

codex alimentarius commission



FOOD AND AGRICULTURE
ORGANIZATION
OF THE UNITED NATIONS

WORLD
HEALTH
ORGANIZATION



JOINT OFFICE: Viale delle Terme di Caracalla 00153 ROME Tel: 39 06 57051 www.codexalimentarius.net Email: codex@fao.org Facsimile: 39 06 5705 4593

Agenda Item 14(a)

CX/CF 07/1/17
February 2007

E

JOINT FAO/WHO FOOD STANDARDS PROGRAMME CODEX COMMITTEE ON CONTAMINANTS IN FOODS

First Session

Beijing, China, 16 - 20 April 2007

DISCUSSION PAPER ON DEOXYNIVALENOL

Background

1. The 36th Session of the Codex Committee on Food Additives and Contaminants (CCFAC) agreed to discontinue consideration of maximum levels for deoxynivalenol (DON) for the time being. Instead, it agreed to request information on: the occurrence of DON in cereals; the influence of processing, decontamination, and sorting, in lowering DON levels; national levels or guidelines for DON; and sampling procedures and methods of analysis, for consideration at its next Session (1).
2. The 37th Session of the CCFAC noted that more data on the occurrence of DON in cereals and processed cereal products were already available or would soon be made available on a more global basis. The Committee therefore decided to ask JECFA to conduct an exposure assessment based on the new data. In this regard, the Committee reconfirmed the importance to take into account processed foods and the effects of processing on the level of DON (2).
3. The Committee also decided to establish an electronic working group led by the United States to develop a discussion paper to provide comprehensive relevant data, including the occurrence of DON and the effects of processing on the levels of DON, for consideration at its next session. The members of the Working Group included: Belgium, Canada, European Community, Finland, France, Germany, Japan, Republic of Korea, Netherlands, United Kingdom and the International Council of Grocery Manufacturers Associations. This document titled "Discussion Paper on Deoxynivalenol" was prepared as requested, circulated and discussed at the 38th CCFAC in April 2006.
4. The 38th CCFAC agreed to re-establish the electronic Working Group led by the United States to revise and update the Discussion Paper with (a) more data from regions where data on DON are missing or inadequate, (b) additional data, especially on DON levels in maize, (c) information on the effect on levels of seasonal variation, and (d) information on the effect of processing on DON levels in foods (3). In addition, the Committee recommended that the discussion paper give a detailed indication of the information that could become available in the near future, including the timing, in order to expedite the possibility for JECFA to schedule an assessment of DON. The re-established Working Group led by the United States includes Australia, Belgium, Canada, European Community, France, Germany, Japan, Netherlands, Republic of Korea and the United Kingdom.

Introduction

5. Deoxynivalenol (DON; vomitoxin; “Rd-toxin”; 12,13-epoxy-3,7,15-trihydroxytrichothec-9-en-8-one ; CAS no. 51481-10-8), belongs to a class of sesquiterpenoid mycotoxins that are referred to as trichothecenes. The trichothecenes are produced by several fungi of the genus *Fusarium*, especially *F. graminearum* and *F. culmorum* which are pathogens of wheat, rye, barley, corn (maize) and other cereal grains. The worldwide distribution of the two species of fungi appears to be related to temperature; *F. graminearum* occurring predominately in warmer climates (4). The trichothecenes are the largest group of toxins produced by fungi of the genus *Fusarium* (5, 6).

6. The trichothecenes are subdivided into four groups referred to as types A-D based on their molecular structure (7). Types A and B are the predominant ones widely distributed in cereals and feed as natural contaminants. Type A trichothecenes include T-2 toxin and HT-2 toxin, while type B trichothecenes include deoxynivalenol, nivalenol and their 3- and 15-acetylated derivatives. Type A trichothecenes are characterized by the presence of a saturated carbon at C-8, while type B trichothecenes have a carbonyl at the same position (8). Deoxynivalenol is found most frequently in cereal grains; however, other trichothecenes might co-occur with DON (9). Among the trichothecenes, type A members are more toxic than type B members (10).

7. *F. graminearum* and *F. culmorum* are found world-wide in soil and are responsible for the *Fusarium* head blight disease (FHB) in cereals resulting in the production of DON. Studies have shown that the severity of *Fusarium* head blight depends mainly on climatic effects (temperature, rainfall, humidity) (11). These fungi usually infect susceptible grain crops in the field when there is cool wet weather at the silking or anthesis stage of grain development (12). The incidence and severity of DON contamination is known to vary from region to region, from one season to the next, and also in different cereal species (13, 14). These variations may be associated with periods of heavy rainfall between the anthesis stage and the time of harvest; this can facilitate *F. graminearum* infections and accumulation of DON (15). The infection of cereal grains by these fungi can result in the production of a mixture of several structurally related trichothecenes in addition to DON; the individual trichothecenes making up the mixture may vary by country as well as by region. DON is water-soluble, very stable during storage and milling, relatively heat stable, shown to survive most processing and cooking procedures, and is not completely destroyed by fermentation (16, 17). DON in contaminated grain survives brewing processes and is transmitted into beer (18,19).

Toxicology

8. The International Agency for Research on Cancer (IARC) reviewed the toxicity of DON and some other toxins derived from *Fusarium graminearum* and *Fusarium culmorum* in 1993 (20). The IARC concluded that there is *inadequate evidence* in humans for the carcinogenicity of toxins derived from *Fusarium graminearum*, *inadequate evidence* in experimental animals for the carcinogenicity of DON and that the toxins derived from *F. graminearum* and *F. culmorum* were not classifiable as to their carcinogenicity to humans (Group 3).

9. Risk assessments and toxicological reviews for DON have been conducted by the Nordic Council of Ministers, the Joint FAO/WHO Expert Committee on Food Additives (JECFA), the European Commission’s Scientific Committee on Food (SCF), the European Food Safety Authority (EFSA) and other investigators (21-25). A provisional maximum tolerable daily intake (PMTDI) of 1 µg DON/kg bw was established by JECFA (22). The genotoxic potential of DON has not been fully explored. Important toxicity data and other considerations relevant for the characterization of hazards associated with trichothecenes was recently summarized (26-28).

10. Exposure of some domestic animals to DON results in loss of appetite, feed refusal and vomiting, accompanied by a decrease in weight gain (24). Many feeding studies have been conducted on swine because this species appears to be more sensitive to DON than poultry, ruminants and other livestock. Reduced feed consumption and lower weight gain have been reported as the principal clinical effects observed in swine following ingestion of low dietary levels (< 2ppm feed) of DON in naturally contaminated feedstuffs; higher doses may induce vomiting and complete feed refusal (29). In several studies involving pigs, where the toxicological responses in animals that were given feed spiked with a known quantity of pure DON were compared with responses noted in animals given naturally contaminated feed containing an equivalent quantity of DON, it was observed that naturally contaminated feed had a stronger effect on the feed intake

and weight gain parameters than the pure toxin (24). It is possible that immunosuppression, hematological and histological changes seen in animals fed naturally contaminated feed containing DON may be intensified by co-contamination with other mycotoxins. Toxicological evaluations of the fate of trichothecenes in laboratory animals as well as other animal species, have been published (22, 30, 31). A major conclusion from these evaluations was that trichothecenes are rapidly excreted from animals, and therefore there is no significant carry-over of these toxins from animal derived products to humans.

11. Human poisonings associated with the consumption of trichothecene- contaminated grains (wheat, barley, corn) have been reported in Korea, Japan, India, Colombia, China and South Africa (16, 22,27,32,33). Symptoms including nausea, vomiting, diarrhea, abdominal pain, headache and dizziness were observed. The precise role of DON in those outbreaks is not certain. A review of epidemiological data available as of 2001 did not permit the establishment of a level below which no acute effects of DON would be expected to occur (22). The epidemiological data available point toward trichothecenes or specifically DON-contaminated grain products as a potential causative factor of the acute human toxicosis.

12. A review of the toxicity of DON and its potential effects on humans was recently published (27). The primary human safety concern for DON should be its potential for inducing acute gastroenteritis with vomiting. In addition, there is potential for chronic effects on growth, immune function and reproduction based on results from animal studies. A urinary biomarker that can be used for DON in both humans and animals was recently developed; the use of this biomarker should facilitate epidemiological studies of the adverse toxicological effects associated with this mycotoxin (34). The co-occurrence of several *Fusarium* toxins, along with zearalenone, has been observed in cereal grains. The presence of other toxins in grains is of concern because of the possible interactions of these toxins and their combined impact on the toxicological responses observed in animals and humans (23, 35, 36).

Sampling

13. It is difficult to estimate accurately and precisely the mycotoxin concentration in a large lot of grain because of the large variability associated with the mycotoxin test procedure. A mycotoxin test procedure generally consists of 3 steps: (a) an aggregate sample (consisting of incremental samples taken from various locations within a lot) is taken from the lot, (b) the whole aggregate sample is ground in a mill to reduce particle size and to ensure homogenization/mixing of the sample before a subsample is removed for extraction by solvents, and (c) the mycotoxin is extracted from the comminuted subsample and quantified by analytical techniques (37). The variance associated with the mycotoxin test procedure that measures DON in cereals is the sum of sampling, sample preparation, and analytical variances (38).

14. Statistical studies (38) have shown that the coefficients of variation associated with testing wheat are relatively small compared with testing other commodities for mycotoxins, such as aflatoxins in peanuts. Although other factors may be involved, the smaller variability (relative to that for other mycotoxins and other food commodities) is due in part to the kernel count of wheat per unit mass (about 30 kernels per gram), which is about 10 times larger than that for shelled corn and 30 times larger than that for shelled peanuts. This suggests that the distribution of DON contamination among wheat kernels may be less skewed than for aflatoxin in other commodities.

15. The variability associated with a mycotoxin test procedure can be reduced by increasing sample size, the degree of sample comminution, subsample size, and the number of aliquots quantified (37). A knowledge of the variability associated with sampling procedures involving cereal grains, when coupled with the availability of validated analytical methods and appropriate information on the distribution and tolerance or guideline levels for DON, can be used to: (1) provide an estimation of errors in the evaluation of DON concentration in lots of cereal products, (2) design sampling plans for DON in cereals and (3) select sample size or number of samples needed to reduce the total variability of an entire test procedure.

Analytical

16. Many analytical methods have been developed for the detection and quantification of DON and other trichothecenes. These toxins can be separated and analyzed by thin-layer chromatography, various liquid, gas and supercritical fluid chromatographic procedures, as well as by immunochemical methods. There are comprehensive reviews of current methodologies that can be used for the detection and analysis of DON and other trichothecenes in cereals (39-42).

17. Thin-layer chromatographic procedures for the detection and analysis of DON are reliable and cost-effective for use in laboratories with a restricted budget (43-46). Liquid chromatographic procedures alone or coupled with mass spectrometry and various detectors are becoming more common in many laboratories for the analysis of DON and other type B- and A- trichothecenes (47-61). Gas chromatographic procedures can also be used for the determination of many trichothecenes. The toxins or their derivatives are mostly detected by flame ionization, electron-capture detectors or by coupling to a mass spectrometer (19, 62-66). Whenever possible and affordable, it is recommended that thin-layer chromatographic methods for the determination of DON be replaced gradually by advanced chromatographic techniques, ideally coupled to a mass spectrometric detector.

18. Immunochemical methods have received a lot of attention lately because they can be used for rapid screening purposes under field conditions or in the laboratory and some can also effectively complement gas and liquid chromatographic procedures that are widely used in routine monitoring. These methods are simpler and less labor intensive. Comprehensive reviews of immunochemical and other rapid methods that can be used for trichothecenes have been published (67-70). Information on mycotoxin test kit data bases can be obtained from the homepages of AOAC International (www.aoac.org) and on the European Mycotoxin Awareness Network (EMAN) (www.mycotoxins.org).

19. Quantitative analytical methods that are to be used for monitoring and enforcement purposes should be validated or collaboratively studied to ensure that the analytical results obtained give an accurate measure of the analyte involved. Methods that have been developed and validated for the extraction and detection of DON in whole grain kernels cannot effectively be used for processed cereal grain products without further modifications in most cases. In conducting surveys involving milled or processed cereal products, it is essential that recovery studies be done for each type of product analyzed in order to determine if changes in DON levels represent accurate levels of the toxin or represent poor recovery of the toxin from the product matrix. Certified reference materials (CRMs) are available for DON in wheat and maize and these can be used to demonstrate that the method used provides accurate results (41, 71). Two commercially available crystalline DON analytical standards used by some laboratories throughout the world have been determined to be >96 and >98% pure, respectively (72, 73). The use of stable isotopes of trichothecenes as internal standards in LC/MS analytical procedures for trichothecenes is currently being explored (74-76).

20. Current AOAC official methods of analysis for DON in wheat include thin-layer chromatography (method 986.17) and gas chromatography (method 986.18). A liquid chromatographic method for determination of DON in whole wheat flour, white flour and bran has been subjected to an interlaboratory study and adopted as a peer-verified method by AOAC International (77). An international interlaboratory study has been conducted to compare different methods for DON analysis (41, 78). Due to the natural co-occurrence of DON with other trichothecenes and zearalenone, emphasis should be placed on the development and validation of methods capable of detecting multi-mycotoxin residues. Liquid chromatography/ tandem mass spectrometry methods have recently been made available for the rapid and simultaneous determination of zearalenone as well as trichothecenes (51, 79).

DON in cereal grains

21. The world-wide occurrence of DON in cereal grains has been well documented and reviewed in the literature (10, 16, 80, 81). Wheat, barley and corn together account for two-thirds of the world production of cereals and are the crops most susceptible to *Fusarium* disease and trichothecene contamination (82). DON has been found in other cereal grains including rye, oats, rice, and their products in many regions of the world (10). Studies have shown that at low levels of contamination, DON and its acetylated derivatives are produced and remain localized in the outer parts of the grain kernels to a large extent (57, 83). However, at higher levels of contamination, the toxins may be more evenly distributed throughout the kernel (84). Zearalenone is known to co-occur with DON and other trichothecenes since it is produced by the same *Fusarium* species. A "masked" naturally occurring glucoside of DON (DON-3-glucoside) was recently found in *Fusarium*-infected wheat and maize (85). The identity of the compound was confirmed by NMR and its presence in naturally contaminated maize and wheat ranged from 4 to 12% of the DON concentration.

22. Analytical data on DON, resulting from surveys conducted on cereals and cereal products in many parts of the world between 1990 and 2000, have been summarized (42). DON was investigated in 15,187 samples; 48.1% were from wheat (mean 531 µg/kg, 61.6% positives), 20% from corn (mean 230 µg/kg, 60.3% positives) and 18.5% from wheat products (mean 314 µg/kg, 52.2% positives). DON was also detected at lower levels in other cereals such as barley, oats, rye, rice and in processed food products such as beer, cooked pancakes, baby and infant foods, breakfast and mixed grain cereals, noodles and cookies.
23. In a survey conducted by the EU involving 12 member countries, a total of 11,022 samples of various food and food raw materials were analyzed for DON, 4,166 for nivalenol, and 5,675 for 3- and 15-acetyl DON (86). Fifty-seven percent of the samples analyzed for DON were positive, 16% analyzed for nivalenol were positive and 28% of the acetylated derivatives were positive. The percentage contamination of the positive samples associated with each toxin were as follows: a) DON – 89% corn, 61% wheat/ products, (b) nivalenol- 35% corn, 21% oats and 14% wheat/products and (c) acetyl DON- 27% maize and 8% wheat products. Among cereals analyzed in this survey, maize showed the highest level of contamination with trichothecenes. Seven percent of the raw cereals and flour contained DON levels at 750 µg/kg or higher and 6% of the cereal products contained DON levels at 500 µg/kg or higher. Wheat and wheat containing products such as pasta and bread represented the major sources of intake of the four trichothecenes that were measured.
24. In a survey of stored grain from the 1999 harvest in the UK, DON was detected in 88% of the 320 samples of wheat, barley and oats analyzed; 83% were below 100 µg/kg, the maximum level was 600 µg/kg. In samples with DON levels exceeding 150 µg/kg, nivalenol was also detected at 50 µg/kg or higher (87).
25. In Croatia, 465 samples of feed grain were collected and analyzed over a 7 year period from manufacturers and farm storage facilities (88). DON was detected at levels ranging from 50 to 340 µg/kg in 41.2 % of the samples. The majority of the samples were for poultry feed.
26. Two hundred and seventy-two oat samples were analyzed after harvest in an area of southwest Germany over a five year period (89). DON was the major toxin with incidences at 49-85% and mean levels in positive samples of 52-302 µg/kg. A correlation was noted between the occurrence and levels of the trichothecenes in oats and the amount of heavy precipitation during the summer months preceding crop harvest.
27. Thirty-eight Finnish grain samples (14 wheat, 22 barley, 1 rye and 1 oats) were collected from different parts of Finland during 2001 and 2002 (66). The mean levels of DON, 3-acetyl DON, and nivalenol in the samples were 272, 17 and 150 µg/kg, respectively.
28. A survey for DON in cereal grains harvested over a 3 year period was conducted in Russia (90). DON was detected in 69% of 2,166 stored wheat samples from the major *Fusarium* epidemic region of Russia. The contamination levels ranged from 100 to 8,600 µg/kg and a positive correlation was noted between DON concentration and the percentage of *Fusarium*-damaged wheat kernels. DON was detected in 11% of 1,908 freshly harvested food wheat samples; the concentration levels of DON ranged from 50 to 6,650 µg/kg. The incidence and levels of DON in freshly harvested barley and rye were significantly lower than in wheat.
29. A total of 843 commercial animal feed and foodstuffs samples were collected and analyzed for trichothecenes over a 3-year period in Saudi Arabia (91). DON was the most frequent trichothecene detected (13% of all the positive samples examined) and the levels ranged from <2 to 4,000 µg/kg. It was present in 21% of the maize samples and in 18% of the poultry feed, but the highest concentration was recorded in barley samples where it had a mean concentration of 2,553 µg/kg.
30. One hundred samples of wheat were analyzed in Brazil in 1995; the mean DON level was 618 µg/kg (92). In 2003, 297 wheat samples were analyzed and 24.9% were contaminated with DON; the mean contamination level was 603 and the maximum level was 8,504 µg/kg. Five hundred and sixty-three barley samples were analyzed between 1998 and 2003 in Brazil. The mean level of DON contamination was 114 µg/kg and the maximum level was 5,715 µg/kg.

31. A total of 125 samples of a group consisting of wheat, oats, corn, corn byproducts, corn plants and corn silage were collected from various sources in Germany during 2000 and 2001 and analyzed for 16 *Fusarium* toxins; 94% contained DON (93). The incidence of DON contamination was 100% for all commodities examined except for wheat (95%) and oats (71%). The mean levels of DON in corn, corn byproducts, corn plants and corn silage were 849, 1626, 598 and 2919 µg/kg, respectively; the maximum levels were 3820, 6682, 818 and 3944 µg/kg, respectively.

32. Two thousand five hundred and twenty-four wheat samples were collected and analyzed for DON in the U.S from 1994 to 2003 (94). Forty-one percent contained DON at levels less than 500 µg/kg; 18.6 % contained >500 to 1,000 µg/kg; 39.8% contained >1,000 to 6,000 µg/kg and 0.6 % contained > 6,000 µg/kg. DON levels varied significantly from year to year.

33. Two thousand one hundred and six barley samples were collected and analyzed for DON in the U.S. from 1993 to 2003 (94). Thirty-eight percent contained DON levels less than 490 µg/kg; 14.5 % contained 490 to 990 µg/kg; 28.5 % contained >990 to 4,990 µg/kg and 18.6% contained 4,990 to >5,000 µg/kg. DON levels varied significantly from year to year.

34. During the first four years (2002-2005) of a five year project in the UK to look at agronomic factors affecting the production of *Fusarium* toxins, including DON, 1,473 wheat samples were analyzed (95). Ninety-seven percent contained less than the EU limit of 1,250 µg/kg DON. Location was a major but not always a consistent factor associated with changes in DON levels from year to year. The risk of DON contamination was high when a wheat crop followed a maize crop. No difference was noted between DON levels in wheat samples obtained from organic and conventional crops. During the same period 630 and 488 samples of barley and oats were analyzed for DON (95). The incidence and levels of DON in barley and oats was low with only one sample of barley containing DON above the EU limit of 1250 µg/kg.

35. Two thousand,nine hundred and twenty-four wheat samples were analyzed for DON in the Netherlands from 1998-2004 (96). The mean DON content of the wheat was 580 µg/kg in 1998, which was the year with the highest *Fusarium* contamination. In the other years, the average DON levels ranged between 190 and 317 µg/kg. The effect of different DON limits on the mean DON content of wheat lots approved for sale to the milling industries was also calculated.

36. In Japan, 136 samples of husked wheat were examined in 2001 and 2002 (97). Seventy- seven percent of the samples analyzed in 2001 contained quantifiable levels of DON with an average of 286 µg/kg. Ninety-five percent of the samples analyzed in 2002 contained quantifiable levels of DON with an average of 184 µg/kg. Additionally, eight hundred and thirty-eight samples of husked domestically produced wheat were analyzed in 2002, 2003, 2004 and 2005; 39% contained quantifiable DON with maximum levels of 2,100, 580, 930 and 230 µg/kg, respectively (98, 99). The 90th percentile levels were 570, 260, 140 and 42 µg/kg, respectively.

37. Two hundred and ten samples of domestically produced barley were examined for DON in 2002, 2003, 2004 and 2005 in Japan; 50% contained DON with maximal levels of 4,800, 3,700, 1,800 and 460 µg/kg, respectively (98, 99). The 90th percentile levels were 670, 930, 680 and 190 µg/kg, respectively.

38. A survey on the occurrence of DON in UK-grown wheat for flour production was carried out from 2003 to 2005 indicated that the toxin occurred frequently in the UK wheat crop (100). The toxin was detected above the limit of quantification (LOQ) of 10 µg/kg in 88, 89.5 and 90% of the 60 samples in 2003, 50 samples in 2004 and 45 samples in 2005, respectively. The mean DON levels in 2003, 2004 and 2005 were 123, 114 and 113 µg/kg, respectively and the maximum levels found were 1250, 1119 and 775 µg/kg, respectively.

39. A survey on the occurrence of DON in UK malting-barley was carried out in 2003 and 2004 (101). The toxin was detected above the LOQ of 5 µg/kg in 10 and 40% of the 19 samples in 2003 and 40 samples in 2004, respectively. The mean DON levels in 2003 and 2004 were 5.8 and 6.9 µg/kg, respectively and the maximum levels found were 13 and 28 µg/kg, respectively. Deoxynivalenol was detected above the LOQ in 5 and 10% of batches of malt produced from the above barley in 2003 and 2004, respectively. The mean and maximum levels found were 6.1 and 27 µg/kg respectively in 2003 and 4.0 and 17 µg/kg, respectively in 2004.

40. An annual survey of grain quality was conducted in Germany addressing a set of mycotoxins and undesirable compounds in unprocessed wheat, rye and other cereals (102). Statistical sampling plans were developed each year anew to ensure the significance of the data. Preceding crop and variety of the samples were known. From 2001 to 2006, 5,387 samples, taken from fields spread all over the country, were analyzed for DON (2,443 wheat, 1,463 rye, 736 triticale, 470 barley and 275 oat samples) (103-105). While DON was detected in 74 % of the wheat samples, it was present in 64 % of the rye samples, when viewed over the six-year period. Nevertheless, DON detection frequencies and concentrations have varied over the years. For example, the median for DON in wheat was 109 µg/kg in 2004 and 36 µg/kg in 2005. Different climatic conditions are responsible for this variation to a large extent.

DON in maize(corn)

41. *F. graminearum* is mainly responsible for *Fusarium* ear rot of maize (corn) resulting in the production of DON and other trichothecenes (106). It has been reported that maize grain is usually the most contaminated with *Fusarium* mycotoxins of all agricultural commodities (9). Amounts of fungal metabolites like DON and zearalenone accumulating in maize kernels are usually significantly higher than amounts present in kernels of wheat or barley infected by the same fungal species; this is a consequence of a higher number of toxigenic *Fusarium* species being able to infect maize ears (9). *F. graminearum* and *F. moniliforme* are the main *Fusarium* species on maize in warmer regions of the world; *F. culmorum* and *F. subglutinans* prevail in cooler regions of the world.

42. High levels of DON have been reported in maize in many countries (10). In studies conducted prior to 1999, levels of up to 927 mg/kg of DON were reported in Poland, 8.5 mg/kg in New Zealand, 4.09 mg/kg in Canada and up to 1.83 mg/kg in South Africa.

43. Maize samples collected from storage bins and feed mills in Northern Italy between 1995 and 1999 were surveyed for various mycotoxins, including DON (107). The DON level was much higher in 1996 than in other years. During that year the extreme rainy weather conditions delayed the harvesting operations but favored the growth of DON producing fungi and the synthesis of the toxin. Specifically, during 1996, 7.7% of the samples contained DON levels <500 µg/kg, 16.4 % contained levels ranging from 500 to 1000 µg/kg, and 75.9 % contained levels >1000 µg/kg. Over the five year period the mean and maximum DON contents ranged from 7 to 30% and from 13 to 33%, respectively, when expressed in percent of the 1996 contents.

44. Forty-six freshly harvested maize samples from Central and Northern Italy were collected and analyzed for type B trichothecenes during 2002 (47). Damp climate, cool temperatures and delayed harvested times were believed to have been the contributing factors resulting in high levels of DON contamination. DON was the most abundant (up to 3430 µg/kg) and detected at a frequency of 40%; 26% of the samples contained 15-acetyl DON at levels up to 3500 µg/kg. The co-occurrence of DON and acetyl DON was observed in 23% of the samples.

45. In a study of maize samples, collected from farms located in different Italian areas immediately after harvest, less than 27% of the 93 samples analyzed were positive for trichothecenes (108). DON was the predominant toxin; its concentration ranged from 4 to 871 µg/kg. In highly contaminated samples, appreciable amounts of the monoacetates were detected in addition to DON.

46. Maize, designated for chicken and pig feed, were sampled from bulk lots and analyzed for DON during 1992-1997 in Brazil (92). The mean DON levels increased from 14.7 µg/kg in 1992 to 637 µg/kg in 1996, but decreased to 559 µg/kg in 1997. One hundred and ninety-five samples of maize for food purposes were analyzed from 1994-1995 in Brazil; 6.2% were contaminated with DON at levels ranging from 102-542 µg/kg. A total of 2,123 samples of maize were analyzed between 2002 and 2006. The mean levels of DON contamination were 82 (2002), 67 (2003), 20 (2004), 60 (2005) and 119 (2006) µg/kg.

47. A survey of maize and wheat collected from farms in the foothills of the Nepal Himalaya Mountains was conducted in 1997 (109). DON and NIV levels greater than 1000 µg/kg were found in 16% of the 76 samples of maize, and levels greater than 2000 µg/kg were found in 12 % of the samples. No DON nor NIV were detected in Nepalese wheat above the detection level of 1000 µg/kg.

48. Sixteen corn samples collected from Indonesia were analyzed for trichothecenes and other toxins (110). DON, nivalenol and zearalenone were each detected in two (12%) samples; 21, 32, and 49 $\mu\text{g}/\text{kg}$, and 169, 11 and 12 $\mu\text{g}/\text{kg}$, respectively. The investigators were of the opinion that this was the first report on the natural occurrence of *Fusarium* mycotoxins in maize from Indonesia, and also the first report of DON in corn from hot areas of Southeast Asia.

Studies to reduce DON levels in grains

49. Various physical procedures including cleaning, washing, sieving, density segregation, dehulling, fractionation by specific gravity table and polishing have been used singly, or in combination with milling procedures to reduce the level of DON in grains. The effectiveness of these procedures depended on the extent of contamination and the distribution of the toxin throughout the grain (111,112). Milling procedures to remove DON from wheat and other grains usually rely substantially on physical separation of the more heavily contaminated outer layers of the grain kernels.

50. Dry-milling is a process by which the components of cereal grains are separated into fractions based on particle size. The different fractions, such as flour and meal, retain most of the characteristics of the original grain (17). Studies have shown that in milled wheat, higher concentrations of trichothecenes are found in the bran fraction than in the original wheat with lower concentrations in white flour (112-115). Flour processing techniques can reduce DON levels by approximately a factor of 2 or greater. The extent to which dry-milling can reduce the DON level in flour is dependent on the extent of the fungal penetration into the endosperm of the wheat kernel and the distribution of DON within the kernels. The extent of fungal penetration is dependent on the cultivar of wheat involved (116, 117). Dry-milling of DON contaminated maize results in the DON being concentrated in the germ meal fraction (118).

51. Wet-milling is a process widely used for maize to obtain starch which can be used for the production of syrups and other products for human consumption. Since DON is highly soluble in water it is partitioned into the aqueous phase during the wet-milling process with negligible accumulation in the solid residue used for food products (17,119).

52. Many liquid and gaseous chemicals have been tested for their effectiveness in reducing DON levels in contaminated grain. Most of the chemicals tested resulted in little or no significant reduction in the levels of DON (111,112). Sodium bisulfite was found to reduce DON levels in maize but it cannot be used directly for human food because it affects the rheological properties of flour and the DON adduct formed is not stable and hydrolyzes back to DON under certain processing conditions (116).

53. Ionizing radiation, extrusion and thermal processing procedures have been developed that can reduce DON levels to some extent under very specific conditions. However, there is no single method currently available that can completely remove all DON from cereals (111, 120,121). There is still a need to determine more precisely the optimal extrusion processing conditions to eliminate or remove DON in a practical approach. Aqueous ozone has been shown to degrade many trichothecenes to simpler products, however, the identity and toxicity of the resulting products have not been fully studied (122).

54. Reductions in DON levels were achieved in naturally contaminated wheat kernels processed in superheated steam (123). The largest reductions occurred at 160 and 185°C. Reductions of up to 52% were achieved at 185°C and 6 minute processing time and were due only to thermal degradation and not to solubilization and extraction.

DON in processed products

55. The process of converting raw and milled grains into food for human consumption has significant effects on the levels of DON in the finished products. Humans are exposed to DON contamination mainly as a result of contamination of finished products. DON is a relatively heat stable molecule; it is stable at 120°C, moderately stable at 180°C and partially stable at 210°C. DON is water-soluble and stable under weakly acid conditions but not stable in alkali (22).

56. A total of 190 samples of common wheat, durum wheat and rye flours were collected from mills and retail markets in Denmark between 1998 and 2001 and analyzed for DON and nivalenol (14). DON had an incidence rate of 78% over all samples for all years. The contamination level varied from year to year. The highest incidence and DON levels were found in wheat and rye samples from the 1998 harvest and this was attributed to the unusually cool and wet growing season that year. The mean concentrations in the wheat and rye flours were 191 and 99 $\mu\text{g}/\text{kg}$, respectively. DON was found in approximately 50% of the rye samples collected between 1998 and 2000, with a mean concentration of 49 $\mu\text{g}/\text{kg}$. Durum wheat flour showed the highest DON contamination level and all samples collected during 2000 and 2001 contained DON with means and medians above 100 $\mu\text{g}/\text{kg}$. Over 70% of the samples contained more than 500 $\mu\text{g}/\text{kg}$ DON and the highest observed concentration was 2,591 $\mu\text{g}/\text{kg}$.
57. Of 60 wheat samples analyzed in Argentina for DON, 93.3% were contaminated, and the average level was 1,798 $\mu\text{g}/\text{kg}$ (124). Sixty-one samples of wheat flour were analyzed and the average DON level was 1309 $\mu\text{g}/\text{kg}$; the average DON level in 42 different bakery products was 464 $\mu\text{g}/\text{kg}$. This survey was done on samples from the 1993/94 wheat crop which experienced a rainy season.
58. During the years 2001-2004, a total of 4,965 food samples, purchased from the German market, were analyzed for DON (125). DON was found in most foods containing cereals with an incidence of greater than 50%. For foods such as breads, rolls, and pasta, the incidence of DON contamination was typically 70-90%. The highest DON contamination was found in durum wheat and products thereof. Median DON levels were about 2-10 times higher than those of other frequently contaminated cereals (soft wheat, maize and products thereof) with maximum DON levels of 2,000-3,000 $\mu\text{g}/\text{kg}$. The mean and median average levels for DON in most products, with few exceptions, were well below the maximum permitted levels in the EU (200-750 $\mu\text{g}/\text{kg}$). Relatively minor qualitative and quantitative differences were noted in the contamination of foods with DON between years. Regional differences were not noted, although incidence and levels were much lower in products resulting from organic farming than in those from conventional production.
59. A total of 562 wheat-based products from the 1993 crop year in the United States were collected and analyzed (126). The percent of samples with DON contamination greater than 1,000 $\mu\text{g}/\text{kg}$ in bran, white flour, whole wheat flour, and miscellaneous test samples were 12, 10, 16 and 5, respectively. About 52, 50, 40 and 27% of the same test samples were contaminated with DON at levels > 100 $\mu\text{g}/\text{kg}$. The 1993 wheat crop in the Midwestern part of the United States experienced cool, wet conditions during the spring and summer months thus resulting in elevated levels of DON in the crop harvested that year.
60. A total of 728 wheat-based products were examined in the United States between 2000 and 2004 (94). The percent of samples with DON contamination greater than 1,000 $\mu\text{g}/\text{kg}$ in wheat bran, wheat flour and other milled wheat products were 17.5, 1.0 and 1.8, respectively. The percent of the same test samples containing greater than 100 $\mu\text{g}/\text{kg}$ were 31, 37 and 36, respectively.
61. Adult cereal-based breakfast foods from the Canadian retail market were surveyed for several mycotoxins over a 3-year period beginning in 1999/2000. Depending on the year, DON was detected in 40 to 59% of samples with mean levels ranging from 10 to 70 $\mu\text{g}/\text{kg}$ (127).
62. Cereal-based infant foods from the Canadian retail market were surveyed for several mycotoxins over a 3-year period. DON was detected in 63% of the samples, with mean levels ranging from 32 -150 $\mu\text{g}/\text{kg}$ (128). In a similar survey of cereal-based infant and baby foods in southwest Germany, DON was detected in 60% of the samples with DON levels ranging from 15 -314 $\mu\text{g}/\text{kg}$ (129).
63. A survey of 101 commercially available breads on the German market in 1999, revealed incidence levels of DON, nivalenol and 3-acetyldeoxynivalenol of 92, 5 and 8%, respectively; the median levels in positive samples were 134, 25 and 40 $\mu\text{g}/\text{kg}$, respectively (130). DON levels were lower in bread samples produced from cereals organically grown compared to cereals conventionally grown. Similar results were obtained in Belgium where it was observed that the DON levels in organically grown wheat was lower than in conventionally grown wheat in 2002 and 2003; it was also observed that DON levels above the limit of quantitation occurred more frequently in conventional wheat flours than in organically produced ones but levels of contamination were equivalent in the two types of flours (59). There are many factors that may contribute to the differences observed between organically and conventionally grown cereals and their milled and finished products (131).

64. Two hundred and nineteen samples of grain-based food, pseudocereals and gluten-free food were collected from food and health food stores during 2000 and 2001 in Germany (132). These samples were analyzed for 13 trichothecene toxins including DON, 3-acetyl DON, 15-acetyl DON and nivalenol; the incidences of contamination were 57, 1, 13, and 10% respectively. The highest level of DON was 389 µg/kg while the levels of the other toxins were less than 100 µg/kg.

65. Sixty samples of white and whole grain wheat flour were collected during 1999 from mills and food stores in an area of southwest Germany (83). DON was the predominant toxin found. Based on total samples, the incidence of DON, nivalenol, 3-acetyl DON, 15-acetyl DON, HT-2, T-2 and zearalenone were 98, 12, 2, 3, 7, 2 and 38%, respectively; the median levels of the positive samples were 199, 25, 11, 15, 12, 4 and 3 µg/kg, respectively.

66. Three hundred and thirty-five retail oats samples were collected and analyzed over a 6 month period in 2003 by the Food Standards Agency in the UK (133). A total of 6 trichothecenes were detected at levels ranging from 10 to 404 µg/kg in 52% of the samples; DON, T-2 and HT-2 toxins were among the most common.

67. A survey consisting of 377 retail cereal samples were analyzed for a number of trichothecenes in the UK during 2003 (134). Two hundred and ninety-eight samples (79%) contained detectable toxins; DON and nivalenol were the most frequently occurring. The highest levels of DON occurred mainly in maize-based breakfast cereals and snacks, at 2,261 and 879 µg/kg, respectively.

68. During 2002-2003, 164 samples of wheat flour were analyzed in Japan (97). Eighty-five percent of the samples analyzed in 2002 contained DON with an average level of 138 µg/kg. Seventy-four percent of the 2003 samples contained DON; the average level was 43 µg/kg. The reduction of the DON level in flour samples during 2003 was attributed to the provisional maximum level for DON in husked wheat that was established at 1.1 mg/kg by the government of Japan in May 2002.

69. The controlling regulatory agencies in The Netherlands analyzed samples of cereals and cereal products for DON during the years 2001 to 2004 (96). Of 447 samples of unprocessed cereals analyzed, 19% contained DON levels less than 100 µg/kg, 52% contained levels between 100 and 750 µg/kg, 2% were between 750 and 1000 µg/kg and 3% were above 1000 µg/kg. Two hundred and thirty-nine samples of self rising flour were analyzed for DON; 91% had DON levels less than 100 µg/kg and 9% had levels less than 750 µg/kg. Four hundred and forty-seven pasta samples were also analyzed for DON; 95% of the samples contained less than 500 µg/kg of DON, 4% contained between 500 and 750 µg/kg and 3% contained greater than 750 µg/kg DON. The co-occurrence of ochratoxin A was noted in some samples of the flour and unprocessed cereals.

70. Two hundred samples of wheat bran (farelo) were analyzed in Brazil during 1996-1997. The mean concentration levels of DON were 1412 (1996) and 1506 (1997) µg/kg (92). Seventy-eight samples of wheat flour (farinha) were also analyzed between 2001 and 2004; 34.6 % were contaminated with DON and the mean level was 284 µg/kg and the maximum level was 794 µg/kg. Seventy-two samples of beer were analyzed; 5.25% were contaminated with DON and the levels of contamination ranged from 50-336 µg/kg.

71. A total of 218 maize products including sweet corn, corn on the cob, baby food, corn oil, corn flour, polenta, maize meal, maize pasta, maize based snacks and tortillas were analyzed for DON in the UK during 2003 (95). Levels were low in most of the samples analyzed and only 5 samples (2 maize meals, 2 breakfast cereals and 1 polenta) contained DON above 500 µg/kg. DON was detected at levels between 50 and 500 µg/kg in 36 samples. DON was not detected above 50 µg/kg in the remaining samples.

72. Wheat bran and maize designated for use as raw materials for poultry feed in Kuwait were analyzed for DON (135). DON was detected in 79% of the wheat bran samples and in 91% of the maize samples analyzed at levels up to 220 and 350 µg/kg, respectively. DON levels in feed prepared as broiler starter and broiler finisher ranged from 220 to 1200 µg/kg and the extent of contamination was 79 and 100%, respectively.

73. One hundred and fifty-six samples of breakfast cereals (corn-, oats-, wheat- and rice-based cereals as well as mixed grain cereals) were collected from the Canadian retail marketplace over a three-year period (136). DON was the most frequently detected mycotoxin; it was detected in 40% of all samples analyzed.

74. A total of 78 samples of corn-based products were collected from food shops and supermarkets in Sao Paulo, Brazil between November 2001 and January 2002 (137). DON and nivalenol were detected in only one of 11 samples of pre-cooked corn flour analyzed. The DON and nivalenol levels were estimated to be 167 and 166 µg/kg, respectively. One of 6 samples of corn grits contained HT-2 and T-2 toxins at levels of 555 and 767 µg/kg, respectively.

75. DON was detected in 6 of 68 commercially processed cereal samples in Turkey (138). The maximum detectable amount was 2.67 µg/g (ppm) in a corn flour sample; lower levels were found in dried corn and macaroni.

76. A survey consisting of 685 food control samples of European origin were analyzed in parallel for DON and for T-2 toxin (49). DON was most common and present above 20 µg/kg in 50% of the analyzed samples. Maximum levels of DON in grains (wheat, rye, barley), oats, bran and corn/corn products were 2,580, 2,380, 2,690 and 1,950 µg/kg, respectively. The highest amounts of T-2 toxin were observed in maize (8.4 µg/kg), oats or oat based products (266 µg/kg), respectively. The authors concluded that the content of DON is always some orders of magnitudes higher or at least in the same order compared with the contamination with T-2 toxin, regardless of products or matrices.

Studies associated with the reduction of DON levels in processed foods

77. Comprehensive reviews on the influence and effectiveness of various processing procedures used to reduce the DON levels in milled cereal grain products, beer and finished bakery products have been published (111, 134, 139-141).

78. A study was recently conducted to compare DON levels in organically and conventionally produced beers available in the Belgian market (141). DON levels were found to range from 2 to 22 µg/l (mean = 6 µg/l) in conventional beers, while organic beers ranged from 2 to 14 µg/l (mean = 4 µg/l). The overall incidences of DON were 67 and 80% in conventionally and organically produced beers, respectively.

79. A recent study was conducted in Argentina to determine the distribution of DON in various fractions resulting from the milling of naturally contaminated wheat using an industrial milling process (115). The levels of DON in the raw wheat, flour, bran and gluten were 1,928, 994, 4,680 and 293 µg/kg, respectively.

80. The stability of DON in wheat flour was evaluated during the fermentation stage of bread-making on a pilot scale in Argentina (142). Using flour containing 150 µg/kg of DON, fermenting the dough at 50°C resulted in the DON level being reduced by 56% for Vienna bread and 49 % for French bread. The investigators concluded that the DON reduction during bread making might be due to some process that occurs during yeast fermentation and not solely to thermal decomposition.

81. A detailed study to determine the influence of *Fusarium* infection of wheat on the baking quality of wheat, was conducted in Germany (143). The results indicated that high *Fusarium* infection levels accompanied by high DON levels did not necessarily deteriorate the baking quality of DON contaminated wheat.

82. A study was initiated to determine the extent of reduction in levels of DON that might occur during various steps in the processing of unclean naturally contaminated durum wheat to the production of cooked spaghetti (13). With respect to the unclean wheat, the average levels of DON decreased to 77% in cleaned wheat, 37% in semolina, 33% in raw spaghetti and 20% in the cooked spaghetti. Average DON levels in the screenings, bran and fine middlings were 4.1, 1.6 and 0.6 fold respectively, relative to the unclean wheat. The results from this study conservatively suggest that the cooked pasta retains 25% or less of the level of DON found in grains.

83. The retention of DON in processes involving milling and finished food production using naturally contaminated wheat was studied in Japan. The milled flour retained 40-55% of the original concentration of DON in the raw wheat and almost 200% of the DON concentration was found in the bran fraction (144). Chemical and biological analysis revealed that the DON level in baked bread was retained at the same DON level as in the flour, but, in Japanese-style wheat noodles, only 30% of the DON was detected in the cooked noodles, and the cooking water solids retained more than 40% of the original DON level. From this data and data on the consumption of wheat products in Japan that was obtained from the National Nutrition Survey, it is estimated that the exposure to DON from the final products of wheat flour would be 60-70% of the DON level in the contaminated flour (145).

Regulatory status

84. At least 37 countries have established regulatory limits or guidance levels for DON in foods and feeds (146). The guideline levels for cereal and finished cereal products intended for human consumption range from 100 µg/kg to 2,000 µg/kg. The guidance levels for DON in the diets of swine, poultry and cattle range from 500 µg/kg to 10,000 µg/kg depending on the age of the animal species.

Risk Management

85. The incidence and levels of DON in cereal crops throughout the world vary considerably depending on many factors including environmental conditions, cultivar of cereal planted, and traditional agronomic practices employed in different countries. In order to manage the risk associated with DON contamination in cereals, an integrated risk management system approach is needed. This includes pre-harvest management (including Good Agricultural Practices), harvesting management (to include time of harvest, control of temperature and moisture during transportation and storage of grains) and post-harvest management (including Good Manufacturing Practices, decontamination and diversion strategies) with appropriate controls at each level (147). More information is needed on the year-to-year variability of the levels of DON in cereals grown in many parts of the world as well as the consumption patterns of various populations.

86. In 2003, the Codex Alimentarius Commission adopted a *Code of Practice for The Prevention and Reduction of Mycotoxin Contamination in Cereals, Including Annexes on Ochratoxin A, Zearalenone, Fumonisin and Trichothecenes*. (CAC/RCP 51-2003). The implementation of the practices pointed out in that document, along with advances in post-harvest techniques, proper drying and storage conditions followed by good manufacturing practices (GMPs), may substantially reduce the levels of DON in the food supply.

87. Comprehensive reviews of cropping systems and pre-and post-harvesting agricultural practices that may reduce or prevent *Fusarium* contamination of cereal crops have recently been published in the scientific literature (11, 148-151).

Research Currently Underway

88. Research efforts in some countries are currently focusing on possible ways of reducing the DON levels in finished food products that are prepared from raw cereals that may be contaminated with DON. In studies of this nature, it is important that naturally contaminated (field infected) cereal grains be used as the starting material because the pattern of *Fusarium* infection in the kernels is crucial to the subsequent fate of the toxins during processing (134).

89. The Ministry of Agriculture, Forestry and Fisheries in Japan has funded a preliminary research project aiming at detecting and sorting DON-contaminated wheat kernels (152) The results of the project obtained as of 2006 indicate that a full color sorter, using visible light, is effective in removing the contaminated kernels and thus reducing the DON content. A combination method of visible and near-infrared lights for the more rapid and accurate sorting will be examined within the next three years as a new research project.

90. In cooperation with the cereal industry in the UK, work is being carried out to measure the levels of *Fusarium* mycotoxins including DON in raw cereals (wheat, maize and oats) and then determine how the key stages of food processing affect the contamination of the final food product (95). The research is aimed at determining the factors that affect toxin levels at each process stage by using laboratory and pilot scale studies and sampling from production plants. The knowledge generated from this project is intended to assist industry to further reduce mycotoxin contamination. This work is also considering the formation of metabolites and bound residues and any toxicological implications arising from those. It is anticipated that the final results and findings of this work will be available towards the end of 2008.

91. Canada is developing new HPLC and ELISA methods for DON based on the modification of existing methods (153).

Conclusions and Recommendations

92. Codex member states should be encouraged to continue to submit data from surveys of levels of DON in cereal products in their countries, using validated analytical methods, and over a period of several years to reflect seasonal variations. These data would be used, taking into account regional differences in food consumption patterns, to determine exposure estimates and for use in developing an appropriate international standard for DON in wheat.

93. Research involving the breeding of cereal cultivars (wheat specifically) that are resistant to the growth of *F. graminearum* and *F. culmorum* and the resulting *Fusarium* head blight disease that can develop in wheat, should be encouraged as well as strategies that can be implemented to help prevent the production of trichothecenes in cereal grains.

94. Research on methods to prevent and/or reduce contamination of cereal grains in the field, during storage and processing by *Fusarium* species should be encouraged and continued. There is need for a better understanding of the *Fusarium*-grain interactions in symptomatic and asymptomatic infections of grains in the field. Studies are needed to identify and determine the toxicity of products resulting from the degradation and chemical modification of DON and other trichothecenes as a result of various processing procedures.

95. The Codex Committee on Contaminants in Foods should defer development of international standards until a new overview of exposure data, including more regional data on incidences and levels of DON in cereals over a period of several years, becomes available along with adequate information on consumption patterns for various countries.

96. The toxicity of the 3-acetyl and the 15-acetyl DON that occur along with DON need to be investigated with respect to their contribution to overall DON toxicity since they often occur at levels of 10-20% of the level of DON.

97. In view of the natural co-occurrence of DON, other trichothecenes and zearalenone, more emphasis should be placed on the development and validation of methods capable of detecting multi-mycotoxin residues.

References

1. ALINORM 04/27/12, para.158.
2. ALINORM 05/28/12, para. 149,150.
3. ALINORM 06/29/12, para. 137,138.
4. Miller J.D., Greenhalgh, R., Wang, Y-Z. and Lu, M. Trichothecene chemotypes of three *Fusarium* species. *Mycologia* 83: 121-130, 1991. .
5. Krska, R., Baumgartner S. and Josephs, R. The state-of-the-art in the analysis of type-A and type-B trichothecene mycotoxins in cereals. *Fresenius J. Analytical Chemistry* 371:285-289, 2001.
6. Yoshizawa, T. and Jin, Y-Z. Natural occurrence of acetylated derivatives of deoxynivalenol and nivalenol in wheat and barley in Japan. *Food Addit. Contam.* 12: 689-694, 1995.
7. Ueno, Y. Trichothecenes-Chemical Biological and Toxicological Aspects. Elsevier Science Publishers, New York, pp.7-111, 1983
8. Ueno, Y. Trichothecenes in food. IN: Krogh, P. (ED). *Mycotoxins in Food. Food Science and Technology.* Academic Press, London. pp. 123-147, 1987.
9. Chelkowski, J. Distribution of *Fusarium* species and their mycotoxins in cereal grains. IN: *Mycotoxins in Agriculture and Food Safety.* K.K. Sinha and D. Bhatnager (EDS). Marcel Dekker, Inc. New York, N.Y. pp 45-64, 1998.
10. Placinta, C.M., D'Mello, J.P.F., and Macdonald, A.M.C. A review of worldwide contamination of cereal grains and animal feed with *Fusarium* mycotoxins. *Animal Feed Science Technology* 78:21-37, 1999.
11. Champeil, A., Fourbet, J.F., Dore, T. and Rossignol, L. Influence of cropping system on *Fusarium* head blight and mycotoxin levels in winter wheat. *Crop Protection* 23:531-537 2004.
12. Sutton, J.C. Epidemiology of wheat head blight and maize ear rot caused by *Fusarium graminearum*. *Can. J. Plant Pathol.* 4:195-209, 1982.
13. Visconti, A., Haidukowski, E.M., Pascale, M. and Silvestri, M. Reduction of deoxynivalenol during durum wheat processing and spaghetti cooking. *Toxicology Letters* 153:181-189, 2004.
14. Rasmussen, P.H., Ghorbani, F. and Berg, T. Deoxynivalenol and other *Fusarium* toxins in wheat and rye flours on the Danish market. *Food Addit. Contam.* 20(4):396-404, 2003.
15. Trigo-Stockli, D.M., Sanchez-Mariflez, R.I., Cortez-Rocha, M.O. and Pedersen, J.R. Comparison of the distribution and occurrence of *Fusarium graminearum* and deoxynivalenol in hard red winter wheat for 1993 -1996. *Cereal Chem.* 75(6):841-846, 1998.
16. Scott, P.M. Trichothecenes in grains. *Cereal Foods World* 35:661, 1990.
17. Bennett, G.A. and Richard, J.L. Influence of processing on *Fusarium* mycotoxins in contaminated grains. *Food Technology* 50(5): 235-238, 1996.
18. Scott, P.M. Mycotoxins transmitted into beer from contaminated grains during brewing. *J. AOAC Int.* 79(4):875-881, 1996.
19. Schothorst, R.C. and Jekel, A.A. Determination of trichothecenes in beer by capillary gas chromatography with flame ionization detection. *Food Chem.* 82:475-479, 2003.
20. IARC. Toxins derived from *Fusarium graminearum*, *F. culmorum*, and *F. crookwellense*: zearalenone, deoxynivalenol, nivalenol and fusarenone X. IARC Monograph on the Evaluation of Carcinogenic Risks of Chemicals to Humans, pp. 397-444, 1993.
21. Eriksen, G.S. and Alexander, J. (EDS). *Fusarium* toxins in cereals-a risk assessment. Nordic Council of Ministers. TemaNord 502, pp. 1-115, Copenhagen, 1998.

22. Canady, R.A., Coker, R.D., Egan, S.K., Krska, R., Kuiper-Goodman, T., Olsen, M., Pestka, J., Resnik, S., and Schlatter, J. Deoxynivalenol. IN: Safety Evaluation of Certain Mycotoxins in Food. WHO Food Additive Series 47. pp. 419-555, World Health Organization, Geneva 2001.
23. SCF (Scientific Committee on Food) Opinion of the Scientific Committee on Food on *Fusarium* toxins. Part 6: Group evaluation of T-2 toxin, HT-2 toxin, nivalenol and deoxynivalenol, adopted on 26 February 2002. European Commission SCF/CS/CNTM/MYC/27, Final.
http://www.europa.eu.int/comm/food/fs/sc/scf/out123_en.pdf
24. The EFSA Journal. Opinion of the Scientific Panel on Contaminants in the Food Chain on a request from the Commission related to Deoxynivalenol (DON) as undesirable substance in animal feed (Question N° EFSA-Q-2003-036) Adopted on 2 June, 2004.
25. Pieters, M.N., Freijer, J.L., Baars, A.J., Fiolet, D.C.M., van Klaveren, J. and Slob, W. Risk assessment of deoxynivalenol in food. Concentration limits, exposure and effects. *Adv. Exp. Med. Biol.* 504:235-248, 2002.
26. Schlatter, J. Toxicity data relevant for hazard characterization. *Toxicology Letters* 153:83-89 2004.
27. Pestka, J.J. and Smolinski, A.T. Deoxynivalenol: toxicology and potential effects on humans. *J. Toxicol. Environ. Health, Part B*: 39-69, 2005.
28. Cavret, S. and Lecoecur, S. Fusariotoxin transfer in animal. *Food and chem. toxicol.* 44:444-453, 2006.
29. Rotter, B.A. and Prelusky, D.B. Toxicology of deoxynivalenol (vomitoxin). *J. Toxicol. Environ. Health.* 48:1-34, 1996.
30. Eriksen, G.S. and Pettersson, H. Toxicological evaluation of trichothecenes in animal feed. *Animal Feed Science and Technology* 114: 205-2239, 2004.
31. Prelusky, D.B. Residues in food products of animal origin. IN: Miller, J.D., Trenholm, H.L. (EDS). *Mycotoxins in Grain. Compounds Other Than Aflatoxin.* Eagen Press, St. Paul, MN, pp 405-414, 1994.
32. Li, F-Q., Li, Y.W., Luo, X.Y., and Yoshizawa, T. *Fusarium* toxins in wheat from an area in Henan Province, PR China, with a previous human red mold intoxication episode. *Food Addit. Contam.* 19(2):163-167, 2002.
33. Henry, S.H. and Bosch, F.X. Foodborne disease and mycotoxin epidemiology. IN: Hui, Y.H., Smith, R.A., and Spoerke, D.G. (EDS). *Foodborne Disease Handbook,* Marcel Dekker, Inc. New York:, pp.593-626, 2000.
34. Meko, F.A., Turner, P.C., Ashcroft, A.E., Miller, J.D., Qiao, Y.L., Roth, M.J. and Wild, C.P. Development of a urinary biomarker of human exposure to deoxynivalenol. *Food Chem. Toxicol.* 41:265-273, 2003.
35. Speijers, G.J.A., and Speijers, M.H.M. Combined toxic effects of mycotoxins. *Toxicology Letters* 153:91-98, 2004.
36. Sudakin, D.L. Trichothecenes in the environment: relevance to human health. *Toxicology Letters.* 143, 97-107, 2003.
37. Whitaker, T.B. Sampling techniques. IN: *Methods in Molecular Biology, Vol. 157: Mycotoxin Protocols.* M.W. Trucksess and A.E. Pohland (EDS). Humana Press Inc. Totowa, N.J. pp. 11-24, 2000.
38. Whitaker, T.B., Hagler, W.M., Giesbrecht, F.G. and Johansson, A.S. Sampling, sample preparation, and analytical variability associated with testing wheat for deoxynivalenol. *J. AOAC Int.* 83(5):1285-1292, 2000.
39. Lombaert, G.A. Methods for the determination of deoxynivalenol and other trichothecenes in foods. *Adv. Exp. Med. Biol.* 504: 141-153, 2002.
40. Koch, P. State of the art of trichothecenes analysis. *Toxicology Letters* 153: 109-112, 2004.
41. Krska, R., Welzig, E., Berthiller, F., Molinelli, A. and Mizaikoff, B. Advances in the analysis of mycotoxins and its quality assurance. *Food Addit. Contam.* 22(4): 345-353, 2005.

42. Samar, M.M. and Resnik, S.L. Analytical methods for trichothecenes surveillance- An overview over the period 1990-2000. *Food Sci. Tech. Int.* 8(5):257-268, 2002.
43. Scott, P.M. Mycotoxin methodology. *Food Addit. Contam.* 12(3): 395-403, 1995.
44. Langseth, W. and Rundberget, T. Instrumental methods for determination of nonmacrocylic trichothecenes in cereals, foodstuffs and cultures. *J.Chromatogr. A*, 815:103-121, 1998.
45. Lin, L., Zhang, J., Wang, P., Wang, Y. and Chen, J. Thin-layer chromatography of mycotoxins and comparison with other chromatographic methods. *J. Chromatogr. A* 815:3-20, 1998.
46. Yoshizawa, Y. Chromatographic methods for trichothecenes. IN: *Methods in Molecular Biology*, Vol. 157: *Mycotoxin Protocols*. M.W. Trucksess and A.E. Pohland (EDS). Humana Press Inc. Totowa, N.J. pp.115-129. 2000.
47. Cavaliere, C., D'Ascenzo, G., Foglia, P., Pastorini, E., Samperi, R. and Lagana, A. Determination of type B trichothecenes and macrocyclic lactone mycotoxins in field contaminated maize. *Food Chem.* 92: 559-568, 2005.
48. Berthiller, F., Schuhmacher, R., Buttinger, G. and Krska, R. Rapid simultaneous determination of major type A- and B- trichothecenes as well as zearalenone in maize by high performance liquid chromatography-tandem mass spectrometry. *J. Chromatogr. A.* 1062:209-216, 2005.
49. Biselli, S. and Hummert, C. Development of a multicomponent method for *Fusarium* toxins using LC-MS/MS and its application during a survey for the content of T-2 toxin and deoxynivalenol in various feed and food samples. *Food Addit. Contam.* 22(8): 752-760, 2005.
50. MacDonald, S.J., Chan, D., Brereton, P., Damant, A. and Wood, R. Determination of deoxynivalenol in cereals and cereal products by immunoaffinity column cleanup with liquid chromatography: interlaboratory study. *J.AOAC Int.* 88(4): 1197-1204, 2005.
51. Cavaliere, C., Foglia, P., Pastorini, E., Samperi, R. and Lagana, A. Development of a multiresidue method for analysis of major fusarium mycotoxins in corn meal using liquid chromatography/tandem mass spectrometry. *Rapid Commun. Mass Spectrom.* 19(14):2085-2093, 2005.
52. Razzazi-Fazeli, E., Bohm, J., Jarukamjorn, K. and Zentek, J. Simultaneous determination of major B-trichothecenes and the de-epoxy-metabolite of deoxynivalenol in pig urine and maize using high-performance liquid chromatography-mass spectrometry. *J. Chromatogr. B. Analyt. Technol. Biomed. Life Sci.* 796:21-33, 2003.
53. Royer, D., Humpf, H-U. and Guy, P.A. Quantitative analysis of *Fusarium* mycotoxins in maize using accelerated solvent extraction before liquid chromatography/atmospheric pressure chemical ionization tandem mass spectrometry. *Food Addit. Contam* 21(7):678-692, 2004.
54. Plattner, R.D. and Maragos, C.M. Determination of deoxynivalenol and nivalenol in corn and wheat by liquid chromatography with electrospray mass spectrometry. *J..AOAC Int.* 86(1):61-65, 2003.
55. Abramovic, B., Jajic, I., Juric, V. and Gaal, F.F. Optimization of the determination of deoxynivalenol in corn by liquid chromatography and a comparison of two clean-up principles. *J. Serb. Chem. Soc.* 70(7): 1005-1013, 2005.
56. Tanaka, H., Takino, M., Sugita-Konishi, Y. and Tanaka, T. Development of a liquid chromatography/time-of-flight mass spectrometric method for the simultaneous determination of trichothecenes, zearalenone and aflatoxins in foodstuffs. *Rapid Commun. Mass Spectrom.* 20 (9):1422-1428, 2006.
57. Klotzel, M., Lauber, U. and Humpf, H-U. A new solid phase extraction clean-up method for the determination of 12 type A and B trichothecenes in cereals and cereal-based food by LC-MS/MS. *Mol. Nutr. Food Res.* 50:261-269, 2006.
58. Stroka, J., Derbyshire, M., Mischke, C., Ambrosio, M., Kroeger, K. Arranz, I., Sizoo, E. and van Egmond, H. Liquid chromatographic determination of deoxynivalenol in baby food and animal feed: interlaboratory study. *J AOAC Int* 89:1012, 2006.

59. Pussemier, L., Pierard, J.-Y., Anselme, M., Tangni, E.K., Motte, J.-C. and Larondelle, Y. Development and application of analytical methods for the determination of mycotoxins in organic and conventional wheat. *Food Addit. Contam.* 23(11): 1208-1218, 2006.
60. Sugita-Konishi, Y., Tanaka, T., Tabata, S., Nakajima, M., Nouno, M., Nakaie, Y., Chonan, T., Aoyagi, M., Kibune, N., Mizuno, K., Ishikuro, E., Kanamaru, N., Minamisawa, M., Aita, N., Kushiro, M., Tanaka, K. and Takatori, K. Validation of an HPLC analytical method coupled to a multifunctional clean-up column for the determination of deoxynivalenol. *Mycopathologia* 161:239-243, 2006.
61. Tanaka, H., Takino, M., Sugita-Konishi, Y., Tanaka, T., Development of a liquid chromatography/time of flight mass spectrometric method for the simultaneous determination of trichothecenes, zearalenone and aflatoxins in foodstuffs, *Rapid Commun Mass Spectrom.* 20, 1422-1428, 2006.
62. Eke, Z., Kende, A. and Torkos, K. Simultaneous detection of A and B trichothecenes by gas chromatography with flame ionization or mass selective detection. *Microchemical J.* 78:211-216, 2004.
63. Eke, Z. and Torkos, K. *N,N*-dimethyl-trimethylsilyl-carbamate as a derivatizing agent in gas chromatography of trichothecene mycotoxins. *Microchemical J.* 77:43-46, 2004.
64. Melchert, H-U. and Pabel, E. Reliable identification and quantification of trichothecenes and other mycotoxins by electron impact and chemical ionization-gas chromatography-mass spectrometry, using an ion-trap system in the multiple mass spectrometry mode. Candidate reference method for complex matrices. *J. chromatogr. A* 1056:195-199, 2004.
65. Olsson, j., Borjesson, T., Lundstedt, T. and Schnurer, J. Detection and quantification of ochratoxin A and deoxynivalenol in barley grains by GC-MS and electronic nose. *Int. J. food Microbiol.* 72:203-214, 2002.
66. Jestoi, M., Ritieni, A. and Rizzo, A. Analysis of the *Fusarium* mycotoxins fusaproliferin and trichothecenes in grains using gas chromatography-mass spectrometry. *J. Agric. Food Chem.* 52:1464-1469, 2004.
67. Wilson, D.M., Sydenham, E.W., Lombaert, G.A., Trucksess, M.W., Abramson, D. and Bennett, G.A.. Mycotoxin analytical techniques. IN: *Mycotoxins in Agriculture and Food Safety*. K.K. Sinha and D. Bhatnager (EDS). Marcel Dekker, Inc. New York, N.Y. pp135-182, 1998.
68. Schneider, E., Curtui, V., Seidler, C., Dietrich, R., Usleber, E. and Martlbauer, E. Rapid methods for deoxynivalenol and other trichothecenes. *Toxicology Letters* 153:113-121, 2004.
69. Zheng, M.Z., Richard, J.L. and Binder, J. A review of rapid methods for the analysis of mycotoxins. *Mycopathologia* 161:261-273, 2006.
70. Yoshizawa, T., Kohno, H., Ikeda, K., Shinoda, T., Yokohama, H., Morita, K., Kusada, O. and Kobayaashi, Y. A practical method for measuring deoxynivalenol, nivalenol, and T-2 + HT-2 toxin foods by an enzyme-linked immunosorbent assay using monoclonal antibodies. *Biosci. Biotechnol. Biochem.* 68(10):2076-2085, 2004.
71. Josephs, R.D., Derbyshire, M., Stroka, J., Emons, H. and Anklam, E. Trichothecenes: reference materials and method validation. *Toxicology Letters* 153:123-132 2004.
72. Krska, R., Sente, E., Freudenschuss, M., Hametner, C. and Zoller, P. Purity assessment of commercially available crystalline deoxynivalenol. *J. AOAC Int.* 87(4): 909-919 2004.
73. Krska, R., Schothorst, R.C., van Egmond, H.P., Josephs, R.D., Lepschy, J., Pettersson, H., Chan, D., Berthiller, F., Schuhmacher, R., Kandler, W., Parich, A. and Welzig, E. Processing and purity assessment of standards for the analysis of type-B trichothecene mycotoxins. *Anal. Bioanal. Chem.* 382(8):1848-1858, 2005.
74. Bretz, M., Beyer, M., Cramer, B. and Humpf, H-U. Stable isotope dilution analysis of the *Fusarium* mycotoxins deoxynivalenol and 3-acetyldeoxynivalenol. *Mol. Nutr. Food Res.* 50:251-260,2006.
75. Bretz, M., Beyer, M., Cramer, B. and Humpf, H-U. Synthesis of stable isotope labeled 3-acetyldeoxynivalenol. *Mol. Nutr. Food Res.* 49:1151-1153, 2005.
76. Haubl, G., Berthiller, F., Krska, R. and Schuhmacher, R. Suitability of a fully ¹³C isotope labeled internal standard for the determination of the mycotoxin deoxynivalenol by LC-MS/MS without clean up. *Anal. Bioanal. Chem.* 384:692-696, 2006.

77. Trucksess, M., Page, S., Wood, G. and Cho, T. Determination of deoxynivalenol in white flour, whole wheat flour, and bran by solid phase extraction/liquid chromatography: Interlaboratory study. *J. AOAC Int.* 81:880-886. 1998.
78. Joseph, R.D., Schuhmacher, R. and Krska, R. International interlaboratory study for the determination of the *Fusarium* mycotoxins zearalenone and deoxynivalenol in agricultural commodities. *Food Addit. Contam.* 18(5):417-430, 2001.
79. Biselli, S., Hartig, L., Wegner, H. and Hummert, C. Analysis of *Fusarium* toxins using LC-MS-MS. Application to various food and feed matrices. *LC-GC North America*, 23 (4): 404-416, 2005.
80. Tanaka, T., Hasegawa, A., Yamamoto, S., Lee, U-S., Sugiura, Y. and Ueno, Y. Worldwide contamination of cereals by the *Fusarium* mycotoxins nivalenol, deoxynivalenol, and zearalenone. 1. Survey of 19 countries. *J. Agric. Food Chem.* 36: 979-983, 1988.
81. Jelinek, C.F., Pohland, A.E. and Wood, G.E. Worldwide occurrence of mycotoxins in foods and feeds-an update. *J. Assoc. Off. Anal. Chem.* 72(2):223-230, 1989.
82. Abramson, D. Mycotoxin formation and environmental factors. IN: *Mycotoxins in Agriculture and Food Safety*. K.K.Sinha and D. Bhatnagar (EDS). Marcel Dekker, Inc., New York, pp.255-277, 1998.
83. Schollenberger, M., Jara, H.T., Suchy, S., Drochner, W. and Muller, H-M. *Fusarium* toxins in wheat collected in an area in southwest Germany. *Int. J. Food Microbiol.* 72:85-89, 2002.
84. Scott, P.M., Kanhere, S.R., Dexter, J.E., Brennan, P.W. and Trenholm, H.L. Distribution of the trichothecene mycotoxin deoxynivalenol (vomitoxin) during the milling of naturally contaminated hard red spring wheat and its fate in baked products. *Food Addit. Contam.* 1(4): 313-323, 1984.
85. Berthiller, F., Dall'Asta, C., Schuhmacher, R., Lemmens, M., Adam, G. and Krska, R. Masked mycotoxins: determination of a deoxynivalenol glucoside in artificially and naturally contaminated wheat by liquid chromatography- tandem mass spectrometry. *J. Agric. Food Chem.* 53:3421-3425, 2005.
86. Schothorst, R.C. and van Egmond, H.P. Report from SCOOP task 3.2.10 "collection of occurrence data of *Fusarium* toxins in food and assessment of dietary intake by the population of EU member states" Subtask:trichothecenes. *Toxicology Letters* 153: 133-143, 2004.
87. MacDonald, S., Prickett, T.J., Wildey, K.B. and Chan, D. Survey of ochratoxin A and deoxynivalenol in stored grains from the 1999 harvest in the UK. *Food Addit. Contam.* 21(2):172-181, 2004.
88. Sokolovic, M. and Simpraga, B. Survey of trichothecene mycotoxins in grains and animal feed in Croatia by thin-layer chromatography. *Food Control* 17:733-740, 2006.
89. Muller, H-M., Reimann, J., Schumacher, U. and Schwadorf, K. Natural occurrence of *Fusarium* toxins in oats harvested during five years in an area of southwest Germany. *Food Addit. Contam.* 15(7):801-806, 1998.
90. Tutelyan, V.A. Deoxynivalenol in cereals in Russia. *Toxicology Letters* 153:173-179 2004.
91. Al-Julaifi, M.Z, and Al-Falih, A.M. Detection of trichothecenes in animal feeds and foodstuffs during the years 1997 to 2000 in Saudi Arabia. *J. Food Prot.* 64(10):1603-1606, 2001.
92. Martinelli, M.A. Privileged Communication. Brazil, 2006.
93. Schollenberger, M., Muller, H-M., Ruffle, M., Suchy, S., Plank, S. and Drochner, W. Natural occurrence of 16 *Fusarium* toxins in grains and feedstuffs of plant origin from Germany. *Mycopathologia* 161:43-52, 2006.
94. Wood, G.E. Privileged Communication, U.S. 2005.
95. Matthews, W. Privileged Communication. U.K. 2005.
96. Tas, W. Privileged Communication. The Netherlands, 2005.
97. Fukushima, K. Privileged Communication, Japan, 2005.
98. Fukushima, K. Privileged Communication. Japan, 2006.

99. Ministry of Agriculture, Forestry and Fisheries of Japan. Domestically produced cereals survey (in Japanese). April, 27, 2003, May 9, 2003, March 17, 2005 and May 23, 2006. http://www.maff.go.jp/syohi_anzen/kabi/chosa_kekka.html Survey.
100. Salmon, S. Monitoring of contaminants in wheat grain. Home Grown Cereals Authority Project Report No. 386, HGCA, London, 2006.
101. Baxter, D. Review of food safety issues relating to the supply and market acceptability of UK malting barley and UK malt. Home grown Cereals Authority Project Report No. 380, HGCA, London, 2006.
102. Lindhauer, M., Münzing, K., Seling, S., Betsche, T., Kersting, H.J., Masloff, S., Seifert, M. High-quality cereals through continuous quality controls. Special yield and quality assessment as an advisory instrument for agricultural and consumer policy and its target groups. Research report 2: 21-25, 2005 (www.bmvel-forschung.de).
103. Masloff, S., Betsche, T. and Wolff, J. Evaluation of processing suitability of bread cereals in official responsibility. Mycotoxin Research 21: 94-96, 2005.
104. Masloff, S. Undesirable substances. Special yield and quality assessment 2004. Series: data analyses, BMELV, Germany, pp. 42-43, 2004 (www.bfel.de).
105. Masloff, S. Undesirable substances. Special yield and quality assessment 2005. Series: data analyses, BMELV, Germany, pp. 42-43, 2005 (www.bfel.de).
106. Abramson, D. Mycotoxin formation and environmental factors. IN: Mycotoxins in Agriculture and Food Safety. K.K.Sinha and D. Bhatnager (EDS). Marcel Dekker, Inc. New York, N.Y. pp 255-277. 1998.
107. Pietri, A., Bertuzzi, T., Pallaroni, L. and Piva, G. Occurrence of mycotoxins and ergosterol in maize harvested over 5 years in Northern Italy. Food Addit. Contam. 21(5):479-487, 2004.
108. Lagana, A., Curini, R., D'Ascenzo, G., De Leva, I., Faberi, A. and Pastorini, E. Liquid chromatography/tandem mass spectrometry for the identification and determination of trichothecenes in maize. Rapid Commun. Mass Spectrom. 17(10):1037-1043, 2003.
109. Desjardins, A. E., Manandhar, G., Plattner, R.D., Maragos, C.M., Shrestha, K. and McCormick, S.P. Occurrence of *Fusarium* species and mycotoxins in Nepalese maize and wheat and the effect of traditional processing methods on mycotoxin levels. J. Agric. Food Chem. 48:1377-1383, 2000.
110. Sardjono, A.N., Yamashita, A. and Yoshizawa, T. Natural co-occurrence of aflatoxins and *Fusarium* mycotoxins (fumonisins, deoxynivalenol, nivalenol and zearalenone) in corn from Indonesia. Food Addit. Contam. 15(4):377-384, 1998.
111. Jackson, L.S. and Bullerman, L.B. Effect of processing on *Fusarium* mycotoxins. Adv. Exp. Med. Biol. 459:243-261, 1999.
112. Charmley, L.L. and Prelusky, D.B. Decontamination of *Fusarium* mycotoxins. IN: Miller, J.D., Trenholm, H.L. (EDS). Mycotoxins in Grain. Compounds Other Than Aflatoxin. Eagen Press, St. Paul, MN, pp. 421-435, 1994.
113. Trigo-Stockli, D.M., Deyoe, C.W., Satumbaga, R.F. and Pedersen, J.R. Distribution of deoxynivalenol and zearalenone in milled fractions of wheat. Cereal Chem. 73(3):388-391, 1996.
114. Lee, U-S., Jang, H-S., Tanaka, T., Oh, Y-J., Cho, C-M and Ueno, Y. Effect of milling on decontamination of *Fusarium* mycotoxins nivalenol, deoxynivalenol, and zearalenone in Korean wheat. J. Agric. Food Chem. 35:126-129, 1987.
115. Samar, M.M., Fontan, C.F., Resnik, S.L., Pacin, A.M. and Castillo, M.D. Distribution of deoxynivalenol in wheat, wheat flour, bran, and gluten, and variability associated with the test procedure. J. AOAC Int. 86(3):551-556, 2003.
116. Young, J.C., Subryan, L.M., Potts, D., McLaren, M.E. and Gobran, F.H. Reduction in levels of deoxynivalenol in contaminated wheat by chemical and physical treatment. J. Agric. Food Chem. 34:461-465, 1986.

117. Nowicki, T.W., Gaba, D.G., Dexter, J.E., Matsuo, R.R. and Clear, R.M. Retention of DON in wheat during processing and cooking of spaghetti and noodles. *J. Cereal Science* 8:189-202, 1988.
118. Patey, A.L. and Gilbert, J. Fate of *Fusarium* mycotoxins in cereals during food processing and methods for their detoxification. IN: Chelkowski, J. (ED), *Fusarium: Mycotoxins, Taxonomy, and Pathogenicity*. Elsevier: New York. pp.399-420, 1989.
119. Lauren, D.R. and Ringrose, M.A. Determination of the fate of three *Fusarium* mycotoxins through wet-milling of maize using an improved HPLC analytical technique. *Food Addit. Contam.* 14(5):435-443, 1997.
120. Castells, M., Marin, S., Sanchis, V. and Ramos, A.J. Fate of mycotoxins in cereals during extrusion cooking: a review. *Food Addit. Contam.* 22(2):150-157. 2005.
121. Cetin, Y. and Bullerman, L.B. Confirmation of reduced toxicity of deoxynivalenol in extrusion-processed corn grits by the MTT bioassay. *J. Agric. Food Chem.* 54:1949-1955, 2006.
122. Young, J.C., Zhu, H. and Zhou, T. Degradation of trichothecene mycotoxins by aqueous ozone. *Food Chem. Toxicol.* 44:417-424, 2006.
123. Pronyk, C., Cenkowski, S. and Abramson, D. Superheated steam reduction of deoxynivalenol in naturally contaminated wheat kernels. *Food Control* 17:789-796, 2006.
124. Pacin, A.M.; Resnik, S.L.; Neira, M.S.; Molto, G. and Martinez, E. Natural occurrence of deoxynivalenol in wheat, wheat flour and bakery products in Argentina. *Food Addit. Contam.* 14(4): 327-331, 1997.
125. Curtui, V., Brockmeyer, A., Dietrich, R., Kappenstein, O., Klaffke, H., Lepschy, J., Maertlbauer, E., Schneider, E., Seidler, C., Thielert, G., Usleber, E., Weber, R. and Wolff, J. German research project "Analysis and occurrence of important *Fusarium* toxins (Deoxynivalenol, Zearalenone) and dietary intake of these toxins by the German consumer". SANCO/2004/2884.
126. Trucksess, M.W., Ready, D.E., Pender, M.K., Ligmond, C.A., Wood, G.E. and Page, S.W.. Determination and survey of deoxynivalenol in white flour, whole wheat flour, and bran. *J. AOAC Int.* 79(4): 883-887, 1996.
127. Health Canada, Privileged Communication. 2005.
128. Lombaert, G.A., Pellaers, P., Roscoe, V., Mankotia, M., Neil, R. and Scott, P.M. Mycotoxins in infant cereal foods from the Canadian retail market. *Food Addit. Contam.* 20(5):494-504 2003.
129. Schollenberger, M., Suchy, S., Jara, H.T., Drochner, W. and Muller, H-M. A survey of *Fusarium* toxins in cereal-based foods marketed in an area of southwest Germany. *Mycopathologia* 147:49-57, 1999.
130. Schollenberger, M., Drochner, W., Ruffle, M., Suchy, S., Terry-Jara, H. and Muller, H-M. Trichothecene toxins in different groups of conventional and organic bread of the German market. *J. Food Comp. Anal.* 18: 69-78, 2005.
131. Pussemier, L., Larondelle, Y., Van Peteghem, C. and Huyghebaert, A. Chemical safety of conventionally and organically produced foodstuffs, a tentative comparison under Belgium conditions. *Food Control* 17:14-21, 2006.
132. Schollenberger, M., Muller, H.-M., Ruffle, M., Suchy, S., Planck, S. and Drochner, W. Survey of *Fusarium* toxins in foodstuffs of plant origin marketed in Germany. *Int. J. Food Microbiol.* 97: 317-326, 2005.
133. United Kingdom Food Standards Agency. Retail oat products survey. February 6, 2004, <http://food.gov.uk/multimedia/webpage/174922>
134. Hazel, C.M. and Patel, S, Influence of processing on trichothecene levels. *Toxicology Letters* 153:51-59 2004.
135. Beg, M.U., Al-Mutairi, M., Beg, K.R., Al-Mazeedi, H.M., Ali, L.N. and Saeed, T. Mycotoxins in poultry feed in Kuwait. *Arch. Environ. Contam. Toxicol.* 50: 594-602, 2006.

136. Lombaert, G.A., Roscoe, V.A., Huzel, V., Neumann, G., Melietio, J., Kitchen, D., Kotello, S., Krakalovich, T., Trelka, R.W. and Scott, P.M. Mycotoxins in breakfast cereals from the Canadian retail market: Three-year survey. AGFD Abstract 121, 232nd ACS National Meeting, San Francisco, CA September 2006.
137. Milanez, T.V., Valente-Soares, L.M. and Baptista, G.G. Occurrence of trichothecene mycotoxins in Brazilian corn-based food products. *Food Control* 17:293-298, 2006.
138. Omurtag, G.Z. and Beyoglu, D. Occurrence of deoxynivalenol (vomitoxin) in processed cereals and pulses in Turkey. *Food Addit. Contam.* 20(4): 405-409, 2003.
139. Trigo-Stockli, D.M. Effect of processing on deoxynivalenol and other trichothecenes. *Adv. Exp. Med. Biol.* 504:181-188, 2002.
140. Wolf-Hall, C.E. and Schwarz, P.B. Mycotoxin and fermentation-beer production. *Adv. Exp. Med. Biol.* 504:217-226, 2002.
141. Anselme, M., Tangni, E.K., Pussemier, L., Motte, J.-C., Van Hove, F., Schneider, Y.-J., Van Peteghem, C. and Larondelle, Y. Comparison of ochratoxin A and deoxynivalenol in organically and conventionally produced beers sold on the Belgian market. *Food Addit. Contam.* 23(9):910-918, 2006.
142. Samar. M.M., Neira, M.S., Resnik, S.L. and Pacin, A. Effects of fermentation on naturally occurring deoxynivalenol (DON) in Argentinean bread processing technology. *Food Addit. Contam.* 18(11): 1004-1010, 2001.
143. Prange, A., Birzele, B., Kramer, J., Meier, A., Modrow, H. and Kohler, P. *Fusarium*-inoculated wheat: deoxynivalenol contents and baking quality in relation to infection time. *Food Control* 16: 739-745, 2005.
144. Sugita-Konishi, Y. Privileged Communication. Japan, 2005.
145. Sugita-Konishi, Y., Park, B.J., Kobayashi-Hattori, K., Tanaka, T., Chonan, T., Yoshikawa, K. and Kumagai, S. Effect of cooking process on the deoxynivalenol content and its subsequent cytotoxicity in wheat products. *Biosci. Biotech. Biochem.* 70:1764-1768. 2006.
146. FAO Worldwide regulations for mycotoxins in food and feed in 2003, FAO Food and Nutrition Paper 81. Food and Agriculture Organization, Rome, Italy ISBN 92-5-105162-3. 2004.
147. Lopez-Garcia, R. and Park, D.L. Effectiveness of postharvest procedures in management of mycotoxin hazards. IN: *Mycotoxins in Agriculture and Food Safety*. K.K. Sinha and D. Bhatnagar (EDS). Marcel Dekker, Inc. New York, pp. 407-433, 1998.
148. Schrodter, R. Influence of harvest and storage conditions on trichothecenes levels in various cereals. *Toxicology Letters* 153:47-49, 2004.
149. Aldred, D. and Magan, N. Prevention strategies for trichothecenes. *Toxicology Letters* 153:165-171 2004.
150. Edwards, S.G. Influence of agricultural practices on *Fusarium* infection of cereals and subsequent contamination of grain by trichothecene mycotoxins. *Toxicology Letters* 153:29-35, 2004.
151. Snijders, C.H.A. Resistance in wheat to *Fusarium* infection and trichothecene formation. *Toxicology Ltrs.* 153:37-46, 2004.
152. Takafumi, I. Improvement on Wheat Flour Quality-Production of Quality Wheat Flour by Colour Sorting and Debranning. Abstract of Australasian Milling Conference – 9th Biennial Conference of the Flour Millers' Council of Australia and the Stock Feed Manufacturers' Council of Australia pp 133-138, 2006
153. Health Canada. Privileged Communication. 2006.