshore advection by the Ekman layer. They may migrate vertically between the two flow regimes, associated with the upwelling process, daily (Parrish *et al.*, 1981; Myers and Drinkwater, 1989; Landaeta and Castro, 2002) or ontogenetically (Gorbunova *et al.*, 1986), or spawning may take place during upwelling relaxation or adult fish may spawn demersal eggs (Parrish *et al.*, 1981). In the case of ontogenetic migration, early fish larvae are transported offshore by the Ekman layer, to the upwelling front where late larvae migrate to the deep, onshore flow, to reach the neritic region again. Moreover, fish larvae of offshore spawning species may use the deep onshore currents to reach the inshore nursery areas (Smith and Suthers, 1999; Landaeta and Castro, 2002, 2012).

Nevertheless, the natural barriers accounted by the upwelling front for the offshore transport of fish larvae may be broken down by the upwelling filaments. These are extensions of coastally upwelled water that reach hundreds of kilometers offshore. They are associated with narrow, elongated currents arising on the continental shelf and that extend long distances into the sea (Brink, 1983). Upwelling filaments are offshore transport mechanisms for nutrients and plankton, fish larvae included (Nelson *et al.*, 1998, Rodriguez *et al.*, 1999; Rodriguez *et al.*, 2004). In the case of the northwestern African coastal upwelling, numerous upwelling filaments are distributed along the coastal-offshore upwelling boundary. Nevertheless, only two, Cape Ghir and Cape Blanc giant filaments, remain as major, permanent features that stretch several hundreds of kilometers offshore (Aristegui *et al.*, 2004). Another important filament is the one that forms just north of Cape Bojador and extends 150 Km towards Gran Canaria (Barton *et al.*, 1998). This filament has been demonstrated to transport fish larvae, which can reach the Canary archipelago, and probably contributes to the replenishment of the fish populations of the Canary Islands waters (Rodriguez *et al.*, 1999, Rodriguez *et al.*, 2004; Brochier *et al.*, 2011).

The coastal upwelling regions only cover 1 percent of the total area of the world's oceans, but they provide about 50 percent of the fish captures of the world fisheries. Fish catches in the Northwest African upwelling region fluctuated between 1.3 Mt and 2.6 Mt over the period

1970-2000. Fisheries landings are mostly based on pelagic fish species (Figure 16), dominated by *Sardina pilchardus*, (Figure 17) that accounts on average for 70 percent of the total catches (Arístegui *et al.*, 2004).



Figure 16. Annual catches of pelagic and demersal fish species in the Canary Current region, from 1970 to 2000

Source: FAO-CECAF (Eastern Central Atlantic) capture production 1970-2000.



Figure 17. Contribution of the European sardine (*Sardina pilchardus*) to the total pelagic catches in the Canary Current from 1970 to 2000

Source: FAO-CECAF (Eastern Central Atlantic) capture production 1970-2000

2. ILLUSTRATIONS OF REPRESENTATIVE LARVAE OF BONY FISH FAMILIES INCLUDED IN THIS GUIDE





ORDER SCOMBRIFORMES





ORDER PLEURONECTIFOMES





Uranoscopidae



ORDER LOPHIIFORMES



Lophiidae
3. IDENTIFICATION SHEETS

Ariosoma balearicum (Delaroche, 1809)

Bandtooth conger - Congre des Baléares

Habitat: neritic, benthic, between 20 and 100 m depth

Distribution: Atlantic and western Indian oceans. Eastern Atlantic, from Angola to Portugal, and the Mediterranean Sea Meristic characters Myomeres: 120-137 Vertebrae: 120-136 Dorsal fin: NA Anal fin: NA

Spawning season: August to September (Mediterranean Sea)

YOLK-SAC LARVAE

Hatch size: < 5.0 mm Body: elongate Yolk sac: elongate Oil globule location: no information Anus: no information Preanus length: no information Pigmentation: unpigmented

LARVAE

PHOTOS

Colour: transparent

EGGS

Habitat: pelagic

Shape: spherical

Perivitelline space: large

Yolk: segmented; unpigmented

Oil globules: one; unpigmented

Body: very elongate, compressed dorso-laterally, and transparent (leptocephalus larvae); deepest region located slightly behind mid-length; tail moderately pointed; dorsal-fin origin located near posterior end of body; maximum body size 200 mm SL

Chorion: double capsule, smooth; diam. about 2.0 mm

Head: small (characteristic of leptocephalus larvae) with short snout; 4 prominent needle-like teeth in early larvae, in both jaws Eye: round and small Gut: straight, narrow and very long, almost reaching caudal region Preanus length: about 95% SL Air bladder: absent Spination: none

Pigmentation: early larvae (8.0 mm SL), 5-6 melanophores on each side of gut; at about 10 mm SL, 5 large melanophores appear dorsally on body (dorsal and ventral melanophores disappear with development); notochord tip pigmented; late larvae, lateral minute melanophores outlining myosepta, immediately below midline, forming a series of short diagonal lines from head to tail; small, ventral row of melanophores below gut anteriorly, switching to top of gut, behind liver; a series of small melanophores on dorsal midline, from head to tail; head unpigmented **Length at flexion:** flexion does not occur **Length at transformation:** unknown

by J.M. Rodriguez



Leptocephalus larva (anterior region), not sized



Leptocephalus larva (end of tail), not sized

CONGRIDAE

Ariosoma balearicum (Delaroche, 1809)



Illustrations' sources: A-D: L. Rodríguez (**A**, **B:** redrawn from D'Ancona, 1931b; **C**, **D:** redrawn from Smith, 1979)

CONGRIDAE

ANGUILLIFORMES

and 55 m depth Distribution: eastern Atlantic Ocean, from Dakar (Senegal) to the North Sea, and the Mediterranean Sea

Spawning season: September to May

Anal fin: 17-21

Meristic characters

Myomeres: 50-53

Vertebrae: 50-53

Dorsal fin: 17-18

European pilchard – Sardine commune



Body: very elongate and slender (clupeid shape*)

Oil globule location: at post-ventral edge of yolk sac

Anus: far behind yolk sac, reaches finfold border

small melanophores from head to tail; a ventral

Pigmentation: no dorsal melanophores; ventral

melanophores aligned on both sides of body,

Pigmentation: two parallel, dorsal rows of

EGGS

Chorion: smooth; diam. 1.30-1.90 mm Oil globules: one; diam. 0.14-0.18 mm; unpigmented Colour: transparent

YOLK-SAC LARVAE

Yolk sac: ovoid

Hatch size: 3.3-4.0 mm

Preanus length: about 83% TL

Habitat: pelagic Shape: spherical Perivitelline space: large Yolk: segmented; unpigmented

melanophore on caudal region

LARVAE

Body: very elongate and slender (clupeid shape*); head length included at least six times in TL; dorsal fin located ahead of anus, migrates forward during development Head: small, short and somewhat high; mouth terminal and relatively large **Eye:** round and relatively large Gut: straight, tube-like; differentiated into two sections

Preanus length: about 80% SL Air bladder: present in late larvae Spination: none

above gut; ventral rows of melanophores in posterior section of gut; caudal fin pigmented; some melanophores between anus and caudal fin (this character helps to distinguish this species from Sardinella aurita) Length at flexion: 10.0 mm Length at transformation: unknown

*Clupeid shape: body elongate and slender and long, tube-like gut

PHOTOS



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CLUPEIDA



CLUPEIFORMES

Sardinella aurita Valencienr	nes, 1847	Round sardinella – Allache		
Habitat: neritic, coastal, pelagic, between 0 and 300 m depth Distribution: Atlantic Ocean and the Mediterranean Sea; eastern Atlantic from South Africa to Cadiz Spawning season: April to November	Meristic character Myomeres: 45-48 Vertebrae: 45-49 Dorsal fin: 17-20 Anal fin: 16-20	ers		
EGGS	Fig. A	YOLK–SAC LARVAE Fig. B		
Habitat: pelagic Shape: spherical Chorion: smooth; diam. 1.20-1.35 mm Perivitelline space: large Yolk: segmented; unpigmented Oil globules: one; diam. 0.10-0.12 mm; unpigmented Colour: transparent		 Hatch size: about 2.5 mm Body: elongate and slender (clupeid shape) Yolk sac: spherical Oil globule: at ventral edge of yolk sac Anus: far behind yolk sac, reaches border of finfold Preanus length: about 85% SL Pigmentation: two parallel, dorsal rows of small melanophores, extending from head to tail 		
LARVAE		Figs. C-F		
 Body: elongate and slender (clupeid shape*); head length included less than 6 times in TL; dorsal fin located ahead of anus migrates forward during development Head: relatively small; mouth large, terminal, extends to mid-eye level Eye: round and relatively large Gut: straight, tube-like; differentiated into two sections Preanus length: ranges between 83 and 89% SL Air bladder: present in late larvae 		Spination: none Pigmentation: no dorsal melanophores; ventral melanophores aligned on both sides of body, above gut; ventral rows of melanophores in posterior section of gut; caudal fin pigmented; no melanophores between anus and caudal fin in early larvae (this character helps to distinguish this species from <i>S. pilchardus</i>) Length at flexion: 7.5-9.5 mm Length at transformation: 16.0-23.0 mm		

***Clupeid shape:** body elongate and slender and long, tube-like gut

PHOTOS

CLUPEIFORMES

by J.M. Rodriguez



CLUPEIDAE



Illustrations' sources: A: D'Ancona (1931a); B-F: L. Rodríguez (B: redrawn from D'Ancona, 1931a; C-F: redrawn from Conan and Fagetti, 1971)

Engraulis encrasicolus Lir	European anchovy - Anchois		
Habitat: neritic, coastal, pelagic, euryhaline Distribution: eastern Atlantic Ocean, from South Africa to Norway, and the Mediterranean Sea Spawning season: spring and summer	Meristic character Myomeres: 47 Vertebrae: 45-47 Dorsal fin: 16-18 Anal fin: 16-18	S I I I I I I I I I I I I I I I I I I I	
EGGS	Fig. A	YOLK-SAC LARVAE Fig. B	
Habitat: pelagic Shape: ovoid Chorion: smooth; diam. 1.2-1.9 x 0.5-1.2 mm Perivitelline space: small Yolk: segmented; unpigmented Oil globules: none Colour: transparent		Hatch size: 3.0-4.0 mm Body: elongate and slender (clupeid shape) Yolk sac: very elongated Anus: close behind yolk sac, reaches finfold border Preanus length: about 75% SL Pigmentation: recently hatched larvae unpigmented, late hatched larvae start to develop pigmentation of larvae	
LARVAE		Figs. C-G	
 Body: elongate and slender (clupeid shape); dorsal fin over anus Head: relatively small; mouth large, terminal, extends to middle of eye Eye: round and relatively large Gut: tube-like; differentiated into two sections; forms a small curve above gas bladder in late larvae Preanus length: about 75% SL in early larvae, decreases during development Air bladder: prominent in late larvae 		Spination: none Pigmentation: no dorsal melanophores; ventral melanophores aligned on both sides of body, above gut; ventral rows of melanophores in posterior section of gut; caudal fin and air bladder pigmented; some ventral melanophores between anus and caudal fin Length at flexion: 7.5-9.5 mm Length at transformation: 35.0-40.0 mm	

PHOTOS

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ENGRAULIDAE

30

Engraulis encrasicolus Linnaeus, 1758



Illustrations' sources: A-G: L. Rodríguez (**A**, **B:** redrawn from D'Ancona, 1931c; **C-E:** redrawn from Alemany, 1997; **F**, **G:** redrawn from Fage, 1920)

ENGRAULIDAE

CLUPEIFORMES

32

ARGENTINIDAE

ARGENTINIFORMES

Argentina sphyraena Linnaeus, 1758			Argentine – Petite argentine	
Habitat: neritic and upper slope, demersal, between 50 and 700 m depth Distribution: eastern Atlantic Ocean, from Western Sahara to northern Norway, and the Mediterranean Sea Spawning season: winter and spring	Meristic character Myomeres: 53-55 Vertebrae: 53-55 Dorsal fin: 10 Anal fin: 12 Adipose fin: prese after transformatio	s nt n		
EGGS	Fig. A	YO	LK–SAC LARVAE Fig. B	
Habitat: pelagic Shape: spherical Chorion: smooth; diam. 1.30-1.90 mm Perivitelline space: small Yolk: segmented; pigmented Oil globules: one; diam. 0.37-0.47 mm; pigmented Colour: transparent		Hatch size: about 7.5 mm Body: elongate and slender Yolk sac: elongate Oil globule location: at post-ventral edge of yolk sac Anus: far away from yolk sac, reaches finfold border Preanus length: about 67% TL Pigmentation: large stellate melanophores on yolk sac and groups of melanophores along dorsal side of gut; caudal–dorsal and ventral groups of melanophores; oil globule pigmented		
LARVAE			Figs. C-F	
Body: long and slender (clupeid shape) Head: relatively small; mouth terminal and small Eye: round and relatively large Gut: long, tube-like, slightly wavy Preanus length: about 72% SL Air bladder: absent Spination: none		Pigmentation: 6 groups of melanophores, approximately equidistant, situated along ventral region of trunk and tail (5 above gut and one on tail); 2 opposing groups of caudal melanophores; tips of upper and lower jaw pigmented; caudal fin pigmented; a melanophore on dorsal surface of terminal gut Length at flexion: begins at about 13.0 mm Length at transformation: unknown		
PHOTOS			by J.M. Rodriguez	
		11	6.0 mm SL	











ARGENTINIDAE

34

Glossanodon leioglossus (Valenciennes, 1848) Smalltoothed argentine - Argentine à petites dents

Habitat: demersal, outer shelf and slope, between 50 and 700 m depth Distribution: eastern Atlantic Ocean, from Mauritania to southern Spain, and the western Mediterranean Sea Spawning season: September to March (Mediterranean Sea) Meristic characters Myomeres: 51-52 Vertebrae: 51-52 Dorsal fin: 13-14 Anal fin: 11-12 Adipose fin: present



YOLK-SAC LARVAE

Fig. A

EGGS

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 1.44-1.52 mm Perivitelline space: small Yolk: segmented Oil globules: one; diam. 0.36 mm; unpigmented Colour: transparent Hatch size: unknown Body: elongate and slender Yolk sac: ovoid Oil globule location: at mid-ventral edge of yolk sac Anus: far away from yolk sac, reaches finfold border Preanus length: about 80% SL Pigmentation: a large patch in caudal region, over primordial fin; 3 large melanophores over gut, yolk sac unpigmented

LARVAE

Body: long and slender (clupeid shape); dorsal fin located at about middle of body; anal fin located at end of body **Head:** relatively small; mouth terminal and oblique **Eye:** round and large **Gut:** long, tube-like; end of gut detached from body, over anal fin

Preanus length: about 85% SL **Air bladder:** absent **Spination:** none

PHOTOS

Pigmentation: 8 strips of pigment along lateroventral sides of body, first one anterior to pectoral fin, one prolonged over terminal gut; a large melanophore at caudal end, spreading through caudal fin in late larvae; 2 large spots on head, one over upper jaw and another behind eye, over opercle

Length at flexion: unknown Length at transformation: unknown

by J.M. Rodriguez



9.5 mm SL

ARGENTINIFORMES





BATHYLAGIDAE

Bathylagoides argyrogaster (Norman, 1930)

Silver deepsea smelt

Habitat: oceanic, mesopelagic, usually between 200 and 300 m depth

Distribution: worldwide in tropical and subtropical waters. Eastern Atlantic, from South Africa to Mauritania Meristic characters Myomeres: 43-44 Vertebrae: 43-44 Dorsal fin: 12-13 Anal fin: 15-16

Spawning season: unknown

EGGS

Undescribed

Undescribed

YOLK-SAC LARVAE

Figs. A-E

LARVAE

Body: elongate; primordial finfold persists throughout all larval stages **Head:** rather elongate, increases in length with development; snout blunt and relatively wide; jaws reach anterior margin of eye; teeth develop in lower jaw at 6.0 mm SL (6 teeth in larvae of 10.0 mm SL) **Eye:** sessile; slightly oval in early larvae, becomes rounded and smaller with development **Gut:** elongate, tube-like **Preanus length:** increases from 80% SL in early larvae to 90% SL in late larvae **Air bladder:** absent **Spination:** none **Pigmentation:** early larvae, 7-8 ventral melanophores, dorso-lateral to gut, between pectoral-fin base and anus, increasing to 10 with development; ventral streak of melanophores on notochord tip, complemented by a dorsal one in some larvae; body unpigmented; late larvae, melanophores appear over body and head, increasing in number with development; gut unpigmented; dorsal and ventral melanophores over urostyle; caudal-fin rays pigmented **Length at flexion:** 8.5-10.2 mm SL **Length at transformation:** unknown

PHOTOS





4.5 mm SL

4.8 mm SL



7.7 mm SL

Bathylagoides argyrogaster (Norman, 1930)



BATHYLAGIDAE

BATHYLAGIDAE



10.0 mm SL



14.2 mm SL

protruding

very long before transformation

tube-like; moderately wavy



Nansenia oblita (Facciolà, 1887) Habitat: mesopelagic, oceanic, **Meristic characters** between 300 and 500 m depth Myomeres: 42-45 Distribution: North Atlantic Ocean Vertebrae: 42-45 and western Mediterranean Sea; Dorsal fin: 10-11 eastern Atlantic from about 24° N to Anal fin: 9-10 southwest of Ireland Spawning season: winter (Mediterranean Sea) YOLK-SAC LARVAE EGGS Fig. A Fig. Habitat: pelagic Hatch size: about 4.0 mm Shape: spherical **Body:** relatively elongate Chorion: with 'pustules' on inner surface; diam. Yolk sac: large and ovoid Oil globule: at ventral side of yolk sac 1.48-1.50 mm Perivitelline space: small Anus: detached from yolk sac reaches finfold Yolk: segmented border Oil globules: one; diam. 0.40-0.43 mm; Preanus length: about 77% SL unpigmented **Pigmentation:** unpigmented Colour: no information LARVAE Body: moderately elongate and laterally Spination: none compressed in early larvae, becomes rounded in Pigmentation: row of melanophores above and late larvae below notochord; scattered melanophores on head Head: relatively large; mouth terminal and small; appear as a horizontal bar through eye; rows of snout rounded

Eye: round and relatively large **Gut:** elongate, tube-like in early larvae; coiled anteriorly and detached from body at its end in late larvae

Preanus length: about 75% TL **Air bladder:** absent

PHOTOS

Pigmentation: row of melanophores above and below notochord; scattered melanophores on head appear as a horizontal bar through eye; rows of small melanophores along ventral side of trunk and tail, from pectoral–fin base to level of anus; row of melanophores along lower jaw; pigment spreads over most of body during development, except on caudal peduncle Length at flexion: 7.0-11.0 mm SL

Length at transformation: > 20.0 mm SL

by J.M. Rodriguez



3.4 mm SL



3.5 mm SL

40

MICROSTOMATIDAE

Nansenia oblita (Facciolà, 1887)



42



LARVAE

opposite

Head: relatively small; mouth terminal and large, extending well posterior to eye in late larvae Eye: relatively small and slightly oval Gut: elongate, tube-like, forming a slight curve above air bladder in late larvae

Preanus length: > 50% SL

Air bladder: prominent in late larvae, located over posterior gut

Spination: none

Pigmentation: a prominent melanophore on ventral side of caudal peduncle; a melanophore at middle of caudal-fin base; a pair of melanophores along lateral sides of gut: a melanophore over anus;

dorsum of air bladder pigmented; a series of 9-13 equidistant melanophores along postanal, ventral region, gradually becoming internal with development; 4-13 melanophores (number increases with development) on myosepta above gut; a few spots on head and lower jaw; none or one melanophore over notochord tip; postanal, dorsal melanophores appear at 6.5 mm behind dorsal-fin origin, spreading forward with development; air bladder pigmented Length at flexion: 5.0-6.0 mm SL Length at transformation: between 13.0 mm and 22.0 mm SL

Note: pigmentation is the primary character to distinguish species of the genus Cyclothone

PHOTOS

by J.M. Rodriguez







STOMIIFORMES

6.7 mm SL

STOMIIFORMES



Illustrations' sources: A-D: L. Rodríguez (redrawn from Watson, 1996a)

43

44



Undescribed

YOLK-SAC LARVAE

Figs. A

Body: elongate and slender; dorsal and anal fins

located at same level Head: relatively small; mouth terminal and large, extending well posterior to eye in late larvae

Eye: round and small **Gut:** elongate, tube-like, forming a slight curve

above air bladder

Preanus length: about 50% SL

Air bladder: prominent (absent in very early larvae), located over posterior of gut

Spination: none

Pigmentation: prominent spot on ventral side of caudal peduncle; three pairs of melanophores along lateral sides of gut: one close to pectoral-fin base,

another about at level of first 1/3 of gut, and a third one over anus; dorsum of air bladder pigmented; a series of 9 to 12 equidistant melanophores along postanal, ventral region in early larvae; late larvae show a regular ventral, postanal row of internal melanophores in correspondence with external postanal row; row of 5-6 internal melanophores on ventrolateral anterior region of body, over gut, following myosepta, and about 3 internal melanophores on upper part of caudal peduncle; dorsum of air bladder pigmented Length at flexion: about 4.8 mm Length at transformation: > 12.0 mm

PHOTOS

by J.M. Rodriguez



3.6 mm SL



5.0 mm SL



10.5 mm SL

Garrick



GONOSTOMATIDAE

STOMIIFORMES

YOLK-SAC LARVAE

Atlantic fangjaw

GONOSTOMATIDAE

Meristic characters Myomeres: 37-40 Distribution: eastern Atlantic Ocean, Vertebrae: 38 from the Gulf of Guinea to Morocco. Dorsal fin: 16-18 Anal fin: 28-30

6

EGGS

Ocean)

Undescribed

LARVAE

PHOTOS

Undescribed

Body: slender; anal-fin origin anterior to dorsal-fin origin; pectoral fin forms on peduncle Head: relatively small; mouth large and slightly oblique; forehead and snout convex; teeth on maxilla in larvae > 12.0 mm SL Eye: oval and moderately large Gut: tubular, forms a loop below gas bladder; terminal gut makes a right angle with body in early larvae; anus protruding Preanus length: about 55-60% SL Air bladder: present and large Spination: none

Gonostoma atlanticum Norman, 1930

Habitat: oceanic, mesopelagic,

between 50 and 1 350 m depth

Spawning season: August to September (northwestern Atlantic

Absent from the Mediterranean Sea

Pigmentation: in early larvae, 5-13 melanophores along ventral tail region; a pair of ventral melanophores on gut, close to pectoral-fin base and two lateral, at beginning of gut loop: a melanophore on terminal gut; melanophores over gas bladder in larvae > 5.0 mm SL; sparse melanophores on caudal fin; in late larvae, add a row of melanophores along anal-fin base; a row of melanophores on each side of gut; some melanophores on caudal peduncle Length at flexion: about 4.5-6.0 mm SL Length at transformation: about 15.0-21.0 mm SL

by J.M. Rodriguez







STOMIFORMES





47

Gonostoma denudatum Rafinesque, 1810

Habitat: oceanic, mesopelagic, between 100 and 700 m depth Distribution: Atlantic Ocean, from Angola to Portugal, and the Mediterranean Sea

Spawning season: unknown

Meristic characters Myomeres: 38-39 Vertebrae: 39 Dorsal fin: 14-15 Anal fin: 28-30 Adipose fin: present



EGGS

Undescribed

Undescribed

YOLK-SAC LARVAE

Figs. A-E

LARVAE
Body: slender; dorsal- and anal-fin origin at about

the same level

Head: relatively small; mouth large and slightly oblique; snout relatively short **Eye:** slightly oval, and moderately large

Gut: tubular, forms a loop under air bladder; anus protruding; terminal gut makes a right angle with body in early larvae

Preanus length: just > 50% SL **Air bladder:** prominent **Spination:** none

Pigmentation: early larvae, 0-2 melanophores on

gut loop; a series of melanophores along ventral margin of tail; usually a large melanophore behind eye; dorsum of air bladder pigmented; a melanophore over terminal gut; late larvae add a series of melanophores along lower part of body, from pectoral fin to air bladder; a diagonal streak of pigment over caudal peduncle; two pairs of melanophores on dorsum, one under dorsal-fin base and another behind; a single dorsal melanophore posterior to head **Length at flexion:** about 7.0 mm **Length at transformation:** unknown

PHOTOS



A, B: J.M. Rodriguez; C: S. Isari



6.7 mm SL

6.9 mm SL



12.2 mm SL

GONOSTOMATIDAE

STOMIIFORMES

Gonostoma denudatum Rafinesque, 1810



Ichthyococcus ovatus (Cocco, 1838) Lightfish Habitat: oceanic, mesopelagic, **Meristic characters** between 0 and 750 m depth Myomeres: 38-42 **Distribution:** subtropical waters Vertebrae: 38-42 of the Atlantic Ocean and the Dorsal fin: 11-12 Mediterranean Sea (uncommon north Anal fin: 15-17 of the Gulf of Cadiz) Adipose fin: present Spawning season: late spring and early summer YOLK-SAC LARVAE EGGS Undescribed

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 0.80 mm Perivitelline space: small Yolk: segmented; unpigmented Oil globules: single; diam. 0.24 mm Colour: transparent

LARVAE

Body: elongate, slender, and rounded; dorsal and ventral margins of body almost parallel; lower pectoral-fin rays very elongate in early larvae; anal-fin origin well posterior to dorsal-fin origin; pelvic fin under dorsal-fin origin

Head: long; snout broad, depressed and pointed; mouth larger in early larvae **Eve:** oval

Gut: tubular, slender and almost constant in diameter throughout its length, detached from body at its end, extending above anal fin

PHOTOS

Preanus length: about 80% SL Air bladder: absent Spination: none Pigmentation: melanophores distributed on miomeres below midline of body; a few spots on snout, on detached section of gut, and on caudal peduncle; pectoral fins pigmented Length at flexion: unknown Length at transformation: unknown

by J.M. Rodriguez



STOMIIFORMES

51



Literature: Badcock (1984b), Fahay (2007), Jespersen and Tåning (1926), Olivar and Fortuño (1991), Richards (2006f) Illustrations' sources: A: Sanzo (1930); B, D, E: Jespersen and Tåning (1926); C: L. Rodríguez (redrawn from Ahlstrom *et al.*, 1984)

52 Pollichthys mauli (Poll, 1953) **Stareye lightfish** PHOSICHTHYIDAE Habitat: oceanic, mesopelagic, **Meristic characters** between 100 and 500 m depth Myomeres: 40-44 Distribution: worldwide in tropical Vertebrae: 40-44 and subtropical waters. Eastern Dorsal fin: 10-12 Atlantic, from 34°S to 60°N (absent Anal fin: 25-26 from the Mediterranean Sea) Adipose fin: present Spawning season: peaks in summer YOLK-SAC LARVAE EGGS Undescribed Undescribed LARVAE Body: very elongate and slender (clupeid shaped); Preanus length: increases with development from anal fin located very posteriorly in early larvae, 80% SL in early larvae to 88% SL in late larvae migrates forward to under dorsal fin in late larvae; Air bladder: small in early larvae, visible at 5.0 mm pectoral fins pedunculated SL, just behind mid-body Head: small and relatively depressed; mouth large; Spination: none snout sharply pointed; teeth appear on maxilla at Pigmentation: none until transformation* 3.5 mm SL Length at flexion: 3.6-7.0 mm SL Eye: oval, slightly directed forward, with ventral Length at transformation: 16.0-18.0 mm SL mass of choroid tissue, becomes rounded with development *Some early larvae collected in the Eastern Atlantic Gut: tube-like, very elongate, and straight Ocean have the dorsum of the air bladder pigmented PHOTOS 18.5 mm SL 19.0 mm SL init inter and the second 20.0 mm SL

Pollichthys mauli (Poll, 1953)



Vinciguerria attenuata (Cocco, 1838) Habitat: oceanic, meso- to bathypelagic, **Meristic characters** between 100 and 2 000 m depth Myomeres: 40-41 **Distribution:** worldwide in tropical Vertebrae: 40-41 to temperate waters Dorsal fin: 13-15 Spawning season: spring to summer

(Mediterranean Sea)

Anal fin: 14-16

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YOLK-SAC LARVAE

Silvery lightfish

EGGS

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 0.84-0.92 mm Perivitelline space: small Yolk: unsegmented; pigmented Oil globules: one; diam. 0.18-0.20 mm; unpigmented Colour: transparent

Undescribed

LARVAE

Body: very elongate and slender (clupeid shaped); body depth does not exceed 6-9% of body length; anal-fin origin located under 5th-6th dorsal-fin ray (middle), both fins located at level of anus Head: relatively elongated; mouth large; snout flattened and concave; thin teeth appear in early larvae

Eve: oval and semi-stalked Gut: tube-like, elongate, forming a small curve above air bladder Preanus length: about 75% SL, decreases throughout development

Air bladder: not present in early larvae, prominent in late larvae, located at about mid-body Spination: none Pigmentation: 6-8 stellate melanophores on both sides of body in larvae < 6.0 mm SL; dorsum of airbladder pigmented; median caudal spot, prominent in late larvae; there may be a melanophore on terminal gut

Length at flexion: unknown Length at transformation: unknown

Remarks: adipose fin present in late larvae

PHOTOS

by J.M. Rodriguez





4.6 mm SL







55

Vinciguerria attenuata (Cocco, 1838)



Illustrations' sources: A, B, D: Alemany (1997); C, E, F: Jespersen and Tåning (1926)



10.1 mm SL

6.6 mm SL

56



Illustrations' sources: A-G: L. Rodríguez (**A**, **B:** redrawn from Gorbunova, 1968; **C:** redrawn from Jespersen and Tåning, 1926; **D**, **E:** redrawn from Gorbunova 1981; **F**, **G:** redrawn from Okiyama, 1988)

PHOSICHTHYIDAE



PHOTOS

by J.M. Rodriguez





10.2 mm SL



11.0 mm SL
Vinciguerria poweriae (Cocco, 1838)



redrawn from Ahlstrom et al., 1984)

PHOSICHTHYIDAE

STOMIIFORMES

	Argyropelecus hemigymnus Cocco, 1829			Half–naked hatchetfish		
	Habitat: oceanic, mesopelagic, between 100 and 800 m depth Distribution: eastern Atlantic Ocean, from South Africa to north of the British Isles, and the western Mediterranean Sea Spawning season: throughout the year	Meristic character Myomeres: 38-39 Vertebrae: 36-41 Dorsal fin: 8-9 Anal fin: 11-12	rs			
	EGGS	Fig. A	YO	LK-SAC LARVAE Fig. B		
)	Habitat: pelagic Shape: spherical Chorion: smooth with inner membrane; diam. 0.92- 1.04 mm Perivitelline space: small Yolk: segmented; unpigmented Oil globules: one; diam. 0.26-0.28 mm; unpigmented Colour: transparent		Hatch size: about 2.5 mm Body: elongate Yolk sac: large, ovoid, projected beyond snout Oil globule location: at ventral side of yolk sac Anus: detached from yolk sac, reaches border of finfold Preanus length: about 50% SL Pigmentation: unpigmented			
	LARVAE	ARVAE		Figs. C-F		
	Body: very elongate and slender in early larvae, uffers a strong shrinkage during transformation especially anterior part of body), gut shortens, and lead deepens Head: large with blunt snout; mouth large, xtending to middle of eye Eye: oval and narrow in early larvae, becomes lightly telescopic and directed dorsally in late arvae Sut: swollen and detached from body Preanus length: about 50% SL in larvae between .0 and 9.0 mm, becoming shorter throughout levelopment		Air k Spin Pign later oper subs area Leng Leng	pladder: present in late larvae ation: none nentation: unpigmented before flexion; in larvae, stomach, frontal areas next to eyes, cle below eye and air bladder pigmented; equently pigment increases on head and gut with at flexion: between 10.0 and 11.0 mm with at transformation: 7.8-12.0 mm		

PHOTOS

STOMIIFORMES

by J.M. Rodriguez



4.3 mm SL





8.8 mm SL

STOMIIFORMES

Argyropelecus hemigymnus Cocco, 1829



Illustrations' sources: A, B: Sanzo (1931b); C, E, F: Alemany (1997); D: Jespersen and Taning (1926)

Habitat: oceanic, mesopelagic, between 10 and 400 m depth Distribution: Atlantic Ocean; eastern Atlantic Ocean, from Senegal to Norway, and the Mediterranean Sea Spawning season: throughout the year	Meristic characters Myomeres: 33-35 Vertebrae: 33-35 Dorsal fin: 10-11 Anal fin: 19-22	
EGGS	Fig. A	YOLK-SAC LARVAE Fig. E
Habitat: pelagic Shape: spherical Chorion: ornamented with hexagon diam. 1.30-1.53 mm Perivitelline space: small Yolk: segmented; unpigmented Oil globules: one; diam. 0.24-0.28 mr Colour: transparent	al sculptures; n; unpigmented	Hatch size: 3.0 mm Body: relatively elongate and slender Yolk sac: oval, very large; projected beyond snout Oil globule location: at ventral side of yolk sac Anus: detached from yolk sac, reaches border of finfold Preanus length: 60% SL Pigmentation: unpigmented

LARVAE

Body: elongate and slender **Head:** relatively large; mouth large, reaching posterior edge of eye in pre-transformation larvae **Eye:** oval, becoming round in late larvae **Gut:** tube-like, elongate, forming a small curve over air bladder **Preanus length:** about 50% SL **Air bladder:** prominent, located at about middle

Maurolicus muelleri (Gmelin, 1789)

Spination: none

Pigmentation: early larvae unpigmented; pigmentation limited to dorsum of air bladder (and photophores) in late larvae; Atlantic specimens may have a row of 4-10 melanophores along anal-fin base **Length at flexion:** 4.0-6.0 mm **Length at transformation:** attained at 13.0-14.0 mm

PHOTOS

of gut



0

7.6 mm SL

10.5 mm SL

by J.M. Rodriguez



6.5 mm SL

62

along along Lengti r: prominent, located at about middle DS

STOMIIFORMES

Silvery lightfish

STOMIIFORMES

Maurolicus muelleri (Gmelin, 1789) **B.** 3.0 mm Α. 9 0 C. 2.9 mm SL Early larvae unpigmented Dorsum of swimbladder pigmented **D.** 5.8 mm SL E. 6.4 mm SL At about 6 mm SL, a photophore under eye and 2 in ventral region, at level of swimbladder F. 10.0 mm SL Literature: Badcock (1984c), Fahay (1983), Jespersen and Tåning (1926), Sanzo (1935)

Illustrations' sources: A, B: Sanzo (1935); C, D: L. Rodríguez (C: redrawn from Alemany, 1997; D: redrawn from Halbeisen, 1988); E, F: Jespersen and Tåning (1926)

Chauliodus sloani Schneider, 1801 Sloane's viperfish Habitat: oceanic, from near the **Meristic characters** surface (at night) to 1 800 m depth Myomeres: 54-62 Distribution: warm and temperate Vertebrae: 54-62 waters of the Atlantic, Pacific and Dorsal fin: 5-7 Indian oceans, and the Mediterranean Anal fin: 10-13 Sea Adipose fin: dorsal and Spawning season: all year round, anal peaking in late winter and early spring YOLK-SAC LARVAE EGGS Fig. A Fig. Habitat: pelagic Hatch size: about 7.2 mm Shape: spherical **Body:** very elongate and slender Chorion: smooth, double membrane; diam. 2.24-Yolk sac: very elongate Anus: slightly behind yolk sac, reaches finfold 2.52 mm Perivitelline space: small border Preanus length: about 90% SL Yolk: segmented; unpigmented Oil globules: none Pigmentation: reduced to caudal region of finfold Colour: transparent LARVAE Figs. C-E Preanus length: about 90% SL Body: very elongate and slender; body depth about 8% SL; remains of preanal finfold still present after Air bladder: absent

8% SL; remains of preanal finfold still present after transformation; dorsal fin located anterior to pelvic fins; pelvic fins located slightly anterior to midbody; ventral adipose fin located before anal fin **Head:** small, relatively short and somewhat tilted down; snout pointed; mouth terminal and relatively large

Air bladder: absent Spination: none Pigmentation: very little pigment or none until transformation Length at flexion: unknown Length at transformation: 44.0 mm SL (transformation involves shrinkage from about 44.0 mm SL to about 27.0 mm SL)

Eye: oval, small **Gut:** straight, tube-like

PHOTOS

by J.M. Rodriguez





16.2 mm SL

STOMIIDAE



Literature: Fahay (2007), Gibbs (1984a), Richards (2006b), Sanzo (1931c) Illustrations' sources: A-E: L. Rodriguez (redrawn from Sanzo, 1931c) Ц

STOMIIDA

Meristic characters Habitat: oceanic, mesopelagic, from 30 to more than 1 900 m depth Myomeres: about 57-69 Distribution: worldwide in tropical Vertebrae: 57-59 and subtropical waters. Eastern Dorsal fin: 11-14 Atlantic, from South Africa to Anal fin: 13-19 Portugal, except the Gulf of Guinea (absent from the Mediterranean Sea) Spawning season: unknown YOLK-SAC LARVAE EGGS Undescribed Undescribed LARVAE Body: elongate and slender; dorsal and anal fins Spination: none

opposite, far posterior on body; pectoral fins present from early larvae on; well-developed dorsal finfold present, even after transformation

Echiostoma barbatum Lowe, 1843

Head: large and depressed (duck-billed shaped in early larvae)

Eye: slightly oval, becomes smaller and relatively round with development

Gut: voluminous, very elongate; terminal gut well developed and detached from body, over anal fin **Preanus length:** > 90% SL

Air bladder: none

Pigmentation: parallel rows of melanophores along dorsum of body (one per myomere) and on ventro-lateral sides of body (1-5 per myoseptum); a patch of melanophores on dorsal surface of terminal gut; melanophores on top of head, on isthmus and on edge of dorsal finfold (early larvae); scattered melanophores on lower lobe of caudal fin

Length at flexion: about 8.0 mm Length at transformation: unknown





16.0 mm SL



16.0 mm SL (dorsal view)

Threadfin dragonfish

STOMIIDAE

Echiostoma barbatum Lowe, 1843



Photostomias guernei Collett, 1889

Habitat: oceanic, bathypelagic, from 1 100 to 3 100 m depth Distribution: temperate and subtropical

waters of the Atlantic, Pacific and Indian oceans. Eastern Atlantic from 3°58'N to southern Portugal (absent from the Mediterranean Sea) Meristic characters Myomeres: about 52-58 Vertebrae: 52-58 Dorsal fin: 22-29 Anal fin: 25-33



Spawning season: unknown

EGGS

Undescribed

Undescribed

YOLK-SAC LARVAE

Figs. A-B

LARVAE

Body: elongate and slender; dorsal and anal fins opposite, far posterior on body; pectoral fins present from early larvae on; finfold well developed, present even in late larvae **Head:** elongate and depressed; snout pointed;

mouth oblique and large; lower jaw slightly protruding

Eye: small and oval, stands out above head **Gut:** foregut voluminous in early larvae; terminal gut tube-like, elongated and detached from body, below caudal fin

Preanus length: > 100% SL in late larvae (gut can extend beyond last vertebra) Air bladder: none Spination: none Pigmentation: opposite series of prominent spots along dorsal and ventral margins of body; tip of lower jaw pigmented; a prominent melanophore on pectoral-fin base; melanophores on trailing section of gut in late larvae Length at flexion: unknown Length at transformation: 30.0 mm SL

PHOTOS





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STOMIIDAE

Stomias boa (Risso, 1810)

Boa dragonfish

Habitat: oceanic, mesopelagic, to more than 1 000 m depth

Distribution: Atlantic Ocean, eastern Atlantic between 20°N and 65°N, and the western Mediterranean Sea **Spawning season:** probably all year round Meristic characters Myomeres: 78 Vertebrae: 78 Dorsal fin: 17-22 Anal fin: 18-22



EGGS

Undescribed

LARVAE

Body: very elongate and slender; dorsal and anal fins far posterior on body; pectoral fins present from early larvae on; pelvic fins absent **Head:** long with prominent jaws (duck-billed shape); lower jaw forms a sharply marked angle **Eye:** oval and small

Gut: tube-like, very elongate and slightly trailing; terminal section slightly detached from body **Preanus length:** about 87-90% SL **Air bladder:** absent

PHOTOS

Undescribed

Spination: none

YOLK-SAC LARVAE

Pigmentation: row of small melanophores over gut, between pectoral and caudal–fin base; some melanophores appear in dorsal–caudal region in larvae of about 6.0 mm SL, extending forward during development; dorsal and ventral series of melanophores disappear in late larvae **Length at flexion:** unknown **Length at transformation:** about 44.0 mm

by J.M. Rodriguez





8.7 mm SL



22.8 mm SL

Stomias boa (Risso, 1810)



STOMIIDAE

Benthalbella infans Zugmayer, 1911

Zugmayer's pearleye

Habitat: oceanic, meso- to bathypelagic,
at depths greater than 500 mMeristic charactersDistribution: in tropical and subtropical
regions of all oceans. Eastern Atlantic,Vertebrae: 55-58Dorsal fin: 8-10

mainly between 20°N and 30°N. Absent from the Mediterranean Sea Myomeres: 54-56 Vertebrae: 55-58 Dorsal fin: 8-10 Anal fin: 19-25 Adipose fin: present

Spawning season: all year round (western Atlantic Ocean)

EGGS

Undescribed

Undescribed

YOLK-SAC LARVAE

Figs. A-F

LARVAE

Body: elongate, moderately deep anteriorly, tapers gradually to caudal peduncle; translucent, almost transparent; dorsal-fin base short **Head:** large; snout large and pointed; jaws large and gurved, with backed teach on tangua

and curved, with hooked teeth on tongue **Eye:** oval, narrower in early larvae, with a ventral bulb of whitish tissue (similar to larvae of some myctophid species); lenses, in lateral position in early larvae, migrate dorsally in late larvae **Gut:** tubular, detached from body Preanus length: about 60% SL Air bladder: absent Spination: none Pigmentation: body unpigmented until transformation Length at flexion: about 15.0 mm Length at transformation: > 50.0 mm SL, abrupt

PHOTOS

by J.M. Rodriguez











10.7 mm SL

72



Illustrations' sources: A-F: L. Rodriguez (**A**, **B:** original; **C-F:** redrawn from Merrett *et al.*, 1973)

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Head: relatively small; snout pointed, elongate, with large, curved jaws; hooked teeth on tongue Eye: slender with pigmented mass of choroid tissue (like a sliver) ventrally; lens migrates to dorsal margin of eye during transformation Gut: tube-like, relatively elongate

advance) and another mid-ventral (immediately behind anal fin) melanophore in caudal peduncle; a slash-like bar of pigment at middle of caudal fin

Length at flexion: about 8.0-10.0 mm SL Length at transformation: gradual, begins at 16.0 mm, completed at 50.0 mm SL

PHOTOS







11.8 mm SL

Dana pearleye

SCOPELARCHIDAE

AULOPIFORMES

Scopelarchoides danae Johnson, 1974



<u>76</u>



8.2 mm SL



16.7 mm SL

Staring pearleye

Meristic characters Myomeres: 46-51

Scopelarchus guentheri Alcock 1896

Habitat: oceanic, mesopelagic,

SCOPELARCHIDAE

77

Scopelarchus guentheri Alcock 1896

from Johnson, 1974)



Habitat: neritic, demersal, between **Meristic characters** one and 400 m depth Myomeres: 58-59 Distribution: Atlantic Ocean and the Vertebrae: 56-58 Mediterranean Sea. Eastern Atlantic, Dorsal fin: 12 from Cape Verde to Morocco Anal fin: 10 Spawning season: unknown EGGS YOLK-SAC LARVAE Habitat: pelagic Hatch size: 4.0-4.5 mm SL **Body:** elongate and slender Shape: spherical Yolk sac: elongated, swollen anteriorly, extends to Chorion: sculptured with hexagonal structures; third ventral melanophore diam. 1.10-1.35 mm Anus: detached from yolk sac, reaches finfold Perivitelline space: absent border Yolk: unsegmented; unpigmented Preanus length: about 80% SL Oil globules: none Pigmentation: 5 large ventral melanophores, 4 Colour: transparent over gut and one in caudal region; melanophores arranged radially over caudal region of primordial fin

LARVAE

Body: elongate and slender (clupeid shaped) Head: relatively small; relatively small mouth; snout pointed Eye: oval and relatively large Gut: tube-like, elongate and straight Preanus length: about 80% SL Air bladder: absent Spination: none

Pigmentation: 5 (6 in late larvae) large melanophores along ventral side of trunk and tail, approximately equidistant, 4 (5 in late larvae) over gut and another in caudal region; melanophores arranged radially over caudal end of primordial fin in early larvae, and over caudal-fin rays in late larvae

PHOTOS

by J.M. Rodriguez



3.4 mm SL



Synodus saurus (Linnaeus, 1758)

Atlantic lizardfish

Length at flexion: unknown Length at transformation: unknown

Synodus saurus (Linnaeus, 1758)



Illustrations' sources: A-F: L. Rodríguez (**A**, **D:** redrawn from Tortonese, 1956c; **B**, **C:** redrawn from Alemany, 1997; **E**, **F:** redrawn from Tåning, 1918)

SYNODONTIDAE

ш

PARALEPIDIDA



PARALEPIDIDAE

AULOPIFORMES

Arctozenus risso (Bonaparte, 1840)



Illustrations' sources: A-E: Ege (1930)

Lestidiops jayakari (Boulenger, 1889) Habitat: oceanic, meso- to bathypelagic, between 50 and 2 000 m depth

Distribution: worldwide, in tropical to temperate waters (except the southeastern Pacific)

Spawning season: throughout the year

EGGS

Undescribed

LARVAE

PHOTOS

Body: elongate and slender, pelvic fins located well anterior to dorsal fin

Head: short and deep (duck-billed shape) in early larvae, becomes longer and pointed during development Eye: round and large

Gut: triangular and short in early larvae, becomes longer and tube-like throughout development Preanus length: increases during development from about 25% SL to about 60% SL Air bladder: absent

Spination: none

Undescribed

YOLK-SAC LARVAE

Pigmentation: two short rows of small melanophores (congener species, L. sphyrenoides, has 3) in postanal, ventral region; two rows of small melanophores in caudal region, one above and the other below urostyle since early larvae; up to 12 peritoneal patches of pigment form during development (absent in early larvae) Length at flexion: about 12.0-16.0 mm **Length at transformation:** > 40.0-45.0 mm

by J.M. Rodriguez



Meristic characters

Vertebrae: 76-85

Dorsal fin: 9-10

Anal fin: 28-33

Adipose fin: present

Myomeres: about 76-85









AULOPIFORMES

PARALEPIDIDAE

Lestidiops jayakari (Boulenger, 1889)



Literature: Ditty (2006d), Ege (1930), Fahay (2007), Froese and Pauly (2022), Olivar and Fortuno (1991), Post (1 Illustrations' sources: A, B: Alemany (1997); C-G: Ege (1930) **PARALEPIDIDAE**

PARALEPIDIDAE

Lestidiops sphyrenoides (Risso, 1820) Habitat: oceanic, epi- to mesopelagic, **Meristic characters** to 400 m depth Myomeres: 84-94 Distribution: eastern Atlantic Ocean, Vertebrae: 84-94 from Mauritania to France, and the Dorsal fin: 9-11 Mediterranean Sea Anal fin: 28-31 Spawning season: throughout the Adipose fin: present year EGGS YOLK-SAC LARVAE Undescribed Undescribed

LARVAE

Body: elongate and slender; dorsal fin located at level of anus; anal fin located at end of tail; pelvic fins slightly ahead of anus; remains of primordial fin in very late larvae

Head: short and deep (duck-billed shape) in early larvae, becomes longer and pointed during development

Eye: round and large

PHOTOS

Gut: triangular and short in early larvae, becomes longer and tube-like during development **Preanus length:** increases during development from about 23% SL to about 59% SL **Air bladder:** absent

Spination: none

Pigmentation: three short rows of small melanophores (congener species, *L. jayakari*, has two) in postanal, ventral region; two rows of small melanophores in caudal region, one above and other below urostyle since early larvae; up to 12 peritoneal patches of pigment form during development (2 in larvae of 7.5 mm); melanophores close to nostril; lower jaw pigmented; melanophores on top of head in late larvae

Length at flexion: unknown Length at transformation: unknown

by J.M. Rodriguez



17.0 mm SL



AULOPIFORMES

Lestidiops sphyrenoides (Risso, 1820)



Illustrations' sources: A-G: Ege (1930)



EGGS

Undescribed

LARVAE

Meristic characters Myomeres: 91-98 Vertebrae: 91-93 Dorsal fin: 9 Anal fin: 40-44 YOLK-SAC LARVAE Undescribed Body: elongate and slender; pelvic fins form well Air bladder: absent anterior to dorsal fin Spination: none Head: fairly large and deep (duck-billed shaped) in finfold, from early larvae on; in addition, late

small larvae, becomes longer, with pointed snout, throughout development

Eye: round and large

Gut: triangular and short in early larvae, becomes longer and tube-like throughout development **Preanus length:** increases during development, anus reaches its final position in larvae of about 20.0 mm SL Pigmentation: numerous melanophores on caudal larvae have two parallel rows of melanophores in caudal region, one above and the other below urostyle; up to 8 peritoneal patches of pigment form during development (one in early larvae) Length at flexion: about 13.0-17.0 mm SL Length at transformation: from about 40.0 mm SL

PHOTOS

by S. Isari





Detail of caudal region

PARALEPIDIDAE

PARALEPIDIDAE





88

Paralepis coregonoides Risso, 1820

Habitat: oceanic, mesopelagic, between 50 and 1 000 m depth Distribution: North Atlantic Ocean, and the Mediterranean Sea; eastern Atlantic Ocean, from Morocco to 65° N Spawning season: March to September Meristic characters Myomeres: 70-74 Vertebrae: 70 Dorsal fin: 9-11 Anal fin: 22-26 Adipose fin: present



EGGS

Undescribed

Undescribed

YOLK-SAC LARVAE

Figs. A-G

LARVAE

Body: elongate and slender; pectoral fins present from early larvae on; pelvic fins form under dorsal fin at about midbody

Head: small and deep; snout relatively short (duckbilled shaped) in early larvae, becomes longer and pointed during development

Eye: large and oval in early larvae, becomes round in late larvae

Gut: very short

Preanus length: increases with development from about 30% TL to about 36% TL **Air bladder:** absent

Spination: none

Pigmentation: postanal region unpigmented in larvae up to about 5.0 mm; larvae > 5.0 mm show a deep internal melanophore above notochord, close to its end and a peritoneal patch; a long internal series of melanophores above and a short one below notochord in late larvae; up to 3 peritoneal patches appear during development (before transformation)

Length at flexion: about 10.0-15.0 mm (no SL) Length at transformation: >20.0-25.0 mm (no SL)



by J.M. Rodriguez









15.0 mm SL

PARALEPIDIDAE

Paralepis coregonoides Risso, 1820





6.5 mm SL



7.4 mm SL

EVERMANNELLIDAE

Evermannella balbo (Risso, 1820)



91

Benthosema glaciale (Reinhardt, 1837)

Glacier lantern fish - Lanterne glaciaire

Habitat: mesopelagic, between 0 and 1 400 m depth

Distribution: eastern Atlantic Ocean from Guinea (Mauritanian Upwelling Region) to Norway, and the Mediterranean Sea.

Meristic characters Myomeres: 34-36 Vertebrae: 34-36 Dorsal fin: 12-14 Anal fin: 17-19 Adipose fin: present



Spawning season: spring and summer

EGGS

Undescribed

Undescribed

YOLK-SAC LARVAE

LARVAE

Body: moderately elongate; a gap between anus and anal fin in early larvae, closes in larvae of about 8.0 mm Head: moderate, with a pointy snout

Eye: oval, with a mass of choroid tissue ventrally Gut: curved downward in early larvae Preanus length: about 50% SL Air bladder: absent Spination: none

Pigmentation: melanophores at posterior edge of opercle, at tip of snout and lower jaw; 3 ventral melanophores: on cleithral symphysis, lateral side of gut, and on terminal gut, respectively; several melanophores on postanal, ventral region, decreasing in number to a single one over mid anal fin in late larvae; abdominal region and pectoralfin rays pigmented; premature preopercular photophore

Length at flexion: 5.0-7.0 mm Length at transformation: about 11.0 mm



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MYCTOPHIDAE

Benthosema glaciale (Reinhardt, 1837)



<u>MYCTOPHIDAE</u>

Benthosema suborbitale (Gilbert, 1913)

Smallfin lanternfish

Habitat: oceanic, mesopelagic, between 10 and 750 m depth Distribution: Atlantic Ocean. Eastern

Atlantic, from Morocco to South Africa. Absent from the Mediterranean Sea Myomeres: 33-35 Vertebrae: 33-35 Dorsal fin: 11-14 Anal fin: 16-19 Adipose fin: present

Meristic characters

Spawning season: peaks in spring and summer (off Hawaii, USA)

EGGS

Undescribed

Undescribed

Figs. A-E

LARVAE

Body: initially elongate, becomes short and deep; gap between anus and anal-fin origin closes between 9.0 and 10.0 mm SL

Head: moderate with slightly pointed snout **Eye:** elliptical with a lunate mass of choroid tissue ventrally

Gut: short, bulbous anteriorly with narrow posterior section; large terminal gut, acutely deflected ventrally

Preanus length: less than 50% SL **Air bladder:** absent

PHOTOS

Spination: none

Pigmentation: most of body unpigmented; several melanophores on ventral surface of head; 2 melanophores anterior to lower and upper portion of pectoral-fin base **Length at flexion:** 5.2-6.5 mm

Length at transformation: about 10.0 mm SL

YOLK-SAC LARVAE

Note: larvae similar to those of *Electrona risso*, which have a longer preanus length, pigment on pectoral-fin rays, and lack blotches at pectoral-fin base



MYCTOPHIDAE
Benthosema suborbitale (Gilbert, 1913)



Illustrations' sources: A-E: L. Rodríguez (redrawn from Moser and Ahlstrom, 1996b)

MYCTOPHIDAE

Ceratoscopelus maderensis (Lowe, 1839)

Myomeres: 37

Vertebrae: 37

Dorsal fin: 13-15

Anal fin: 13-15

Habitat: oceanic, mesopelagic, between 100 and 1 000 m depth Distribution: North Atlantic Ocean and the Mediterranean Sea. Eastern Atlantic, from 20°N to 50°N

Spawning season: spring and summer

S (Lowe, 1839) Madeira lantern fish – Lanterne de Madère Meristic characters

YOLK-SAC LARVAE



EGGS

Undescribed

LARVAE

Undescribed

Figs. A-F

by J.M. Rodriguez

Body: elongate, moderately slender

Head: relatively small; snout pointed, short in early larvae, lengthens during development; mouth large reaches posterior border of eye in late larvae Eye: round and large Gut: tube-like, moderately slender and gently

curved downward at about its midway **Preanus length:** increases from > 50% SL in early larvae to > 60% SL in late larvae **Air bladder:** absent **Spination:** no spination **Pigmentation:** single melanophores on lateral side of gut and on terminal gut; a continuous line of melanophores between anus and caudal fin decreasing in number during development (3 in late larvae); 3-4 melanophores on dorsal side of caudal peduncle (not present in early larvae); no melanophores at cleithral symphysis **Length at flexion:** about 6.0 mm **Length at transformation:** about 16.0 mm

PHOTOS









7.9 mm SL



98

Dasyscopelus asper (Richardson, 1845)

Habitat: oceanic, mesopelagic, between 0 and 750 m depth Distribution: Atlantic, Pacific and Indian oceans. Eastern Atlantic, from South Africa to Mauritania Spawning season: unknown

Meristic characters Myomeres: 35-38 Vertebrae: 35-38 Dorsal fin: 12-14 Anal fin: 17-19 Adipose fin: present

Prickly lanternfish



EGGS

Undescribed

Undescribed

Figs. A-E

by S. Isari

LARVAE

Body: elongate in early larvae, becomes deeper and stout with development

Head: large and broad; snout pointed in early larvae, rounded in late larvae; snout and forehead concave (duck-billed shaped) in early larvae, becomes convex with development; mouth large and oblique; jaws with prominent teeth since early larvae

Eye: oval, with a ventral, unpigmented mass of choroid tissue

Gut: bulky anteriorly, with thinner terminal section; anus protruding

Preanus length: increases from about 40-48% SL in early larvae, to 56-64% SL in late larvae

Air bladder: absent Spination: none

YOLK-SAC LARVAE

Spination: none

Pigmentation: early larvae, one melanophore at tip of upper and lower jaws, on forehead (between eyes), at base of pectoral fin, and over terminal gut; pigmentation increases with development, adding a melanophore at dorsal-fin origin and another at end of adipose fin; a few internal melanophores on epaxial myosepta; a large melanophore at base of caudal fin

Length at flexion: 4.5-6.0 mm SL Length at transformation: 11.0-13.0 mm SL

PHOTOS







7.0 mm SL

99

MYCTOPHIDAE

MYCTOPHIFORMES

Dasyscopelus asper (Richardson, 1845)



Literature: Fahay (2007), Froese and Pauly (2022), Hulley (1990), Moser and Ahlstrom (1974, 1996b), Moser and Watson (2001, 2006)

Illustrations' sources: A-E: L. Rodríguez (A, B, D, E: redrawn from Ozawa, 1986; C: redrawn from Moser and Ahlstrom, 1974)

Dasyscopelus selenops (Tåning, 1928)

MYCTOPHIDAE

Habitat: oceanic, mesopelagic, between 40 and 450 m depth Distribution: Atlantic, Pacific and Indian oceans. Eastern Atlantic, from South Africa to Morocco (absent from

south Africa to Morocco (absent from the Mauritanian upwelling region) Spawning season: spring (off Hawaii, USA)

Myomeres: 34-35 Vertebrae: 34-35 Dorsal fin: 12-14 Anal fin: 17-19 Adipose fin: present

Meristic characters

Wisner's lantern fish

YOLK-SAC LARVAE

EGGS

Undescribed

Undescribed

Figs. A-D

by J.M. Rodriguez

LARVAE

Body: relatively slender in early larvae, soon becomes stout and deep, especially through head and trunk

Head: relatively large, broad and deep; snout pointed; mouth large and moderately oblique **Eye:** elliptical, with a large mass of choroid tissue ventrally

Gut: triangular, narrow in terminal section; anus protruding

Preanus length: 50-53% SL in early larvae, increases to 61-64% SL in postflexion stage **Air bladder:** absent

Spination: none

PHOTOS

Pigmentation: tail unpigmented; upper and lower jaw-tips pigmented; midline of pigment anterior to forebrain in early larvae, becomes a pair of spots lateral to mid-brain with development; a single group of melanophores over opercular margin; few spots over mid-gut and on pectoral-fin base; scattered melanophores on nostril; a stellate melanophore ventrally on trunk; pectoral-fin rays with scattered pigment; choroid tissue mass pigmented

Length at flexion: 4.5-6.0 mm SL Length at transformation: 10.0-13.0 mm SL



3.9 mm SL









Dasyscopelus selenops (Tåning, 1928)



Illustrations' sources: A-D: L. Rodríguez (A, B, D: redrawn from Moser and Watson, 2001; C: redrawn from Moser and Ahlstrom, 1974)

MYCTOPHIDAE



Diaphus holti Tåning, 1918



MYCTOPHIDAE

Distribution: Atlantic, western Pacific and western Indian oceans, and the western Mediterranean Sea. Eastern Atlantic, from South Africa to the British Isles

Meristic characters Myomeres: 35 Vertebrae: 35 Dorsal fin: 14-16 Anal fin: 14-16 Adipose fin: present

Spawning season: unknown

EGGS

Undescribed

LARVAE

PHOTOS

Body: relatively deep, similar to *D. rafinesquii* Head: large and somewhat bulbous; snout short Eye: round and moderately large

Gut: thicker anteriorly, more narrow posteriorly; anus slightly protruding

Preanus length: increases from about 50% SL, in early larvae, to about 60% SL, in late larvae Air bladder: absent Spination: none

Pigmentation: a single, large ventral melanophore, at posterior margin of anal fin (at about mid-tail); a pair of melanophores over anus in early larvae; melanophores over lateral sides of gut and snout (see photo) appear in late larvae; rest of body unpigmented; no melanophore on ventral side of gut, posterior to cleithral symphysis, in early larvae

YOLK-SAC LARVAE

Undescribed

Length at flexion: about 5.0 mm SL Length at transformation: about 11.0 mm SL

by J.M. Rodriguez

Bluntnose lanternfish





8.1 mm SL

Diaphus metopoclampus (Cocco, 1829)



MYCTOPHIFORMES

Illustrations' sources: A-E: L. Rodríguez (redrawn from Sparta, 1952)



PHOTOS

by J.M. Rodriguez















Diogenichthys atlanticus (Taning, 1928)

Habitat: oceanic, mesopelagic, between 18 and 1 250 m depth Distribution: Atlantic, Pacific and Indian oceans and the western Mediterranean Sea. Eastern Atlantic Ocean, from 48°S to 50°N

Meristic characters Myomeres: 31-35 Vertebrae: 31-35 Dorsal fin: 10-12 Anal fin: 14-18 Adipose fin: present

Spawning season: unknown

EGGS

Undescribed

LARVAE

Body: elongate and slender, becoming somewhat compressed with development

Head: large, rounded in early and late stages of development, slightly concave at middle stages; snout pointed, becoming round and shorter in late larvae; a symphyseal barbel forms at about 5.0 mm, and disappears before transformation

Eye: elliptical, becoming rounded in late larvae; choroid tissue absent

Gut: slightly sigmoid, thicker anteriorly; terminal gut makes almost a right angle with body in early larvae; anus moderately protruding

Preanus length: about 50-60% SL

PHOTOS

Air bladder: absent

YOLK-SAC LARVAE

Spination: none

Undescribed

Pigmentation: in early larvae, a ventro-lateral pair of melanophores just posterior to cleitrhum; dorsolateral pairs of melanophores on terminal gut; two lateral pairs on mid-gut; about 3 melanophores in ventral series, posterior to anus; in late larvae, number of melanophores on lateral side of gut and ventral series, posterior to anus, increases; melanophores on jaw barbel appear; a large spot develops at base of caudal fin

Length at flexion: 6.0-9.0 mm SL Length at transformation: 13.5-15.5 mm SL

by J.M. Rodriguez

Atlantic lanternfish

Figs. A-E



5.4 mm SL







Not sized

Diogenichthys atlanticus (Tåning, 1928)



Illustrations' sources: A-E: L. Rodríguez (redrawn from Moser and Ahlstrom, 1970)

MYCTOPHIDAE

110

MYCTOPHIDAE

Gonichthys cocco Cocco, 1829 Cocco's lanternfish Habitat: oceanic, mesopelagic, **Meristic characters** between 0 and 1 000 m depth Myomeres: 40-41 Distribution: Atlantic Ocean and the Vertebrae: 40-41 Mediterranean Sea. Eastern Atlantic, Dorsal fin: 20-23 s d from South Africa to Morocco Anal fin: 13-16 Spawning season: May to June Adipose fin: present (Mediterranean Sea) YOLK-SAC LARVAE EGGS Undescribed Undescribed LARVAE Body: deep, highly laterally compressed; large

Body: deep, highly laterally compressed; large anterior finfold present until transformation **Head:** large; snout large and pointed in early larvae, becoming blunt in later larvae; jaws large, slightly oblique **Eye:** oblique, with a large conical mass of choroid

Gut: strongly sigmoid; anus protruding **Preanus length:** increases from 50 to 60% SL throughout development

Air bladder: absent Spination: none **Pigmentation:** 2 large dorsal and 2 opposing, large ventral melanophores behind anus; an additional melanophore on dorsum of body, anterior to dorsal fin, appearing after flexion; a series of melanophores along margins of upper and lower jaw; a large spot at caudal-fin base; later larvae with melanophores above and on ventral surface of gut, on lateral sides of head, and on snout; pectoral-fin rays pigmented **Length at flexion:** 5.0-7.5 mm SL **Length at transformation:** > 12.0 mm SL

PHOTOS





4.5 mm SL

6.1 mm SL

by J.M. Rodriguez



9.3 mm SL

MYCTOPHIFORMES

Gonichthys cocco Cocco, 1829



2006), Tåning (1918)











Hygophum benoiti (Cocco, 1838)



Illustrations' sources: A-D: L. Rodríguez (redrawn from Olivar and Palomera, 1994); E: Tåning (1918)

	114				
	Hygophum hygomii (Lütken, 1892)		Bermuda lanternfish - Lanterne des Bermudes		
AYCTOPHIDAE	Habitat: oceanic, mesopelagic, between 0 and 700 m depth Distribution: Atlantic Ocean, and the Mediterranean Sea. Eastern Atlantic, from South Africa to Portugal Spawning season: peaks in summer and autumn (Mediterranean Sea)	Meristic character Myomeres: 36-38 Vertebrae: 36-38 Dorsal fin: 12-14 Anal fin: 19-21 Adipose fin: prese	's ent		
	EGGS		YOLK-SAC LARVAE		
	Undescribed		Undescribed		
	LARVAE Figs. A-E				
	Body: moderately slender, slightly increasing in depth with development; remains of prominent finfold, between head and dorsal fin, up to pre- transformation stage Head: relatively large; snout pointed in early larvae, becoming rounded with development Eye: moderately elliptical with a prominent conical mass of choroid tissue ventrally Gut: thick, slightly curved, with visible transverse folds; anus situated at anterior margin of anal fin, protruding Preanus length: < 60% SL Air bladder: absent		Spination: none Pigmentation: a ventral series of melanophores at cleithral symphysis and on isthmus; melanophores rarely found at tip of snout, lower jaw, and on caudal-fin rays; 1-2 melanophores on lateral sides of gut; a prominent melanophore over anus; postanal ventral melanophores reduced to one (or none) over middle of anal-fin base; finfold borders unpigmented; no melanophores over ventral rays of caudal fin; pectoral-fin rays pigmented Length at flexion: 6.0-7.0 mm SL Length at transformation: 13.0-14.5 mm SL		
	Larvae of the two species of the <i>Hygophum</i> genus, <i>H. benoiti</i> and <i>H. hygomii</i> , are very similar and their differentiation, based mainly on their pigmentation patterns, is problematic. PHOTOS by J.M. Rodriguez				
	4.9 mm SL				
				A	



9.3 mm SL

6.6 mm SL

Hygophum hygomii (Lütken, 1892)



Illustrations' sources: A-C: L. Rodríguez (redrawn from Olivar and Palomera, 1994); D, E: Tåning (1918)



Hygophum macrochir (Günther, 1864)



Hygophum reinhardtii (Lütken, 1892) **Reinhardt's lanternfish** Habitat: oceanic, mesopelagic, **Meristic characters** between 10 and 900 m depth Myomeres: 38-40 Distribution: Atlantic, Pacific and Vertebrae: 38-40 southern Indian oceans. Eastern Dorsal fin: 13-15 Atlantic from Namibia to Morocco Anal fin: 21-25 (absent from the Mediterranean Sea) Adipose fin: present Spawning season: unknown YOLK-SAC LARVAE EGGS Undescribed Undescribed LARVAE **Body:** elongate and thin, more elongate than larvae Pigmentation: 2 ventral melanophores on of congener species; body depth 10% SL, deepens isthmus; ventral melanophores just posterior to before transformation cleitrhum, along mid-section of gut and on anus; Head: flattened in early larvae; snout pointed in 2 postanal melanophores at ventral mid-tail; 1-2 early larvae, becomes rounded before transformation melanophores on dorsum of tail end (disappear in Eye: strongly elliptical on short stalks; prominent late larvae); a melanophore at base of caudal fin; mass of ventral choroid tissue present in late larvae, melanophores increase in number Gut: elongate and thin, nearly straight along lateral sides of gut and ventral tail, and these **Preanus length:** increases from 55% to 65% latter migrate onto miosepta throughout development Length at flexion: 8.8-10.3 mm SL Air bladder: absent Length at transformation: 14.9-16.4 mm SL Spination: none PHOTOS by J.M. Rodriguez 4.8 mm SL 6.0 mm SL 10.1 mm SL

12.4 mm SL

Hygophum reinhardtii (Lütken, 1892)



Literature: Fahay (2007), Froese and Pauly (2022), Hulley (1984), Moser and Ahlstrom (1970, 1974), Moser and Watson (2001, 2006)

MYCTOPHIDAE



6.3 mm SL



Hygophum taaningi Becker, 1965

and Ahlstrom, 1974)





peduncle; body becomes shallower throughout development Head: large; snout pointed; jaws large; teeth

protruding in upper jaw from early larvae on Eye: slightly elliptical

Gut: triangular and thick; anus protruding Preanus length: increases from about 60% SL, in early larvae, to about 70% SL, in later larvae

PHOTOS

Air bladder: present

Spination: two series of large preopercular spines, one on edge and one on lateral ridge Pigmentation: a pair of melanophores on head and another pair over anus; upper and lower jaw tips pigmented; a large melanophore at dorsal-fin end;

air bladder pigmented Length at flexion: 5.0-6.0 mm SL

Length at transformation: > 15.0 mm SL

by J.M. Rodriguez



Not sized



Not sized (dorsal view)



7.3 mm SL

Lampanyctus ater Tåning, 1928



Illustrations' sources: A-E: L. Rodríguez (redrawn from Moser and Watson, 2001)

Lampanyctus crocodilus (Risso, 1810)

Jewel lanternfish – Lanterne crocodile

Habitat: oceanic, mesopelagic, between 0 and 1 000 m depth Distribution: North Atlantic Ocean and the Mediterranean Sea. Eastern Atlantic, from the Mauritanian

and the Mediterranean Sea. Eastern Atlantic, from the Mauritanian upwelling region to 65°N **Spawning season:** March to August Meristic characters Myomeres: 35-36 Vertebrae: 35-36 Dorsal fin: 13-15 Anal fin: 16-18 Adipose fin: present

EGGS

Undescribed

Undescribed

YOLK-SAC LARVAE

Figs. A-F

LARVAE

Body: initially elongate, soon deepens, especially at level of pectoral region

Head: large with pointed snout; well–developed protruding teeth in upper jaw from early larvae on **Eye:** round and moderately large

Gut: thicker anteriorly; anus makes almost a right angle with body in early larvae; anus slightly protruding

Preanus length: increases from about 25% SL in early larvae, to about 60% SL in late larvae **Air bladder:** present in late larvae **Spination:** none **Pigmentation:** single large melanophore on top of head (absent in early larvae); a single melanophore on body dorsum, between dorsal and adipose fins, from flexion stage on; single melanophores at tip of lower jaw and over anus; peritoneal pigment develops in late larvae; pectoral–fin base and rays pigmented; melanophores embedded in myosepta anteriorly on trunk in late larvae **Length at flexion:** about 6.0-7.0 mm SL **Length at transformation:** about 20.0 mm SL

PHOTOS





5.5 mm SL





6.5 mm SL (dorsal view)

by J.M. Rodriguez

Lampanyctus crocodilus (Risso, 1810)



MYCTOPHIDAE

Lampanyctus pusillus (Risso, 1810) Habitat: oceanic, mesopelagic, **Meristic characters** between 25 and 1 000 m depth Myomeres: 30-32 Distribution: Atlantic Ocean and the Vertebrae: 30-32 Mediterranean Sea. Eastern Atlantic, Dorsal fin: 11-13 from the Mauritanian upwelling Anal fin: 13-16 region to 65°N Adipose fin: present Spawning season: summer and autumn (Mediterranean Sea) YOLK-SAC LARVAE EGGS

Undescribed

LARVAE

PHOTOS

Body: elongate in early larvae, becomes short and plump throughout development

Head: large; mouth large, reaches posterior edge of eye in late larvae; snout blunt and rounded; welldeveloped teeth in upper jaw from early larvae on Eye: round and large

Gut: thick and bulbous; terminal section of gut protruding, forms almost a right angle with body Preanus length: increases from about 30% SL, in early larvae, to about 65% SL, in late larvae Air bladder: present

Spination: none

Undescribed

Pigmentation: 1-3 melanophores from snout to top of head; melanophores at tip of lower jaw, over opercle, pectoral-fin base, anterior of gut, and over terminal gut; internal pigment on air bladder; ventral melanophores on abdominal region; late larvae, add paired series of melanophores along dorsum and a row of melanophores along lateral midline

Length at flexion: about 4.0-6.0 mm SL Length at transformation: about 12.0 mm SL







3.0 mm SL

3.4 mm SL



4.3 mm SL

Pigmy lanternfish

MYCTOPHIDAE

127

Lampanyctus pusillus (Risso, 1810)



Illustrations' sources: A-C: Alemany (1997); D-F: Tåning (1918)

Lepidophanes gaussi (Brauer, 1906)

Gauss' lanternfish

Habitat: oceanic, mesopelagic, between 100 and 1 000 m depth

Distribution: Atlantic Ocean (absent from the Mediterranean Sea). Eastern Atlantic, between 13°S and 27°S and between 18°N and 42°N Meristic characters Myomeres: 35-36 Vertebrae: 35-36 Dorsal fin: 12-15 Anal fin: 13-15 Adipose fin: present

Spawning season: year-round (western Atlantic Ocean)

EGGS

Undescribed

Undescribed

YOLK-SAC LARVAE

Figs. A-D

LARVAE

PHOTOS

Body: elongate and slender; dorsal and ventral margins of body almost parallel Head: moderately small; snout moderately pointed, concave in early larvae Eye: round and large Gut: tube-like, moderately slender and almost of constant diameter along all its length Preanus length: increases from > 59% SL in early larvae to 64% SL in postflexion stage Air bladder: prominent Spination: none **Pigmentation:** 6 postanal ventral melanophores in early larvae, reduced to 2 large spots in late larvae, one over end of anal fin, the other close to caudal peduncle; one large dorsal melanophore in caudal region in early larvae, 2 in late larvae: one under adipose fin, the other close to caudal peduncle, opposite to posterior ventral ones; dorsum of air bladder pigmented; a dorso-lateral melanophore on each side of terminal gut; a melanophore in occipital region in late larvae **Length at flexion:** 5.3-5.6 mm SL

Length at transformation: about 13.0 mm SL

by J.M. Rodriguez



9.8 mm SL



130

MYCTOPHIDAE

Lobianchia dofleini (Zugmayer, 1911) Habitat: oceanic, mesopelagic, **Meristic characters** between 40 and 750 m depth Myomeres: 33-35 Distribution: eastern Atlantic Ocean Vertebrae: 33-35 between 40°S and 50°N, and the Dorsal fin: 15-17 Mediterranean Sea **Anal fin:** 13-15 Spawning season: throughout the Adipose fin: present year with a peak between February and June (Mediterranean Sea) YOLK-SAC LARVAE EGGS Undescribed Undescribed

LARVAE

Body: elongate in early larvae, becomes short and thick; pectoral fins large with elongate upper rays Head: large and broad; snout relatively long and pointed; forehead slightly concave in early larvae Eye: small and round in early larvae, becomes slightly elliptical with development Gut: fairly curved and thick; terminal gut prominent Preanus length: increases from about 40% SL, in early larvae, to about 60% SL, in late larvae Spination: none

Pigmentation: melanophores over lateral and ventral side of gut, on ventral midline anterior to cleithral symphysis, and along base of anal fin; a melanophore on each side of terminal gut; base and rays of pectoral fins pigmented; melanophores appear on dorsum of body in late larvae; air bladder pigmented

Length at flexion: 5.0-6.0 mm SL **Length at transformation:** about 10.0-11.0 mm SL



MYCTOPHIFORMES

Air bladder: present

Dofleini's lanternfish - Lanterne de Dofleini




Lobiancha gemellarii (Cocco, 1838) Habitat: oceanic, mesopelagic, between 25 and 500 m depth Myome

between 25 and 500 m depth Distribution: Atlantic Ocean and the Mediterranean Sea. Eastern Atlantic, from Mauritania to Ireland Spawning season: peaks in late autumn and winter (Bermuda) Meristic characters Myomeres: 34-35 Vertebrae: 34-35 Dorsal fin: 16-18 Anal fin: 13-15 Adipose fin: present

Gemellar's lanternfish



EGGS

Undescribed

Undescribed

Figs. A-E

LARVAE

PHOTOS

Body: deep and stout, mainly in anterior part (especially in late larvae); upper pectoral-fin rays strongly developed

Head: deep and broad; snout pointed in early larvae, becomes rounded with development; prominent teeth since early larvae

Eye: large and slightly oval, with lunate mass of choroid tissue ventrally, becomes rounded with development

Gut: thick anteriorly, with narrow terminal section **Preanus length:** increases from about 52% SL, in early larvae, to about 60% SL in late larvae

Air bladder: present

YOLK-SAC LARVAE

Spination: none

Pigmentation: early larvae, melanophores anterior to pectoral fin and on pectoral-fin base, on foregut, and a pair on terminal gut; pectoral-fin rays with scattered pigment; a melanophore at end of analfin base; air bladder pigmented; late larvae add pigment on anterior region of body, and a large melanophore at caudal-fin base (2 in later larval stages)

Length at flexion: 5.0-6.0 mm SL Length at transformation: 12.0-14.0 mm SL

8.1 mm SL

by J.M. Rodriguez



MYCTOPHIFORMES

MYCTOPHIDAE

MYCTOPHIDAE

Lobiancha gemellarii (Cocco, 1838)



Illustrations' sources: A-E: L. Rodríguez (**A**, **B**, **E:** redrawn from Moser and Ahlstrom, 1996b; **C:** redrawn from Moser and Ahlstrom, 1974; **D:** redrawn from Tåning, 1918)

MYCTOPHIDAE



section Preanus length: increases from 48-56% SL in early larvae, to 60-67% SL in late larvae Air bladder: absent

Spination: none

PHOTOS

melanophores on dorsal margin, near adipose fin, and on ventral margin, over middle of anal-fin base; large melanophores at caudal-fin base in late larvae

Length at flexion: 4.2-6.0 mm SL Length at transformation: 11.5-13.0 mm SL

by J.M. Rodriguez



6.4 mm SL







Myctophum nitidulum Garman, 1899 **Pearlyspotted lanternfish** MYCTOPHIDAE Habitat: oceanic, mesopelagic, **Meristic characters** between 0 and 950 m depth Myomeres: 36-39 Distribution: Atlantic, Pacific, and Vertebrae: 36-39 Indian oceans. Eastern Atlantic, from Dorsal fin: 12-14 South Africa to Morocco Anal fin: 18-21 Spawning season: peaks in early Adipose fin: present spring YOLK-SAC LARVAE EGGS Undescribed Undescribed

LARVAE

Body: stout, deepest anteriorly; pectoral fin precocious, with large base

Head: very large and wide; snout pointed, mouth large and moderately oblique

Eye: slightly elliptical, on short stalk, with a fairly prominent, conical mass of choroid tissue ventrally **Gut:** large, triangular, with prominent terminal section

Preanus length: increases from about 50% SL in early larvae to 60% SL in late larvae **Air bladder:** absent

Spination: none

Pigmentation: early larvae, melanophores at tip of lower jaw; one at midway along postanal, ventral region; 2 pairs on anterior surface of gut and a dorso-lateral pair on terminal section of gut; late larvae, one melanophore near nostril, one behind eye, one at angle of opercle; a series of melanophores on isthmus; rows of melanophores on upper and lower jaws; 4 opposing melanophores on dorsal and ventral margins of tail; two parallel lines of melanophores on anterior, ventral surface of gut; melanophores on caudal-fin base in transforming larvae **Length at flexion:** 6.5-7.0 mm SL **Length at transformation:** about 11.0 mm SL

PHOTOS



4.2 mm SL

5.6 mm SL

by S. Isari





9.4 mm SL

Myctophum nitidulum Garman, 1899



Literature: Fahay (2007), Froese and Pauly (2022), Hulley (1984), Moser and Ahlstrom (1970, 1974, 1996b), Moser and Watson (2001, 2006), Olivar and Fortuño (1991)

Illustrations' sources: A-F: L. Rodríguez (A-C: redrawn from Moser and Ahlstrom, 1996b; D-F: redrawn from Moser and Watson, 2001)

MYCTOPHIDAE

MYCTOPHIDAE

Myctophum punctatum Rafinesque, 1810

Habitat: oceanic, mesopelagic, between 40 and 1 000 m depth

Distribution: eastern Atlantic Ocean, from the Mauritanian upwelling region to 65°N, and the Mediterranean Sea Myomeres: 40 Vertebrae: 40 Dorsal fin: 13 Anal fin: 19-21 Adipose fin: present

Meristic characters

Spawning season: late winter to early spring

EGGS

Undescribed

Undescribed

Figs. A-E

LARVAE

Body: elongate in early larvae, becoming slightly deeper during development; pectoral fins large
Head: large and flat; forehead slightly concave in early larvae (duck-billed shaped); snout pointed and broad
Eye: elliptical, stalked in early larvae, with tapered choroid mass ventrally
Gut: elongate (tube-like) in early larvae
Preanus length: about 50-60% SL

Air bladder: absent

Spination: none

PHOTOS

Pigmentation: melanophores on edges of both jaws and on upper part of opercle; several ventral, postanal melanophores; ventral series of melanophores from head to anus; single, well developed melanophores on dorsal and ventral tail end; melanophore on caudal-fin base; posterior rays of dorsal and anal fins may be pigmented; rays and base of pectoral fins pigmented; dorsal melanophores on tail in late larvae **Length at flexion:** about 7.0 mm **Length at transformation:** about 21.0-22.0 mm

YOLK-SAC LARVAE

by J.M. Rodriguez



4.8 mm SL





8.6 mm SL



6.7 mm SL (dorsal view)



Spotted lanternfish





MYCTOPHIDAE

Notolychnus valdiviae (Brauer, 1904)

Habitat: oceanic, mesopelagic, between 25 and 700 m depth

Distribution: worldwide in tropical, subtropical, and temperate waters. Eastern Atlantic, from South Africa to the British Isles (absent from the Mediterranean Sea)

Spawning season: peaks in early summer

EGGS

Undescribed

YOLK-SAC LARVAE

Undescribed

LARVAE

Body: elongate and slender, with long caudal peduncle

Head: moderately large; snout pointed, mainly in early larvae, becomes somewhat elongated with development

Eye: relatively narrow, becomes irregularly oval; a crescent mass of choroid-like tissue appears on dorsal surface of eye at about 4.0 mm SL and on ventral surface at about 6.0 mm SL

Gut: short, thick anteriorly, tapers gradually with slow curvature; anus slightly protruding Preanus length: increases from 35% SL in early larvae to 50% SL in late larvae

Air bladder: prominent Spination: none

Pigmentation: early larvae, a melanophore on lateral sides of mid-gut; a pair of melanophores over terminal gut; postanal, ventral line of 1-4 melanophores; gas bladder pigmented; late larvae, up to 3 melanophores laterally on gut; usually 3-4 (range from 2 to 7) postanal, ventral melanophores, displaced to either side of anal-fin base; a melanophore at mid-base of caudal fin; head and dorsum of body unpigmented Length at flexion: 4.4-6.2 mm SL Length at transformation: 10.0-10.8 mm SL

PHOTOS

by J.M. Rodriguez



Topside lampfish

Meristic characters Myomeres: 27-31 Vertebrae: 27-31 Dorsal fin: 10-12 **Anal fin: 12-15** Adipose fin: present



Notolychnus valdiviae (Brauer, 1904)



Illustrations' sources: A-E: L. Rodríguez (redrawn from Moser and Ahlstrom, 1996b)

Notoscopelus resplendens (Richardson, 1845)

Patchwork lanternfish

Habitat: oceanic, meso-bathypelagic, between 300 and 2 100 m depth Distribution: eastern Atlantic Ocean (absent from the Mediterranean Sea). Spawning season: unknown Meristic characters Myomeres: 35-38 Vertebrae: 35-38 Dorsal fin: 21-24 Anal fin: 17-20 Adipose fin: present



EGGS

Undescribed

Undescribed

YOLK-SAC LARVAE

Figs. A-D

LARVAE

Body: elongate and slender in early larvae becomes deeper and laterally compressed throughout development Head: large, deep and compressed; snout rounded Eye: round and large Gut: short, thick in its anterior part; terminal section of gut protruding Preanus length: increases from about 40% SL to 60% SL Air bladder: prominent

PHOTOS

Spination: none

Pigmentation: increases with development; early larvae, melanophores at jaw tips and on top of head; peritoneum and dorsum of swimbladder pigmented; a melanophore over terminal gut; late larvae, two parallel dorsal rows of melanophores, two ventral and a lateral, over body midline **Length at flexion:** 5.0-6.5 mm **Length at transformation:** about 20.0 mm SL

by J.M. Rodriguez













Notoscopelus resplendens (Richardson, 1845)



Literature: Badcock and Merret (1976), Fahay (2007), Froese and Pauly (2022), Moser and Ahlstrom (1996b) Illustrations' sources: A-D: L. Rodríguez (A-C: redrawn from Moser and Ahlstrom, 1996b; D: redrawn from Moser and Ahlstrom, 1972) MYCTOPHIDAE

Symbolophorus veranyi (Moreau, 1888) Large scale lantern fish - Lanterne à grandes écailles

Meristic characters

Vertebrae: 39-40

Dorsal fin: 12-13

Anal fin: 21-23

Myomeres: about 39-40

Habitat: oceanic, mesopelagic, between 100 and 800 m depth Distribution: Atlantic Ocean. Eastern

Atlantic, from 26°N to 54°N, and the Mediterranean Sea

Spawning season: unknown

YOLK-SAC LARVAE

Undescribed

EGGS

Undescribed

Figs. A-G

LARVAE Body: moderately elongate, somewhat deeper anteriorly; pectoral fins very large with large base Head: moderately large; snout pointed, flattened

and concave in early larvae, becomes rounded during development

Eye: elliptical, stalked in early larvae, with tapered choroid mass ventrally **Gut:** relatively elongate, globose anteriorly, narrows towards its end

Preanus length: > 60% SL

PHOTOS

Air bladder: absent

Spination: none

Pigmentation: melanophores on tips of snout and lower jaw; large spot on posterior edge of opercle; few preanal, ventral melanophores; row of postanal ventral melanophores; row of spots over gut; pectoral-fin rays pigmented (heavier at ray base); pigmentation decreases towards end of larval period

Length at flexion: < 8.0 mm SL Length at transformation: about 20.0 mm

by J.M. Rodriguez





6.6 mm SL (dorsal view)

Symbolophorus veranyi (Moreau, 1888)



MYCTOPHIDAE

Zeus faber Linnaeus, 1758

John dory - Saint Pierre



Oil globule location: at posterior, ventral edge of yolk sac

Anus: located close behind yolk sac, reaches finfold border

Preanus length: about 40% SL

Pigmentation: head, yolk sac and base of finfold covered with large stellate melanophores that extend to finfold border forming 2 patches close behind mid-tail; caudal region unpigmented; eye pigmented

LARVAE

unpigmented

Colour: transparent

Perivitelline space: small

Yolk: unsegmented; pigmented

Oil globules: one; diam. 0.35-0.40 mm;

Body: relatively short, rhomboid and very high, tapering to narrow tail; pelvic fins, well developed from early larvae on, have 6 rays **Head:** large and very high

Mouth: very large, oblique, reaching to level of mid-eye

Eye: rounded, large and well forward on head **Gut:** triangular and swollen

Preanus length: increases through development, from about 50% SL in early larvae to about 70% SL in late larvae

Air bladder: absent

Spination: none

Pigmentation: similar to yolk-sac larvae; pelvic fins heavily pigmented; anterior half of dorsal fin pigmented; caudal region unpigmented **Length at flexion:** begins at about 6.0 mm **Length at transformation:** unknown

PHOTOS

by J.M. Rodriguez



4.3 mm SL



Illustrations' sources: A, B, D, E: Sanzo (1956a); C: Alemany (1997)

ZEIFORMES

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LOTIDA

Gaidropsarus biscayensis (Collett, 1890)

Habitat: neritic and upper-slope, benthopelagic, between 80 and 600 m depth

Distribution: eastern Atlantic Ocean, from Morocco to the Bay of Biscay, and the western Mediterranean Sea **Spawning season:** February Meristic characters Myomeres: 45-46 Vertebrae: 45-46 2nd dorsal fin: 51-54 Anal fin: 42-47

Mediterranean bigeye rockling - Motelle



EGGS

Undescribed

Undescribed

YOLK-SAC LARVAE

Figs. A-E

LARVAE

PHOTOS

Body: deep and stubby, with caudal region slender in early larvae; pelvic fins large Gut: relatively short, globose and triangular Head: large and rounded Eye: round and large Preanus length: decreases from about 60% SL in early larvae to about 46% SL in late larvae Air bladder: absent Spination: 2 prominent cephalic spines on each side of head

Pigmentation: melanophores on top of head and on lateral surface of trunk; dorsal side of peritoneum strongly pigmented; melanophores on lateral surface of trunk increase in number throughout development and widen joining those on head, forming a continuous bar; upper and lower jaw tips pigmented; pelvic fins heavily pigmented; most postanal region unpigmented in early larvae, reduced to caudal region in later larvae **Length at flexion:** starts at about 4.0-4.5 mm **Length at transformation:** unknown

by J.M. Rodriguez





LOTIDAE

GADIFORMES

Gaidropsarus mediterraneus (Linnaeus, 1758) Shore rockling – Motelle de Méditerranée

LOTIDAE

Habitat: neritic and upper slope, demersal, between 1 and 450 m depth Distribution: eastern Atlantic Ocean, from north of Morocco to Norway, and the Mediterranean Sea Spawning season: September to March (Mediterranean Sea) Meristic characters Myomeres: 46-49 Vertebrae: 46-49 1st dorsal fin: an elongate flexible ray 2nd dorsal fin: 51-63 Anal fin: 43-53

Fig. A



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EGGS

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 0.66-0.72 mm Perivitelline space: small Yolk: unsegmented; unpigmented Oil globules: one; diam. 0.15-0.19 mm; pigmented Colour: transparent

YOLK-SAC LARVAE

Hatch size: 1.8-1.9 mm Body: relatively elongate and slender Yolk sac: elongated

Oil globule location: at posterior edge of yolk sac **Anus:** close to yolk sac, opens on lateral side of finfold

Preanus length: about 50% SL

Pigmentation: melanophores on head, tip of upper jaw and dorsum of trunk; a band of pigment at about mid-tail; a melanophore on ventral side of caudal region; oil globule pigmented

LARVAE

Body: relatively short with a large and deep abdominal region; pelvic fins large since early larvae Head: large and rounded Eye: round and large Gut: triangular, globose Preanus length: < 50% SL Air bladder: present Spination: 2 large cephalic spines on each side of head Figs. C-F

Fig.

Pigmentation: a band of pigment at about midtail that with development gradually changes to a mid-dorsal pigmented area which progressively enlarges, first anteriorly, later posteriorly; pigment patch on ventral side of caudal fin; melanophores on neck, head, lower jaw and peritoneal region; pelvic fins heavily pigmented **Length at flexion:** 4.0-4.5 mm **Length at transformation:** unknown

PHOTOS



LOTIDAE

GADIFORMES





GADIDAE

Gadiculus argenteus Guichenot, 1850

Habitat: oceanic, bathypelagic, between 200 and 1 000 m depth

Distribution: eastern Atlantic Ocean, from Morocco to North Cape (Norway), and the western Mediterranean Sea
 Myomeres: about 37-40

 Vertebrae: 37-40

 1st dorsal fin: 8-12

 2nd dorsal fin: 10-14

 3rd dorsal fin: 11-15

 1st anal fin: 15-18

 2nd anal fin: 12-16

Meristic characters

Silvery put - Merlan argenté



Spawning season: January and February

EGGS

Undescribed

LARVAE

Undescribed

Figs. A-D

Body: short and plump, with deep abdominal region and tapered tail (spindle-shaped, typical of gadid species) Head: large Eye: round and relatively large Gut: relatively short, globose and triangular Preanus length: < 50% SL Air bladder: absent Spination: none

Pigmentation: 2 opposing groups of melanophores (dorsal and ventral) at about mid-tail in early larvae, growing on lateral sides of body to form a continuous bar in late larvae; some melanophores on head; upper and lower jaw-tips and peritoneal region pigmented; caudal fin pigmented; pelvic fins unpigmented **Length at flexion:** unknown

Length at transformation: unknown

YOLK-SAC LARVAE

PHOTOS

by J.M. Rodriguez













GADIDAE

GADIFORMES

Gadiculus argenteus Guichenot, 1850



Literature: Cohen *et al.* (1990), D'Ancona (1933a), Froese and Pauly (2022), Russell (1976), Sabatés (1988), Svetovidov (1986a) Illustrations' sources: A-D: D'Ancona (1933a)

Micromesistius poutassou (Risso, 1827)

Habitat: oceanic, meso-benthopelagic over the continental slope, from 150 m to 3 000 m depth

Distribution: North Atlantic Ocean, and the Mediterranean Sea. Eastern Atlantic, from Cape Bojador to the Barents Sea

Spawning season: January to March

Fig. A

Meristic characters

Myomeres: 55-58

Vertebrae: 55-58

1st dorsal fin: 12-14 2nd dorsal fin: 12-14

3rd dorsal fin: 23-28

1st anal fin: 33-39

2nd anal fin: 24-27

EGGS

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 1.00-1.14 mm Perivitelline space: small Yolk: unsegmented; unpigmented Oil globules: none Colour: transparent

YOLK–SAC LARVAE Hatch size: 2.0–2.2 mm

Body: relatively elongate and slender **Yolk sac:** oval, and relatively large **Anus:** close to yolk sac opens on lateral side of finfold **Preanus length:** about 50% SL

Pigmentation: melanophores over head; scattered melanophores over body, except caudal region; posteriorly, dorsal and ventral bars of melanophores appear on trunk and a cap of melanophores on head; yolk sac unpigmented

LARVAE

Body: relatively long with a relatively large head, deep abdominal region and tapered tail (spindleshaped, typical of Gadid species) Gut: triangular Head: large; mouth oblique and relatively small Eye: round and large Preanus length: decreases with development to about 40% SL after flexion Air bladder: absent **Pigmentation:** dorsal (from head) and ventral (from anus) rows of paired melanophores to about mid– tail (extending backwards during development), with dorsal rows extending more backwards than ventral ones; melanophores on head; peritoneal region pigmented; no melanophores on lateral sides of trunk in larvae < 6.0 mm; end of tail and caudal region unpigmented **Length at flexion:** 8.0-13.0 mm

Length at transformation: > 20.0 mm

PHOTOS

Spination: none

by J.M. Rodriguez

Fig.



2.4 mm SL





6.0 mm SL

8.0 mm SL

GADIDAE

Blue Whiting(=Poutassou) - Merlan bleu

Micromesistius poutassou (Risso, 1827)



Illustrations' sources: A-C, G: Seaton and Bailey (1971); D, E: Schmidt (1905); F: D'Ancona (1933a)

GADIDAE

GADIFORMES

GADIDAE

Trisopterus luscus (Linnaeus, 1758) Pouting (=Bib) – Tacaud commun Habitat: benthopelagic, between 50 **Meristic characters** and 100 m depth Myomeres: 47-49 Distribution: eastern Atlantic Ocean, Vertebrae: 47-49 1st dorsal fin: 11-14 from Morocco to the North Sea, and 2nd dorsal fin: 20-24 the Mediterranean Sea 3rd dorsal fin: 18-20 Spawning season: December to April 1st anal fin: 30-34 2nd anal fin: 19-22 YOLK-SAC LARVAE Fig. Hatch size: about 3.0 mm

EGGS

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 0.90-1.23 mm Perivitelline space: small Oil globules: none Yolk: unsegmented; unpigmented Colour: transparent

Body: relatively elongate and slender Yolk sac: oval, and relatively large Anus: close behind yolk sac, opens on lateral side of finfold Preanus length: about 50% SL

Pigmentation: melanophores distributed over anterior region of body: last third of body and yolk sac unpigmented

LARVAE

Body: relatively short (shorter than that of *M. poutassou*) with deep abdominal region and tapered tail (spindle-shaped, typical of gadid species)

Head: large and deep; mouth oblique Gut: relatively short, globose and triangular Eye: round and relatively large Preanus length: about 50% SL Air bladder: absent Spination: none

PHOTOS

Pigmentation: dorsal (from head) and ventral (from anus) rows of melanophores, ending at same level, at about mid-tail; peritoneum, ventral side of abdomen and occipital region pigmented; 1-2 melanophores on lateral sides of body in early larvae, increasing in number with development; posterior mid-tail and caudal region unpigmented Length at flexion: unknown Length at transformation: unknown

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3.7 mm SL

4.6 mm SL



7.8 mm SL

GADIDAE

B. 3.0 mm

A. C. 3.8 mm One or two melanophores on lateral sides of body in early larvae Posterior mid-tail region unpigmented **D.** 4.5 mm Dorsal and ventral rows of paired melanophores end at same level 40%

Trisopterus luscus (Linnaeus, 1758)

E. 7.3 mm

1211/101

minn



GADIFORMES

Literature: Cohen et al. (1990), D'Ancona (1933a), Russell (1976), Svetovidov (1986a) Illustrations' sources: A, B, E: D'Ancona (1933a); C, D, F: Munk and Nielsen (2005)

Trisopterus minutus (Linnaeus, 1758)

Habitat: neritic, demersal, between 10 **Meristic characters** and 200 m depth Myomeres: 47-48 Distribution: eastern Atlantic Ocean, Vertebrae: 47-48 from Morocco to Norway, and the 1st dorsal fin: 12-13 2nd dorsal fin: 23-27 western Mediterranean Sea 3rd dorsal fin: 20-25 Spawning season: December to 1st anal fin: 25-29 March 2nd anal fin: 15-20 EGGS

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 0.95-1.03 mm Perivitelline space: small Yolk: unsegmented; pigmented Oil globules: none Colour: transparent

Poor cod - Capelan de Méditerranée

YOLK-SAC LARVAE

Fig.

Hatch size: about 2.5 mm Body: elongate and slender Yolk sac: oval Anus: close behind yolk sac, opens on lateral side of finfold Preanus length: about 43% SL Pigmentation: dorsal and ventral rows of melanophores; eyes pigmented

LARVAE

Body: relatively long (slender than that of *T. luscus*) and with a tapered tail (spindle-shaped, typical of gadid species) Gut: relatively short, globose and triangular Head: relatively large; mouth oblique Eye: round and large Preanus length: about 48% SL Air bladder: absent Spination: none

Pigmentation: single dorsal and ventral rows (about 9 in each row) of large melanophores, extending posteriorly for approximately same distance from caudal-fin base; a melanophore on each side of head, above eye; peritoneum and upper jaw pigmented; a few large preanal ventral melanophores; some melanophores appear above and below urostyle at about 6.0 mm; lateral sides of body unpigmented Length at flexion: unknown Length at transformation: unknown

PHOTOS

by J.M. Rodriguez





2.6 mm SL



8.2 mm SL

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Trisopterus minutus (Linnaeus, 1758)



Merluccius merluccius (Linnaeus, 1758)

Habitat: neritic and slope, benthopelagic, demersal, between 30 and 1 000 m depth

Distribution: eastern Atlantic Ocean, from Mauritania to Norway, and the Mediterranean Sea

Myomeres: 49-54 Vertebrae: 49-54 1st dorsal fin: I, 7-10 2nd dorsal fin: 36-40 Anal fin: 36-40

Meristic characters

European hake – Merlu européen

Spawning season: January to March (Morocco)

Fig. A

EGGS

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 0.94-1.03 mm Perivitelline space: small Yolk: unsegmented; pigmented Oil globules: one; diam. 0.25-0.28 mm; pigmented Colour: transparent

YOLK-SAC LARVAE

Hatch size: about 3.0 mm Body: relatively elongate and slender Yolk sac: elongated

Oil globule location: at posterior edge of yolk sac Anus: close behind yolk sac, opens on lateral side of finfold

Preanus length: about 37% SL Pigmentation: 3 postanal, large, stellate melanophores; melanophores on head; peritoneal region and oil globule pigmented

LARVAE

Body: relatively short with a large and deep abdominal region and tapered tail; pelvic fins appear in 4.0-6.0 mm larvae Head: large and deep; mouth large, moderately oblique

Eye: round and relatively large Gut: triangular, globose and short Preanus length: about 45% SL Air bladder: absent

PHOTOS

Spination: none

Pigmentation: 3 stellate melanophores in lateralpostanal region; peritoneal region and pelvic fins pigmented; melanophores on snout, lower jaw and on neck; ventral side of tail unpigmented **Length at flexion:** < 9.1 mm SL Length at transformation: unknown

by J.M. Rodriguez

Fig.





3.8 mm SL



7.0 mm SL





MERLUCCHDAE

Merluccius merluccius (Linnaeus, 1758)



Illustrations' sources: A: D'Ancona (1933a); **B-G:** L. Rodríguez (**B**, **G:** redrawn from D'Ancona, 1933a; **C-F:** redrawn from Palomera *et al.*, 2005)

GADIFORMES

MELAMPHAIDAE

Melamphaes simus Ebeling, 1962

Habitat: oceanic, meso- to bathypelagic, below 150 m depth Distribution: tropical and subtropical regions of the Atlantic, Pacific and Indian oceans.

Spawning season: unknown

EGGS

Undescribed

Undescribed

Meristic characters

Dorsal fin: III + 15-17

Myomeres: 28-30

Vertebrae: 28-30

Anal fin: I + 9-10



LARVAE

Body: elongate in early larvae, remains elongate throughout development; pelvic and pectoral fins well developed in early larvae; pelvic-fin origin moves backward with development; pelvic-fin rays long and fragile in early larvae, reduce in size with development

Head: relatively large; mouth slightly oblique **Eye:** round and moderately large

Gut: thicker and triangular; terminal gut makes almost a right angle with body in early larvae; anus slightly protruding

Preanus length: increases from 29–35% SL in preflexion larvae to 61–64% SL in juveniles

Air bladder: absent

Spination: none

Pigmentation: a single melanophore on ventral mid-tail in early larvae; dorsum of body unpigmented in preflexion stage; pelvic-fin rays pigmented; throughout development, ventral spots expand to form a line between anal and caudalfin base, and a dorsal line of melanophores that expands embracing most of body appears in late larvae; peritoneal region pigmented; melanophores over head and at caudal peduncle in late larvae **Length at flexion:** 5.1-7.3 mm

Length at transformation: about 12.0-13.0 mm

by J.M. Rodriguez



4.5 mm SL



8.1 mm SL

PHOTOS



Literature: Ebeling and Weed (1963), Ebeling (1986), Fahay (2007), Frias-Torres (2006), Froese and Pauly (2022), Sandknop and Watson (1996)

Illustrations' sources: A-D: L. Rodríguez (redrawn from Sandknop and Watson (1996)

Parophidion vassali (Risso, 1810) Habitat: neritic, demersal, between 0 **Meristic characters** and 150 m depth Myomeres: 71-73 Distribution: eastern Atlantic Ocean, Vertebrae: 71-73 from Mauritania to Portugal, and the Dorsal fin: 122-128 Mediterranean Sea Anal fin: 107-110 Spawning season: summer (Mediterranean Sea) YOLK-SAC LARVAE EGGS Fig. A Fig. Habitat: pelagic Hatch size: about 3.6 mm Body: relatively elongate and slender Shape: spherical Yolk sac: elongated, reaches snout Chorion: smooth; diam. 0.96 mm Anus: detached from yolk sac, reaches border of Perivitelline space: small finfold, forming a right angle with gut Yolk: unsegmented; unpigmented Preanus length: about 50% SL Oil globules: none Pigmentation: melanophores over ventral profile Colour: transparent

LARVAE

Body: elongate and slender Head: relatively long and deep; jaws in angle pointing down Eve: round and small Gut: short, forms a swelling (probably a loop, like in other Ophidiidae species), midway along its length Preanus length: about 42% SL Air bladder: absent Spination: none

Pigmentation: a ventral series of dotted melanophores extending from pectoral fin to urostyle end; ventral melanophores over gut; a series of melanophores on dorsal posterior half of tail; late larvae add melanophores on lower jaw and posterior edge of operculum; lateral sides of body unpigmented Length at flexion: unknown Length at transformation: unknown

of body and laterally at about mid-body

PHOTOS

by J.M. Rodriguez











Echiodon dentatus (Cuvier, 1829) Habitat: oceanic, demersal, between **Meristic characters** 120 and 3 250 m depth Myomeres: many Distribution: eastern Atlantic Ocean, Vertebrae: NA from the equator to North of Spain, Dorsal fin: 144 and the Mediterranean Sea Anal fin: 165 Spawning season: winter (Mediterranean Sea) YOLK-SAC LARVAE EGGS Fig. Habitat: pelagic Hatch size: about 3.80 mm Body: elongate and slender; incipient dorsal Shape: ovoid appendage situated above yolk sac **Chorion:** smooth, embedded in slime; size 1.43 x **Yolk sac:** large, slightly ovoid (almost circular) 0.89 mm **Oil globule location:** at anterior edge of yolk sac Perivitelline space: small Anus: close behind yolk sac, reaches finfold border Yolk: unsegmented; pigmented Preanus length: about 36% SL Oil globules: one; diam. 0.13-0.17 mm; pigmented Pigmentation: a melanophore at base of dorsal Colour: transparent appendage; some melanophores on dorsal surface of yolk sac; 2 melanophores in ventral tail region; opposing dorsal and ventral melanophores at caudal region; oil globule and yolk sac pigmented LARVAE Body: very elongate and slender, tapering to a Pigmentation: melanophores on snout, on lower filamentous tip; a large dorsal appendage situated a jaw and over gut; 3 relatively large and other small little posterior to anus postanal ventral melanophores; opposing dorsal Head: relatively long and curved downward; snout and ventral melanophores at caudal region; in late concave larvae, ventral melanophores migrate to sides of Eve: round tail, dorsal melanophore on caudal region may Gut: short, coiled disappear and ventral melanophore reduce in size Preanus length: about 20% SL and increase in number, up to 3; dorsal appendage Air bladder: prominent strongly pigmented Spination: none Length at flexion: unknown

PHOTOS

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not sized

Length at transformation: unknown



not sized

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CARAPIDAE



Cubiceps pauciradiatus G	Bigeye cigarfish			
Habitat: oceanic, epi- mesopelagic, between 58 and 1000 m depth Distribution: worldwide in tropical and subtropical waters Spawning season: unknown	Meristic characters Myomeres: 30-31 Vertebrae: 30-31 Dorsal fin: XI-XII+15-2 Anal fin: II + 14-17		Contraction	
EGGS	Fig. A	YO	LK-SAC LARVAE Fig. B	
Habitat: pelagic Shape: spherical Chorion: smooth; diam. 0.70-0.80 mm Perivitelline space: small Yolk: unsegmented; pigmented Oil globules: one; diam. 0.14-0.20; pigmented Colour: transparent		Hatch size: < 2.1 mm SL Body: slender; gut straight Yolk sac: no information Oil globule location: anterior in yolk sac Anus: detached from yolk sac, reaches finfold border Preanus length: < 50% SL Pigmentation: tip of jaws, peritoneal region, oil globule and yolk sac pigmented		
LARVAE			Figs. C-F	
 Body: initially elongate, soon deepens, becoming moderately stocky with development Head: moderately large; mouth relatively large and oblique Eye: round and relatively large Gut: initially straight, becomes bulky and triangular after coiling Preanus length: increases from 52-59% SL to 59-65% SL throughout development Air bladder: absent 		Spination: 4-5 weak preopercular spines (forming at about 5.0 mm SL); a few spines on lateral ridge (forming at about 8.5 mm SL) are reabsorbed at about 11.2 mm SL Pigmentation: melanophores on head; tips of jaws and peritoneal region pigmented; melanophores form three stripes (on dorsum, middle and ventral sides of body) at about mid-tail; pigmentation increases with development Length at flexion: 3.8-4.5 mm Length at transformation: unknown		

PHOTOS

SCOMBRIFORMES

by J.M. Rodriguez



4.0 mm SL

NOMEIDAE

NOMEIDAE

Cubiceps pauciradiatus Günther, 1872



A.



D. Ventral view



Melanophores form three stripes, on dorsum, middle and ventral sides of tail





Literature: Ahlstrom *et al.* (1976), Fahay (2007), Froese and Pauly (2022), Lamkin (2006), Olivar and Fortuño (1991) Illustrations' sources: A-F: L. Rodríguez (redrawn from Ahlstrom *et al.*, 1976)

Auxis rochei (Risso, 1810)

Habitat: neritic-oceanic, epipelagic **Distribution:** worldwide, in tropical and temperate waters Spawning season: November to August

Meristic characters Myomeres: 39 Vertebrae: 39 1st dorsal fin: XX-XXII 2nd dorsal fin: 10-12 Dorsal finlets: 8 Anal-fin: 11-14 Anal finlets: 7

Fig. A

Bullet tuna - Bonitou

EGGS

Habitat: pelagic Shape: spherical Chorion: smooth; diam 0.82-1.10 mm Perivitelline space: small Yolk: unsegmented; unpigmented Oil globules: one; diam. 0.24-0.29 mm; pigmented Colour: transparent

YOLK-SAC LARVAE

Hatch size: about 2.14 mm Body: elongate Yolk sac: no information Oil globule location: no information Anus: no information Preanus length: about 37% SL **Pigmentation:** melanophores on head; a row of melanophores on postanal ventral region; gut pigmented

LARVAE

Body: moderately elongate in early larvae, becoming deeper with development Head: slightly bigger than in Scomber larvae, showing a fairly blunt profile; jaws short (compared with other tuna species) Eve: round and large Gut: triangular and compact Preanus length: increases from 37 to 50% SL during development Air bladder: absent Spination: preopercular spines on edge and lateral ridge well developed with spine at angle longer; 2-3 post-temporal spines in late larvae

Pigmentation: early larvae similar to yolk–sac larvae; late larvae, origin of ventral postanal row of melanophores moves backwards and melanophores decrease in number; melanophores on top of head; peritoneal pigment increases; 2 dorsal melanophores on caudal peduncle that spread forward with development; a melanophore at cleithral symphysis Length at flexion: 4.5-6.0 mm SL

Length at transformation: unknown

PHOTOS

by J.M. Rodriguez











SCOMBRIDA

SCOMBRIFORMES

Auxis rochei (Risso, 1810)



SCOMBRIDAE

SCOMBRIFORMES

Euthynnus alletteratus (Rafinesque, 1810)

Little tunny - Thonine commune

Habitat: neritic, epipelagic in coastal waters

Distribution: Atlantic Ocean, in tropical and temperate waters, and the Mediterranean Sea

Spawning season: April to November

Meristic characters Myomeres: 39 Vertebrae: 39 1st dorsal fin: XIII-XV 2nd dorsal fin: 11-12 Dorsal finlets: 8 Anal fin: 11-13 Anal finlets: 7-8

EGGS

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 0.84-1.08 mm Perivitelline space: no information Yolk: unsegmented Oil globules: one; diam. 0.28 mm **Colour:** transparent

Undescribed

YOLK-SAC LARVAE

LARVAE

Body: moderately stubby (deeper than similarsized Auxis larvae)

Head: large, with a large and pointed snout; mouth large, reaching beyond mid-point of eye; each jaw with 14 relatively large teeth, since early larvae Eye: round and large

Gut: triangular and compact

Preanus length: increases from 50 to 70% SL during development

Air bladder: absent

Spination: preopercular spines well developed on edge and lateral ridge, with spine at angle longer; 1-2 post-temporal spines

Pigmentation: row of melanophores over lower jaw that increases in number and length with development; tip of upper jaw pigmented; around 11 postanal, ventral, regularly spaced melanophores; a melanophore at base of caudal fin; melanophores on top of head, increase in number with development and extend anteriorly to area in front of eyes; peritoneum pigmented; dorsal fin pigmented; lateral sides of trunk and tail unpigmented until later larvae Length at flexion: 5.5-7.5 mm SL Length at transformation: unknown

PHOTOS





4.3 mm SL



9.3 mm SL

SCOMBRIDA

172

by S. Isari



Literature: Collette (1986b), Collette and Nauen (1983), Fahay (2007), Matsumoto (1959, 1962), Richards (2006g) Illustrations' sources: A-E: Matsumoto (1959)

Sarda sarda (Bloch, 1793)

SCOMBRIDAE

Habitat: neritic, epipelagic Distribution: tropical and temperate Atlantic Ocean. Eastern Atlantic, from South Africa to Norway, and the Mediterranean Sea

Chorion: smooth; diam. 1.20-1.32 mm

0.02-0.24 mm when multiple; pigmented

Spawning season: December to July

Meristic characters Myomeres: 50-55 Vertebrae: 50-55 1st dorsal fin: XX–XXIII 2nd dorsal fin: 13–18 Dorsal finlets: 7-9 Anal fin: 14-16 Anal finlets: 6-8

YOLK-SAC LARVAE

Hatch size: about 4.0 mm Body: elongate and slender Yolk sac: large, ovoid

Oil globules location: at posterior edge of yolk sac **Anus:** detached from yolk sac, reaches finfold border **Preanus length:** about 45% SL

Pigmentation: some small melanophores on head; some isolated melanophores on lateral and ventral sides of body; anterior, dorsal and ventral finfold borders pigmented; yolk sac and oil globule(s) pigmented

LARVAE

Colour: transparent

EGGS

Habitat: pelagic

Shape: spherical

Perivitelline space: small

Yolk: unsegmented; pigmented

Body: moderately elongate in early larvae, deepest through pectoral region with development **Head:** large with pointed snout; mouth large, extends to midpoint of eye, with large jaws; prominent teeth from early larvae on **Eye:** round and large **Gut:** triangular and compact

Oil globules: 1-5; diam. 0.28–0.32 mm, when single;

Preanus length: increases from about 50% SL to > 60% SL

Air bladder: absent

Spination: supraoccipital crest with a single spine; two series of preopercular spines, internal one larger; an opercular spine in upper part of

operculum; supraocular crest with several strong spines; postemporal spines

Pigmentation: a series of large spots along ventral tail region that move up internally, between myomeres, with development; top of head and peritoneal region well–pigmented; melanophores at cleithral symphysis, at snout and lower jaw tip; a group of melanophores in lateral caudal region in late larvae; caudal–fin base pigmented **Length at flexion:** 6.0-7.0 mm SL **Length at transformation:** unknown

PHOTOS

SCOMBRIFORMES

by J.M. Rodriguez

Figs. C-F



Atlantic bonito – Bonite à dos rayé

SCOMBRIDAE



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SCOMBRIDA

Scomber colias Gmelin, 1789

Habitat: neritic, epipelagic

Distribution: warm and temperate waters of the Atlantic Ocean and the Mediterranean Sea. Eastern Atlantic, from South Africa to the Bay of Biscay **Spawning season:** April to August Meristic characters Myomeres: 31 Vertebrae: 31 1st dorsal fin: IX–XIII 2nd dorsal fin: 11–12 Dorsal finlets: 5 Anal fin: I + 11-14 Anal finlets: 5



Atlantic chub mackerel - Maquereau espagnol

EGGS

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 1.04-1.14 mm Perivitelline space: small Yolk: unsegmented; pigmented Oil globules: one; diam. 0.26-0.27 mm; pigmented Colour: transparent

YOLK-SAC LARVAE

Hatch size: about 3.0 mm Body: relatively elongate and slender Yolk sac: large, ovoid

Oil globule location: at posterior edge of yolk sac **Anus:** close behind yolk sac, reaches finfold border **Preanus length:** about 50% SL

Pigmentation: melanophores irregularly distributed along dorsal and ventral contours of body; melanophores on snout and behind eye; yolk sac and oil globule pigmented

LARVAE

Body: moderately elongate, becoming stubby with development (stubbier than comparable sizes of *Scomber scombrus*) **Head:** moderate, snout rounded; mouth moderate;

teeth prominent from 4.0 mm larvae on Eye: round and large Gut: triangular and compact

Preanus length: increases from about 50% SL to about 60% SL during development (larger than in *S. scombrus*) **Air bladder:** absent Figs. C-F

Fig.

Spination: none

Pigmentation: a postanal ventral row of melanophores, beginning some distance from anus; 1-2 dorsal melanophores on caudal peduncle in larvae < 7.0 mm; peritoneum pigmented; melanophores at caudal–fin base, on head and on body sides (absent in preflexion stages); no melanophores at cleithral symphysis **Length at flexion:** 5.0-7.0 mm **Length at transformation:** unknown

PHOTOS

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4.9 mm SL





7.3 mm SL



SCOMBRIDAE

SCOMBRIFORMES

Scomber scombrus Linnaeus, 1758

Habitat: neritic, pelagic and benthopelagic

Distribution: Atlantic Ocean and the Mediterranean Sea. Eastern Atlantic, from Morocco to Norway Spawning season: spring

Meristic characters Myomeres: 31 Vertebrae: 31 1st dorsal fin: X + 13-15 2nd dorsal fin: 11 Dorsal finlets: 5 Anal fin: II + 11 Anal finlets: 5

Atlantic mackerel – Maquereau commun

EGGS

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 1.0-1.38 mm Perivitelline space: small Yolk: unsegmented; unpigmented Oil globules: one; diam. 0.28-0.35 mm; unpigmented Colour: transparent

YOLK-SAC LARVAE

and behind eye; oil globule pigmented

Hatch size: about 2.4 mm **Body:** relatively elongate Yolk sac: large, ovoid Oil globule location: at posterior edge of yolk sac Anus: close behind yolk sac, reaches finfold border Preanus length: about 40% SL Pigmentation: some melanophores on head; double rows of irregularly distributed melanophores (up to 14 each) along dorsal and ventral contours of body; melanophores on snout

LARVAE

Body: moderately elongate, becoming stubby with development (slimmer than comparable sizes of S. colias)

Head: moderate, snout rounded; mouth moderate; teeth prominent from larvae 4.0 mm long Eye: round and large Gut: triangular and compact Preanus length: increases from about 40% SL to 60% SL during development Air bladder: absent

Spination: none

Pigmentation: postanal rows of dorsal and ventral melanophores; peritoneum pigmented; some melanophores on urostyle, along caudal-fin base, on head and sometimes on snout and lower jaw; no melanophores on body sides in preflexion larvae; a melanophore at cleithral symphysis Length at flexion: 5.0-7.0 mm Length at transformation: unknown

PHOTOS

by J.M. Rodriguez





3.7 mm SL



7.0 mm SL

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SCOMBRIDA



Illustrations' sources: A-G: Padoa (1956n)

SCOMBRIFORMES

Thunnus alalunga (Bonnaterre, 1788)

Habitat: epipelagic and mesopelagic, Merist

oceanic Distribution: cosmopolitan in tropical and temperate waters. Eastern Atlantic, from South Africa to Great Britain Spawning season: summer (Mediterranean Sea) Meristic characters Myomeres: 39 Vertebrae: 39 1st dorsal fin: XI-XIV 2nd dorsal fin: 12-16 Dorsal finlets: 7-9 Anal fin: 11-16 Anal finlets: 7-8

EGGS

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 0.84-0.94 mm Perivitelline space: small Yolk: unsegmented; pigmented Oil globules: one; diam. 0.25-0.28 mm; pigmented Colour: transparent

YOLK–SAC LARVAE Hatch size: about 2.7 mm SL

Body: elongate **Yolk sac:** oval and large **Oil globule location:** at posterior end of yolk sac **Anus:** detached from yolk sac, reaches finfold border

Preanus length: about 39% SL **Pigmentation:** melanophores on yolk and on gut; dotted melanophores dorsal and ventral to urostyle

LARVAE

Body: moderately stocky, deepest through pectoral region, tapering to a narrow caudal peduncle Head: large with a pointed snout and large jaws; prominent teeth since early larvae Eye: round and large Gut: compact and triangular Preanus length: increases from about 40% SL to 55% SL during development Air bladder: absent Spination: a supraoccipital spine; several strong spines along edge and a few smaller on lateral ridge of opercle; 1-2 small post-temporal spines **Pigmentation:** a melanophore ventral to urostyle in very early larvae; peritoneal region strongly pigmented, melanophores extend over lateral sides of gut with development; occipital region pigmented (strongly in late larvae); tips of jaws pigmented in larvae > 7.0 mm; melanophores appear on 1st dorsal fin at > 5.0 mm; tail unpigmented **Length at flexion:** about 5.0-7.0 mm SL

Length at flexion: about 5.0-7.0 mm SL **Length at transformation:** unknown

PHOTOS







5.2 mm SL

by J.M. Rodriguez



SCOMBRIDAE

SCOMBRIFORMES





Literature: Alemany (1997), Collette (2016), Fahay (2007), Matsumoto *et al.* (1972), Olivar and Fortuño (1991), Richards (2006g), Padoa (1956n)

Illustrations' sources: A: Padoa (1956n); B, C: Alemany (1997); D-G: Ueyanagi (1969)

Thunnus thynnus (Linnaeus, 1758) Atlantic bluefin tuna - Thon rouge de l'Atlantique

Habitat: epipelagic and mesopelagic, oceanic, but seasonally close to the shore

Distribution: Atlantic Ocean and the Mediterranean Sea. Eastern Atlantic, from South Africa to Norway Spawning season: June and July (Mediterranean Sea)

Meristic characters Myomeres: 39 Vertebrae: 39 1st dorsal fin: XI-XIV 2nd dorsal fin: 12-16 Dorsal finlets: 8-10 Anal fin: 11-16 Anal finlets: 7-9

Oil globule location: at posterior end of yolk sac

Pigmentation: melanophores from snout to

midbrain; 3 large clusters of melanophores in dorsal midline; a line of melanophores above gut and on ventral tail region; a ventral cluster of melanophores in caudal region (opposite to last dorsal one); melanophores along lateral midline of

Anus: detached from yolk sac, reaches finfold border

YOLK-SAC LARVAE

trunk

Hatch size: about 2.8 mm TL

Preanus length: about 34% SL

Yolk sac: oval and large

Body: elongate

Fig. B

EGGS

Habitat: pelagic Shape: spherical

Chorion: smooth; diam. 1.10-1.12 mm Perivitelline space: small Yolk: unsegmented; unpigmented Oil globules: one; diam. 0.25-0.28 mm; pigmented Colour: transparent

LARVAE

Body: stout, deepest through pectoral region, tapering to a narrow caudal peduncle Head: large with pointed snout and large jaws; prominent teeth since early larvae Eye: round and large Gut: compact and triangular Preanus length: increases from about 40% SL to 55% SL throughout development Air bladder: absent Spination: several strong spines along edge and a few smaller ones on lateral ridge of opercle; 1-2 posttemporal spines develop at about 7.0 mm SL

Pigmentation: about 4 dotted postanal-ventral melanophores in early larvae, increasing in number with development; a single melanophore on dorsal side of mid-tail in early larvae, migrates to dorsal fin in larvae > 5.0 mm; peritoneum strongly pigmented, melanophores extend over lateral sides of gut with development; melanophores on top of head; tips of upper and lower jaws pigmented; melanophores along lateral midline and on dorsal side of tail in late larvae Length at flexion: 5.0-7.0 mm SL Length at transformation: unknown

A: F. by F. de la Gándara; B, C: by J.M. Rodriguez

PHOTOS



6.7 mm SL

[1]

SCOMBRIDA



Illustrations' sources: A: Padoa (1956n); B, F: Miyashita et al. (2001); C-E: Richards (2006g)

	Brama brama (Bonnaterre, 1788)			Atlantic pomfret – Grande castagnole		
RAMIDAE	 Habitat: oceanic, epi- to mesopelagic, to 1 000 m depth Distribution: southern Pacific, Indian and Atlantic oceans, and the Mediterranean Sea. Eastern Atlantic, from South Africa to Norway Spawning season: August to September (Mediterranean Sea) 	Meristic character Myomeres: 41-43 Vertebrae: 41-43 Dorsal fin: 35-38 Anal fin: 29-32	'S			
	EGGS	Fig. A	YO	LK-SAC LARVAE Fig. B		
	Habitat: pelagic Shape: spherical Chorion: smooth; diam. 1.50-1.60 mm Perivitelline space: small Yolk: unsegmented; pigmented Oil globules: one, oval; diam. 0.40 x 0.32 mm; pigmented Colour: transparent Education: transparent Colour: transparent Education: transparent Education: transparent Education: transparent Education: transparent Education: transparent Education: transparent Education: transparent Solut: round and bulky Shape: specially through pectoral region, with development Head: moderately large with "angry face"; mouth oblique; teeth present since early larvae Eye: round in early larvae becomes oval in late larvae Gut: round and bulky		Hatch size: about 4.8 mm Body: elongate and fairly slender; pectoral and caudal fins apparent Yolk sac: ovoid Oil globule location: at posterior, ventral side of yolk sac Anus: close behind yolk sac, reaches finfold border Preanus length: about 30% SL Pigmentation: head, yolk sac, oil globule, pectoral and caudal fins pigmented; 2 prominent pigment patches in tail, one dorsal and another ventral extending into finfold			
				Figs. C-G		
			 Preanus length: about 30% SL increases during development Air bladder: absent Spination: a series of spines along preopercle edge Pigmentation: head and gut heavily pigmented; pectoral and caudal fins pigmented; tail unpigmented Length at flexion: 5.0-6.5 mm SL Length at transformation: about 10.0 mm SL 			
	РНОТОЅ			by J.M. Rodriguez		



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Illustrations' sources: A, B: Padoa (1956d); C-F: Alemany (1997); G: Schmidt (1918)



3.6 mm SL



11.6 mm SL

4.8 mm SL



Lepidopus caudatus (Euphrasen, 1788) Silver scabbardfish - Sabre argenté Habitat: neritic and upper slope, **Meristic characters** benthopelagic, between 42 and 620 m Myomeres: 105-114 depth Vertebrae: 105-114 Distribution: eastern Atlantic Ocean, Dorsal fin: IX + 90-107 from South Africa to France, and the **Anal fin: II + 60-65** Mediterranean Sea Spawning season: February to April YOLK-SAC LARVAE EGGS Fig. A Habitat: pelagic Hatch size: about 6.0 mm **Body:** elongate and slender; head very small and Shape: spherical rounded; a dorsal protuberance behind head Chorion: smooth; diam. 1.6-1.7 mm Yolk sac: ovoid, unpigmented Perivitelline space: small **Oil globule location:** at posterior edge of yolk sac Yolk: unsegmented; unpigmented Anus: close behind yolk sac, reaches finfold border Oil globules: one; diam. 0.40 mm; pigmented Preanus length: about 25% SL Colour: transparent **Pigmentation:** some dorsal and ventral melanophores on tail end; 2 large dorsal melanophores with a ventral one in between; some melanophores on head LARVAE Body: very elongate and slender; first dorsal-fin **Pigmentation:** dorsal and ventral melanophores spine highly developed in early larvae in yolk-sac larvae migrate to finfold; ventral caudal melanophores spread forward and dorsal Head: large; snout long and pointed (head duckmelanophores disappear; melanophores on head, billed shaped) Eye: round and relatively small snout, and post-occipital region; peritoneum and Gut: triangular lateral sides of gut pigmented; melanophores **Preanus length:** increases from 26% SL in early on dorsal-fin base extend backwards with larvae to about 50% in flexion larvae development Air bladder: absent Length at flexion: about 14.0 mm SL

Spination: none

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by J.M. Rodriguez



8.9 mm SL



9.9 mm SL

Length at transformation: unknown

TRICHIURIDA





compressed with development; dorsal- and anal-fin spines serrated anteriorly; pelvic and caudal fins absent

Head: relatively large, increases in length with development and becomes moderately concave; mouth large, reaching anterior edge of eye; snout short and pointed in early larvae, increases in length with development; teeth in upper jaw develop early Eye: relatively large and round

Gut: large and triangular, increases in size with development

Preanus length: increases from 31% SL to 55% SL in late larvae, then decreases to about 48% SL in juveniles

PHOTOS

SCOMBRIFORMES

Spination: a low supraocular crest forms in late larvae; several small preopercular spines on posterior edge and on lateral ridge Pigmentation: early larvae, opposing clusters of melanophores on dorsal and ventral finfold edges, and scattered melanophores on top of head; late larvae, a series of melanophores forms on either side of dorsal midline, increasing in number backward with development; few melanophores may occur on sides of head and on lower jaw Length at flexion: flexion does not occur Length at transformation: unknown



A. 5.8 mm SL

B. 6.3 mm SL

D. 7.6 mm SL

E. 12.5 mm SL

Flexion does not occur,

urostyle remains straight

[©] SCOMBRIFORMES

E. 17.0 mm SL Literature: Fahay (2007), Froese and Pauly (2022), Padoa (1956q), Parin (1986), Nakamura and Parin (1993), Richards (2006d)

Body elongate and slender and laterally compressed

Dorsal and ventral clusters of melanophores on finfold border

New York

Small preopercular spines

A series of melanophores on either side of dorsal midline

Trichiurus lepturus Linnaeus, 1758

Serrated anteriorly

C. Detail of dorsal-fin spine

Illustrations' sources: A-F: L. Rodríguez (**A**, **C**, **D**, **E:** redrawn from Okiyama, 1988; **B**, **F:** redrawn from Collette *et al.*, 1984)



LARVAE

Body: elongate, laterally compressed Head: moderate, rounded dorsally, with short snout Eye: slightly oval and large Gut: triangular Preanus length: 30-40% SL in early larvae, increases slightly during development Air bladder: absent Spination: none

Pigmentation: increases during development; early larvae show an occipital melanophore, peritoneum strongly pigmented, a row of postanal ventral melanophores, a short row of mid-lateral melanophores on tail, and a single melanophore under urostyle; dorsum of tail unpigmented; late larvae show several melanophores over head and on dorsum of trunk and tail: gut, except its ventral side, pigmented

Length at flexion: 4.5-5.2 mm SL Length at transformation: unknown



MULLIDA

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MULLIDAE



Illustrations' sources: A-E: Alemany (1997)



MULLIDAE

SYGNATHIFORMES



Illustrations' sources: A, B: Montalenti (1937b); C-F: Alemany (1997)

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Callionymus lyra Linnaeus, 1758 **Dragonet** - **Dragonet** lyre Habitat: neritic and upper shelf break, **Meristic characters** demersal, to about 200 m depth Myomeres: NA Distribution: eastern Atlantic Ocean, Vertebrae: NA from Mauritania to Norway, and the 1st dorsal fin: IV Mediterranean Sea 2nd dorsal fin: 9 Spawning season: February to April Anal fin: I + 9 (Mediterranean Sea) Female YOLK-SAC LARVAE EGGS Fig. Habitat: pelagic Hatch size: about 2.0 mm Body: slender Shape: spherical Chorion: sculptured with hexagonal structures in Yolk sac: ovoid, relatively large some species; diam. 0.66-0.97 mm Anus: close behind yolk sac, does not reach finfold Perivitelline space: small border in recently hatched larvae **Preanus length:** < 50% SL Yolk: segmented; pigmented

Pigmentation: melanophores scattered over body, finfold and yolk sac

and pigmentation patterns become species-specific.

Spination: a preopercular spine bifurcated **Pigmentation:** body, except caudal region,

Air bladder: present

strongly pigmented

Length at flexion: unknown

ichthyoplankton samples.

Length at transformation: unknown

Figs. C-I

LARVAE

Oil globules: none

Colour: transparent

Body: short and tapering; urostyle long, strongly developed and curved up **Head:** large and high **Eye:** round and large **Gut:** triangular and thick **Preanus length:** about 50% SL

Four species of the genus *Callionymus*: *C. filamentosus* (ELS not described), *C. lyra*, *C. maculatus* and *C. reticulatus* live off the north African coast. The identification at the species level is only possible (although quite difficult) in larvae larger than 3.0 mm, when preopercular spines develop, and pelvic fins

Here we include the description of the early stages of *C. lyra* and illustrations of the other species because *Callionymus* larvae are very frequent in

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2.5 mm SL Callionymus sp.







5.9 mm SL

CALLIONYMIDAE

SYGNATHIFORMES

Callionymus lyra Linnaeus, 1758



Illustrations' sources: A, B: Padoa (1956e); C-H: L. Rodríguez (C, D, F-H: redrawn from Demir, 1976; E, redrawn from Fage, 1918); I: Fage (1918)

CENTRISCIDAE



LARVAE

Body: relatively short and high in late larvae Head: relatively small; snout short and pointed in early larvae, long and concave in late larvae, showing a 'duck-billed' profile Eye: round and large Gut: elongated and bulky **Preanus length:** < 50% SL Air bladder: present Spination: supraorbital and occipital crests develop in larvae of 4.0–6.0 mm; spines on preopercle edge;

spinous scales develop along lateral line in larvae of about 4.2 mm and cover entire body by 6.2 mm Pigmentation: early larvae similar to yolksac larvae; late larvae show a lateral line of melanophores in postanal region; pigmentation increases during development spreading onto flanks, covering most of body, except caudal peduncle; melanophores on top of head Length at flexion: 6.0 mm Length at transformation: 13.0-15.0 mm

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Figs. C-F









5.1 mm SL

5.9 mm SL



APOGONIDAE

Apogon imberbis (Linnaeus,	1758)		Cardinal fish – Coq			
Habitat: neritic, demersal, to 200 m depth Distribution: eastern Atlantic Ocean, from northern Angola to Morocco, and the Mediterranean Sea Spawning season: June to September (Mediterranean Sea)	Meristic characters Myomeres: 24 Vertebrae: 24 Dorsal fin: IX-X + 23- Anal fin: III + 22-24					
EGGS	Fig. A	YO	LK-SAC LARVAE Fig. B			
Habitat: pelagic Shape: quasi spherical Chorion: with filaments; diam. 0.77 mm Perivitelline space: small Yolk: segmented; unpigmented Oil globules: one; diam. 0.23 mm; red pigmented in live eggs Colour: transparent Note: eggs are gathered in groups joined by filaments that start from a single point in the chorion		Hatch size: about 2.5 mm Body: relatively elongate and slender Yolk sac: spherical Oil globule location: mid-dorsal region of yolk sac Anus: detached from yolk sac, does not reach finfold border Preanus length: about 67% SL Pigmentation: body and yolk sac unpigmented; oil globule red colored in live individuals				
LARVAE			Figs. C-G			
Body: relatively short and high; caudal peduncle long; 2 well developed dorsal fins in late larvae Head: moderately large Eye: round and large Gut: triangular Preanus length: 50% SL Air bladder: present Spination: a series of preopercular spines in larvae > 2.4 mm SL; at 6.5 mm, preoperculum with one long, thin spine in upper part, and 3 shorter, thin spines at bottom; supraorbital crest doubly serrated, weakly marked		Pigmentation: ventral side of trunk and tail, from pectoral–fin base to mid–tail, pigmented; some melanophores over head and a pair under gut; a group of 1-5 dorsal melanophores over tail opposed to ventral ones in larvae < 5.0 mm; pigmentation diminishes during development; live or newly formalin-preserved specimens strongly red colored Length at flexion: begins at 3.0 mm Length at transformation: unknown				
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3.5 mm SL



3.7 mm SL

Apogon imberbis (Linnaeus, 1758)



APOGONIDAE

GOBIIDAE









GOBIIFORMES

3.2 mm SL


Illustrations' sources: A-F: L. Rodríguez (A, E: redrawn from Padoa, 1956i; B, F: redrawn from Lebour 1919; C, D: redrawn from Alemany, 1997)

GOBIIDAE



4.5 mm SL

GOBIIFORMES



GOBIIDAE

GOBIIFORMES

Habitat: neritic, demersal, inshore, **Meristic characters** between 0 and 12 m depth Myomeres: 31 Distribution: eastern Atlantic Ocean, Vertebrae: 31 from Mauritania to Norway, and the 1st dorsal fin: VI Mediterranean Sea 2nd dorsal fin: I + 8-9 Spawning season: February to Anal fin: I+8-9 September YOLK-SAC LARVAE EGGS Fig. Habitat: demersal Hatch size: about 3.0 mm Body: elongate; mouth open; air bladder apparent Shape: pear-shaped, with rounded apex Yolk sac: rounded and relatively small Chorion: smooth; size 1.0–0.7 x 0.65–0.8 mm Anus: detached from yolk sac, reaches finfold Perivitelline space: small border Yolk: unsegmented; pigmented Preanus length: about 50% SL Oil globules: many; diam. to 0.08 mm Pigmentation: row of melanophores along **Colour:** translucent ventral side of trunk and tail; yolk sac and ventral side of gut strongly pigmented; large branched melanophore on dorsal mid-tail region; air bladder and eyes pigmented LARVAE Pigmentation: a row of postanal ventral

Body: relatively elongate and slender Head: relatively small; mouth small, oblique Eye: round Gut: elongated Preanus length: about 50% SL Air bladder: prominent Spination: none **Pigmentation:** a row of postanal ventral melanophores with a large one, ramified, at about mid-tail; a large branched melanophore on dorsal mid-tail region, opposite to ventral one; about 4 melanophores on ventral side of gut; a melanophore over anus; air bladder pigmented **Length at flexion:** unknown **Length at transformation:** unknown

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3.2 mm SL



4.0 mm SL

Pomatoschistus microps (Krøyer, 1838)

Common goby - Gobie commun



Pomatoschistus microps (Krøyer, 1838)



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BOTHIDA

Arnoglossus laterna (Walbaum, 1792) Mediterranean scaldfish - Arnoglosse de Méditerranée

Habitat: neritic, demersal, between 10 and 200 m depth

Distribution: eastern Atlantic Ocean, from Angola to Norway, and the Mediterranean Sea

Spawning season: April to August (North Sea and the Mediterranean Sea)

Meristic characters Myomeres: NA Vertebrae: NA Dorsal fin: 81-93 Anal fin: 75-82

Fig.



EGGS

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 0.60-0.76 mm Perivitelline space: small Yolk: unsegmented; pigmented Oil globules: one; diam. 0.11-0.15 mm; pigmented Colour: transparent

YOLK-SAC LARVAE Fig.

Hatch size: about 2.5 mm Body: elongate and slender Yolk sac: ovoid, elongated

Oil globule location: at posterior edge of yolk sac **Anus:** close behind yolk sac, reaches finfold border **Preanus length:** < 50% SL

Pigmentation: melanophores on head and dorsal anterior region of body; two opposite bars of melanophores a about mid-tail; dorsal primordial fin, yolk sac and oil globule pigmented; caudal region unpigmented

LARVAE

Body: initially elongate, becomes deeper, mainly through abdominal region, and laterally compressed with development; a tentacle over head develops from primordial fin (disappears at transformation) **Head:** moderately large with a snub nose; mouth small

Eye: round and relatively small

Gut: tightly coiled, extends beyond ventral margin of body

Preanus length: decreases with development from about 47% SL to 40% SL

Air bladder: present from about 4.5 mm

Figs. C-F

Spination: aggregates of simple spines on anal-fin base and over liver; 5-6 spines near base of tentacle Pigmentation: two rows of ventral melanophores, one on each side of body, from anus to caudal end; two opposite bars of melanophores at about mid-tail; melanophores along abdominal ventral contour of body, along lower jaw and over terminal gut; air bladder pigmented; primordialfin border pigmented in early larvae Length at flexion: unknown Length at transformation: about 18.0 mm (North Sea)

PHOTOS

by J.M. Rodriguez







BOTHIDAE

	Arnoglossus thori Kyle, 1913		Thor's scaldfish - Arnoglosse tacheté	
OTHIDAE	Habitat: neritic, demersal, between 15 and 100 m depth Distribution: eastern Atlantic, from Sierra Leone to Ireland, and the Mediterranean Sea Spawning season: April to July	Meristic character Myomeres: NA Vertebrae: NA Dorsal fin: 84-92 Anal fin: 63-69	rs	
m	EGGS		YOLK–SAC LARVAE Fig. A	
	Undescribed LARVAE Body: initially elongate, becomes deeper, mainly through abdominal region and laterally compressed with development; 1-2 tentacles develop over head from primordial fin and disappear at transformation Head: moderately large, with a snub nose; mouth small Eye: relatively small and round Gut: tightly coiled, extends beyond ventral margin of body Preanus length: about 43% SL Air bladder: present since early larvae Spination: aggregates of spined plates over		Hatch size: about 2.3 mm Body: elongate and slender Yolk sac: ovoid, very elongated Oil globule location: at posterior edge of yolk sac Anus: close behind yolk sac, reaches finfold border Preanus length: < 50% SL Pigmentation: two opposite accumulations of melanophores on postanal region that spread onto primordial fin, one at about mid-tail and another at caudal region; melanophores on head and on dorsal anterior region of body; oil globule pigmented	
				Figs. B-F
			abdo at ba Pigm behin a pat larva mela grou row o dorsa over Leng Leng	minal region, aligned along liver; some spines se of tentacle/s nentation: early larvae, a bar of melanophores nd mid-tail; a melanophore under urostyle; ch of pigment over coiled region of gut; late e, tail bar pigment reduced to a group of nophores over dorsal fin and an opposite p over anal fin, a short ventral line, and a of melanophores develops along anal fin and al-fin base, respectively; some melanophores gut loop; air bladder pigmented th at flexion: completed at 14.5 mm SL th at transformation: 21.0-25.0 mm SL

PHOTOS



5.9 mm SL

8.1 mm SL

by J.M. Rodriguez

Arnoglossus thori Kyle, 1913



Literature: Aldebert *et al.* (1990), Alemany (1997), Nielsen (1986a), Padoa (1956c), Russell (1976), Sabatés (1988) Illustrations' sources: A, F: Padoa (1956c); B-E: L. Rodríguez (redrawn from Alemany, 1997) BOTHIDAE

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BOTHIDA



3.9 mm SL



BOTHIDAE

Bothus podas (Delaroche, 1809)



CYNOGLOSSIDAE

Symphurus nigrescens Rafinesque, 1810 Tong Habitat: neritic and slope, demersal, Meristic characters

Myomeres: 48

Vertebrae: 48

Dorsal fin: 83-94

Anal fin: 71-78

Tonguesole - Plagusie sombre

Habitat: neritic and slope, demersal, between 20 and 1 140 m depth

Distribution: eastern Atlantic Ocean, from Angola to Portugal, and the Mediterranean Sea

Spawning season: January to May (Mediterranean Sea)

YOLK-SAC LARVAE

Undescribed

EGGS

LARVAE

Body: relatively short with deep abdominal region; laterally compressed from early larvae; first 4 dorsal-fin rays considerably enlarged (these rays enlarge in a sequential order, from anterior to posterior); pectoral fins large **Head:** large; snout rounded; mouth small and oblique **Eye:** round and small **Gut:** coiled, largely protruding; finger–shaped prolongation of gut at posterior ventral side of gut

loop; anus moderately protruding **Preanus length:** < 50% SL **Air bladder:** present

PHOTOS

Undescribed

Spination: none

Pigmentation: melanophores on ventral side of gut, on gut prolongation and over anus; a group of ventral melanophores on anterior tail and 2 isolated melanophores on ventral caudal region; opposing dorsal and ventral groups of melanophores at about mid–tail; groups of small melanophores along ventral tail region in later larvae; air bladder pigmented Length at flexion: unknown Length at transformation: unknown

by J.M. Rodriguez

not sized



not sized



6.0 mm SL



PLEURONECTIFORMES

Symphurus nigrescens Rafinesque, 1810



PLEURONECTIDAE







4.3 mm SL



7.3 mm SL

Platichthys flesus (Linnaeus, 1758)



PLEURONECTIDAE

SCOPHTHALMIDAE

Lepidorhombus boscii (Risso, 1810) Four-spot megrim - Cardine à quatre taches Habitat: neritic and upper slope, **Meristic characters** demersal, between 7 and 800 m depth Myomeres: 41 Distribution: eastern Atlantic Ocean, Vertebrae: 41 from Cape Bojador to the British Isles, Dorsal fin: 82-89 and the Mediterranean Sea Anal fin: 65-71 Spawning season: March to June All And All An YOLK–SAC LARVAE EGGS Undescribed Undescribed LARVAE **Body:** relatively elongate and slender in early larvae, becomes considerably deep, especially through abdominal region, and laterally compressed with development eye, at 6.0 mm Head: relatively small; mouth becomes almost vertical with development; mouth protractile in late larvae

Eye: round and relatively small, begins migrating to left side of body when larva is about 8.9 mm long Gut: spherical, coiled, extends beyond ventral margin of body

Preanus length: increases with development from 20.2% SL in early larvae to 44.2% in late larvae Air bladder: present

Spination: two rows of preopercular spines (larger spines in posterior row) visible in 5.0 mm larvae; a spine appears on post-temporal region, behind

Pigmentation: 4 groups of melanophores evenly arranged along dorsal side of body, and 3 (2 in early larvae) postanal ventral melanophores extending to finfold and later to dorsal and anal fins; gut, head, frontal region, tips of upper and lower jaws and air bladder pigmented; melanophores on dorsal and ventral surface of gut and on cleithral symphysis; pelvic fins pigmented Length at flexion: 6.0-9.8 mm SL Length at transformation: unknown

PHOTOS



LEURONECTIFORMES

3.2 mm SL



6.9 mm SL

by J.M. Rodriguez



5.2 mm SL





depth

Lepidorhombus whiffiagonis (Walbaum, 1792)

Megrim – Cardine franche

Habitat: neritic and upper slope, demersal, between 100 and 700 m

Distribution: eastern Atlantic Ocean, from Cape Bojador to Iceland, and the western Mediterranean Sea

Spawning season: March to June

Meristic characters Myomeres: 42 Vertebrae: 42 Dorsal fin: 85-94 Anal fin: 64-74

Fig. A



EGGS

Habitat: pelagic

YOLK-SAC LARVAE

Preanus length: about 45% SL

pigmented; oil globule pigmented

Hatch size: about 4.05 mm

Body: elongated

finfold border

Yolk sac: large, ovoid

Shape: spherical Chorion: smooth; diam. 1.02-1.22 mm Perivitelline space: small Yolk: unsegmented; pigmented Oil globules: one; diam. 0.25-0.30 mm; pigmented Colour: transparent

by J.M. Rodriguez

LARVAE

Body: relatively elongate and slender in early larvae, becomes deep, especially through abdominal region, and laterally compressed with development Head: relatively small; mouth oblique and relatively large; snout pointed in early larvae becomes concave throughout development Eye: round and small; asymmetry begins in larvae 12.7 mm long Gut: spherical, coiled, extends beyond ventral margin of body

Preanus length: about 50% SL Air bladder: absent

Spination: 2 large otocystic spines on each side of head

Oil globule location: at posterior edge of yolk sac

Anus: slightly detached from yolk sac, reaches

Pigmentation: a few melanophores on head; trunk and tail, except caudal region, covered with uniformly scattered melanophores; dorsal and ventral margins of finfold, except caudal region,

Pigmentation: melanophores evenly scattered over dorsal and anal fins; two parallel rows of large melanophores along dorsal (from head) and ventral (from anus) contours of body; peritoneum and ventral side of gut pigmented; some melanophores on head and on upper and lower jaws

Length at flexion: < 11.7 mm Length at transformation: unknown

PHOTOS

3.8 mm SL 3.7 mm SL (dorsal view) 4.6 mm SL 5.1 mm SL

SCOPHTHALMIDAE





SCOPHTHALMIDAE

Scophthalmus rhombus (1	Linnaeus, 1758)	Brill - Barbue		
Habitat: neritic, demersal, between 5 and 50 m depth Distribution: eastern Atlantic Ocean, from Morocco to Iceland, and the Mediterranean Sea Spawning season: March to August (Mediterranean Sea)	Meristic characters Myomeres: 25-36 Vertebrae: 35-36 Dorsal fin: 74-80 Anal fin: 55-62	s		
EGGS	Fig. A	YOLK-SAC LARVAE Fig. B		
Habitat: pelagic Shape: spherical Chorion: smooth; diam. 1.24-1.50 mm Perivitelline space: small Yolk: unsegmented; pigmented Oil globules: one; diam. 0.16-0.25 mm; pigmented Colour: transparent		Hatch size: about 3.8 mm TL Body: elongated and slender Yolk sac: ovoid, pigmented Oil globule location: at ventral, posterior edge of yolk sac Anus: close behind yolk sac, reaches finfold border Preanus length: about 50% SL Pigmentation: body (except caudal region) and yolk sac pigmented; melanophores extend onto finfold to form a wide postanal bar and a dorsal patch above gut		
LARVAE		Figs. C-F		
Body: relatively elongate and slende larvae, becomes considerably deep, gut region, laterally compressed and development Head: moderately large Eye: round, begins migration to left larvae of about 8.9 mm Gut: spherical, coiled, extends beyon margin of body Preanus length: about 50% SL	er in early especially in d ovoid with side of body in nd ventral	Air bladder: present, disappears after metamorphosis Spination: opercular spines and a ridge above eye from larvae 6.0 mm long Pigmentation: body covered with melanophores (caudal region free of pigment in early larvae); body pigment extends onto dorsal and anal fins forming almost regular bands in later larvae Length at flexion: unknown Length at transformation: unknown		

PHOTOS

PLEURONECTIFORMES

by J.M. Rodriguez





2.8 mm SL

3.1 mm SL



4.1 mm SL

SCOPHTHALMIDAE





Zeugopterus regius (Bonnaterre, 1788) Habitat: neritic, demersal, between 0 and 180 m depth **Distribution:** eastern Atlantic Ocean, from Morocco to the British Isles, and the Mediterranean Sea Spawning season: May to August (British Isles)

Meristic characters Myomeres: NA Vertebrae: NA Dorsal fin: 77 Anal fin: 66

Eckström's topknot



EGGS

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 0.90-0.99 mm Perivitelline space: small Yolk: unsegmented; pigmented Oil globules: one; diam. 0.16-0.18 mm; pigmented Colour: transparent

Body: elongate and slender Yolk sac: ovoid Oil globule location: ventral, posterior in yolk sac Anus: detached from yolk sac, reaches finfold border **Preanus length:** > 50% SL

Pigmentation: body, yolk sac and finfold covered with small melanophores; oil globule pigmented

LARVAE

Body: relatively elongate and slender in early larvae, becomes relatively deep, especially in gut area, laterally compressed and ovoid with development

Head: relatively large; mouth relatively elongated in early larvae becomes almost vertical with development

Eye: round and relatively small;

Gut: triangular, coiled, extends beyond ventral margin of body

PHOTOS

Preanus length: about 50% SL Air bladder: absent

Hatch size: about 2.4 mm

Spination: 2 large otocystic spines on each side of head

Pigmentation: body and fins covered with many small, uniformly distributed melanophores Length at flexion: urostyle turned up at 8.0 mm Length at transformation: unknown

by J.M. Rodriguez









6.7 mm SL

SCOPHTHALMIDAE





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SOLEIDA

Buglossidium luteum (Risso, 1810) Solenette - Petite sole jaune Habitat: neritic, upper slope, **Meristic characters** demersal, between 5 and 450 m depth Myomeres: 36-38 Distribution: eastern Atlantic Ocean, Vertebrae: 36-38 from Angola to Scotland, and the Dorsal fin: 65-78 Mediterranean Sea Anal fin: 49-63 Spawning season: March to June (Bay of Biscay) YOLK-SAC LARVAE EGGS Fig. Habitat: pelagic, Hatch size: about 2.0 mm Body: slender Shape: spherical Yolk sac: large, ovoid Chorion: smooth; diam. 0.64-0.94 mm

Chorion: smooth; diam. 0.64-0.94 mi Perivitelline space: small Yolk: segmented; pigmented Oil globules: 12-15; unpigmented Colour: transparent

Oil globule location: regularly distributed over yolk sac

Anus: close behind yolk sac, does not reach finfold border

Preanus length: about 42% SL

Pigmentation: melanophores irregularly distributed over body in recently hatched larvae; late yolk-sac larvae have 4 spots on dorsal and 1-2 on ventral body margins; some melanophores on primordial fin margins; yolk sac pigmented

LARVAE

Body: initially elongated becomes short and laterally compressed with development Head: moderately large; snout rounded Eye: round; asymmetry begins at about 6.0 mm SL, almost completed at 8.0 mm SL Gut: coiled, extends quite beyond ventral margin of body Preanus length: < 50% SL Air bladder: prominent

Spination: none

Pigmentation: evenly spaced melanophores along dorsal (9–13) and ventral (8–11) body contours; top of head, ventral abdominal region, air bladder and pectoral fins pigmented **Length at flexion:** unknown

Length at transformation: probably completed at about 10.0 mm

PHOTOS

by J.M. Rodriguez

Figs. C-F





7.4 mm SL



Buglossidium luteum (Risso, 1810)



Microchirus variegatus (Donovan, 1808)Thickback sole - Sole-pedrix communeHabitat: neritic and slope, demersal,
between 80 and 400 m depthMeristic characters
Myomeres: 36-40
Vertebrae: 36-40
Dorsal fin: 63-80
Anal fin: 47-64Meristic und the sole - Sole-pedrix commune

Fig. A

YOLK–SAC LARVAE Fig. F

EGGS

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 1.16-1.42 mm Perivitelline space: small Yolk: segmented; pigmented Oil globules: 30-50 of different sizes; unpigmented Colour: transparent Hatch size: about 2.5 mm Body: relatively elongate and slender Yolk sac: large, rounded Oil globule location: regularly distributed over yolk sac Anus: close behind yolk sac, reaches finfold border Preanus length: < 50% SL

Pigmentation: body, primordial fin and yolk sac covered with small, stellate melanophores; rows of stellate melanophores along dorsal and ventral margins of primordial fin

LARVAE

Body: short with a prominent head and deep abdominal region Head: moderately large and rounded; mouth small and oblique Eye: round and small Gut: spherical, tightly coiled, extends far beyond ventral margin of body Preanus length: about 50% SL Air bladder: absent Spination: none

Pigmentation: body and fins covered with small stellate or simple rather evenly spaced melanophores; dorsal (about 70) and ventral (about 50) rows of relatively large melanophores along body contours, these melanophores are larger and tend to merge and form continuous rows in late larvae **Length at flexion:** unknown **Length at transformation:** 9.0-12.0 mm

PHOTOS

LEURONECTIFORMES

Length at transfo

by J.M. Rodriguez







Pegusa lascaris (Risso, 1810)

Habitat: neritic and upper slope,

demersal, between 0 and 350 m depth

Distribution: eastern Atlantic Ocean,

Spawning season: May to September

from South Africa to the North Sea,

and the Mediterranean Sea

(Atlantic Iberian peninsula)

Sand sole - Sole-pole

Meristic characters Myomeres: 45-47 Vertebrae: 45-47 Dorsal fin: 70-90 Anal fin: 58-75



YOLK-SAC LARVAE Fig. 1

EGGS

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 1.28-1.38 mm Perivitelline space: small Yolk: segmented; may be slightly pigmented Oil globules: up to 50 or more Colour: transparent

Hatch size: < 4.0 mm

Body: relatively elongate and slender; dorsal finfold extends well over head (larva with a hooded appearance); air bladder visible in late yolk-sac larvae Yolk sac: ovoid Oil globules location: some in small groups, some scattered over yolk sac Anus: slightly detached from yolk sac, reaches finfold border Preanus length: < 50% SL Pigmentation: irregularly scattered melanophores

over head, body and finfold in early yolk-sac larvae; melanophores aggregated over body and finfold forming a postanal band in late yolk-sac larvae; scattered melanophores on yolk sac

Figs. C-F

LARVAE

Body: relatively short with deep abdominal region; laterally compressed; dorsal finfold extends well over head (larva with a hooded appearance) in early larvae

Head: moderately large; mouth small; lower jaw protruding

Eye: round and small

Gut: spherical, coiled, extends far beyond ventral margin of body **Preanus length:** about 45% SL Air bladder: prominent since early larvae Spination: none

Pigmentation: similar to yolk–sac larvae in early larvae; melanophores form an anterior band on dorsal fin, above anus in late larvae; pigmentation increases through development; air bladder pigmented

Length at flexion: begins at about 7.5 mm **Length at transformation:** unknown

PHOTOS

by J.M. Rodriguez





8.8 mm SL

PLEURONECTIFORMES



Illustrations' sources: A: Padoa (1956o); B-F: Nichols (1976)

Habitat: neritic, demersal, between 0 **Meristic characters** and 150 m depth Myomeres: 46-52 Vertebrae: 46-52 Dorsal fin: 69-97 Anal fin: 53-79 YOLK-SAC LARVAE EGGS Fig. A Habitat: pelagic Hatch size: 2.5-3.8 mm Body: relatively elongate and slender Shape: spherical Yolk sac: spherical and large Chorion: smooth; diam. 0.95-1.58 mm Perivitelline space: small posterior periphery of yolk sac Yolk: segmented; pigmented Oil globules: many, small, aggregated in clusters; finfold border unpigmented

Colour: transparent

Oil globule location: in clusters on dorsal and **Anus:** slightly detached from yolk sac, reaches Preanus length: 50% SL Pigmentation: body (except caudal region), primordial fin and yolk sac covered with small

branched melanophores; rows of melanophores along dorsal body contour (9-14), extending from head to caudal region and along ventral body contour (4-12), extending from abdominal region to caudal region

LARVAE

Body: short with deep abdominal region Head: moderately large Eye: round and small Gut: spherical, coiled, extends far beyond ventral margin of body Preanus length: about 50% SL **Air bladder:** visible in larvae > 4.0 mm SL Spination: none

Pigmentation: similar to yolk-sac larvae in early larvae; body covered with large stellate melanophores; dorsal and ventral rows of melanophores almost forming an unbroken line of pigment in late larvae Length at flexion: unknown Length at transformation: 7.0-9.5 mm

PHOTOS

LEURONECTIFORMES

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by J.M. Rodriguez



4.5 mm SL



Distribution: eastern Atlantic Ocean, from Senegal to the North Sea, and the Mediterranean Sea Spawning season: January to April

Solea solea (Linnaeus, 1758)

(Mediterranean Sea)



PLEURONECTIFORMES



Illustrations' sources: A-F: Padoa (1956o), Quero *et al.* (1986b), Russel

CARANGIDAE

Chloroscombrus chrysurus (Linnaeus, 1766)

Atlantic bumper - Sepater

Habitat: neritic, benthopelagic, between 0 and 110 m depth

Distribution: Atlantic Ocean (some records from the Mediterranean Sea). Eastern Atlantic, from Angola to Spain Meristic characters Myomeres: 24 Vertebrae: 24 1st dorsal fin: VIII 2nd dorsal fin: I + 25-28 1st anal fin: II 2nd anal fin: I + 25-28

Spawning season: probably summer (Gulf of Mexico)

EGGS

YOLK-SAC LARVAE Undescribed

LARVAE

Undescribed

Body: moderately deep

Head: moderately large, snout moderately pointed; mouth oblique reaches anterior part of eye

Eye: round and relatively large

Gut: triangular

Preanus length: > 50% SL

Air bladder: present

Spination: supraoccipital crest appears at 1.8 mm SL; preopercular spines present, with a prominent spine at preopercle angle; a small supraocular spine on a weak ridge; tiny post-temporal and supracleithral spines

Pigmentation: early larvae, 3-4 dorsal melanophores over trunk; a melanophore on top of head; tips of jaws, peritoneum and dorsal surface of air bladder pigmented; a melanophore at angle of lower jaw; melanophores anterior to cleithral symphysis and on ventral surface of gut; row of postanal ventral melanophores; short series of melanophores on lateral midline (absent in very early larvae); late larvae, pigmentation increases with development extending over most of body; ventrolateral melanophores aligned with myosepta Length at flexion: 4.0-5.0 mm SL Length at transformation: 10.0-15.0 mm SL

PHOTOS



3.2 mm SL

4.9 mm SL



Chloroscombrus chrysurus (Linnaeus, 1766)



Illustrations' sources: A-G: L. Rodríguez (A-C: redrawn from Laroche *et al.*, 2006; D-G: redrawn from Sánchez-Ramírez and Flores-Coto, 1993)

CARANGIDAE

CARANGIFORMES

Decapterus punctatus Cuvier, 1829

Round scad - Comète quiaquia

CARANGIDA Habitat: neritic, benthopelagic, **Meristic characters** between 0 and 100 m depth Myomeres: 25 Distribution: Atlantic Ocean. Eastern Vertebrae: 25 Atlantic, from South Africa to 1st dorsal fin: VIII Morocco (unconfirmed records from 2nd dorsal fin: I + 30-34 the Mediterranean Sea) 1st anal fin: II Spawning season: year-round 2nd anal fin: I + 26-29 (western Atlantic Ocean) YOLK-SAC LARVAE



EGGS

Undescribed

Undescribed

LARVAE

Body: deep in early larvae, becomes shallower with development

Head: large, deep, and relatively pointed; snout relatively concave in early larvae, becomes straight with development; mouth oblique

Eye: round and large, increases in relative size with development

Gut: triangular

Preanus length: around 50% SL Air bladder: absent

PHOTOS

Spination: an orbital and a prominent supraoccipital crest; preopercular spines present, with a long spine at preopercle angle Pigmentation: light pigmentation over most of body until late postflexion; a few melanophores on top of head and jaws, infrequently on snout and cheeks; parallel rows of melanophores along bases of dorsal and anal fins; row of melanophores along midline of tail; peritoneum pigmented Length at flexion: 4.0-6.0 mm SL Length at transformation: about 9.0-15.0 mm SL

by S. Isari





3.8 mm SL

6.2 mm SL



10.8 mm SL

CARANGIFORMES





CARANGIDAE

Selene setapinnis (Mitchill, 1	Atlantic moonfish - Musso atlantique	
 Habitat: neritic, inshore, between 0 and 50 m depth Distribution: western Atlantic Ocean. Replaced by <i>S. dorsalis</i>* in the Eastern Atlantic Ocean, from South Africa to Morocco Spawning season: unknown 	Meristic characters Myomeres: 24 Vertebrae: 24 1 st dorsal fin: VIII 2 nd dorsal fin: I + 21-24 1 st anal fin: II 2 nd anal fin: I + 16-19	
EGGS	YO	OLK-SAC LARVAE
Undescribed	Und	escribed

LARVAE

Body: moderately deep during preflexion stage, increases with development to 72.4% SL in postflexion stage; dorsal-fin spines and pelvic fin very long

Head: moderately large, stout, rounded, increases in length with development; snout relatively concave in early larvae; mouth small and oblique **Eye:** round and relatively small

Gut: triangular

Preanus length: decreases from about 51-57% SL in preflexion larvae to 49.6% SL in postflexion larvae **Air bladder:** present

Spination: supraoccipital crest precocious and small, disappears in larvae \geq 5.25 mm SL; low supraocular ridge with a simple spine; 5-11

preopercular spines, with spine at angle slightly longer; single, small post-temporal spine; single, small supracleithral spine present in larvae $\geq 4.0 \text{ mm SL}$

Pigmentation: relatively scant in all larval stages; early larvae, head generally unpigmented; a postanal, ventral row of melanophores, which disappears in late larvae; pigment on dorso-lateral part of body consists of scattered spots on nape and under first and second dorsal fins; peritoneum and air bladder pigmented; pelvic fins and first dorsal-fin membrane pigmented from 3.4 mm SL; first dorsal-fin rays pigmented by 4.25 mm SL **Length at flexion:** about 4.3-6.3 mm SL **Length at transformation:** about 11.0 mm SL

**S. setapinnis* was cited for Mauritania by Maigret and Ly (1986). *S. setapinnis* and *S. dorsalis* have not been adequately studied and may prove to be conspecific (Maigret and Ly, 1986).

PHOTOS



3.9 mm SL

4.7 mm SL

by S. Isari
CARANGIDAE



Literature: Aboussouan (1975), Fahay (2007), Froese and Pauly (2022), Katsuragawa (1997), Laroche *et al.* (2006), Sánchez-Ramírez and Flores-Coto, (1993)

Illustrations' sources: A-F: L. Rodríguez (A, B: redrawn from Katsuragawa, 1997; C-E: redrawn from Sanchez-Ramirez and Flores-Coto, 1993; F: redrawn from Laroche *et al.*, 2006)

CARANGIFORMES

Trachurus mediterraneus (Steindachner, 1868) Chinchard à queue jaune Habitat: neritic and upper slope, **Meristic characters** benthopelagic, between 40 and 500 m Myomeres: 24 depth Vertebrae: 24 Distribution: eastern Atlantic Ocean, 1st dorsal fin: VIII from Mauritania to the Bay of Biscay, **2**nd **dorsal fin:** I + 29-35 and the Mediterranean Sea 1st anal fin: II Spawning season: spring and 2nd anal fin: I + 26-39 summer YOLK-SAC LARVAE EGGS Habitat: pelagic Hatch size: may be about 2.0 mm Body: relatively slender Shape: spherical Yolk sac: large, ovoid, projected a little beyond Chorion: smooth; diam. 1.00-1.04 mm snout Perivitelline space: small Oil globule location: at anterior end of yolk sac Yolk: segmented; unpigmented Anus: detached from yolk sac, reaches finfold border Oil globules: one; diam. 0.24 mm; pigmented Preanus length: around 50% SL Colour: transparent Pigmentation: 3-4 dorsal, preanal melanophores, some along ventral profile of trunk and tail; oil globule pigmented; finfold unpigmented LARVAE Figs. B-F **Body:** relatively slender in early larvae, gradually Spination: an occipital crest and two series of deepens and tapers to relatively narrow caudal preopercular spines peduncle; very similar to Trachurus trachurus **Pigmentation:** 3-4 large preanal, dorsal (differences between the two species mainly lie on melanophores; peritoneal region pigmented; a melanophore over terminal gut; a postanal row pigmentation patterns) Head: large, moderately pointed; mouth oblique of dotted melanophores; during development, Eye: round and relatively large melanophores appear over lateral body walls, over Gut: triangular head, dorsum of tail, under gut and lower jaw; air Preanus length: about 50% SL bladder pigmented Air bladder: present

Length at flexion: almost completed at 4.8 mm SL Length at transformation: unknown

PHOTOS

CARANGIFORMES

by J.M. Rodriguez

Mediterranean horse mackerel









3.1 mm SL

CARANGIDA



Illustrations' sources: A-F: L. Rodríguez **(A:** redrawn from Alemany, 1997; **B-F:** redrawn from Demir, 1961)

Trachurus trachurus (Linnaeus, 1758) Atlantic horse mackerel – Chinchard d'Europe Habitat: neritic, benthopelagic, **Meristic characters** between 40 and 500 m depth (usually Myomeres: 24 100-200 m depth) Vertebrae: 24 Distribution: eastern Atlantic Ocean, 1st dorsal fin: VIII from Cape Verde to Norway, and the 2nd dorsal fin: I +29-33 Mediterranean Sea 1st anal fin: II Spawning season: probably all year-2nd anal fin: I + 24-29 round

Fig. A

EGGS

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 0.81-1.04 mm Perivitelline space: small Yolk: segmented; unpigmented Oil globules: one; diam. 0.19-0.23 mm; pigmented Colour: transparent

YOLK-SAC LARVAE

Hatch size: about 2.2 mm Body: relatively elongate and slender Yolk sac: ovoid, projected beyond snout Oil globule location: at anterior end of yolk sac Anus: detached from yolk sac, reaches finfold border Preanus length: > 50% SL

Pigmentation: melanophores are irregularly spread over body, except in caudal region; melanophores on dorsal finfold; oil globule pigmented

LARVAE

Body: relatively elongate in early larvae, gradually deepens and tapers to narrow caudal peduncle; very similar to *T. mediterraneus* (differences between two species mainly lie on pigmentation patterns) **Head:** large, snout moderately pointed; mouth oblique **Eye:** round and relatively large **Gut:** triangular

Preanus length: > 50% SL

Air bladder: present Spination: an occipital crest and two series of preopercular spines with spine at angle in outer series longer

Pigmentation: dorsal and ventral body rows of melanophores; dorsal row of about 10 melanophores, ends at about mid tail; ventral row consists of 4-5 large postanal melanophores, followed by small melanophores extending to nearly notochord end; melanophores on lower jaw, ventral surface of abdomen and over head; peritoneum and air bladder pigmented **Length at flexion:** may begin at about 5.0 mm SL **Length at transformation:** unknown

PHOTOS

by J.M. Rodriguez





2.7 mm SL

3.4 mm SL



5.3 mm SL

CARANGIDAE

Trachurus trachurus (Linnaeus, 1758)



CARANGIFORMES

Coryphaena hippurus Linnaeus, 1758 Common dolphinfish – Coryphène commune Habitat: neritic and oceanic, pelagic **Meristic characters Distribution:** worldwide, in tropical Myomeres: 30-31 and subtropical waters, and the Vertebrae: 30-31 Mediterranean Sea Dorsal fin: 58-66 Spawning season: May to September Anal fin: 25-31

YOLK-SAC LARVAE

Hatch size: about 4.0 mm TL

Preanus length: about 60% SL

Body: elongate

Yolk sac: ovoid

yolk sac

border

Fig.

EGGS Fig. Habitat: pelagic Shape: spherical Chorion: smooth; diam. 1.3 mm Perivitelline space: small Yolk: segmented; pigmented Oil globules: one; diam. 0.3-0.4 mm; pigmented Colour: transparent

Body: elongate Head: moderately blunt; mouth oblique Eye: round Gut: elongate, tubular in early larvae **Preanus length:** about 60% SL Air bladder: absent Spination: preopercular, supraocular, post-temporal, ptreotic and articular spines (see illustration H)

Pigmentation: body heavily pigmented; melanophores arranged in bars in late larvae; caudal region unpigmented in early larvae; pelvicfin rays pigmented in larvae > 8.0 mm SL Length at flexion: 7.5-9.0 mm SL Length at transformation: 25.0-30.0 mm SL

Oil globule location: at ventral, posterior end of

Pigmentation: body (except caudal region), yolk

sac and oil globule strongly pigmented

Anus: detached from yolk sac, reaches finfold

PHOTOS

LARVAE

by J.M. Rodriguez



3.8 mm SL



mid-sized larva

CORYPHAENIDAE



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CORYPHAENIDAE

CARANGIFORMES

SPHYRAENIDAE



SPHYRAENIDAE

Sphyraena guachancho Cuvier, 1829



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Xiphias gladius Linnaeus, 1758 Swordfish - Espadon Habitat: oceanic, pelagic and **Meristic characters** mesopelagic, to 800 m depth Myomeres: 24 Distribution: worldwide in tropical-Vertebrae: 24 temperate waters 1st dorsal fin: 38-45 Spawning season: year-round 2nd dorsal fin: 4-5 Anal fin: 12-16 EGGS YOLK-SAC LARVAE Habitat: pelagic Hatch size: about 4.2 mm Body: elongate Shape: spherical Yolk sac: elongated, large Chorion: smooth; diam. 1.6-1.8 mm Oil globule location: at ventral, posterior edge of Perivitelline space: small yolk sac Yolk: segmented; pigmented Anus: detached from yolk sac, reaches finfold Oil globules: one; diam. 0.40-0.52 mm; pigmented border Colour: transparent Preanus length: about 67% SL Pigmentation: body and finfold covered with small melanophores; oil globule and yolk sac pigmented LARVAE Body: relatively elongate preopercular spines large, often slightly curved, Head: shallow; mouth very large with very with 2 spines at angle longer; supraocular crest with strong spines; 1-2 strong pteroptic spines; elongated jaws; teeth well developed from larvae of 2 post-temporal spines, increasing to 3; spinous 6.0 mm SL scales appear at 12.0-15.0 mm SL Eye: round and large Pigmentation: scattered melanophores over body, Gut: bulky and elongated except caudal peduncle Preanus length: about 70-80% SL, increases with Length at flexion: 8.0-12.0 mm SL development Length at transformation: unknown Air bladder: absent

PHOTOS

Spination: frontal crest on dorsum, anterior to eye;

by J.M. Rodriguez



3.7 mm SL

Not sized



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Ľ XIPHIIDA



XIPHIIDAE

<i>Chromis chromis</i> (Linnaeus, 1758)			Damselfish - Castagnole	
Habitat: neritic, in midwater, between 3 and 35 m depth Distribution: eastern Atlantic Ocean, from Angola to Portugal, and the Mediterranean Sea Spawning season: summer	Meristic character Myomeres: 26 Vertebrae: 26 Dorsal fin: XIV + 8 Anal fin: II + 9-11	r s 3-11		
EGGS	Fig. A	YO	LK–SAC LARVAE Fig. B	
Habitat: demersal, attached to hard substrates Shape: elliptical Chorion: with adhesive filaments; diam. 0.85–0.90 x 0.70–0.72 mm Perivitelline space: small Yolk: segmented; pigmented Oil globules: one; 0.20 mm; unpigmented Colour: transparent		Hatch size: about 2.6 mm TL Body: elongate and slender Yolk sac: ovoid, pigmented Oil globule location: at anterior, ventral edge of yolk sac Anus: slightly detached from yolk sac, reaches finfold border Preanus length: about 33% SL Pigmentation: row of postanal ventral melanophores; occipital melanophores; yolk sac pigmented		
LARVAE			Figs. C-F	
Body: elongate and laterally compressed in early larvae, deepens after flexion becoming moderately stocky Head: deep with rounded snout in early larvae, becomes slightly pointed in late larvae; mouth protractile and oblique Eye: round and large Gut: triangular and bulky; anus moderately protruding Preanus length: < 50% SL Air bladder: absent		Pign larva mela a ver mela dors a do: mela Leng comj Leng	nentation: similar to yolk-sac larva in early ne; with development, postanal-ventral nophores reduce and concentrate, forming ntral bar at about mid-tail; in late larvae, nophores appear on mid-lateral line and on um, at same level of ventral group forming rsum-ventral bar; peritoneum pigmented; nophores on head gth at flexion: begins at 3.60 mm SL and is pleted at 4.75 mm SL gth at transformation: unknown	

Air bladder: absent Spination: none

<u>PHOTOS</u>

CICHLIFORMES

by J.M. Rodriguez





2.4 mm SL

3.9 mm SL



POMACENTRIDAE



Literature: Fage (1918), Froese and Pauly (2022), Padoa (1956m), Quignard and Pras (1986b), Sabatés (1988) Illustrations' sources: A, B: Padoa (1956m); C, D: Alemany (1997); E, F: Fage (1918)

BELONIDAE

Belone belone (Linnaeus, 1761) Garfish - Orphi				
Habitat: neritic, epipelagic Distribution: eastern Atlantic Ocean, from Cape Verde to Norway, and the Mediterranean Sea Spawning season: February to May (Mediterranean Sea)	Meristic character Myomeres: 75-84 Vertebrae: 75-84 Dorsal fin: 16-20 Anal fin: 19-23	°S	All and a second	
EGGS	Fig. A	YO	LK–SAC LARVAE Fig. B	
Habitat: demersal Shape: spherical Chorion: with filaments; diam. 3.0-3.5 mm Perivitelline space: small Yolk: unsegmented; pigmented Oil globules: none Colour: transparent		Hatc deve Body finfo Heac beyo Yolk Anus bord Preas Pigm mela pigm	h size: about 9.0 mm in an advanced stage of lopment y: elongate and relatively slender; no dorsal ld; large ventral finfold d: relatively small; lower jaw slightly projected nd upper jaw; mouth open sac: elongated s: detached from yolk sac, reaches finfold er nus length: about 72% SL nentation: body completely covered with nophores; yolk sac barely visible due to nentation; eye pigmented	
LARVAE			Figs. C-E	
Body: elongate and slender Head: small; mouth small, lower jaw very large, strongly projected beyond upper jaw Eye: round and large		Air b Spin Pign arrar	bladder: small ation: none nentation: strongly pigmented; melanophores nged along myomeres in dorsal half of body	

PHOTOS

Gut: elongate, tube-like **Preanus length:** about 77% SL

by J.M. Rodriguez



Length at flexion: unknown

Length at transformation: unknown

13.0 mm SL



BELONIFORMES

Not sized

BELONIDAE



Illustrations' sources: A, C-E: modified from Russell (1976); B: D'Ancona (1931e)

<i>Scomberesox saurus</i> (Walbaum, 1792)			Atlantic saury - Balaou atlantiq	ue
Habitat: oceanic, epipelagic (usually in the very upper layer) Distribution: North Atlantic Ocean. Eastern Atlantic, from Morocco to Iceland, and the Mediterranean Sea Spawning season: year-round in waters between 16.5 °C and 23.5 °C (western Atlantic Ocean)	Meristic character Myomeres: 64-68 Vertebrae: 64-68 Dorsal fin: 9-12 Dorsal finlets: 5-6 Anal fin: 12-13 Anal finlets: 5-7	s	CO STA	¢
EGGS	Fig. A	YO	LK-SAC LARVAE Fig.	B
EGGSFig. AHabitat: pelagicShape: slightly ovalChorion: with rigid, uniformly-spaced bristles;max. diam. 2.15-2.76 mmPerivitelline space: smallYolk: unsegmented; unpigmentedOil globules: noneColour: transparent		Hatch size: 6.0-8.5 mm Body: relatively elongate and slender; caudal fin well developed and flexion underway at hatching Yolk sac: elongated Anus: detached from yolk sac, reaches finfold border Pigmentation: body unpigmented at hatching; melanophores develop early; dense, deep- blue coloring (black in preserved organisms) over entire body, excluding fins and yolk; eyes pigmented at hatching Preanus length: about 60% SL		
LARVAE			Figs. C	-F
Body: elongate, slender and cylindrical; dorsal and		Preanus length: 60-70% TL		

Body: elongate, slender and cylindrical; dorsal and anal fins opposite, located posteriorly on body; preanal finfold persistent; finlets form in larvae of about 25.0 mm SL **Head:** relatively large; snout very short; mouth small and oblique Preanus length: 60-70% TL Air bladder: absent Spination: none Pigmentation: dorsum dark-blue in live larvae (black in preserved larvae), flanks silvery Length at flexion: 4.4 mm Length at transformation: about 25.0 mm

Gut: elongate, tube-like

Eye: round and large

PHOTOS





by J.M. Rodriguez



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BELONIFORMES



Illustrations' sources: A, B: L. Rodríguez (A: redrawn from Collette *et al.*, 1984a; **B:** redrawn from Hardy and Collette, 2003); **C:** Sanzo (1940); **D-F:** D'Ancona (1931e)

2

SCOMBERESOCIDAE

BELONIFORMES



LARVAE

Body: moderately elongate and slender in early larvae, deepens through pectoral and abdominal regions with development; pectoral fins large (reaching anus) and rounded; 12 pectoral-fin rays **Head:** moderately large and rounded; mouth small; teeth apparent in late larvae **Eye:** round and large **Gut:** short and triangular

Preanus length: increases with development to about 50% SL

PHOTOS

Air bladder: absent Spination: none

Pigmentation: some melanophores on head; peritoneum heavily pigmented; 5-6 ventral melanophores at posterior half of postanal region; pectoral fins large, rounded, heavily pigmented with melanophores located between fin rays **Length at flexion:** completed at 7.5 mm **Length at transformation:** unknown

by J.M. Rodriguez

Figs. B-F



3.5 mm SL



3.9 mm SL

3.6 mm SL



Literature: Fives (1986), Ford (1922), Padoa (1956b), Russell (1976), Zander (1986) Illustrations' sources: A: Cipria (1938); B, D-F: Ford (1922); C: Alemany (1997) ш

BLENNIIDA



Preanus length: increases with development to about 50% SL

Length at transformation: unknown

PHOTOS

by J.M. Rodriguez







5.6 mm SL



9.0 mm SL

Lipophrys pholis (Linnaeus, 1758)



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BLENNIIDA

Parablennius gattorugine (Linnaeus, 1758) **Tompot blenny** Habitat: neritic, demersal, between 3 Meristic characters and 32 m depth Myomeres: NA Distribution: eastern Atlantic Ocean, Vertebrae: NA from South Guinea to the Bay of Dorsal fin: XIII + 18-19 Biscay, and the Mediterranean Sea Anal fin: II + 19-20 Spawning season: March to May EGGS YOLK-SAC LARVAE Habitat: demersal, adherent, in a single layer Hatch size: about 4.5 mm Body: elongate; pectoral fins apparent and rounded Shape: hemispherical Yolk sac: no information Chorion: diam. 1.6 mm Anus: no information Perivitelline space: small **Preanus length:** < 30% SL Yolk: unsegmented Pigmentation: dorsal side of gut, from behind eye Oil globules: none to anus, pigmented; a few melanophores on snout Colour: purple and gold in live eggs and top of head; pectoral fins unpigmented; eye

pigmented

LARVAE

Body: moderately elongate and slab-sided; caudal peduncle deep; dorsal and ventral margins nearly parallel; pectoral fins with 14 rays, fairly long and pointed in late larvae Head: relatively small; mouth small; teeth apparent in late larvae Eye: round and relatively large Gut: triangular and short **Preanus length:** < 30% SL Air bladder: absent

Spination: none

PHOTOS

BLENNIIFORMES

Pigmentation: dorso-lateral side of gut strongly pigmented; postanal, ventral row of 19 to 21 regularly spaced melanophores, beginning some distance behind anus and reaching caudal peduncle; melanophores on head and snout; some caudal melanophores; lateral row of melanophores above notochord in late larvae; no melanophores over anus; pectoral-fin base pigmented; dorsal row of melanophores in later larvae, while lateral row disappears

Length at flexion: unknown Length at transformation: unknown

by J.M. Rodriguez

Figs. A-D



3.8 mm SL







10.5 mm SL



BLENNIIDAE





Illustrations' sources: A, B: L. Rodríguez (redrawn from Fives, 1986); C, D: Ford (1922)

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BLENNIIDA

Habitat: neritic, demersal, between 0
and 25 m depthMeristic characters
Myomeres: NA
Vertebrae: NA
Dorsal fin: XII + 21
Anal fin: II + 23Meristic characters
Myomeres: NA
Dorsal fin: XII + 21
Anal fin: II + 23EGGSYOLK–SACLARVAE
Undescribed

LARVAE

Body: elongate and slab-sided; caudal peduncle deep; dorsal and ventral margins nearly parallel; pectoral fins small and rounded Head: relatively small; mouth small; teeth in larvae > 6.0 mm SL Eye: round and relatively large Gut: triangular and short Preanus length: 36% of SL in early larvae, decreases to 32% in late larvae Air bladder: absent

Parablennius pilicornis (Cuvier, 1829)

Spination: 6-8 preopercular spines in larvae > 4.0 mm SL

Ringneck blenny

Pigmentation: stellate melanophores over head (only one in early larvae); peritoneal region heavily pigmented; a single melanophore ventrally over anus; ventral row of melanophores from anus to notochord tip; a melanophore on caudal–fin base in late larvae; dorsal pigmentation on tail appears in larvae of about 8.5 mm SL, spreading forward **Length at flexion:** 5.0-6.0 mm SL **Length at transformation:** unknown

PHOTOS by J.M. Rodriguez 2.8 nm SL 2.8 nm SL Image: State of the state

Parablennius pilicornis (Cuvier, 1829)



Science (1986) 4: 193-201 with permission – ©NISC (Pty) Ltd

Literature: Froese and Pauly (2022), Olivar (1986), Olivar and Fortuño (1991) Illustrations' sources: A-F: Olivar (1986) BLENNIIDAE

BLENNIIFORMES

	Diplecogaster bimaculata (Bonnaterr	e, 1788) Two-spotted clingfish Lépadogastère à deux tâches			
BIESOCIDAE	Habitat: neritic, demersal, between 5 and 200 m depthMeristic c MyomereDistribution: eastern Atlantic Ocean, from Morocco to Norway, and the 	haracters s: NA : NA : 5-7 I-6			
\mathbf{O}	EGGS Fig	g. A YOLK–SAC LARVAE Fig. B			
	Habitat: demersal, attached to empty shells Shape: flattened, oval-shaped Chorion: smooth, size 1.37-1.54 mm long, 1.08 mm wide, 0.62-0.70 mm high Perivitelline space: small Yolk: unsegmented; pigmented Oil globules: one; diam. 0.24-0.28 mm Colour: slightly dark	Hatch size: may be about 4.3 mm Body: elongate; mouth well formed at hatching Yolk sac: circular and small Oil globule location: no information Anus: detached from yolk sac, reaches finfold border Preanus length: about 65% SL Pigmentation: rounded stellate melanophores arranged in fairly regular rows along sides of body and gut; no melanophores on head; caudal region free of pigment; yolk sac pigmented with melanophores along surface of yolk; eye pigmented			
	LARVAE	Figs. C-D			
	Body: elongate and slender; pelvic fins develog a sucker at about 6.0 mm Head: moderate in size, pointed; mouth termin Eye: round and relatively small Gut: long, tube-like Preanus length: about 70% SL Air bladder: absent	p into Spination: none Pigmentation: similar to yolk-sac larvae, melanophores extend backwards on caudal region with development Length at flexion: unknown Length at transformation: unknown			
	PHOTOS	by J.M. Rodriguez			
		4.9 mm SL			
ORMES					
		6.4 mm SL			



6.4 mm SL (dorsal view)





MUGILIDAE

Mugil cephalus Linnaeus, 1758 Flathead grey mullet - Mulet à grosse tête Habitat: neritic, pelagic, inshore, **Meristic characters** entering estuaries and lagoons Myomeres: 24 Distribution: cosmopolitan in coastal Vertebrae: 24 waters of tropical, subtropical and Dorsal fin: V + 7-9 temperate seas Anal fin: III + 8-9 Spawning season: July to October EGGS YOLK-SAC LARVAE Fig. A Fig. Habitat: pelagic Hatch size: about 2.5 mm **Body:** elongated Shape: spherical Yolk sac: ovoid Chorion: smooth; diam. 0.72 mm Oil globule location: at posterior edge of yolk sac Perivitelline space: small Anus: detached from yolk sac, reaches finfold Yolk: unsegmented; pigmented border Oil globules: one; diam. 0.28 mm; pigmented Preanus length: about 60% SL Colour: transparent Pigmentation: heavily pigmented, except caudal region; dorsal and ventral rows of about 4 dotted melanophores at caudal region; yolk sac pigmented with melanophores located around oil globule; oil globule strongly pigmented LARVAE **Body:** relatively stubby **Pigmentation:** strongly pigmented, except lateral Head: moderate with an oblique mouth sides of head and caudal region, where dotted Eye: round and large melanophores of yolk-sack larvae persist; pigment Gut: large and bulky is heaviest on dorsum, dorsal surface of gut and Preanus length: up to 70% SL on postanal ventral region; mid-lateral row of Air bladder: apparent, even in early larvae melanophores apparent Spination: none Length at flexion: about 4.0-5.0 mm Length at transformation: 10.0 mm PHOTOS by J.M. Rodriguez





7.5 mm SL



2.3 mm SL

3.3 mm SL



MUGILIFORMES



Dicentrarchus labrax (Linnaeus, 1758) European seabass – Bar européen Habitat: neritic, demersal, between 10 and 100 m depth Meristic characters Distribution: eastern Atlantic Ocean, from Senegal to Norway, and the Mediterranean Sea Meristin KIII-IX 2nd dorsal fin: XIII-IX 2nd dorsal fin: I + 12-13 Spawning season: January to March Anal fin: III + 10-12

(Mediterranean Sea)

Fig. A

YOLK-SAC LARVAE

Fig. B

Figs. C-G

EGGS

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 1.20-1.51 mm Perivitelline space: small Yolk: unsegmented; pigmented Oil globules: one; diam. 0.36-0.46 mm; pigmented Colour: transparent

Hatch size: 3.5-4.0 mm

Body: elongate Yolk sac: ovoid Oil globule location: at ventral side of yolk sac Anus: detached from yolk sac, reaches finfold border Preanus length: > 50% SL Pigmentation: strongly pigmented with melanophores forming bands situated over yolk sac, at level of anus, mid-tail and caudal region; ventral row of melanophores (above gut) and postanal

LARVAE

PHOTOS

Body: very elongate and narrow Head: small; mouth terminal and small Eye: round Gut: elongate, tube-like, forming a slight curve above air bladder Preanus length: about 50% SL Air bladder: present, prominent in late larvae Spination: none **Pigmentation:** continuous line of melanophores stretching from snout to caudal–fin base; dorsal melanophores restricted to tail end; tip of snout and lower jaw pigmented; melanophores on ventral side of head and gut; air bladder pigmented **Length at flexion:** about 6.0 mm SL

region; yolk sac and oil globule pigmented

Length at transformation: unknown

by J.M. Rodriguez



5.6 mm SL





Illustrations' sources: A, G: Bertolini (1933b); **B, C:** Kennedy and Fitzmaurice (1966); **D-F:** modified from Russell (1976)

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SPARIDAE

Boops boops (Linnaeus, 1758)

Habitat: neritic, benthopelagic,

Distribution: eastern Atlantic Ocean,

from Angola to Norway, and the

Spawning season: March to May

between 10 and 200 m depth



Fig.

Meristic characters Myomeres: 24 Vertebrae: 24 Dorsal fin: XIII-XV + 12-16 Anal fin: III + 14-16

Fig. A

EGGS

Mediterranean Sea

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 0.89 mm Perivitelline space: small Yolk: unsegmented: unpigmented Oil globules: one; diam. 0.2 mm; pigmented Colour: transparent

YOLK-SAC LARVAE

Hatch size: about 3.2 mm Body: elongate and slender Yolk sac: elongated Oil globule location: at posterio

Oil globule location: at posterior end of yolk sac **Anus:** close behind yolk sac, reaches finfold border **Preanus length:** about 45% SL

Pigmentation: dorsal row of melanophores to about mid-tail; opposing dorsal and ventral bars of melanophores in posterior mid–tail; ventral series of melanophores posterior to ventral tail bar; a melanophore under urostyle; oil globule pigmented

LARVAE

Body: elongate and slender in early larvae, increases in height during development
Head: small; snout slightly rounded
Eye: round and large
Gut: triangular, terminal section forms a right angle with body
Preanus length: about 40% SL
Air bladder: prominent from early larvae on
Spination: opercular spines in larvae longer than

of melanophores at about mid-tail in early larvae; a row of postanal ventral melanophores (ventral bar disappears with development); peritoneum pigmented; single melanophores over head (several in late larvae), under gut, and under urostyle (may be 2 in late larvae), which migrates to caudal-fin base with development; air bladder pigmented **Length at flexion:** 5.20-5.80 mm **Length at transformation:** unknown

Pigmentation: opposing dorsal and ventral bars

PHOTOS

4.8 mm





Illustrations' sources: A-G: De Gaetani (1937)

PERCIFORMES

Habitat: neritic, littoral, demersal, to
50 m depthMeristic charactersDistribution: eastern Atlantic Ocean,
from Angola to the British Iles, and
the Mediterranean SeaMeristic charactersSpawning season: March to June
(western Mediterranean Sea)Maristic characters

Fig.

Diplodus sargus (Linnaeus, 1758)

YOLK-SAC LARVAE

globule pigmented

Habitat: pelagicHatch size: about 2.5 mmShape: sphericalBody: elongate and slenderChorion: smooth; diam. 0.88-0.97 mmYolk sac: ovoidPerivitelline space: smallOil globule location: at posterior edge of yolk sacYolk: unsegmented; pigmentedOil globules: one; diam. 0.19-0.21 mm; pigmentedOil globules: one; diam. 0.19-0.21 mm; pigmentedPigmentation: scattered melanophores over
dorsum, ventral anterior mid-tail and above gut; oil

LARVAE

EGGS

Body: elongate (the most among species belonging to this genus) and slender in early larvae **Head:** relatively small; mouth small; snout rounded, mainly in later larvae **Eye:** round and relatively large **Gut:** triangular; anus forms a right angle with body **Preanus length:** about 30% SL in early larvae, increases with development **Air bladder:** present Spination: two series of preopercular spines Pigmentation: postanal ventral row of melanophores; peritoneum, ventral region of gut, occipital and shoulder regions pigmented; air bladder pigmented; there may be a dorsal melanophore on mid-tail in larvae about 4.0 mm long Length at flexion: 7.5-8.0 mm

Length at transformation: unknown

The genus *Diplodus* includes several species whose ELS are very similar. Here, the ELS of *D. sargus* are described as an aid to identify *Diplodus* larvae.

PHOTOS







Diplodus sp. 4.5 mm SL

by J.M. Rodriguez

Figs. C-F



Diplodus sp. 5.4 mm SL



Diplodus sargus (Linnaeus, 1758)



Illustrations' sources: A-C: Ranzi (1933); D-F: Brownell (1979)

SPARIDAE

PERCIFORMES

Pagellus acarne (Risso, 1826) Axillary seabream – Pageot acarne Habitat: neritic/upper slope, **Meristic characters** demersal, to 500 m depth Myomeres: 24 Distribution: eastern Atlantic Ocean, Vertebrae: 24 from Senegal to the British Isles, and Dorsal fin: XII-XIII + the Mediterranean Sea 10-12 Spawning season: November to Anal fin: III + 9-10 March EGGS YOLK-SAC LARVAE Fig. A Habitat: pelagic Hatch size: about 2.4 mm Body: elongate Shape: spherical Yolk sac: ovoid Chorion: smooth; diam. 0.9-1.0 mm Oil globule location: at posterior, ventral edge of Perivitelline space: small yolk sac Yolk: unsegmented; unpigmented Anus: slightly detached from yolk sac, reaches Oil globules: one; diam. about 0.20 mm; pigmented finfold border Colour: transparent Preanus length: about 58% SL **Pigmentation:** a dorsal and an opposed ventral melanophore at about mid-tail; a melanophore on ventral side of gut; oil globule pigmented LARVAE **Body:** shorter and stouter than that of *P. bogaraveo* Pigmentation: a large dorsal (in some individuals Head: relatively small; mouth relatively large; snout there may be 2) and an opposed large ventral pointed melanophore (in some, mainly early larvae up Eve: round and large to 4) at about mid-tail; a melanophore on head, **Gut:** triangular; terminal section forms a right angle and on tip of upper jaw; peritoneum, air bladder, lateral side of gut pigmented; 2 (one in early with body Preanus length: about 52% SL larvae) melanophores on caudal-fin base; air Air bladder: present bladder pigmented; pigmentation increases with Spination: two series of preopercular spines, with development. central spine of outer series longer Length at flexion: unknown Length at transformation: unknown

Remark: pigmentation quite variable

by J.M. Rodriguez



6.0 mm SL

11.5 mm SL

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SPARIDA

PHOTOS

PERCIFORMES
Pagellus acarne (Risso, 1826)



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Pagellus bogaraveo (Brünnich, 1768)			Blackspot(=red) seabream - Dorade rose	
Habitat: neritic/upper slope, demersal, to 700 m depth Distribution: eastern Atlantic Ocean, from Mauritania to Norway, and the Mediterranean Sea Spawning season: January to May	Meristic character Myomeres: 24 Vertebrae: 24 Dorsal fin: XII-XII 11-13 Anal fin: III + 11-1	rs I + 2		
EGGS	Fig. A	YO	LK–SAC LARVAE Fig. B	
Habitat: pelagic Shape: spherical Chorion: smooth; diam. about 1.20 mm Perivitelline space: small Yolk: unsegmented; unpigmented Oil globules: one; diam. about 0.28 mm; pigmented Colour: transparent		Hatch size: about 3.8 mm Body: elongate and slender Yolk sac: ovoid Oil globule location: at posterior edge of yolk sac Anus: close behind yolk sac, reaches finfold border Preanus length: about 40% SL Pigmentation: dorsal melanophores between head and end of tail region; two opposing groups of melanophores at posterior tail; oil globule pigmented		
LARVAE			Figs. C-G	
Body: relatively elongate and slender Head: relatively small; mouth small; snout rounded Eye: round and large Gut: triangular; terminal section forms a right angle with body Preanus length: about 40% SL Air bladder: present Spination: two series of preopercular spines		Pigmentation: numerous melanophores over head; large dorsal melanophores (2-4) on tail; about 6 (smaller) ventral melanophores on postanal region; peritoneum and ventral side of gut pigmented; a melanophore on ventral side of caudal peduncle in early larvae; caudal-fin base pigmented in later larvae; air bladder pigmented Length at flexion: unknown Length at transformation: unknown Remark: pigmentation guite variable		

PHOTOS

4.5 mm SL

by J.M. Rodriguez





2.0 mm SL

SPARIDAE

SPARIDAE

Pagellus bogaraveo (Brünnich, 1768)



Habitat: neritic, benthopelagic, between 0 and 250 m depth Distribution: eastern Atlantic Ocean, from Morocco to the British Isles, and the Mediterranean Sea Spawning season: April to June Meristic characters Myomeres: 24 Vertebrae: 24 Dorsal fin rays: XI-XIII+ 9-12 Anal fin rays: III+7-9

Fig. A

EGGS

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 0.89-0.93 mm Perivitelline space: small Yolk: unsegmented; unpigmented Oil globules: one; diam. 0.18-0.20 mm; pigmented Colour: transparent

Pagrus pagrus (Linnaeus, 1758)

YOLK-SAC LARVAE

Hatch size: 1.92-3.04 mm SL Body: slender

Yolk sac: elongate

Oil globule location: at posterior edge of yolk sac **Anus:** close behind yolk sac, reaches finfold border **Preanus length:** 40% SL

Red porgy – Pagre rouge

Pigmentation: melanophores along dorsal and ventral (from anus) midlines of body; head and oil globule pigmented

LARVAE

PHOTOS

Body: relatively short and moderately deep through pectoral region Head: large; mouth moderate Eye: round and large Gut: triangular; terminal section forms a right angle with body in early larvae Preanus length: increases from 40% in early larvae to 60% in late larvae Air bladder: present Spination: occipital crest from early larvae on (specific character among Sparidae species); two series of preopercular spines, posterior series more developed with central spine very long; supraorbital spinous arch

Pigmentation: light; peritoneum pigmented; about 6 postanal, ventral melanophores that reduce to 1-3 during development; single melanophores on head, over terminal section of gut and base of caudal fin; air bladder pigmented **Length at flexion:** 4.60-6.41 mm SL **Length at transformation:** 9.5-13.0 mm SL

by J.M. Rodriguez





3.7 mm SL



7.8 mm SL

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SPARIDAE

PERCIFORMES





Illustrations' sources: A: Machinandiarena *et al.* (2003); **B:** L. Rodríguez (redrawn from Machinandiarena *et al.*, 2003); **C, D:** Alemany (1997); **E, F:** Fage (1918)

 Habitat: nerific, demersal, between 15 and 170 m depth Distribution: eastern Atlantic Ocean, from Morocco to Portugal, and the Mediterranean Sea Spawning season: February to May 	Meristic character Myomeres: NA Vertebrae: NA Dorsal fin: XI-XII - Anal fin: III + 8-10	rs + 10-12				
EGGS	Fig. A	YO	DLK–SAC LARVAE Fig.	B		
Habitat: demersal, attached to objects Shape: slightly elliptical Chorion: smooth; size 0.89 x 0.72 mm Perivitelline space: small Yolk: segmented; unpigmented Oil globules: one; diam. 0.19-0.21 mm; unpigmented Colour: transparent			Hatch size: unknown Body: elongate and slender Yolk sac: spherical Oil globule location: in central-ventral side of yolk sac Anus: close behind yolk sac, reaches finfold border Preanus length: about 30% SL Pigmentation: unpigmented			
LARVAE			Figs. C-0			
Body: elongate and slender in early larvae, becomes more robust with development			nentation: a small occipital melanophore; 2 toneal pigment patches; around 15 postanal			

mo Head: moderately large and pointy Eye: round and large Gut: triangular **Preanus length:** < 50% SL Air bladder: present Spination: two series of preopercular spines

Spicara smaris (Linnaeus, 1758)

Habitat: neritic, demersal, between 15 Meristic characters

hore; 2 ostanal ventral melanophores, between beginning of anal fin to base of caudal fin; 1-3 dotted melanophores under urostyle that move to caudal-fin base in late larvae; 2 melanophores between dorsal and caudal fins in late larvae Length at flexion: almost completed at 6.5 mm SL

Length at transformation: unknown

PHOTOS

by J.M. Rodriguez

Picarel - Picarel



3.4 mm SL



6.8 mm SL

PERCIFORMES

Spicara smaris (Linnaeus, 1758)



Spondyliosoma cantharus (Linnaeus, 1758)

Black seabream - Dorade grise

Habitat: neritic, demersal, from 5 to 300 m depth

Distribution: eastern Atlantic Ocean, from Namibia to Scandinavia, and the Mediterranean Sea

Spawning season: April to June

Meristic characters Myomeres: 24 Vertebrae: 24 Dorsal fin: XI + 11-13 Anal fin: III + 9-11

EGGS

Habitat: demersal Shape: spherical Chorion: smooth; diam. 1.0-1.2 mm Perivitelline space: moderately large Yolk: unsegmented; pigmented Oil globules: one; diam. 0.20-0.25 mm; pigmented Colour: transparent

YOLK-SAC LARVAE

Fig. A

Hatch size: about 3.2 mm SL Body: elongate and slender Yolk sac: rounded Oil globule location: ventral, posterior in yolk sac Anus: slightly detached from yolk sac, reaches finfold border Preanus length: about 33% SL Pigmentation: yolk sac strongly pigmented; row of ventral melanophores from anus to about mid– tail in early yolk–sac larvae, reaches caudal region

later; melanophores on head; yolk sac and oil

globule pigmented

LARVAE

Body: elongate and slender in early larvae, increases in height during development **Head:** relatively small; mouth relatively large **Eye:** round and large **Gut:** triangular; terminal section forms a right angle

with body

Preanus length: < 50% SL until after flexion **Air bladder:** present **Spination:** two preopercular crests of small spines **Pigmentation:** a postanal ventral, row of melanophores; ventral-lateral scattered melanophores developing at about 4.0 mm SL, becoming more numerous during development; peritoneum and ventral side of abdomen pigmented; a melanophore over terminal section of gut; melanophores over head and anterior region of trunk; air bladder pigmented **Length at flexion:** about 6.0 mm SL **Length at transformation:** unknown

PHOTOS

by J.M. Rodriguez



SPARIDA

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Spondyliosoma cantharus (Linnaeus, 1758)



Illustrations' sources: A-C: Camus and Besseau (1986); D, E: Fage (1918)

SPARIDAE

PERCIFORMES

CEPOLIDAE

Cepola macrophthalma (Linnaeus, 1758) Red bandfish – Cépole commune Habitat: neritic, demersal, from 15 to **Meristic characters** about 400 m depth Myomeres: NA Distribution: eastern Atlantic Ocean, Vertebrae: NA from north of Senegal to the British Dorsal fin: 67-69 Isles, and the Mediterranean Sea Anal fin: 60 Spawning season: spring and autumn (Mediterranean Sea) YOLK-SAC LARVAE EGGS The egg description that is available is not Undescribed accurate

LARVAE

Body: rather elongate in early larvae, becomes very deep through head and pectoral regions with development

Head: moderately large **Eye:** round and relatively large

Gut: triangular

Preanus length: about 50% SL

Air bladder: absent

Spination: from 3.0 mm, larvae show 2 opercular spines and an indication of an occipital spine; late larvae show two series of preopercular spines (7-9 spines), those at angle longer; occipital and

supraorbital (with 4-5 teeth) crests, and a row of 6-7 small denticles along lower margin of jaw **Pigmentation:** peritoneal region strongly pigmented; groups of 1-3 ventral melanophores, at about middle of tail and on caudal region; some melanophores under gut, over head and one over terminal gut; pigmentation disappears from tail and increases in anterior region of head and trunk during development

Length at flexion: unknown Length at transformation: unknown

PHOTOS

by J.M. Rodriguez

Figs. A-D















PHOTOS





1.6 mm SL

2.0 mm SL

by J.M. Rodriguez







Tortonese (1986d)

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SERRANIDA





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SERRANIDA



6.2 mm SL

SERRANIDAE

PERCIFORMES

Serranus cabrilla (Linnaeus, 1758)



Literature: Alemany (1997), Bertolini (1933b), Fage (1918), Froese and Pauly (2022), Olivar and Fortuño (1991), Russell (1976), Tortonese (1986) Illustrations' sources: A-C: Bertolini (1933b); D-F: Fage (1918)

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SERRANIDA

Serranus hepatus (Linnaeus, 1758) Brown comber – Serran tambour **Meristic characters** Habitat: neritic, demersal, between 5 and 100 m depth Myomeres: 24 Distribution: eastern Atlantic Ocean, Vertebrae: 24 from Senegal to Portugal, and the Dorsal fin: X + 11-13 Mediterranean Sea Anal fin: III + 6-7 Spawning season: March to August (Mediterranean Sea) YOLK-SAC LARVAE EGGS

Habitat: pelagic Shape: spherical Chorion: smooth; diam. 0.78 mm Perivitelline space: small Yolk: unsegmented Oil globules: one; diam. 0.14 mm Colour: transparent

spines, upper spine much longer

Undescribed

LARVAE

Body: moderately stocky, deepest through pectoral region; pelvic fins weakly developed Head: large, moderately deep and slightly concave; mouth relatively large and slightly oblique Eye: round Gut: relatively elongated and globose **Preanus length:** > 50% SL Air bladder: present Spination: two series of preopercular spines, posterior series with larger spines; 4 opercular

Pigmentation: 2 dorsal and a single ventral melanophore over finfold in early larvae; 3 regularly spaced, postanal, ventral melanophores; a melanophore over anus and another on cleithral symphysis; some melanophores on ventral surface of gut; no dorsal melanophores; peritoneum, air bladder and pelvic fins pigmented Length at flexion: almost completed at 5.7 mm SL Length at transformation: unknown

PHOTOS

PERCIFORMES

by J.M. Rodriguez

2.1 mm SL



6.5 mm SL



5.5 mm SL

SERRANIDAE

Serranus hepatus (Linnaeus, 1758)



Illustrations' sources: A-E: Alemany (1997); F: Fage (1918)

Coris julis (Linnaeus, 1758) **Rainbow wrasse - Girelle** Habitat: neritic, demersal, from one to **Meristic characters** about 60 m depth Myomeres: 25-26 Distribution: eastern Atlantic Ocean, Vertebrae: 25-26 from Gabon to Sweden, and the Dorsal fin: VII + 11-12 Mediterranean Sea Anal fin: III + 11-12 Spawning season: April to August (Mediterranean Sea) Female YOLK-SAC LARVAE EGGS Fig. Fig. Habitat: pelagic Hatch size: about 2.2 mm Body: elongate and slender Shape: spherical Yolk sac: elongated, projected beyond snout Chorion: smooth; diam. 0.60-0.67 mm **Oil globule location:** at anterior end of yolk sac Perivitelline space: small Anus: detached from yolk sac, reaches finfold Yolk: unsegmented; pigmented border Oil globules: one; diam. 0.12-0.16 mm; pigmented Preanus length: about 50% SL Colour: transparent Pigmentation: several dorsal melanophores decrease in number to 2 and migrate to finfold during development; row of ventral melanophores along trunk and tail, ending in a large melanophore LARVAE Body: laterally compressed; elongated in early Pigmentation: 2 dorsal melanophores on finfold larvae, increases in height during development in early larvae which remain on dorsal fin in Head: small with moderately pointed snout; mouth late larvae; 2 large ventral melanophores, one relatively small and oblique over gut, close to gut loop (expands over it with Eye: round development) and another in caudal region (forms Gut: elongate, ends in a prominent loop a bar in late larvae) Preanus length: about 50% SL

Length at flexion: completed at 9.8 mm SL Length at transformation: unknown





2.7 mm SL



Ц LABRIDA

> Air bladder: absent Spination: none

PHOTOS





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Illustrations' sources: A: Sparta (1956a); B-G: L. Rodríguez (redrawn from Sparta, 1956a)

LABRIDAE



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LABRIDA





7.1 mm SL

Dorsal view of head



Labrus bergylta Ascanius, 1767			Ballan wrasse – Vieille commune	
Habitat: neritic, littoral, demersal, down to 20 m depth Distribution: eastern Atlantic Ocean, from Morocco to Norway. Records from the Mediterranean Sea doubtful Spawning season: unknown	Meristic characters Myomeres: 35-40 Vertebrae: 35-40 Dorsal fin: XVIII-X 9-13 Anal fin: III + 8-12			
EGGS		YO	LK–SAC LARVAE	
The egg descriptions available are doubtrul		 Hatch size: 2.8-3.0 mm Body: elongate and slender Yolk sac: elongated Anus: detached from yolk sac, reaches finfold border Preanus length: about 55% SL Pigmentation: melanophores scattered over most of body and yolk sac; posterior mid-tail and cauda regions unpigmented; finfold unpigmented 		
LARVAE			Figs. A-D	
Body: laterally compressed; elongat early larvae, increases in height with Head: relatively small; mouth small snout rounded in early larvae becon throughout development Eye: round Gut: elongate, tube-like in early larv tapering to a relatively narrow, sligh anus Preanus length: about 57% SL	e and slender in a development and oblique; nes pointed vae; bulging, ntly protruding	Spin Pigm mela pigm mid- of bo patte Leng Leng	nation: none nentation: crescent-shaped groups of nophores on head; anterior region of anal fin nented; snout, lateral side of head, posterior tail and caudal region unpigmented; rest ody pigmented; no changes in pigmentation ern throughout development gth at flexion: unknown gth at transformation: unknown	
Air bladder: absent				





2.9 mm SL

Labrus bergylta Ascanius, 1767



LABRIDAE



anus in late larvae Preanus length: about 50% SL Air bladder: absent Spination: none

single melanophores over head and tip of snout Length at flexion: unknown Length at transformation: unknown

PHOTOS

PERCIFORMES

by J.M. Rodriguez



3.0 mm SL



4.3 mm SL







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Illustrations' sources: A-C: modified from Russell (1976); D, E: Fives (1976)

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LABRIDA

Symphodus melops (Linnaeus, 1758) Corkwing wrasse - Crénilabre mélops Habitat: neritic, demersal, from one **Meristic characters** and 30 m depth Myomeres: 33 Distribution: eastern Atlantic Ocean, Vertebrae: 33 from Morocco to Norway, and the Dorsal fin: XV-XVIII + Mediterranean Sea 8-10 Spawning season: March to May **Anal fin:** III + 8-10 (northwestern Mediterranean Sea) YOLK-SAC LARVAE EGGS Fig. Habitat: demersal, attached to seaweeds in nests Hatch size: 2.40-2.85 mm SL Body: elongate Shape: spherical Yolk sac: ovoid, pigmented Ch: no information; diam. 0.80-0.85 mm Anus: detached from yolk sac, reaches finfold border Perivitelline space: no information Preanus length: about 50% SL Yolk: unsegmented; pigmented **Pigmentation:** 2 rows of dorsal melanophores Oil globules: none from behind eye to about mid tail; a row of Colour: no information melanophores above, another below notochord and another along gut; melanophores on primordial fin, behind anus; 3 ventral melanophores at end of tail; yolk sac pigmented; dorsal side of head unpigmented

LARVAE

PHOTOS

Body: laterally compressed; elongate and relatively slender in early larvae, increases in height with development **Head:** small and rounded; mouth small **Eye:** round and large

Gut: elongate, tube-like in early larvae, bulging, tapers to a relatively narrow protruding anus in late larvae **Preanus length:** about 50% SL **Air bladder:** absent **Spination:** none **Pigmentation:** a small number of melanophores on head; 2 parallel rows of dorsal melanophores to about mid tail; lower jaw pigmented; lateral sides of body (except posterior mid-tail region) and anal fin pigmented; 3 or 4 melanophores between anal and caudal fins; a row of melanophores along interspinous area of anal fin

Length at flexion: probably begins at about 4.0 mm SL

Length at transformation: unknown

by J.M. Rodriguez

Figs. B-D







Illustrations' sources: A, B: Quignard (1967); C, D: Fives (1976)

LABRIDAE

PERCIFORMES

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LABRIDAE

<i>Thalassoma pavo</i> (Linnaeus, 1758)			Ornate wrasse - Girelle paon		
Habitat: neritic, demersal, to 150 m depth Distribution: Eastern Atlantic, from Gabon to Portugal, and the Mediterranean Sea Spawning season: June and July (Mediterranean Sea)	Meristic character Myomeres: 25 Vertebrae: 25 Dorsal fin: VIII + Anal fin: III + 10-1	rs 12-13 .2			
EGGS	Fig. A	YO	LK–SAC LARVAE Fig. B		
Habitat: pelagic Shape: spherical Chorion: smooth; diam. 0.62-0.64 mm Perivitelline space: small Yolk: unsegmented; unpigmented Oil globules: one; diam. 0.16 mm; pigmented Colour: transparent		Hatch size: about 1.8 mm SL Body: elongate and slender Yolk sac: elongate, projected beyond snout Oil globule location: at anterior end of yolk sac Anus: close behind yolk sac, reaches finfold border Preanus length: about 69% SL Pigmentation: 7 melanophores over dorsal profile of trunk, close to each other; some dotty melanophores over ventral profile of trunk; oil globule pigmented			
LARVAE			Figs. C-F		
Body: elongate, laterally compressed, deeper through pectoral region Head: small, pointed, with relatively short snout; mouth very small Eye: almost round Gut: initially straight, becomes coiled after flexion Preanus length: decreases from about 59% SL in early larvae to about 52% in late larvae Air bladder: absent		Spination: none Pigmentation: early larvae, reduced to two melanophores, one over terminal gut and another close to ventral tail end; late larvae, body unpigmented; pigmentation restricted to pectoral- fin rays Length at flexion: unknown Length at transformation: unknown			
РНОТОЅ			by J.M. Rodriguez		









LABRIDAE

PERCIFORMES



Literature: Fahay (2007), Gomon and Forsyth (1990), Jones *et al.* (2006), Sparta (1956a) Illustrations' sources: A-C: Sparta (1956a); D-F: L. Rodríguez (D, F: redrawn from Jones *et al.*, 2006; E: redrawn from Richards and Leis, 1984) (Original illustrations D-F correspond to *T. bifasciatum*) ш

LABRIDA







LABRIDAE

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Literature: Fahay (2007), Jones *et al.* (2006), Quignard and Pras (1986a), Richards and Leis (1984), Sparta (1956a) Illustrations' sources: A, B: Sparta (1918a), C-F: L. Rodríguez (C, D, F: redrawn from Jones *et al.*, 2006, E: redrawn from Richards and Leis, 1984)

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TRACHINIDA

Echiichthys vipera (Cuvier, 1829) Lesser weever - Petite vive Habitat: neritic, demersal from a few **Meristic characters** to about 150 m depth Myomeres: 23-24 Distribution: eastern Atlantic Ocean, Vertebrae: 23-24 from Morocco to the North Sea, and 1st dorsal fin: V-VIII the Mediterranean Sea 2nd dorsal fin: 21-24 Spawning season: unknown Anal fin: I + 24-26 EGGS YOLK-SAC LARVAE Fig. A Fig. Habitat: pelagic Hatch size: about 3.3 mm Body: relatively elongate; pelvic fins well developed Shape: spherical Yolk sac: ovoid Chorion: smooth, with inner membrane; diam. 1.00-Oil globules location: ventral in yolk sac 1.37 mm Anus: close behind yolk sac, reaches finfold border Perivitelline space: small Preanus length: about 45% SL Yolk: unsegmented; pigmented Pigmentation: dorsal and ventral rows of Oil globules: 6-30; unpigmented melanophores extending posteriorly a little beyond Colour: transparent mid-tail in newly hatched larvae; pigmentation soon becomes characteristic with melanophores on snout, head and peritoneum and two bars of melanophores, one at level of anus and another at mid-tail LARVAE Body: relatively elongate and slender in early larvae, 4 in late larvae; one opercular spine in early larvae, 2 in late larvae larvae, becomes stout and deep (mainly through Pigmentation: tail bars of pigment disappear at pectoral and abdominal regions) with development; about 4.5 mm; 3-4 melanophores appear on ventral pelvic fins well developed from early larvae tail-caudal region that reduce to one in late larvae; Head: moderately large; mouth small melanophores on head and on shoulder; pelvic Eve: round fins, peritoneum and lateral side of gut strongly **Gut:** triangular and bulky pigmented in late larvae; dorsal fin pigmented Preanus length: about 45% SL Length at flexion: it begins at about 6.0 mm SL Air bladder: absent Spination: 3 preopercular spines in early Length at transformation: unknown PHOTOS by J.M. Rodriguez 2.7 mm SL





3.5 mm SL

7.7 mm SL



TRACHINIDAE

Trachinus draco Linnaeus, 1758

Habitat: neritic, demersal, from a few

Distribution: eastern Atlantic Ocean,

from Morocco to North Sea, and the

Spawning season: unknown

Meristic characters Myomeres: 42 Vertebrae: 42 1st dorsal fin: V-VII 2nd dorsal fin: 29-32 Anal fin: II + 28-34

Fig. A

Greater weever - Grande vive

EGGS

Habitat: pelagic

to about 150 m depth

Mediterranean Sea

Shape: spherical Chorion: smooth with inner membrane; diam. 0.96-1.11 mm Perivitelline space: small

Yolk: unsegmented; pigmented

Oil globules: one; diam. 0.19-0.23 mm; pigmented **Colour:** transparent

YOLK-SAC LARVAE

Hatch size: about 3.0 mm Body: elongate; primordial fin, globose, reaches snout

Yolk sac: ovoid Oil globule location: anterior, ventral in yolk sac Anus: close behind yolk sac, reaches finfold border Preanus length: > 37% SL

Pigmentation: dorsal melanophores on anterior part of body, on snout and 2 behind eye; row of postanal ventral melanophores; a bar of melanophores at mid-tail; a melanophore over anus; melanophores in dorsal region of body disappear during yolk sac development

LARVAE

Body: relatively elongate and slender, becoming stout and deep (mainly through pectoral region) with development; pelvic fins appear at about 4.0 mm SL; primordial fin globose, reaching snout in early larvae Haad: moderately large; mouth relatively large

Head: moderately large; mouth relatively large Eye: round and large Gut: triangular Preanus length: about 40% SL

Air bladder: absent

PHOTOS

Spination: one supraorbital and 5 preopercular spines

Pigmentation: early larvae similar to yolk– sac larvae; during development postanal bar disappears, peritoneal pigment increases, origin of postanal-ventral row of melanophores moves backwards and melanophores decrease in number; pelvic fins strongly pigmented

Length at flexion: between 5.0 and 7.5 mm SL **Length at transformation:** unknown

by J.M. Rodriguez

Figs. C-F





3.3 mm SL

4.3 mm SL





PERCIFORMES
TRACHINIDAE

PERCIFORMES





Illustrations' sources: A-C: Padoa (1956r); D, E: Alemany (1997); F: Fage (1918)

Habitat: neritic and upper slope, **Meristic characters** demersal, between 15 and 400 m Myomeres: NA depth Vertebrae: NA Distribution: eastern Atlantic Ocean, 1st dorsal fin: III-IV from Morocco to the Bay of Biscay, 2nd dorsal fin: 13-21 and the Mediterranean Sea Anal fin: I + 12-13 Spawning season: April to August YOLK-SAC LARVAE EGGS Fig. A Habitat: pelagic Hatch size: about 4.0 mm Shape: spherical Body: relatively elongate Chorion: sculptured with hexagonal structures; Yolk sac: ovoid, relatively small diam. 1.6-2.0 mm Anus: close behind yolk sac, reaches finfold border Perivitelline space: small Pigmentation: body, except caudal region, strongly Yolk: ovoid; pigmented pigmented; tail-end of dorsal and ventral regions Oil globules: none of finfold pigmented; yolk sack pigmented Preanus length: about 60% SL Colour: relatively opaque and white LARVAE Body: short, stocky with an extremely high preanal Air bladder: absent region Spination: a supraorbital crest and 4 protuberances Head: large; mouth large, oblique; snout blunt looking like blunt spines on head Pigmentation: body, except caudal peduncle,

Eye: round and large; migrates to dorsum of head in juvenile stage Gut: rounded and bulky Preanus length: about 60% SL

PHOTOS

by J.M. Rodriguez

2.4 mm SL





4.8 mm SL

PERCIFORMES







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Uranoscopus scaber Linnaeus, 1758

Stargazer - Uranoscope



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TRIGLIDA



PHOTOS









by J.M. Rodriguez



7.0 mm SL

TRIGLIDAE

Eutrigla gurnardus (Linnaeus, 1758)



Illustrations' sources: A-C, E: Padoa (1956p); D: modified from Russell (1976)



Blackbelly rosefish – Sébaste chèvre



EGGS

Habitat: pelagic Shape: ovoid Chorion: smooth; size 0.92–0.98 × 0.88–0.93 mm Perivitellne space: small Yolk: segmented; unpigmented Oil globules: one; diam. 0.2 mm; pigmented Colour: unpigmented

Helicolenus dactylopterus (Delaroche, 1809)

YOLK-SAC LARVAE

Hatch size: 1.9-2.6 mm Body: moderately elongate, in a jelly matrix Yolk sac: ovoid **Oil globule location:** at ventral, posterior edge of yolk sac Anus: slightly detached from yolk sac, reaches finfold border Preanus length: about 50% SL Pigmentation: oil globule, dorsum of body and finfold pigmented

LARVAE

PHOTOS

Body: relatively short; primordial fin prolonged to snout in early larvae; a mass of spongy tissue appears in spiny dorsal region at 4.0 mm Head: large; mouth small in early larvae Eve: round and relatively large Gut: triangular **Preanus length:** increases from about 50% SL to < 65% SL during development Air bladder: absent Spination: 3-4 stout spines along preopercular edge,

and a few small spines on lateral ridge; supraocular

ridge with a small, simple spine; parietal spine well developed with secondary serrations; small pteroptic and post-temporal spines Pigmentation: peritoneum, ventral side of gut, lower-jaw tip and cleithral region pigmented; 3 melanophores form a ventral patch close to caudal region; some dotted melanophores on head and behind eye; pectoral-fin border pigmented Length at flexion: 4.5-6.6 mm Length at transformation: between 19.0 and 35.0

by J.M. Rodriguez











mm

SCORPAENIDAE

SCORPAENIDAE

B. 2.2 mm

Body moderately elongate

C. 2.3 mm SL

D. 3.6 mm

E. 5.6 mm

PERCIFORMES



Pectoral-fin border

pigmented

Literature: Alemany (1997), Fahay (2007), Froese and Pauly (2022), Hardy (2006), Hureau and Litvin Sparta (1956b), Tåning (1961)

Illustrations' sources: A, B: Brownell (1979); C: Alemany (1997); D-F: Tåning (1961)

Helicolenus dactylopterus (Delaroche, 1809)

A.

Finfold extends to

snout in early larvae

Tip of lower jaw

pigmented

Prominent head

armature in late larvae

Habitat: neritic and upper slope, **Meristic characters** demersal, to 800 m depth Myomeres: 24 Distribution: eastern Atlantic Ocean, Vertebrae: 24 from Senegal to the British Isles, and Dorsal fin: XII + 9 the Mediterranean Sea Anal fin: III + 4-5 Spawning season: May to August (Mediterranean Sea) Fig. A

EGGS

Habitat: pelagic Shape: ovoid Chorion: smooth; size 0.92 × 0.84 mm Perivitelline space: small Yolk: unsegmented; pigmented Oil globules: none Colour: transparent

Scorpaena porcus Linnaeus, 1758

YOLK-SAC LARVAE

Hatch size: about 1.4 mm Body: short and stout Eye: ellipsoid Yolk sac: large, ovoid Anus: close behind yolk sac, does not reach finfold border **Preanus length:** > 60% SL **Pigmentation:** body, yolk sac and finfold covered with dotty melanophores

Black scorpionfish - Rascasse brune

LARVAE

Body: short, increases in height during development; primordial fin prolonged to snout in early larvae; pectoral fins very large Head: large and concave; mouth relatively large, reaches middle of eve Eve: round Gut: triangular Preanus length: about 50% SL Air bladder: absent Spination: cephalic, preopercular and opercular

spines; occipital and supraoccipital crests; supraoccipital crests with 2 strong spines finely denticulated

Pigmentation: peritoneum strongly pigmented; a postanal ventral row of melanophores in early larvae, decreasing in number (to 2-4) during development; pectoral-fin borders strogngly pigmented Length at flexion: almost completed at 3.4 mm SL

Length at transformation: unknown

PHOTOS

by J.M. Rodriguez



2.5 mm SL





3.5 mm SL

B. 1.4 mm A. **Body short C.** 1.8 mm SL Head armed with spines and crests D. 2.9 mm SL G. Dorsal view of head (3.4 mm SL individual) Pectoral fin very large with borders strongly pigmented **E.** 4.0 mm H. Detail of pectoral fin (4.4 mm SL individual) **F.** 6.5 mm

Scorpaena porcus Linnaeus, 1758

Literature: Alemany (1994), Froese and Pauly (2022), Hureau and Litvinenko (1986), Sparta (1956b) Illustrations' sources: A, B: Sparta (1956b); C, D, G, H: Alemany (1997); E, F: Fage (1918)

SCORPAENIDAE

SCORPAENIFORMES

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CAPROIDA]

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Capros aper (Linnaeus, 1758) **Boarfish - Sanglier** Habitat: neritic, demersal, between 40 **Meristic characters** and 600 m depth Myomeres: 21-23 Distribution: eastern Atlantic Ocean, Vertebrae: 21-23 from Senegal to Norway, and the Dorsal fin: IX-X + 23-25 Mediterranean Sea Anal fin: III + 22-24 Spawning season: spring and summer (Mediterranean Sea) YOLK-SAC LARVAE EGGS Fig. Fig. Habitat: pelagic Hatch size: about 2.0-2.5 mm Body: elongated Shape: spherical Yolk sac: large, ovoid Chorion: smooth; diam. 0.90-1.01 mm Oil globule location: at posterior edge of yolk sac Perivitelline space: small Anus: detached from yolk sac, reaches finfold Yolk: unsegmented; pigmented border Oil globules: one; diam. 0.15-0.17 mm; Preanus length: about 67% SL unpigmented Pigmentation: body, except caudal region, covered Colour: transparent with large stellate melanophores; back side of yolk sac pigmented (close to oil globule)

LARVAE

Body: elongate and slender in early larvae, acquires a rhomboid shape at about 3.0 mm Head: large and high; mouth becomes almost vertical with development and protractile, in postflexion larvae Eye: round Gut: almost tube-like in early larvae becomes globose and triangular with development Preanus length: about 50% SL Air bladder: apparent in late larvae

Spination: opercular spines and a spinous crest over head appear during development; anterior part of dorsal fin spinous; body covered with small spines in late larvae

Pigmentation: early larvae, a row of ventral melanophores above gut to about middle of tail; dorsal row of 6 large melanophores extending from above mid-gut to level of ventral row end; row of melanophores along ventral surface of gut; some melanophores on head and on mid-lateral side of body; upper and lower jaw tips pigmented; late larvae, body covered with very numerous stellate melanophores, only caudal peduncle remains free of pigment; air bladder pigmented Length at flexion: completed at 6.0 mm Length at transformation: unknown

PHOTOS

2.5 mm SL



by J.M. Rodriguez

CAPROIDAE





ACANTHURIFORMES

LOPHIIDA



Eye: round and relatively large Gut: bulky and rounded Preanus length: around 50% SL

PHOTOS

in larvae larger than 11.0-12.0 mm Length at flexion: unknown Length at transformation: unknown







7.0 mm SL

LOPHIIDAE

LOPHIIFORMES

Lophius piscatorius Linnaeus, 1758



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5. GLOSSARY

Anal fin: Fin (usually single but double in some gadiforms) located on the ventral margin of the tail, behind the anus.

Anus: Orifice and surrounding tissue at end of the gut.

Bathypelagic: Pelagic zone between 1 000 and 4 000 m depth. Bathypelagic fish are those inhabiting the bathypelagic region.

Benthic: Referring to the sea bottom. Benthic species are those living on or in the bottom (substrate).

Benthopelagic: Living and feeding near the bottom as well as in midwater or even near the surface.

Body clupeid shaped: Body elongate and slender and long, tube-like gut

Caudal fin: Median fin situated at the posterior end of the fish.

Caudal peduncle: Narrow part of the tail located between the posterior end of dorsal or anal fin and the base of the caudal fin.

Chorion/shell: Outer membrane of a fish egg.

Choroid tissue: Mass of vascular tissue on the ventral side of the eye in larvae of some myctophids and other meso– and bathypelagic fishes, usually associated with elliptical eyes.

Cleithrum: Vertical bone in the pectoral girdle, at the junction of head and body of the larva.

Cleithral symphysis: Ventral junction of the cleithral bones.

Chromatophore: Cell containing pigment that reflects the light.

Continental shelf: The flattened edge of the continental land mass, between the coast and the continental slope (generally, the continental subtidal zone to a depth of about 200 m).

Continental slope: The sloping edge of the continental land mass, generally beginning at depth of around 200 m.

Demersal: Living on or near the bottom of the sea.

Dorsal fin: Median fin or fins located on the dorsal margin of the body.

Early life history: The early life stages of fishes spanning from egg to juvenile.

Embryo: Early stage of development, from fertilisation to hatching

Epipelagic: The illuminated, uppermost layer of the ocean (from 0 to 200 m depth).

Eye stalks: Movable peduncles of varying length bearing the eyes.

Finfold/primordial fin: Median fold of skin surrounding the body of young larvae, within which the dorsal, caudal and anal fins are developed.

Flexion: Larval stage during which the urostyle bends dorsally.

Gut: Alimentary tube and associated organs.

Gut loop: Loop, fold, or curve found along the axis of the gut.

Head length: Distance from the tip of the snout to the posterior margin of the cleithrum. **Hindgut**: Posterior part of the gut.

Homogeneous: Uniform composition throughout; opposite to segmented in referring to egg yolk.

Ichthyoplankton: Zooplankton fraction including eggs and larval stages of fishes.

Isthmus: Ventral region of the head separating the gill openings of a fish.

Juvenile: A young fish, fundamentally resembling the adult in meristic characters (excluding squamation) but smaller and reproductively inactive.

Larva: Early life-history stage of fishes between the egg and juvenile stages.

Leptocephalus: The flat, transparent, and often large larvae of fishes in the orders Anguilliformes, Elopiformes and Notacanthiformes, characterized by small heads and prominent teeth

Melanophore: A cell containing melanin; a black or brown pigment cell.

Meristic characters: Countable structures occurring in series (e.g. myomeres, vertebrae, fin rays).

Mesopelagic: Occurring in the open ocean at middle depths, usually between 200 and 1 000 m.

Metamorphosis: A marked change in form or structure at the end of the larval stage involving acquisition of adult characters and loss of larval characters; synonymous of transformation.

Myomeres: Muscle segments occurring in series, the number is approximately equal to the number of vertebrae in adults.

Myosepta: Connecting tissue between adjacent myomeres.

Nekton: Motile, marine organisms living in the water column and capable of swimming against currents.

Neritic: Pelagic coastal zone extending from the low tide mark to the edge of the continental shelf.

Neural crest: Region of the neural ridge of the developing embryo that differentiates into many kinds of tissue and cells, including melanophores.

Neustonic: Occurring close to the surface of the ocean.

Notochord: Longitudinal flexible cartilaginous rod of cells forming the supporting axis of the body.

Notochord length (NL): The distance from the tip of the snout to the posterior tip of the notochord.

Occipital crest: A median, bony ridge, usually serrated, located on top of the head, posteriorly. **Oceanic**: Open sea region beyond the edge of the continental shelf.

Oil globule: Spheres of fatty material within the yolk of some fish eggs.

Opercular: Relative to the operculum.

Operculum or Opercle: The bony plate of the gill cover.

Ovoviviparous: Producing eggs that develop within the maternal body.

Pectoral fin: Paired lateral (sometimes ventrolateral) fins located behind the head.

Peduncle: A narrow part or stalk that connects a structure to the body (e.g. caudal peduncle connecting the caudal fin to body).

Pelagic: Free living in the water column, away from the sea bottom.

Pelvic fins: Paired fins, usually located on the ventral edge of the body, in the abdominal region.

Peritoneal: Region of the body associated with the gut or the membrane of the peritoneum.

Peritoneum: The membrane and associated tissue lining the gut cavity.

Perivitelline space: Fluid-filled space between the embryo and chorion or shell of an egg.

Photophores: Luminous organs on some marine (mostly deep-sea) fish larvae.

Plankton: Small free–living organisms, passively floating or weakly swimming that drift with currents.

Pigmentation: Deposition of pigment in body tissues.

Planktonic: Passively floating, drifting, or weakly swimming with prevailing currents. **Postanal**: posterior to the anus.

Postflexion stage: A stage in the development of larvae after the completion of flexion.

Preanal/preanus: Located anterior to the anus; preanal length (synonymous of snout–anus distance) measured from the tip of the snout to the posterior margin of the anus.

Preflexion stage: Larval stage before the beginning of the process of flexion.

Preopercle: Upper anterior bone of the gill cover.

Preopercular: Relative to the preopercle.

Recruitment: The amount of fish added to the exploitable stock each year due to growth and/ or migration into the fishing area.

Sculptured: Egg chorion with ornamentations or surface features of different shapes and textures.

Segmented: Particulate or divided; opposite of homogeneous in referring to egg yolk.

Shell: The membrane that encloses an egg; generally, equivalent to chorion.

Shrinkage: The act or fact of shrinking, to contract or lessen in size.

Swimbladder: Sac filled with air or other gases located in the abdominal region, beneath the backbone.

Snout: Forward part of the head, anterior to the eye.

Stalked eye: Eye situated on a stalk or peduncle.

Standard length (SL): The distance from the tip of the snout to the tip of urostyle.

Stellate melanophore: Star-like pigment spot.

Stock (Fish): A group of individuals in a species occupying a well–defined spatial range independent of other stocks of the same species.

Supraorbital spine: Occurring above the eye.

Tail: The portion of the body posterior to the anus, the postanal region.

Telescopic eye: Elongate, cylindrical eye that protrudes forward or upward within an envelope of skin.

Tentacle: Any of various slender, flexible appendages in larvae.

Total length (TL): Measurement from the tip of the snout to the most posterior part of a larva, including the caudal finfold or caudal fin.

Transformation: The process (synonymous of metamorphosis) at the end of the larval stage, characterized by a marked change in form or structure and involving the acquisition of juvenile or adult characters and the loss of larval characters.

Trunk: Portion of the body between the head and the anus.

Urostyle: The last vertebral elements in fishes, formed by fusion or loss of several vertebrae. **Year class**: All of the fish in a stock that were spawned in a particular year, such as all those

spawned in 1990. Also referred to as a "cohort".

Yolk: Nutritive material of the egg forming a sac–like mass (yolk sac) below the abdominal region of a newly hatched larva.

Yolk-sac larva: Newly hatched larva with yolk sac.

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This guide presents the egg and larval descriptions of 150 species of fishes belonging to 57 families, which are most likely to be present in plankton samples collected in the continental shelf and oceanic waters off northwest Africa. The guide is structured in two parts. The first introductory part describes the different applications of ichthyoplankton studies in fisheries research and management, and fish population ecology, the main sampling strategies, methods and gears, and the problems related to sample representativeness. It also describes the early life history of fishes, and how to identify them. A brief description of the hydrography of the study area is also presented. The second part of the guide features the species identification sheets. Each species sheet includes the following information: an illustration of the adult fish and information on its distribution, habitat, spawning season, and meristic characters; a description of the main features useful towards identifying the egg, yolk-sac and larval stages; and illustrations and photos of different larval stages. Finally, the guide provides a comprehensive list of references.

