SPINETORAM (233)

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EXPLANATION

Spinetoram, a multi-component tetracyclic macrolide in the class of spinosyn insecticides, was developed for the control of lepidopterous larvae, leafminers, and thrips on a variety of crops. Its mode of action is disruption of nicotinic/gamma amino butyric acid-gated chloride channels.

It was identified as a priority new compound at the 39th Session of the CCPR in 2007 (ALINORM 07/30/24-Rev.1) for evaluation by the 2008 JMPR. The Meeting received information on physical and chemical properties, animal and plant metabolism, environmental fate, analytical methods, storage stability, use patterns, supervised trials, processing and farm animal feeding.

IDENTITY

Spinetoram consists of two closely related active ingredients, as shown below, present approximately in a three to one ratio. The term spinetoram is used interchangeably for the combination of these two compounds. Spinetoram is a chemically-modified fermentation product of Saccharopolyspora spinosa.

ISO common name: Spinetoram

(Mixture of two main components as shown below)

Chemical name

IUPAC: (2R,3aR,5aR,5bS,9S,13S,14R,16aS, 16bR)-2-(6-deoxy-3-O-Major

component ethyl-2,4-di-O-methyl-α-L-mannopyranosyloxy)-13-

[(2R,5S,6R)-5-(dimethylamino)tetrahydro-6-methylpyran-2-(XDE-175-J)

yloxy]-9-ethyl-2,3,3a,4,5,5a,5b,6,9,10,11,12,13,14,16a,16b-

hexadecahydro-14-methyl-1*H-as*-indaceno[3,2-

doxacyclododecine-7,15-dione

Minor (2R,3aR,5aS,5bS,9S,13S,14R,16aS, 16bS)-2-(6-deoxy-3-O-

component ethyl-2,4-di-O-methyl-α-L-mannopyranosyloxy)-13-

[(2R,5S,6R)-5-(dimethylamino)tetrahydro-6-methylpyran-2-(XDE-175-L) yloxy]-9-ethyl-2,3,3a,5a,5b,6,9,10,11,12,13,14,16a,16b-

tetradecahydro-4,14-dimethyl-1*H-as*-indaceno[3,2-

doxacyclododecine-7,15-dione

CAS: (2R.3aR.5aR.5bS.9S.13S.14R.16aS. 16bR)-2-[(6-deoxy-3-O-Major

ethyl-2,4-di-O-methyl- α -L-mannopyranosyl)oxy]-13component [[(2R,5S,6R)-5-(dimethylamino)tetrahydro-6-methyl-2H-

(XDE-175-J)

pyran-2-yl]oxy]-9-ethyl-

2,3,3a,4,5,5a,5b,6,9,10,11,12,13,14,16a,16b-hexadecahydro-14-methyl-1*H-as*-indaceno[3,2-*d*]oxacyclododecine-7,15-

Minor (2R,3aR,5aS,5bS,9S,13S,14R,16aS, 16bS)-2-[(6-deoxy-3-Ocomponent ethyl-2,4-di-O-methyl- α -L-mannopyranosyl)oxy]-13-

[(2R,5S,6R)-5-(dimethylamino)tetrahydro-6-methyl-2H-(XDE-175-L)

pyran-2-yl]oxy]-9-ethyl-

2,3,3a,5a,5b,6,9,10,11,12,13,14,16a,16b-tetradecahydro-4,14-dimethyl-1*H-as*-indaceno[3,2-*d*]oxacyclododecine-

7,15-dione

CAS Registry No.: XDE-175-J 187166-40-1

XDE-175-L 187166-15-0

CIPAC No.: 802

Synonyms for active X574175 (2003-2004) substance: XDE-175 (2004-2007)

DE-175 (2007-)

Structural formula: XDE-175-J

XDE-175-L

Molecular formula: XDE-175-J $C_{42}H_{69}NO_{10}$

XDE-175-L $C_{43}H_{69}NO_{10}$

Molecular weight: XDE-175-J 748.02

XDE-175-L 760.03

PHYSICAL AND CHEMICAL PROPERTIES

Pure active ingredient

Property	Results		Reference
	XDE-175-J	XDE-175-L	
Appearance:	White powder	White to yellow crystals	Jennings, 2007, FAPC 73363 & FAPC 73364
Odour:	Odourless	Almond odour	Jennings, 2007, FAPC 73363 & FAPC 73364
Melting point:	143.4 °C	70.8 °C	Madsen & Jennings, 2005 FAPC-052-002 & FAPC- 052-003
Relative density:	1.1495 g/cm ³ at 19.5 °C	1.1807 g/cm ³ at 20.1 °C	Tunink, 2006 NAFST-06- 134

ъ.	D 1		D. C
Property	Results	VDF 175 I	Reference
C-1-1:1:4 :4	XDE-175-J	XDE-175-L	
Solubility in water at 2 Unbuffered:		21.0 mg/I	Comb 2005 NAEST 05
pH 5:	10.0 mg/L 423 mg/L	31.9 mg/L	Comb, 2005 NAFST-05- 071 & NAFST-05-072
рн 3. pH 7:	11.3 mg/L	1.63 g/L 46.7 mg/L	0/1 & NAFS1-03-0/2
pH 9:	11.3 mg/L	1.98 mg/L	
pH 10:	6.27 mg/L	0.706 mg/L	
Vapour pressure:	5.3×10^{-5} Pa at 20 °C	2.1×10^{-5} Pa at 20 °C	Comb, 2005 NAFST-05-
rapour pressure.	3.5 × 10 1 a at 20 C	2.1 × 10 1 a at 20 C	073 & NAFST-05-074
Volatility, Henry's La	w Constant at 20 °C (calcula	tion):	0,0 00 1,111 51 00 0, 1
Unbuffered:	$4.0 \times 10^{-3} \text{ Pa·m}^3/\text{mol}$	$5.0 \times 10^{-4} \text{ Pa·m}^3/\text{mol}$	Huntley, 2005 NAFST-05-
pH 5:	$9.4 \times 10^{-5} \text{ Pa·m}^3/\text{mol}$	$9.8 \times 10^{-3} \text{ Pa·m}^3/\text{mol}$	129
pH 7:	$3.5 \times 10^{-3} \mathrm{Pa \cdot m^3/mol}$	$3.4 \times 10^{-4} \mathrm{Pa \cdot m^3/mol}$	
pH 10:	$6.3 \times 10^{-3} \mathrm{Pa \cdot m^3/mol}$	$2.3 \times 10^{-2} \text{ Pa·m}^3/\text{mol}$	
*	n coefficient at 20 °C (logPo		
pH 5:	2.44	2.94	Comb, 2005 NAFST-05-
pH 7:	4.09	4.49	075 & NAFST-05-076
pH 9:	4.22	4.82	073 & 174 51 03 070
Dissociation constant		2	
Dissociation constant	7.86	7.59	Madsen & Jennings, 2005 FOR-05-043 & FOR-05- 044
Hydrolysis at 20 °C:			
pH 5:	Stable against hydrolysis	Stable	Laughlin et al., 2005
•	Stable		040108
pH 7:	Stable	Stable	
pH 9:		DT50=154 days	
Photolysis:	Numerous (> 70) minor	N-demethyl-175-L and	Yoder, 2005 040079
	breakdown products	numerous (>70) numerous	
	DT50: 0.5 days	minor breakdown products	
	Mass balance:	DT50: 0.3 days	
	$101.1 \pm 3.7\%$	Mass balance: $94.2 \pm 2.0\%$	
Quantum yield at 25 $^{\circ}$			
	0.042	0.066	Yoder, 2005 040079
TD1 1.1.C	. 2505	1 1 2 3	
Theoretical lifetime in	aqueous systems at 25°C (c		X. 1. 2005 040070
	0.5 days	0.3 days	Yoder, 2005 040079
To along and and and all			
Technical material			
Property	Results		Reference
Appearance:	Off-white solid		Madsen & Jennings, 2005 FAPC-052-004
Odour	Musty odour		Madsen & Jennings, 2005 FAPC-052-004
Solvent solubility at	Methanol	> 250 g/L	Comb, 2005 NAFST-05-
20 °C	Acetone	> 250 g/L	078
	Xylene	> 250 g/L	
	1,2-Dichloroethane	> 250 g/L	
	Ethyl acetate	> 250 g/L	
	n-Octanol	132 g/L	
Formulations:	Water dispersible granule	(WG) formulation containing	
	250 g ai/kg		
	Suspension concentrate (S	C) formulation containing 120	

g ai/L

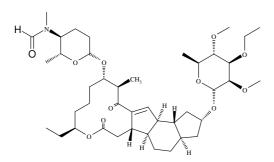
METABOLISM AND ENVIRONMENTAL FATE

The following links manufacturer code number and structure or description of the compounds appearing in the various metabolism and environmental fate studies.

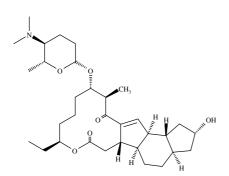
Structure of compounds appearing in metabolism and environmental fate studies

XDE-175-J

Plants, animals, soil, water, rotational crops



N-formyl-175-J Plants



C9-pseudoaglycone-175-J

Plants

N-demethyl-175-J

Plants, animals, soil, water, rotational crops

3-O-deethyl-175-J

Plants

Aglycone-175-J

Plants

C17-pseudoaglycone-175-J

Soil, plants

2'-O-demethyl-175-J

Animals

N-demethyl-N-nitroso-175-J

Soil

XDE-175-L

Plants, animals, soil, water, rotational crops

3'-O-deethyl-175-J

Animals

4'-O-demethyl-175-J

Animals

N-succinyl-175-J

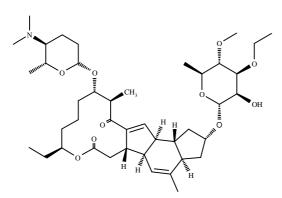
Soil

N-demethyl-175-L

Plants, animals, soil, water, rotational crops

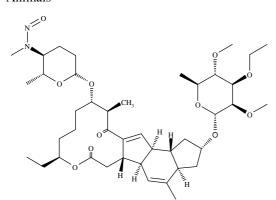
N-formyl -175-L

Plants



2'-O-demethyl-175-L

Animals



N-demethyl-N-nitroso-175-L

Soil

3'-O-deethyl-175-L

Animals

4'-O-demethyl-175-L

Animals

N-succinyl-175-L

Soil

Radio-labelled XDE-175-J and XDE-175-J Used in Metabolism Studies

In plant and animal metabolism studies, XDE-175-J or XDE-175-L which were uniformly labelled with ¹⁴C in the macrolide ring was used. No radioactive carbon was incorporated into either sugar group.

Three forms of $^{14}\text{C-XDE-175-J}$ produced through fermentation in the presence of $^{14}\text{C-acetate}$ were combined to give a mixture of 1:1:1 ratio (w/w/w) with nominal specific activity of 11.62 μ Ci/mg (8.72 mCi/mmol or 429,940 Bq/mg. One form was ethylated at the 3'-position of the rhamnose and the 5,6-position of the macrolide ring then reduced to give ^{14}C -XDE-175-J (INV1947). A second form was likewise reduced but was ethylated using a deuterated alkylating agent to give $^{14}\text{C-XDE-175-J}$ (D5). A third form was ethylated in the same manner as the first but was reduced in

the presence of deuterium to give ¹⁴C-XDE-175-J (D2). These three forms aided in the mass spectrometric analyses of metabolites in confirming that peaks of interest were related to the test materials.

Two forms of $^{14}\text{C-XDE-}175\text{-L}$ were combined to give a mixture of approximately one to one ratio (w/w) with nominal specific activity of 7.57 μ Ci/mg (5.77 mCi/mmol or 280,090 Bq/mg). One form was ethylated at the 3' position of the rhamnose to give $^{14}\text{C-XDE-}175\text{-L}$. A second form was ethylated using a deuterated alkylating agent to give $^{14}\text{C-XDE-}175\text{-L}$ (D5). Since the 5,6-positions of the material cannot be reduced, there is no comparable D2 material formed. These two forms aided in the mass spectrometric analyses of metabolites confirming that peaks of interest were related to the test materials.

The following table shows the position of radioactive carbon in the test materials used.

Table 1 Radio-labelled compounds used in the metabolism studies

Name	Structure
¹⁴ C-XDE-175-J	NN O O O O O O O O O O O O O O O O O O
¹⁴ C-XDE-175-J (D2)	O D D D D D D D D D D D D D D D D D D D
¹⁴ C-XDE-175-J (D5)	
¹⁴ C-XDE-175-L	CH ₃ What is the second of t

Name	Structure
¹⁴ C-XDE-175-L (D5)	O H H H H H H H H H H H H H H H H H H H

Animal metabolism

The Meeting received information on the results of studies on lactating goats and laying hens.

Lactating goats: Two lactating goats (Alpine × Nubian Cross breed, 4 years old, 47-52 kg) were given oral doses of either ¹⁴C-XDE-175-J (14.9 mg/animal/day) or ¹⁴C-XDE-175-L (14.8 mg/animal/day) equivalent to 10–11 ppm in the total diet (on a dry weight basis) once daily via balling gun for five consecutive days (Magnussen, *et. al.*, 2005; Report 040088). Milk was collected twice a day throughout the dosing period and urine and faeces at 24-h intervals. The mean daily milk production of the control, XDE-175-J treated or XDE-175-L treated goat was 1.01, 1.70 or 1.44 L respectively. Twenty-one hours (±1 h) after administering the final dose, the animals were sacrificed and liver, kidney, fat and muscle samples were collected for analysis.

Among excreta, a total of 51.1% of the administered radioactivity was recovered in faeces in the case of ¹⁴C-XDE-175-J and 78.3% in the case of ¹⁴C-XDE-175-L. A total radioactivity recovered in urine was not significant: 0.17% for ¹⁴C-XDE-175-J; and 0.03% for ¹⁴C-XDE-175-L. The cage rinse contained less than 0.01% of the administered dose.

Radioactivity started to appear in milk within 24 h after the first administration and seemed to reach a plateau by the fifth administration. The maximum total radioactive residues expressed in mg-equivalents of respective compound/kg were 0.047 mg/kg in the second milking of day 3 and the first milking of day 4 for ¹⁴C-XDE-175-L and 0.039 mg/kg in the first milking of day 5 for ¹⁴C-XDE-175-L. In the course of the study, no milk sample contained more than 0.07% of the administered dose. Only about 0.3% of the total administered dose of ¹⁴C-XDE-175-J and 0.2% of that of ¹⁴C-XDE-175-L was recovered from all the collected milk samples.

Radioactive residues were relatively low in edible tissues after sacrifice: the concentrations in fat were the highest at 0.235 and 0.119 mg/kg and in other tissues they were, in decreasing order, in liver at 0.116 and 0.099 mg/kg, in kidney at 0.065 and 0.047 mg/kg and in muscle at 0.017 and 0.015 mg/kg for ¹⁴C-XDE-175-J and ¹⁴C-XDE-175-L, respectively (Table 2). For both compounds, only about 0.3% and 0.5% of the total dose was accounted for in the edible tissues for XDE-175-L and XDE-175-J, respectively. The results showed that radioactivity of these compounds were not readily transferred into these edible tissues and that the residues accumulate more in tissues with higher fat content.

Table 2 Total radioactive residues (TRR) in the milk and edible tissues of lactating goats given either ¹⁴C-XDE-175-J or ¹⁴C-XDE-175-L

Tissue		Total Radioactive Residues (mg/kg)					
	Tissue	XDE-175-J	XDE-175-L				
Milk	Day 1 1st	-	-				
	Day 1 2nd	0.001	0.011				
	Day 2 1st	0.012	0.015				
	Day 2 2nd	0.023	0.014				
	Day 3 1st	0.019	0.021				

Tissue	Total Radioactive Residues (mg/kg)					
Tissue	XDE-175-J	XDE-175-L				
Day3 2nd	0.047	0.014				
Day 4 1st	0.047	0.029				
Day 4 2nd	0.037	0.028				
Day 5 1st	0.029	0.039				
Day 5 2nd	0.037	0.029				
Fat	0.235	0.119				
Liver	0.116	0.099				
Kidney	0.065	0.047				
Muscle	0.017	0.015				

Radioactive residues in tissues and milk were extracted under mild condition: fat was extracted with hexane; liver, kidney and muscle with acetonitrile/water; and milk with acetonitrile. The organic extract was subjected to SPE cleanup prior to analysis. In tissues and milk, radioactive residues extracted in organic solvents were $\geq 85\%$ of TRR, except for liver where residues in organic solvent were 57–71% of TRR. Radioactive residues extracted in aqueous layer were $\leq 5\%$ of TRR for all tissues and milk, except for ¹⁴C-XDE-175-J in liver where the aqueous extract contained 18% of TRR. Less than 15% of TRR or 0.025 mg/kg were unextractable from all tissues except liver. From liver, 23–35% of TRR were unextractable.

Characterization and identification of the organo-soluble residues from the muscle, milk and excreta samples showed the primary residue to be XDE-175-J or XDE-175-L, indicating that minimal metabolism had occurred. No residue components other than the unchanged parent compounds were identified in milk, kidney or fat. Radioactive residues in liver consisted of XDE-175-J or XDE-175-L, a metabolite that was tentatively identified as N-demethyl-175-J or –L at very low levels (< 2% of the TRR), and an unidentified metabolite (Metabolite J1 or Metabolite L1) that was more polar than most of the available reference standards. Radioactive residues in muscle also consisted primarily of XDE-175-J or XDE-175-L and much lesser amounts of what seemed to be the same unidentified metabolite as found in liver. Table 3 shows summaries of the characterised radioactive residues. Overall, for both compounds 77–99% of TRR were extracted.

Table 3 Characterization and identification of radioactive residues in the tissues and milk of lactating goats following oral doses of ¹⁴C-XDE-175-J or ¹⁴C-XDE-175-L

Compound	Mus	cle	Fa	ıt	Kidney		Liver		Day 5 Milk ^a	
	mg/kg	% TRR	mg/kg	% TRR	mg/kg	% TRR	mg/kg	% TRR	mg/kg	% TRR
			Oral	Dose of 14	C-XDE-17	5-J				
XDE-175-J	0.007	42.3	0.190	80.8	0.035	53.9	0.035	29.8	0.029	84.4
N-Demethyl-175- J	-	-	-	-	-	-	0.002	1.8	-	-
Metabolite J1	0.001	5.5	-	-	< 0.001	-	0.012	10.0	-	-
Total identified	0.007	42.3	0.190	80.8	0.035	53.9	0.037	31.6	0.029	84.4
Total characterised ^b	0.009	47.2	0.021	8.8	0.021	31.8	0.050	43.3	0.005	14.7
Total extracted	0.016	89.5	0.211	89.6	0.056	85.7	0.087	74.9	0.034	99.1
Total unextractable	0.002	10.7	0.025	10.5	0.009	14.5	0.029	25.2	< 0.001	1.0
Total measured (TRR in ())	(0.017)	100	(0.235)	100	(0.065)	100	(0.116)	100	(0.034)	100

Compound	Mus	cle	Fa	ıt	Kidr	ney	Liv	er	Day 5 N	Milk ^a	
	mg/kg	% TRR	mg/kg	% TRR	mg/kg	% TRR	mg/kg	% TRR	mg/kg	% TRR	
	Oral Dose of ¹⁴ C-XDE-175-L										
XDE-175-L	0.007	45.8	0.086	72.3	0.030	64.3	0.026	25.9	0.042	84.2	
N-Demethyl-175-L	-	-	-	-	-	-	0.001	1.2	-	-	
Metabolite L1	0.003	17.7	-	-	-	-	0.027	26.9	-	-	
Total identified	0.007	45.8	0.086	72.3	0.030	64.3	0.027	27.1	0.042	84.2	
Total characterised	0.008	47.3	0.018	14.8	0.012	21.8	0.049	49.5	0.007	13.6	
Total extracted	0.015	93.1	0.104	87.1	0.042	86.1	0.076	76.6	0.049	97.8	
Total unextractable	0.001	6.9	0.016	12.9	0.007	13.9	0.023	23.4	0.002	2.2	
Total measured (TRR in ())	(0.015)	100	(0.119)	100	(0.047)	100	(0.099)	100	(0.049)	100	

^a Prior to extraction and analysis, aliquots of the Day 5 PM milk sample were re-assayed by liquid scintillation counter to confirm the TRR levels.

<u>Laying hens:</u> Laying hens (Bovan White Leghorn; 24 week old; 1.3-1.6 kg) were given either 14 C-XDE-175-J or 14 C-XDE-175-L, each at 1.25 or 1.75 mg/bird/day, equivalent to 10 ppm in the diet, once daily via balling gun for seven consecutive days with (Smith-Drake, 2005; Report 040087). Each dosing group consisted of ten birds. Eggs were collected twice daily and excreta once daily. The hens were sacrificed within 22 ± 3 h of the final dose and the liver, muscle (breast and thigh), fat (abdominal), and skin with subcutaneous fat was collected for analysis.

The overall recovery of the dose was 95 and 94% for ¹⁴C-XDE-175-J and ¹⁴C-XDE-175-L, respectively. Approximately 93 and 91% of the administered dose of ¹⁴C-XDE-175-J and ¹⁴C-XDE-175-L, respectively, was recovered in the excreta.

Recovery of the administered dose in eggs and tissues was low for both ¹⁴C-XDE-175-J (0.40+0.98%) and ¹⁴C-XDE-175-L (0.93 + 2.1%). TRR in eggs increased over the experimental period and reached a maximum of 0.204 and 0.488 mg/kg for ¹⁴C-XDE-175-J and ¹⁴C-XDE-175-L, respectively, on Day 7. TRR in the tissues were highest in abdominal fat (1.04–2.46 mg/kg), followed by skin with subcutaneous fat (0.66–1.41 mg/kg), liver (0.53–0.90 mg/kg), eggs (0.2–0.49 mg/kg in Day 7) and muscle (0.05–0.11 mg/kg) (Table 4). TRR found in eggs and tissues of hens dosed with ¹⁴C-XDE-175-L were approximately twice as high as those from hens dosed with ¹⁴C-XDE-175-J. There is a tendency for these test compounds to accumulate more in tissues with higher fat content.

Table 4 Total radioactive residues (TRR) in the eggs and edible tissues of hens given either ¹⁴C-XDE-175-J or ¹⁴C-XDE-175-L

	Tissue	Total Radioactive Residues (mg/kg)					
	Tissue	XDE-175-J	XDE-175-L				
	Day 1	0.000	0.000				
	Day 2	0.032	0.036				
Egg	Day 3	0.067	0.123				
	Day 4	0.114	0.225				
	Day 5	0.161	0.340				
	Day 6	0.185	0.420				
	Day 7	0.204	0.488				
1	Abdominal fat	1.04	2.46				
	Skin/Fat	0.661	1.41				

^bCharacterised but not identified.

Tissue	Total Radioactive Residues (mg/kg)					
Tissue	XDE-175-J	XDE-175-L				
Muscle	0.050	0.108				
Liver	0.535	0.902				

Radioactive residues were readily extracted under the mild condition. Following hexane extraction of abdominal fat and skin with fat, 94–97% of TRR were extracted in hexane layer and 3–6% of TRR were unextractable. Partition of radioactive residues in muscle, liver, and egg samples between acetonitrile/water and dichloromethane resulted in 73–88% of TRR in the organic phase, < 1% of TRR in the aqueous phase, and 12–20% of TRR unextractable. Acid extraction of the liver tissue remaining after the above-mentioned extraction further released 7–8% of TRR as organosoluble residues with 4–7% of TRR still not extracted. Table 5 shows summaries of the identified/characterised residues. Unextracted radioactivity was less than 7% of the TRR in all samples except muscle and eggs in which it was 12–20%.

Neither of these two test compound was extensively or rapidly metabolized in hens. Unchanged XDE-175-J or XDE-175-L remained as the primary residue in the egg and tissues.

Table 5 Characterization of radioactive residues in hen tissues following oral doses of ¹⁴C-XDE-175-J or ¹⁴C-XDE-175-L

Compound	Abdon	Abdominal fat Skir		Skin with fat Muscle		Liver		Day 4 Egg		
	mg/kg	% TRR	mg/kg	% TRR	mg/kg	% TRR	mg/kg	% TRR	mg/kg	% TRR
Oral Dose of ¹⁴ C-XDE-175-J										
XDE-175-J	0.723	69.6	0.531	80.2	0.034	67.8	0.069	13.0	0.068	58.4
3'-O-deethyl-175-J	0.019	1.8	-	-	-	-	0.093	17.7	-	-
O-demethyl-175-J	0.059	5.7	0.022	3.3	0.002	3.2	0.034	6.5	-	-
Total identified	0.801	77.1	0.553	83.5	0.036	71.0	0.196	37.2	0.068	58.4
Total characterised ^a	0.205	19.9	0.086	13.0	0.005	10.5	0.294	55.8	0.028	25.6
Total extracted	1.006	97.0	0.639	96.5	0.041	80.5	0.490	93.0	0.096	84.0
Total unextractable	0.032	3.0	0.023	3.5	0.010	19.5	0.037	7.0	0.019	16.0
Total measured (TRR in ())	(1.04)	100	(0.662)	100	(0.050)	100	(0.526)	100	(0.115)	100
			Oral	Dose of 14	C-XDE-17	75-L				
XDE-175-L	1.366	55.5	0.784	55.6	0.048	44.5	0.105	11.7	0.111	48.9
3'-O-deethyl-175-L	0.128	5.2	0.079	5.6	0.006	5.4	0.115	12.8	0.029	12.5
O-demethyl-175-L	0.479	19.5	0.239	16.9	0.020	17.8	0.135	15	0.030	13.4
N-demethyl-175-L	-	-	-	-	-	-	0.015	1.7	-	-
Total identified	1.973	80.2	1.102	78.1	0.074	67.7	0.370	41.2	0.170	74.8
Total characterised ^a	0.414	16.8	0.226	15.9	0.016	14.8	0.493	54.5	0.029	13.2
Total extracted	2.387	97.0	1.328	94.0	0.090	82.5	0.863	95.7	0.199	88
Total unextractable	0.075	3.0	0.084	6.0	0.019	17.5	0.039	4.4	0.027	12
Total measured (TRR in ())	(2.46)	100	(1.41)	100	(0.108)	100	(0.902)	100	(0.226)	100

^a Characterised but not identified.

Unchanged parent compound was the primary residue component in milk and all ruminant tissues as well as eggs and all avian tissues except liver for both XDE-175-J and XDE-175-L. The liver and muscle of the goat treated with XDE-175-J or XDE-175-L contained one unidentified metabolite present at less than 10 or 27% of TRR, respectively. The ruminant liver also contained a

minor amount of N-demethyl-175-J or N-demethyl-175-L (\leq 2% TRR). In hen liver, the O-deethyl-and O-demethyl metabolites were also found.

Metabolism of spinetoram appears to be primarily through dealkylation of the rhamnose sugar to give the O-deethyl and/or O-demethyl (two possible isomers) metabolites. Many other minor metabolites were also observed with each being present at levels lower than 10% of the TRR. Proposed metabolic pathways for spinetoram in livestock are shown in Figure 1 (XDE-175-J) and Figure 2 (XDE-175-L). The metabolism spinetoram in ruminant paralleled that in hens.

Figure 1 Proposed metabolic pathway of XDE-175-J in livestock g, goat; h, hen.

Figure 2 Proposed Metabolic Pathway of XDE-175-L in Livestock g, goat; h, hen.

Plant Metabolism

The Meeting received information on the fate of spinetoram after foliar applications in apples, lettuce and turnips representing the fruits, leafy crops and root crops, respectively.

Apple

A branch with immature fruits of individual Granny Smith apple tree was treated with single foliar application of either ¹⁴C-XDE-175-J or ¹⁴C-XDE-175-L at the rate of 1.8 kg ai/ha (4.8×) or 1.1 kg ai/ha (8.9×), respectively (Byrne, *et. al.*, 2005; Report 040050). Immature apples and leaves were collected 0, 1, 3, 7, and 14 days after treatment (DAT). Covered apples, as dark samples, were also collected 3 DAT and 7 DAT (covered from 3–7 DAT). Mature apples were collected 30 DAT, as well as leaf samples. In addition, one branch on each treated tree was covered with plastic prior to application for testing translocation to an untreated branch. Apple fruits and leaves of the untreated branches were collected 30 DAT.

The collected samples were washed with acetonitrile and dichloromethane. The total radioactive residues (TRR) were determined as the sum of the residues in the washings and the washed fruit or leaves. In the fruit obtained from the 14 C-XDE-175-J treated branch (0–30 DAT), 78–93% of TRR were found in the acetonitrile washings and 4–11% in the dichloromethane washings. Peels contained 2–11% of TRR while pulp contained less than 1% (≤ 0.007 mg/kg). In fruits from 14 C-XDE-175-L treated branch (0-30 DAT), 46–85% of TRR were found in acetonitrile washings and 10–21% in dichloromethane washings. Peel contained 6–33% of TRR while pulp contained less than 4%.

In leaves from the ¹⁴C-XDE-175-J treated branch (0–30 DAT), 67–88% of TRR were recovered in acetonitrile washings and 6–11% of TRR in dichloromethane washings. Washed leaves contained 5–23% of TRR. In leaves from ¹⁴C-XDE-175-L treated branch (0–30 DAT), 55–79% of

TRR were recovered in acetonitrile washings and 10–17% of TRR in dichloromethane washings. Washed leaves contained 11–31% of TRR.

The leaves and peel from both treatments, and pulp from the $^{14}\text{C-XDE-175-L}$ treatment were extracted with acetonitrile:water (80:20, v/v). Only pulp from the $^{14}\text{C-XDE-175-L}$ treated trees was extracted since TRR in pulp from the $^{14}\text{C-XDE-175-J}$ treated trees were $\leq 1\%$ of TRR (≤ 0.007 mg/kg). Radioactive residues unextractable were < 10% of TRR in or on all samples. Pooled acetonitrile washing, dichloromethane washing, and extracts were analysed. Tables 6 and 7 summarize the identified residues. Several minor metabolites assumed to be structurally similar to the parent compound were also detected in the treated apples and leaves, each at $\leq 7.5\%$ of TRR. A multicomponent mixture of extensively degraded compounds represented up to 39–77% of TRR but each component was less than 1% of TRR.

Radioactive residues in the apple fruits from ¹⁴C-XDE-175-L treated tree shows a tendency to be higher when exposed than those in covered (dark) apple samples.

In the translocation experiment, TRR were below the limit of quantification of 0.001 mg/kg in apple fruits and 0.053 mg/kg and 0.009 mg/kg in leaves collected from the ¹⁴C-XDE-175-J and ¹⁴C-XDE-175-L treated trees, respectively indicating that translocation is negligible. As the concentrations in tissues were very low, no further analysis was conducted.

Table 6 Characterization and identification of radioactive residues in apple fruit washings and extracts ^a following application of ¹⁴C-XDE-175-J at 1.8 kg ai/ha or ¹⁴C-XDE-175-L at 1.1 kg ai/ha

Compound	%TRR	mg/kg	%TRR	mg/kg	%TRR	mg/kg	%TRR	mg/kg
	Applica	ation of ¹⁴ C	-XDE-175-	J at 1.8 kg a	ai/ha			
	0 D	DAT	1 D	PAT	3 DAT	(dark)	3 E	OAT
XDE-175-J	82.2	0.715	77.2	1.286	61.4	0.625	54.8	0.729
N-demethyl-175-J	5.0	0.044	7.0	0.116	10.2	0.104	8.4	0.112
N-formyl-175-J	1.7	0.015	4.1	0.068	3.1	0.031	3.6	0.048
C9-pseudoaglycone-175-J	0.5	0.004	0.1	0.002	0.8	0.008	1.3	0.018
3-O-deethyl-175-J	1.2	0.010	3.9	0.065	-	-	-	-
Total identified	90.6	0.788	92.3	1.534	75.5	0.768	68.1	0.907
Total characterised ^b	9.2	0.081	7.2	0.130	24.4	0.250	31.5	0.418
Total extractable	99.8	0.869	99.5	1.664	99.9	1.018	99.6	1.325
Total unextractable	0.4	0.003	0.4	0.006	0.3	0.003	0.9	0.011
Total measured (TRR in ())	100	(0.870)	100	(1.67)	100	(1.02)	101	(1.33)
	7 DAT	(dark)	7 DAT		14 DAT		30 DAT	
XDE-175-J	44.4	0.352	43.3	0.501	44.3	0.485	18.9	0.135
N-demethyl-175-J	8.3	0.066	9.5	0.110	9.1	0.100	6.3	0.044
N-formyl-175-J	5.9	0.047	5.1	0.059	3.0	0.033	3.5	0.024
C9-pseudoaglycone-175-J	1.5	0.011	2.9	0.033	3.8	0.043	1.5	0.010
3-O-deethyl-175-J	0.1	0.001	0.1	0.001	3.2	0.035	2.0	0.014
Total identified	60.2	0.477	60.9	0.704	63.4	0.696	32.2	0.227
Total characterised b	37.7	0.299	38.3	0.445	35.0	0.381	64.3	0.462
Total extracted	97.9	0.776	99.2	1.149	98.4	1.077	96.5	0.689
Total unextractable	1.6	0.013	1.6	0.02	1.3	0.014	3.8	0.027
Total measured (TRR in ())	100	(0.793)	101	(1.16)	100	(1.09)	100	(0.713)
	Applica	ation of ¹⁴ C-	XDE-175-	L at 1.1 kg	ai/ha			
	0.0	DAT	1 D	OAT	3 DAT (dark)		3 DAT	
XDE-175-L	42.7	0.184	18.4	0.115	17.7	0.237	12.9	0.115
N-demethyl-175-L	7.9	0.034	4.0	0.025	3.4	0.046	3.6	0.032

Compound	%TRR	mg/kg	%TRR	mg/kg	%TRR	mg/kg	%TRR	mg/kg
N-formyl-175-L	1.6	0.007	2.3	0.014	3.5	0.047	0.9	0.008
Total identified	52.2	0.225	24.7	0.154	24.6	0.330	17.4	0.155
Total characterised b	45.1	0.194	74.5	0.466	71.6	0.958	75.9	0.672
Total extracted	97.3	0.419	99.2	0.620	96.2	1.288	93.3	0.827
Total unextractable	2.0	0.009	1.2	0.007	3.0	0.041	7.0	0.063
Total measured (TRR in ())	99	(0.431)	100	(0.626)	99	(1.34)	100	(0.888)
	7 DAT (dark)		7 DAT		14 DAT		30 DAT	
XDE-175-L	8.5	0.088	1.3	0.005	-	-	-	-
N-demethyl-175-L	2.7	0.027	1.0	0.004	0.3	0.002	0.4	0.003
N-formyl-175-L	1.0	0.010	1.0	0.004	0.2	0.001	0.4	0.003
Total identified	12.2	0.125	3.3	0.013	0.5	0.003	0.8	0.006
Total characterised	82.6	0.858	86.6	0.307	90.6	0.477	89.5	0.650
Total extracted	94.8	0.983	89.9	0.320	91.1	0.48	90.3	0.656
Total unextractable	4.0	0.042	7.2	0.025	6.9	0.036	9.0	0.066
Total measured (TRR in ())	99	(1.04)	97	(0.356)	98	(0.528)	99	(0.728)

^a Sum of residues of pooled fruit washings and peel/pulp extracts by HPLC. Only the pulp from XDE-175-L treated tree was extracted.

Table 7 Characterization and identification of radioactive residues in apple leaf washings $^{\rm a}$ and extracts following application of $^{\rm 14}$ C-XDE-175-J at 1.8 kg ai/ha or $^{\rm 14}$ C-XDE-175-L at 1.1 kg ai/ha (Byrne, *et. al.*, 2005)

Compound	%TRR	mg/kg	%TRR	mg/kg	%TRR	mg/kg
Ap	plication of ¹⁴ C	C-XDE-175-J	at 1.8 kg ai/ha	1		
	0 D	OAT	1 DAT		3 DAT	
XDE-175-J	80.3	105	78.6	100.0	42.4	71.27
N-demethyl-175-J	9.1	11.9	10.4	13.35	14.8	24.83
N-formyl-175-J	2.4	3.20	2.4	3.07	0.7	1.15
C9-pseudoaglycone-175-J	0.9	1.11	0.2	0.21	2.3	3.86
3-O-deethyl-175-J	0.1	0.10	0.9	1.07	4.2	6.95
Total identified	92.8	122	92.5	117.69	64.4	108.06
Total characterised ^b	9.6	12.8	8.5	10.92	35.1	59.23
Total extracted	102.4	134	101.0	128.61	99.5	167.29
Total unextractable	0.3	0.33	0.2	0.29	0.6	1.03
Total measured (TRR in ())	103	(131)	101	(127)	100	(168)
	7 D	AT	14 DAT		30 DAT	
XDE-175-J	37.6	58.40	24.0	34.36	19.9	27.77
N-demethyl-175-J	12.0	18.70	9.4	12.45	7.5	10.50
N-formyl-175-J	3.4	5.39	3.0	4.34	2.2	3.07
C9-pseudoaglycone-175-J	1.7	2.63	3.6	5.11	3.1	4.37
3-O-deethyl-175-J	2.2	3.40	4.3	6.15	2.5	3.45
Total identified	56.9	88.53	44.3	63.40	35.2	49.15
Total characterised ^b	41.9	65.06	54.1	77.51	61.0	84.98
Total extracted	98.8	153.59	98.4	140.92	96.2	134.13
Total unextractable	0.5	0.78	1.0	1.48	3.3	4.54

^bCharacterised but not identified

Compound	%TRR	mg/kg	%TRR	mg/kg	%TRR	mg/kg
Total measured (TRR in ())	99	(155)	99	(143)	100	(140)
App	plication of ¹⁴ C	C-XDE-175-L	at 1.1 kg ai/ha	ì		
	0 D	OAT	1 D)AT	3 D	OAT
XDE-175-L	26.8	18.56	19.6	9.29	2.0	1.15
N-demethyl-175-L	1.8	1.36	3.2	1.53	0.6	0.37
N-formyl-175-L	2.0	1.50	1.3	0.58	0.7	0.38
Total identified	30.6	21.42	24.1	11.40	3.3	1.90
Total characterised ^b	69.3	42.93	73.8	35.08	95	55.17
Total extracted	99.9	64.35	97.9	46.48	98.3	57.07
Total unextractable	1.2	0.77	1.3	0.60	2.2	1.29
Total measured (TRR in ())	101	(64.4)	99	(47.4)	101	(58.1)
	7 D	OAT	14 DAT		30 DAT	
XDE-175-L	1.3	0.59	0.7	0.24	0.2	0.12
N-demethyl-175-L	0.9	0.41	0.6	0.25	0.2	0.12
N-formyl-175-L	1.2	0.55	1	0.39	0.3	0.18
Total identified	3.4	1.55	2.3	0.89	0.7	0.42
Total characterised ^b	94.6	49.44	94.4	37.26	90.4	50.34
Total extracted	98.0	50.99	96.7	38.14	91.1	50.76
Total unextractable	2.7	1.38	2.4	0.94	6.8	3.77
Total measured (TRR in ())	101	(52.1)	99	(39.4)	98	(55.4)

^a Sum of residues of pooled washings and extracts by HPLC.

Lettuce

Individual pots of red leaf lettuce (variety New Fire Red) were treated with either single or multiple foliar spray applications of ¹⁴C-XDE-175-J or ¹⁴C-XDE-175-L (Magnussen, *et. al.*, 2005; Report 040048). For ¹⁴C-XDE-175-J, single applications were made at rates equivalent to 0.90 kg ai/ha (single application treatment) while the same amount of the test compound was sprayed in three separate applications with the equal rate at weekly intervals (multiple application treatment). For ¹⁴C-XDE-175-L, plants were treated in a similar fashion but at a rate equivalent to 0.30 kg ai/ha. For both compounds, the amounts applied approximately correspond to four times the maximum seasonal rate described on label.

Lettuce receiving a single application was harvested at 0, 0.25, 1, 3, and 7 DAT. Lettuce treated with multiple applications was harvested at 3 and 7 DAT. Identification of residues was not conducted for some of the 7 DAT samples as they were damaged by heat stress.

Collected samples were washed first with dichloromethane followed by acetonitrile in order to remove surface residues. Total radioactive residues (TRR) were determined as sum of radioactive residues in washings and washed lettuce. In both ¹⁴C-XDE-175-J and ¹⁴C-XDE-175-L treated lettuce samples, 76–96% of TRR were found in the solvent washings. From lettuce samples collected 0, 0.25 and 1 day after the last application, approximately 3.4–21% of TRR were found in acetonitrile washings. The radioactive residues remaining after washing in lettuce samples taken 3 DAT were extracted by homogenizing and refluxing with acetonitrile/water (75/25, v/v) followed by partitioning with acetonitrile/dichloromethane (50/50, v/v) at neutral and acidic pH. About 12–19% of TRR were in the organic phase and 0.3–3.4% of TRR in the aqueous phase. Overall, only 0.2–5.2% of TRR remained unextractable in all treated lettuce samples.

The solvent washings and extracts were pooled and analysed. Table 8 summaries the identified residues. Several minor metabolites assumed to be structurally similar to the parent

^bCharacterised but not identified

compound were also observed in the 14 C-XDE-175-J and 14 C-XDE-175-L treated lettuce at or less than 6% of TRR. A multi-component mixture of extensively degraded compounds represented up to 13–78% of TRR, each component at less than 3% of TRR.

Table 8 Characterization and identification of radioactive residues in lettuce following application of 14 C-XDE-175-J at 0.90 kg ai/ha or 14 C-XDE-175-L at 0.30 kg ai/ha

Compound	0 D	AT	0.25	DAT	1 D)AT	3 D	AT	3 D (mult	
	%TRR	mg/kg	%TRR	mg/kg	%TRR	mg/kg	%TRR	mg/kg	%TRR	mg/kg
		App	lication of	¹⁴ C-XDE-	175-J at 0	.90 kg ai/h	a			
XDE-175-J	63.6	31.74	42.1	27.23	31.1	10.71	17.6	6.41	8.5	0.52
N-demethyl-175-J	8.9	4.44	18.0	11.64	19.6	6.75	15.5	5.64	7.2	0.44
N-formyl-175-J	6.6	3.29	7.8	5.05	10.6	3.65	11.2	4.08	14.8	0.90
Total identified	79.1	39.48	67.9	43.92	61.3	21.11	44.3	16.12	30.5	1.86
Total characterised	20.7	10.32	31.6	20.45	37.5	12.90	55.1	20.07	66.6	4.06
Total extracted	99.8	49.80	99.5	64.37	98.8	34.01	88.4	36.19	97.1	5.93
Total unextractable	0.2	0.10	0.5	0.32	1.2	0.41	0.6	0.22	3.0	0.18
Total measured	100	(49.9)	100	(64.7)	100	(34.4)	100	(36.3)	100	(6.11)
(TRR in ())										
		App	lication of	¹⁴ C-XDE-	175-L at 0	.30 kg ai/h	ıa			
XDE-175-L	62.4	6.18	8.2	1.47	11.9	0.91	5.1	0.55	2.8	0.10
N-demethyl-175-L	17.6	2.08	6.7	0.54	7.2	0.55	3.5	0.38	1.5	0.05
N-formyl-175-L	5.9	0.70	5.3	0.43	4.0	0.30	2.0	0.22	1.1	0.04
Total identified	75.9	8.95	30.2	2.45	23.1	1.76	10.6	1.14	5.4	0.18
Total characterised	22.5	2.66	65.8	5.33	71.7	8.97	87.7	9.46	91.2	3.08
Total extracted	98.4	11.61	96	7.77	94.8	7.21	98.3	10.60	96.6	3.28
Total unextractable	1.6	0.19	4.0	0.32	5.2	0.40	1.8	0.19	3.4	0.12
Total measured	100	(11.8)	100	(8.10)	100	(7.61)	100	(10.8)	100	(3.38)
(TRR in ())										

^a Samples received multiple applications of ¹⁴C-XDE-175-J or ¹⁴C-XDE-175-L.

Turnips

Individual pots of Purple Top White Globe turnips were treated with either a single or multiple foliar applications of ¹⁴C-XDE-175-J or ¹⁴C-XDE-175-L (Graper, *et. al.*, 2005; Report 040049). Single applications of XDE-175-J were made at rates equivalent to 0.90 kg ai/ha (single application treatment) while the same amount of the test compound was sprayed in three separate applications with the equal rate at weekly intervals (multiple application treatment). For XDE-175-L, plants were treated in a similar fashion but at a rate equivalent to 0.30 kg ai/ha. For both test compounds, applied approximately correspond to four times the maximum seasonal rate described on label.

Following application, plants treated with a single application were harvested at 0, 0.25, 1, 3 and 7 DAT. Plants that received the multiple applications were harvested at 3 and 7 DAT of the final application (17 and 21 days after the initial application).

Collected turnip tops were washed first with dichloromethane followed by acetonitrile in order to remove surface residues. Total radioactive residues (TRR) were determined as the sum of the residue in the washing and washed tissue. In ¹⁴C-XDE-175-J treated turnip tops, 59–91% of TRR were surface residues found in the solvent washings. In ¹⁴C-XDE-175-L treated turnip tops, surface residues were 39–80% of TRR, slightly smaller percentage than in XDE-175-J treated tops. In treatment with both compounds, the percentage of TRR as surface residues of turnip tops was lower in samples receiving multiple applications than for those receiving a single application.

Washed turnip tops were extracted by homogenizing and refluxing with acetonitrile/water (80/20, v/v) followed by partitioning with dichloromethane at neutral and acidic pH. Approximately 8–35% of TRR were found in organic phase and < 0.1–9% of TRR were in the aqueous phase, but < 1–18% of TRR were unextractable. The remaining residues in 3 DAT turnip top samples receiving multiple applications were extracted with acid and 2–3% of TRR were released as organo-soluble residues and 1–4% of TRR as aqueous residues, leaving 4–5% of TRR still not extracted.

Turnip root samples were extracted by homogenizing and refluxing with acetonitrile/water (80/20, v/v) followed by dichloromethane partitioning at neutral and acidic pH. In turnip root samples from both ¹⁴C-XDE-175-J and ¹⁴C-XDE-175-L treatments, 38–93% of TRR were in organic phase and 2–28% of TRR were in the aqueous phase, but 6–34% of TRR were unextractable.

For metabolite characterization and identification, the washings and residues in organic solvent and aqueous extracts of turnip tops or turnip roots were pooled as composite extracts. For tops, samples harvested at 0.25 DAT and 3 DAT after a single application and those at 3 DAT after multiple applications were used. For roots, samples taken at 3 DAT after a single application were used. Table 9 summarizes residues characterised and identified in turnip tops and roots. Several minor metabolites assumed to be structurally similar to the parent compound were also observed in treated turnip tops, each less than 4% of TRR. A multi-component mixture of extensively degraded compounds represented 10–74% of TRR, each compound at less than 1% TRR.

Table 9 Characterization and identification of radioactive residues in turnip tops and roots following application of ¹⁴C-XDE-175-J at 0.90 kg ai/ha or ¹⁴C-XDE-175-L at 0.30 kg ai/ha

		TOPS					ROOTS	
Compound	0.25	DAT	3 D	AT	3 D		3 DAT	
Compound					(mult	iple) ^a		
	% TRR	mg/kg	% TRR	mg/kg	% TRR	mg/kg	% TRR	mg/kg
	Appl	ication of 14	C-XDE-175	-J at 0.90 k	g ai/ha			
XDE-175-J	35.7	4.20	9.4	1.11	4.9	0.35	22.3	0.03
N-demethyl-175-J	14.4	1.69	8.5	1.01	4.1	0.30	10.0	0.01
N-formyl-175-J	10.4	1.23	11.2	1.32	11.4	0.82	16.6	0.02
C17-pseudoaglycone-175-J	-	-	-	-	0.3	0.02	-	-
Aglycone-175-J	-	-	-	-	0.5	0.03	-	-
Total identified	60.5	7.12	29.1	3.43	21.2	1.53	48.9	0.06
Total characterised	38.9	4.58	68.7	8.08	74.8	5.41	40.8	0.05
Total extracted	99.4	11.71	97.8	11.51	96.0	6.93	89.0	0.11
Total unextractable	0.6	0.08	2.2	0.26	4.0	0.29	10.3	0.01
Total measured	100	(11.8)	100	(11.8)	100	(7.22)	100	(0.12)
(TRR in ())								
	Appli	ication of 14	C-XDE-175	-L at 0.30 k	g ai/ha			
XDE-175-L	17.1	0.91	2.9	0.06	3.0	0.07	14.8	0.005
N-demethyl-175-L	7.4	0.40	0.6	0.01	1.1	0.02	-	-
N-formyl-175-L	3.0	0.16	1.0	0.2	0.5	0.01	3.0	0.001
Total identified	27.5	1.47	4.5	0.10	4.6	0.10	17.8	0.006
Total characterised	69.8	3.73	88.8	1.89	89.2	1.92	62.4	0.019
Total extracted	97.3	5.20	93.3	1.99	93.8	2.03	80.2	0.025
Total unextractable	2.7	0.14	6.6	0.14	6.2	0.13	19.8	0.006
Total measured (TRR in ())	100	(5.34)	100	(2.13)	100	(2.16)	100	(0.031)

^a Samples received multiple applications of ¹⁴C-XDE-175-J or ¹⁴C-XDE-175-L.

A consistent metabolism of spinetoram was observed in the three crops studied-apple, lettuce and turnip-indicating that a common metabolism is expected for not only fruits, leafy vegetables and root vegetables but also other plants.

Based on the results of the submitted metabolism studies, it appears that three metabolic pathways are responsible for the breakdown of spinetoram in plants. The first one involves changes to the N-dimethyl moiety on the forosamine sugar to give the N-demethyl and N-formyl metabolites. Due to the presence of these metabolites at the 0-day PHI, it is thought that their formation may be the result of photolysis. The second involves cleavage of the macrolide ring system at one or more positions, ultimately resulting in a complex residue mixture consisting of numerous components. The third, applicable only to XDE-175-J, involves changes to the rhamnose sugar producing the 3-O-deethyl and C9-pseudoaglycone-175-J metabolites. All the metabolites occurring as a result of changes in forosamine and rhamnose are further degraded via the second pathway. XDE-175-L might also undergo degradation through the third pathway no relevant metabolites were detected perhaps because degradation through the second pathway might be too fast. The proposed metabolic pathways for spinetoram in plants are shown in Figures 3 and 4 for XDE-175-J and XDE-175-L, respectively.

Figure 3 Proposed Metabolic Pathway of XDE-175-J in Plants

Figure 4 Proposed Metabolic Pathway of XDE-175-L in Plants

Environmental fate in soil

The Meeting received information on the aerobic soil metabolism, aqueous photolysis and hydrolysis, and residues in succeeding crops. Since spinetoram is intended for use on root vegetables and supervised trials were conducted on sugar beet, the above mentioned information was reviewed. The Meeting also received information on anaerobic soil metabolism and photolysis in soil but they were not reviewed.

In laboratory studies on aerobic soil metabolism, the same 14 C-labelled and 14 C- and D-labelled active substances were used as in the animal and plant metabolism studies. Unless otherwise indicated, the radiochemical purity was > 97%.

Aerobic soil metabolism

Yoder *et al.* (2005; Report 040068) incubated radio-labelled spinetoram for up to one year under aerobic condition in the dark at 25 °C and 75% of 1/3 bar moisture in four soils in the USA: Commerce loam from Mississippi; Fayette silt loam from Iowa; Kimberlina/Nord sandy loam from California; and Slagle loam from Virginia. XDE-175-J was applied at the rate of 0.21 mg ai/kg soil, equivalent to 375 g ai/ha (1×). XDE-175-L was also applied at 0.21 mg ai/kg soil, or approximately equivalent to 3× its anticipated maximum use rate.

The degradation of spinetoram in European soils under aerobic conditions in the dark was also studied (Smith-Drake, *et. al.*, 2007; Report 050076.1). Radio-labelled spinetoram was incubated at 20 °C for 127 days in four soils: a sandy loam from Longwoods Quarry, Lincolnshire, UK; a loamy sand from Hanhofen, Rheinlan-Pfalz, Germany; a sandy clay loam from Little Shelford, UK; and a sandy clay loam from Altlussheim, Baden-Wurttemberg, Germany. XDE-175-J was applied at the rate of 0.8 mg ai/kg soil, equivalent to 300 g ai/ha. XDE-175-L was applied at the same rate equivalent to 3× the label rate. Radio-labelled spinetoram was also incubated at 10 °C in the dark for 127 days in the sandy loam from Longwoods Quarry.

The results of these studies are summarized in Table 9.

Under aerobic conditions, spinetoram applied to soil was degraded relatively rapidly. In all soils tested, XDE-175-L was degraded faster than XDE-175-J. After one year of incubation, 1.2–2.8% and 0.3–2.9% of applied XDE-175-J and XDE-175-L respectively (dose rate, 0.21 mg/kg soil; 25 °C) remained as the parent in US soils tested. In European soils except the loamy sand, after 127 days of incubation, 2.0–4.9% and 1.4–5.0% of applied XDE-175-J and XDE-175-L respectively (dose rate, 0.80 mg/kg soil; 20 °C) remained as the parent. In the loamy sand, 50% and 33% of the applied XDE-175-J and XDE-175-L respectively (dose rate, 0.80 mg/kg soil; 20 °C) remained as the parent). In the sandy loam maintained at 10 °C, 4.9% and 2.1% of the applied XDE-175-J and XDE-175-L respectively (same dose rate as at 20 °C) remained as the parent.

Carbon dioxide was evolved slowly from all soils and accounted for 5.0-35% and 9.5-32% of the applied XDE-175-J and XDE-175-L respectively after one year at 25 °C, and 0.8-1.1% and 1.2-3.2% of the applied XDE-175-J and XDE-175-L respectively after 127 days at 20 °C.

As major degradation products, N-demethyl 175-J and N-demethyl-175-L were formed and then degraded during the study periods. As minor products (at or less than 10% of the applied dose), N-demethyl-N-nitroso-175-J, N-demethyl-N-nitroso-175-L, N-succinyl-175-L and N-succinyl-175-L were also formed and degraded. Many other degradates were formed but at very low concentrations.

While extractable radioactivity decreased, non-extractable radioactivity steadily increased to reach 22-29% and 32-37% of the applied XDE-175-J and XDE-175-L respectively after one year at 25 °C; and 5-15% and 11-24% of the applied XDE-175-J and XDE-175-L respectively after 127 days at 20 and 10 °C.

The half-life of spinetoram was calculated to be 21 days and 13 days for XDE-175-J and XDE-175-L respectively at 0.21 mg/kg dose rate (25 °C) and 20 days and 14 days for XDE-175-J and XDE-175-L respectively at 0.80 mg/kg dose rate (20 °C). When maintained at 10 °C the half-life was 21 days and 16 days for XDE-175-J and XDE-175-L respectively at 0.80 mg/kg dose rate. In the sterile sandy clay loam from Little Shelford, the half-life of XDE-175-J and XDE-175-L was much longer at 207 and 298 days respectively than in the respective unsterilized soil. The half-life of XDE-175-J and XDE-175-L under anaerobic condition in sandy loam from Longwoods Quarry was 203 days and 122 days respectively, much longer than the half-life under aerobic condition.

The half-life of N-demethyl-175-J and -L ranged 32–273 days and 5–88 days respectively at 25 $^{\circ}$ C and 136–301 days and 59–204 days at 20 $^{\circ}$ C.

Table 9 Results of aerobic soil metabolism studies [(J): XDE-175-J (L): XDE-175-L]

Ref: Yoder et al., 2005; Report 040068 ¹⁴C accountability: Soil: Loam (Commerce) 97-108% (J) Dose rate: 0.21 mg ai/kg soil (both compounds) 90-105% (L) Duration: 276 days (J) and 365 days (L) Mineralization at the end: 7.5% (J) Temperature: 25 ± 2 °C 18% (L) Unextractable at the end: 22% (J) pH: 7.5 Moisture: 75% of 1/3 bar moisture 33% (L) Average half-life of N-demethyl metabolite: Organic carbon: 0.6% Average half-life: 23 days (J) 253 days (J) 17 days (L) 88 days (L)

Spinetoram remaining at the end: 1.2% (J)

0.7% (L)

Metabolite	Max % of dose	Day
XDE-175-J	102	0
N-demethyl-175-J	68	98, 125
XDE-175-L	95	0
N-demethyl-175-L	41	32

N-demethyl metabolite at the end: 45% (J)

Ref: Yoder et al., 2005; Report 040068

Mineralization at the end: 6.9% (J)

Unextractable at the end: 23% (J)

273 days (J)

18 days (L)

Average half-life of N-demethyl metabolite:

N-demethyl metabolite at the end: 41% (J)

¹⁴C accountability:

9.1% (L)

0.8% (L)

Soil: Silt loam (Fayette)

Dose rate: 0.21 mg ai/kg soil (both compounds)

Duration: 366 days (J) and 365 days (L)

Temperature: 25 ± 2 °C

pH: 7.4

Metabolite

XDE-175-J

XDE-175-L

N-demethyl-175-J

N-demethyl-175-L

Moisture: 75% of 1/3 bar moisture

Organic carbon: 1.1%

Average half-life: 29 days (J)

15 days (L)

95

29

Max % of dose

Spinetoram remaining at the end: 1.7%(J)

0.3% (L)

Day	
0	
63	
0	
7, 32	7

62 63 88 0

Soil: Sandy loam (Kimberlina/Nord)

Dose rate: 0.21 mg ai/kg soil (both compounds)

Duration: 366 days (J) and 365 days (L)

Temperature: 25 ± 2 °C

pH: 8.1

Moisture: 75% of 1/3 bar moisture

Organic carbon: 0.7%

Average half-life: 23 days (J)

17 days (L)

Spinetoram remaining at the end: 2.8%(J)

	2.0% (L)	
Metabolite	Max % of dose	Day
XDE-175-J	98	0
N-demethyl-175-J	51	92
XDE-175-L	97	0
N-demethyl-175-L	33	32

Ref: Yoder et al., 2005; Report 040068

¹⁴C accountability: 97-109% (J)

98-107% (L)

92-109% (J)

92-104% (L)

17% (L)

37% (L)

Mineralization at the end: 5.0% (J)

9.5% (L)

Unextractable at the end: 25% (J)

35% (L)

Average half-life of N-demethyl metabolite:

156 days (J) 29 days (L)

N-demethyl metabolite at the end: 23% (J)

6.9% (L)

Soil: Loam (Slagle) Dose rate: 0.21 mg ai/kg soil (both compounds) Duration: 366 days (J) and 365 days (L)

Temperature: 25 ± 2 °C

pH: 5.8

Moisture: 75% of 1/3 bar moisture

Organic carbon: 0.5% Average half-life: 8 days (J)

3 days (L)

Spinetoram remaining at the end: 2.0%(J)

2.9% (L)

Ref: Yoder et al., 2005; Report 040068 ¹⁴C accountability: 90-104% (J) 8-104% (L)

Mineralization at the end: 35% (J)

32% (L)

Unextractable at the end: 29% (J)

32% (L)

Average half-life of N-demethyl metabolite:

32 days (J) 5 days (L)

N-demethyl metabolite at the end: 6.3% (J)

0.6% (L)

Mineralization at the end: 0.9% (J)

Unextractable at the end: 13% (J)

136 days (J)

59 days (L)

end (125 days): 4.2%(J)

Average half-life: 203 days (J)

Average half-life of N-demethyl metabolite:

N-demethyl metabolite at the end: 36%(J)

7.1% (L)

122 days (L)

In anaerobic condition, spinetoram remaining at the

¹⁴C accountability:

Ref: Smith-Drake, et. al., 2007; Report 050076.1

95-103% (J)

94-100% (L)

15% (L)

3.2% (L)

24% (L)

Metabolite	Max % of dose	Day
XDE-175-J	93	0
N-demethyl-175-J	45	14
XDE-175-L	89	0
N-demethyl-175-L	35	123

Soil: Sandy loam (Longwoods Quarry)

Dose rate: 0.80 mg ai/kg soil (both compounds)

Duration: 127 days Temperature: 20 ± 2 °C

pH: 7.9

Moisture: 10.8 g/g-soil at 1/3 bar

Organic carbon: 0.8%

Average half-life: 8 days (J)

7 days (L)

Spinetoram remaining at the end: 2.0% (J)

1.4% (L)

Metabolite	Max % of dose	Day
XDE-175-J	93	0
N-demethyl-175-J	46	29
N-demethyl-N-nitroso-175-J	19	85
XDE-175-L	93	0
N-demethyl-175-L	38	21
N-demethyl-N-nitroso-175-L	13	42
N-succinyl-175-L	13	42

Ref: Smith-Drake, et. al., 2007; Report 050076.1

¹⁴C accountability: 95–103% (J)

94-100% (L)

Mineralization at the end: 0.9% (J)

1.4% (L)

Unextractable at the end: 13% (J)

13% (L)

Average half-life of N-demethyl metabolite:

386 days (J) 110 days (L)

N-demethyl metabolite at the end: 36% (J)

27%(L)

Soil: Sandy loam (Longwoods Quarry)

Dose rate: 0.80 mg ai/kg soil (both compounds)

Duration: 127 days Temperature: 10 ± 2 °C

pH: 7.9

Moisture: 10.8 g/g-soil at 1/3 bar

Organic carbon: 0.8%

Average half-life: 21 days (J)

16 days (L)

Spinetoram remaining at the end: 4.9% (J)

2.1% (L)

Metabolite	Max % of dose	Day
XDE-175-J	93 ^a	0
N-demethyl-175-J	45	43, 85
N-demethyl-N-nitroso-175-	19	71, 99
J		
XDE-175-L	93 1	0
N-demethyl-175-L	31	42
N-demethyl-N-nitroso-175-	16	56
L		

^a Data from 20 °C experiment.

Soil: Loamy sand (Hanhofen)

Dose rate: 0.80 mg ai/kg soil (both compounds)

Duration: 127 days

Temperature: 20 ± 2 °C

pH: 6.0

Moisture: 9.7 g/g-soil at 1/3 bar

Ref: Smith-Drake, et. al., 2007; Report 050076.1

¹⁴C accountability: 95–102% (J)

94–101% (L)

Mineralization at the end: 0.4% (J)

1.2% (L)

Unextractable at the end: 4.7% (J)

11% (L)

Organic carbon: 1.8%

Average half-life: 129 days (J)

71 days (L)

Spinetoram remaining at the end: 50% (J)

33% (L)

	33 /c (L)	
Metabolite	Max % of dose	Day
XDE-175-J	88	0
N-demethyl-175-J	30	113
N-demethyl-N-nitroso-175-	1.6	113
J		
XDE-175-L	92	0
N-demethyl-175-L	20	84
N-demethyl-N-nitroso-175-	3.7	7
L		

Average half-life of N-demethyl metabolite:

180 days (J) 72 days (L)

N-demethyl metabolite at the end: 30% (J)

17% (L)

Soil: Sandy clay loam (Little Shelford)

Dose rate: 0.80 mg ai/kg soil (both compounds)

Duration: 127 days Temperature: 20 ± 2 °C

pH: 7.8

Moisture: 16.9 g/g-soil at 1/3 bar

Organic carbon: 1.2%

Average half-life:

11 days (J) 8 days (L)

Spinetoram remaining at the end: 3.2% (J)

2.1% (L)

Metabolite	Max % of dose	Day
XDE-175-J	91	0
N-demethyl-175-J	57	99
N-demethyl-N-nitroso-	5.4	85
175-J		
XDE-175-L	93	0
N-demethyl-175-L	37	14
N-demethyl-N-nitroso-	2.8	28
175-L		

Ref: Smith-Drake, et. al., 2007; Report 050076.1

¹⁴C accountability: 96-106% (J)

91-101% (L)

Mineralization at the end: 0.8% (J)

2.9% (L)

Unextractable at the end: 15% (J)

24% (L)

Average half-life of N-demethyl metabolite:

292 days (J) 112 days (L)

N-demethyl metabolite at the end: 55% (J)

22% (L)

In the sterile soil, spinetoram remaining at the end

(120 days): 54% (J)

74% (L)

Average half-life: 207 days (J)

298 days (L)

Soil: Sandy clay loam (Altlussheim)

Dose rate: 0.80 mg ai/kg soil (both compounds)

Duration: 127 days Temperature: 20 ± 2 °C

pH: 7.8

Moisture: 20.4 g/g-soil at 1/3 bar

Average half-life:

Organic carbon: 1.3%

15 days (J) 12 days (L)

4.9% (J) Spinetoram remaining at the end:

5.0% (L)

Day Metabolite Max % of dose XDE-175-J 87 0 N-demethyl-175-J 59 85 10 N-demethyl-N-nitroso-57 175-J XDE-175-L 91 0 84, 98 N-demethyl-175-L 42 N-demethyl-N-nitroso-5.6 70 175-L

Ref: Smith-Drake, et. al., 2007; Report 050076.1

¹⁴C accountability: 95-103% (J)

95-100% (L)

Mineralization at the end: 1.1% (J)

2.5% (L)

Unextractable at the end: 12% (J)

17% (L)

Average half-life of N-demethyl metabolite:

301 days (J) 204 days (L)

N-demethyl metabolites at the end: 58% (J)

39% (L)

Figure 5 Proposed Degradation Pathway of XDE-175-J in Soil under Aerobic Conditions

Figure 6 Proposed Degradation Pathway of XDE-175-L in Soil under Aerobic Conditions

Aqueous photolysis

The aqueous photolysis of radiolabelled spinetoram was studied at 25 °C in sterile aqueous tris buffer solution at pH 7 (Yoder, et al., 2005, Report 040079). XDE-175-J was added at approximately 0.3 mg ai/L and XDE-175-L at 0.5 mg ai/L. Acetonitrile (0.5%) was used as a co-solvent. Samples were continuously irradiated using a xenon lamp as a light source. No trapping of CO_2 or organic volatiles took place.

Samples were analysed after 4, 8 and 16 h, and 1, 2, 4, 7, 13 and 19 days of continuous irradiation; 24 h of continuous exposure (1 DAT) was equivalent to 1.8 days of summer sunlight at

40° N latitude. Time 0 samples and dark controls were also analysed. Aliquots of each sample were directly analysed by LSC and HPLC. Characterization of transformation products was done by LC/MS.

A PNAP/pyridine chemical actinometer solution was used to quantify the amount of light that the sample solutions received. Based on actinometer data, the 19 experimental days of continuous irradiation was equivalent to 34 days of irradiation in the summer sun at 40° N latitude.

Material balance was $101.1 \pm 3.7\%$ of applied radioactivity for the irradiated XDE-175-J solutions and $95.8 \pm 3.4\%$ for the dark XDE-175-J controls. Irradiated and dark XDE-175-L solutions had a material balance of $94.2 \pm 2.0\%$ and $96.7 \pm 4.0\%$, respectively. At test termination, greater than 90% of the applied amount remained as parent in the dark controls indicating that negligible transformation of the parent compounds occurred in the dark. No degradates were observed in the dark controls.

In the irradiated solutions, the concentration of XDE-175-J decreased from 98% at 0 DAT to 0% (95% to 0% for XDE-175-L) of the applied amount at study termination. The major transformation product detected in the irradiated XDE-175-J solutions had a molecular weight of 813 g/mol and a chemical formula of $C_{42}H_{71}NO_{14}$. This degradate reached a maximum concentration of 11% of the applied amount at 7 DAT before declining to approximately 1% at study termination. The major product formed from XDE-175-L photodegradation was N-demethyl-L, which reached a maximum concentration of 13% of the applied radioactivity after 4 h of irradiation and declined to less that 1% after 2 DAT. Numerous minor degradate peaks were observed that were not identified, but all were less than 10% of the applied radioactivity from either parent compounds. No loss of radioactivity due to volatiles was observed.

Photodegradation of XDE-175-J resulted in a complex pathway containing numerous degradates, of which only one (Molecular Weight 813) was observed at concentrations greater than 10% of applied. Photoproducts with additional oxygen atoms or hydroxyl groups were observed. As many as 80 peaks, each accounting for less than 5% of the applied radioactivity, were observed at study termination. Photodegradates became increasingly polar over time.

As a major degradate, N-demethyl-175-L was observed in solutions dosed with XDE-175-L. The N-demethyl-175-L degradate and parent XDE-175-L degraded rapidly to numerous increasingly polar compounds. As many as 75 minor peaks (less than 5% of the applied radioactivity) were detected at 19 DAT. The polar degradates formed from the photolysis of both XDE-175-J and XDE-175-L were presumed to be products created by cleavage of the macrolide ring.

Aqueous hydrolysis

Hydrolysis of radiolabelled XDE-175-J and XDE-175-L at 0.5 mg ai/L was studied in the dark at 25 °C in sterile aqueous buffered solutions at pH 5 (acetate), pH 7 (THAM), and pH 9 (borate) for 30 days (Rutherford, *et al.*, 2005, Report 040108). Samples were analysed at 0, 2, 7, 14, 21, and 30 days by diluting the samples with a mixture of acetonitrile and methanol to adjust the concentration of acetonitrile at 20%.

For XDE-175-J-containing solutions, material balance was $98.4 \pm 1.0\%$, $97.6 \pm 1.4\%$ and $97.0 \pm 1.4\%$ of the applied radioactivity at pH 5, pH 7 and pH 9, respectively. The concentration of parent compound remained constant at $96.2 \pm 1.6\%$, $94.5 \pm 2.1\%$, and $91.3 \pm 2.5\%$ of applied radioactivity at pH 5, pH 7, and pH 9, respectively. At pH 9, a minor transformation product, N-demethyl-175-J, with a maximum average concentration of 6.7% of applied radioactivity, was observed on the 30^{th} day of incubation. Volatiles were not trapped. The average unidentified radioactivity was 2.7, 3.3 and 1.6% of the applied amount at pH 5, pH 7 and pH 9, respectively, at study termination.

XDE-175-J was stable to hydrolysis at pH 5 and pH 7. Although a degradate product formed at pH 9, the averaged parent compound concentration did not decrease below 89% of the applied radiocarbon.

For XDE-175-L-containing solutions, material balance was $94.0 \pm 1.2\%$, $96.9 \pm 0.8\%$ and $97.0 \pm 1.3\%$ of the applied radioactivity at pH 5, pH 7, and pH 9, respectively. The concentration of parent compound remained constant at $90.4 \pm 2.4\%$ and $92.6 \pm 2.1\%$ of applied radiocarbon at pH 5 and pH 7, respectively. At test termination, the average concentration of the parent compound had decreased from 92.1 to 81.6% of the initial applied radioactivity at pH 9. At pH 9 the major transformation product was N-demethyl-175-L, with a maximum average concentration of 11.9% of applied radioactivity, observed on the 30th day of incubation. No minor transformation products were detected. Volatiles were not trapped. The average unidentified radioactivity was 4.8, 5.1 and 3.1% of the applied amount at pH 5, pH 7 and pH 9, respectively, at study termination.

XDE-175-L was stable to hydrolysis at pH 5 and pH 7. Using simple first-order kinetics, the degradation rate constant of XDE-175-L at pH 9 was 0.0045 days⁻¹, which corresponds to a half-life of 154 days and a DT₉₀ of 512 days.

In summary, no hydrolysis of XDE-175-J or XDE-175-L occurred at pH 5 or pH 7. At pH 9, N-demethyl-175-J and N-demethyl-175-L were observed as minor transformation products on the 30th day of incubation. No other minor transformation products were detected.

Residues in Succeeding Crops

In an rotational crop study conducted in the USA in 2008 (Graper and Smith, 2008; Report 040086.01), applications of ¹⁴C-XDE-175-J or ¹⁴C-XDE-175-L were made to confined, outdoor plots of sandy loam soil at rates of 405 and 135 g ai/ha, respectively, which were equivalent to the seasonal maximum use rate (based on 3 to 1 ratio in the final formulation). The test plots were aged for 30, 120 and 365 days after treatment (DAT), and radishes, lettuce, and wheat were planted into the plots at each interval (plant-back). Immature and mature radishes, immature and mature lettuce, wheat forage, wheat hay, and mature wheat grain and straw were harvested from the plots.

Total radioactive residues (TRR) in the raw agricultural commodities (RAC), expressed as mg of XDE-175 equivalents per kg plant tissue (mg/kg) are summarized in Table 10.

Table 10 Total radioactive residues determined in rotational crops planted 30, 120, and 365 days after application of radiolabelled spinetoram to soil

			Total radi	ioactive residu	ies (mg-XDE-	·175 equival	ents/kg)			
		RADISH		LETT	TUCE	WHEAT				
	Immature tops	Mature tops	Mature roots	Immature	Mature	Forage	Hay	Straw	Grain	
30 DAT ^a	(68)	(89)	(89)	(82)	(89)	(105)	(140)	(177)	(177)	
Control b	< LOQ	< LOD	< LOD	< LOD	< LOQ	< LOD	< LOD	< LOD	< LOD	
XDE-175-J	0.079	0.014	0.005	0.085	0.027	0.004	0.013 ^c	0.022	< LOQ	
XDE-175- L	0.021	< LOQ	< LOD	0.033	0.016	< LOQ	< LOQ	0.015	< LOQ	
120 DAT ^a	(56)	(69)	(69)	(56)	(69)	(77)	(93)	(128)	(128)	
XDE-175-J	0.013	0.006	< LOQ	0.012	< LOQ	0.011	0.014	0.023	< LOQ	
XDE-175- L	< LOQ	< LOQ	< LOQ	< LOQ	< LOQ	< LOQ	0.008 ^d	0.020	< LOQ	
365 DAT ^a	(31)	(58)	(58)	(31)	(58)	(31)	(129)	(205)	(205)	
XDE-175-J	0.018	0.016	0.006	0.013	0.005	0.013	0.031	0.045	0.008	
XDE-175- L	0.006	< LOQ	< LOQ	0.016	< LOQ	< LOQ	0.021	0.029	0.006	

^a Days between planting and harvest.

^b Limits of detection (LOD): 0.001 and 0.002 mg/kg for XDE-175-J and XDE-175-L, respectively.

Limits of quantification (LOQ): 0.004 and 0.006 mg/kg for XDE-175-J and XDE-175-L, respectively.

^c Measured TRR value of 0.009 mg/kg (40% moisture) adjusted to 0.013 mg/kg (15% moisture).

^d Measured TRR value of 0.007 mg/kg (23.5% moisture) adjusted to 0.008 mg/kg (15% moisture).

No radioactive residues greater than 0.085 mg/kg, in spinetoram-equivalents, were found in any of the crops evaluated. Crop samples that contained TRR of 0.010 mg/kg or higher were subjected to solvent extraction, and unextractable residues in tissue were less than 0.019 mg/kg. The extractable radioactivity was characterised further by solvent partitioning. Residues of no greater than 0.065, 0.004 and 0.007 mg/kg were found in the neutral organic phases, acidic organic phases, and in the extracted aqueous phase, respectively. The organic phases were purified with silica solid phase extraction (SPE), and then eluates were analysed by HPLC. In any immature or mature sample, no single component exceeded 0.024 mg/kg or 0.007 mg/kg, respectively. The N-demethyl-175-J and N-formyl-175-J metabolites and parent XDE-175-J were tentatively identified at 0.024 mg/kg or less in 30 DAT immature samples and at 0.007 mg/kg or less in 30 DAT mature samples.

At 120 DAT and 365 DAT, no radioactive residues were associated with any of XDE-175-L, N-demethyl-175-L or N-formyl-175-L. The results of 120 DAT indicated that no single residue greater than 0.006 mg/kg was present. Of the XDE-175-J treated crop samples, radish (immature tops), radish (mature tops), lettuce (immature), and wheat forage, hay, and straw contained TRR greater than 0.010 mg/kg but the concentrations were too low for identification.

The characterization and identification of radioactive residues in 30 DAT samples following treatment with radiolabelled XDE-175-J is summarized in Table 11. Identification of the residues from XDE-175-L treated crops was not possible due to low concentration of residues.

Table 11 Characterization and identification of radioactive residues in rotational crops following application of radiolabelled spinetoram

Compound	Radish				Lett	uce		Who	eat hay	Whea	t straw	
	Immature Ma		ture Imn		mature Ma		iture					
	%TR R	mg/kg	%TR R	mg/kg	%TR R	mg/kg	%TR R	mg/kg	%TR R	mg/kg	%TR R	mg/kg
Application of ¹⁴ C-XDE-175-J at 0.41 kg ai/ha (30DAT)												
XDE-175-J	16.1	0.012	-	-	14.1	0.012						
N-demethyl- 175-J			-	-								
O-deethyl-175-J	26.8	0.021	-	-	29.3	0.025	25.3	0.007				
N-formyl-175-J			-	-								
Total identified	42.9	0.033	-	-	43.4	0.037	25.3	0.007				
Total characterised	43.2	0.035	75.4	0.010	42.3	0.036	62.7	0.017	55.3	0.007	56.7	0.013
Total extractable	86.1	0.068	75.4	0.010	85.7	0.073	88.0	0.024	55.3	0.007	56.7	0.013
Total unextractable	13.9	0.011	24.6	0.003	14.3	0.012	15.4	0.004	44.7	0.006	43.3	0.010
Total measured (TRR in ())	100	(0.079)	100	(0.014)	100	(0.085)	103.4	(0.027)	100	(0.013)	100	(0.022)
		Aj	pplicatio	n of ¹⁴ C-2	XDE-17	5-L at 0.13	3 kg ai/h	a (30DAT	")		•	•
Total identified	-	-			-	-	-	-			-	-
Total characterised	60.2	0.013			66.3	0.022	66.5	0.010			50.7	0.007
Total extractable	60.2	0.013			66.3	0.022	66.5	0.010			50.7	0.007
Total unextractable	39.8	0.009			33.7	0.011	33.5	0.005			49.3	0.007
Total measured (TRR in ())	100	(0.021)			100	(0.033)	100	(0.015)			100	(0.015)

Compound	Radish	tops, Lettuce,		Wheat forage		Wheat hay		Wheat straw		
	immatur	immature immatu		re						
	%TRR	mg/kg	%TRR	mg/kg	%TRR	mg/kg	%TRR	mg/kg	%TRR	mg/kg
Application of ¹⁴ C-XDE-175-J at 0.41 kg ai/ha (120DAT)										
Total identified	-	-								
Total characterised	71.4	0.009	76.6	0.009	70.7	0.008	53.9	0.007	57.3	0.013
Total extractable	71.4	0.009	76.6	0.009	70.7	0.008	53.9	0.007	57.3	0.013
Total unextractable	28.6	0.004	23.4	0.003	29.3	0.003	46.1	0.006	42.7	0.010
Total measured	100	(0.013)	100	(0.012)	103	(0.011)	100	(0.014)	100	(0.023)
(TRR in ())										

Compound	Radis	h, ture tops	Radish mature tops		Lettuc	*	Wheat	forage	Whea	Wheat hay		Wheat straw	
	%T	mg/kg	%TR	mg/k	%T	mg/kg	%TR	mg/k	%T	mg/kg	%TR	mg/k	
14	RR		R	g	RR		R	g	RR		R	g	
Application of ¹⁴ 0	C-XDE-	-175-J at 0	.41 kg ai	/ha (365E	DAT)								
Total identified	-	-	-	-	-	-	-	-	-	-	-	-	
Total	63.4	0.011	64.7	0.010	71.8	0.010	73.2	0.010	51.1	0.016	58.2	0.026	
characterised													
Total	63.4	0.011	64.7	0.010	71.8	0.010	73.2	0.010	51.1	0.016	58.2	0.026	
extractable													
Total	36.6	0.006	35.3	0.006	28.2	0.004	26.8	0.003	48.9	0.015	41.8	0.019	
unextractable													
Total measured	100	(0.018	100	(0.01	100	(0.013	103	(0.01	100	(0.031	100	(0.04	
(TRR in ()))		6))		3))		5)	
Application of 14	C-XDE-	-175-L at ().13 kg a	i/ha (365I	DAT)								
Total identified					-	-			-	-	-	-	
Total					60.2	0.010			32.6	0.007	54.1	0.015	
characterised													
Total					60.2	0.010			32.6	0.007	54.1	0.015	
extractable													
Total					39.8	0.006			67.4	0.014	45.9	0.013	
unextractable													
Total measured					100	(0.016			100	(0.021	100	(0.02	
(TRR in ())))		9)	

The levels of radioactivity taken up from soil treated with [\$^{14}\$C]spinetoram into the three succeeding crops (radish, lettuce, and wheat) planted 30, 120, or 365 days after treatment, were below 0.085 mg/kg spinetoram equivalents. Since such low radioactive residues were found in analysed fractions of these rotational crop samples, spinetoram is unlikely to be taken up readily by succeeding crops.

METHODS OF RESIDUE ANALYSIS

Analytical methods

Analytical methods for determination of residues of XDE-175-J and XDE-175-L and a set of their metabolites, including N-demethyl-175-J and -L, N-formyl-175-J and -L were developed for a wide range of plant matrices and animal matrices.

After an extraction specific to matrix, and a reasonably standard cleanup, spinetoram and the metabolites were determined by HPLC with positive-ion electron-spray (ESI) tandem mass spectrometry (HPLC-MS/MS). The methods have been extensively validated with numerous recovery tests on a wide range of matrices.

Multi-residue Methods

FDA Multi-Residue Method PAM, Vol. I

The applicability of the multi-residue screen methods outlined in the FDA Pesticide Analytical Manual, Volume I, Third Edition was assessed for spinetoram and its metabolites, XDE-175-J and XDE-175-L, N-demethyl-175-J and -L, and N-formyl-175-J and -L (Peyton, 2005, Report 051013). The test substances were analysed in accordance with selected multiresidue methods (MRMs) described in Protocols A and C of the FDA PAM I. Results of the tests summarized below show that the FDA PAM I multi-residue methods are not suitable for enforcement purposes.

<u>Protocol A:</u> XDE-175-J, XDE-175-L, N-demethyl-XDE-175-J, N-demethyl-XDE-175-L, N-formyl-XDE-175-J and N-formyl-XDE-175-L were each injected at approximately 1000 ng and showed no response in FSD when analysed according to the above procedures outlined in Protocol A. Because no fluorescence was demonstrated for any of the test substances, no further procedures were required for Protocol A

<u>Protocol B</u>: Because the analytes are not acids or phenols, testing through Protocol B was not required.

Protocol C: All test substances were subjected to Protocol C, modules DG-1, DG-5, DG-13, DG-17, and DG-18. GC systems for each module were set up appropriately according to the parameters described in the PAM Volume 1, Section 302. The test substances were analysed using a nitrogen-phosphorous detector set to nitrogen mode detection according to modules DG-5 and DG-17. DG-5 analysed the test substances at 1000 ng. Results for the analytes at 1000 ng were off-scale and therefore were reanalysed at 200 ng. DG-5 analysis resulted in sample peaks that were broad with significant tailing. DG-17 analysis resulted in sample peaks that were less broad but still contained a significantly large tailing factor. Modules DG-5 and DG-17 were unsuccessful for XDE-175 due to broad peak shape and tailing factors observed. Failure of the test substances to produce quantifiable peaks using the nitrogen-specific modules led to analysis of the test substance by more general modules, non-specific for chemical properties, DG-1, DG-13 and DG-18. No peaks were observed for any of the test substances using electron capture detection modules. Protocol C was unsuccessful due to chromatographic peak broadening and tailing and (for electron capture detection) a lack of detectability.

<u>Protocols D, E, and F</u>: Due to the poor sensitivity and poor chromatographic properties of the test substances to detection by methods described in Protocol C, no further analyses were performed by Protocol D and Protocol E.

<u>Protocol G</u>: Because the test substances are not carbamates, testing through Protocol G was not required.

Multi-Residue Enforcement Method, DFG S19

The German multi-residue enforcement method DFG S19 was intended for the determination of pesticides by gas chromatography (GC). Recently it was proposed to extend the S19 based modular multi-method L 00.00-34 (German §35 LMBG) by a module using liquid chromatography with tandem mass spectrometric detection (LC-MS/MS) for the determination of a multitude of active substances.

Various DFG S19 extraction modules were employed: extraction module E 1 was used for apple; extraction module E 2 was assessed for wheat grain; module E 3 was used for whole orange and grape; and module E 7 was assessed for oilseed rape seed. Basic steps of the E 1, E 2, and E 3 extraction modules are adjustment of neutral pH (if necessary) and total water to 100 mL followed by extraction with water/acetone (1/2, v/v) and subsequent partition into the organic phase by addition of sodium chloride and ethyl acetate/cyclohexane (1/1, v/v). Module E 7 uses extraction with acetonitrile/acetone (225/25, v/v) with addition of synthetic calcium silicate (Calflo E) and Celite to eliminate oily plant matrix and facilitate filtration.

The DFG S19 multiresidue method, in conjunction with LC-MS/MS determination, using matrix matched standards for external calibration, was validated for XDE-175-J and XDE-175-L, N-demethyl-175-J and -L, and N-formyl-175-J and -L in three plant matrices, apple, grape, and orange (Class, 2007; Report 061066). Validation was also attempted for wheat grain and for oilseed rape seed but was unsuccessful most likely due to incomplete extraction and excessive matrix effects. Recoveries of spinetoram and its metabolites in apples, oranges and grape using the multiresidue method are summarized in Table 12. Average recoveries ranged between 73% and 102%, with relative standard deviations of $\leq 14\%$ for all analytes and matrices.

Table 12 Procedural recovery for spinetoram and its metabolites from fortified apple, grape, and orange samples, using the multiresidue method, DFG S-19

Analyte	Matrix	Fortification	Recove	ery Rate (%)	RSD	n
Anaryte	Mauix	Level (mg/kg)	mean	range	(%)	n
XDE-175-J	Apple	0.010	83	77–91	8	5
m/z Q1/Q3 748.7/142.1		1.0	84	81–88	3	5
		0.010-1.0	84	77–91	6	10
XDE-175-L	Apple	0.010	83	77–91	8	5
m/z Q1/Q3 760.7/142.1		1.0	79	71–86	8	5
		0.010-1.0	81	71–91	8	10
N-demethyl-175-J	Apple	0.010	82	75–90	7	5
m/z Q1/Q3 734.7/127.9		1.0	79	76–81	3	5
		0.010-1.0	80	75–90	6	10
<i>N</i> -demethyl-175-L	Apple	0.010	82	69–95	13	5
m/z Q1/Q3 746.8/127.9		1.0	73	69–78	5	5
		0.010-1.0	79	69–95	11	10
N-formyl-175-J	Apple	0.010	96	90–100	9	5
m/z Q1/Q3 762.3/156.1		1.0	91	88–92	5	5
		0.010-1.0	93	88–100	8	10
N-formyl-175-L	Apple	0.010	103	85–112	11	5
m/z Q1/Q3 774.3/155.9		1.0	87	84–89	2	5
		0.010-1.0	95	84–112	12	10
XDE-175-J	Apple	0.010	85	79–92	6	5
m/z Q1/Q3 748.7/98.2		1.0	82	79–86	4	5
		0.010–1.0	83	79–92	5	10
XDE-175-L	Apple	0.010	80	77–87	5	5
m/z Q1/Q3 760.7/98.2		1.0	79	71–86	8	5
		0.010-1.0	79	71–87	6	10
N-demethyl-175-J	Apple	0.010	85	77–94	8	5
m/z Q1/Q3 734.7/84.2		1.0	72	69–75	4	5
		0.010–1.0	78	69–94	11	10
<i>N</i> -demethyl-175-L	Apple	0.010	86	76–98	12	5
m/z Q1/Q3 746.8/84.2		1.0	73	69–79	6	5
		0.010-1.0	80	69–98	12	10
N-formyl-175-J	Apple	0.010	105	98–112	5	5
m/z Q1/Q3 762.3/203.0		1.0	89	88–90	1	5
		0.010-1.0	97	88–112	9	10
N-formyl-175-L	Apple	0.010	105	91–115	9	5
m/z Q1/Q3 774.3/203.0		1.0	87	84–89	2	5

Analyte	Matrix	Fortification	Recove	ery Rate (%)	RSD	n
Allaryte	Matrix	Level (mg/kg)	mean	range	(%)	11
		0.010-1.0	96	84–115	12	10
XDE-175-J	Grape	0.010	92	84–96	6	4
m/z Q1/Q3 748.7/142.1		1.0	84	77–91	7	5
		0.010-1.0	87	77–96	7	9
XDE-175-L	Grape	0.010	88	80–92	6	4
m/z Q1/Q3 760.7/142.1		1.0	78	71–84	7	5
		0.010-1.0	82	71–92	9	9
N-demethyl-175-J	Grape	0.010	84	77–91	7	4
m/z Q1/Q3 734.7/127.9		1.0	78	73–83	6	5
		0.010-1.0	81	73–91	7	9
N-demethyl-175-L	Grape	0.010	86	76–100	12	4
m/z Q1/Q3 746.8/127.9		1.0	73	69–77	4	5
		0.010-1.0	79	69–100	12	9
N-formyl-175-J	Grape	0.010	102	88–111	9	5
m/z Q1/Q3 762.3/156.1		1.0	94	90–100	5	5
		0.010-1.0	98	88–111	8	10
N-formyl-175-L	Grape	0.010	97	89–106	8	5
m/z Q1/Q3 774.3/155.9		1.0	94	91–98	3	5
		0.010-1.0	96	89–106	6	10
XDE-175-J	Grape	0.010	90	85–95	5	4
m/z Q1/Q3 748.7/98.2		1.0	82	76–89	7	5
		0.010-1.0	86	76–95	8	9
XDE-175-L	Grape	0.010	84	77–89	7	4
m/z Q1/Q3 760.7/98.2		1.0	76	69–83	8	5
		0.010-1.0	80	69–89	9	9
<i>N</i> -demethyl-175-J	Grape	0.010	85	78–95	9	4
m/z Q1/Q3 734.7/84.2		1.0	77	72–81	5	5
		0.010-1.0	80	72–95	9	9
N-demethyl-175-L	Grape	0.010	91	77–106	13	4
m/z Q1/Q3 746.8/84.2		1.0	75	72–79	4	5
		0.010-1.0	82	72–106	14	9
N-formyl-175-J	Grape	0.010	97	89–106	7	5
m/z Q1/Q3 762.3/203.0		1.0	94	89–100	5	5
		0.010-1.0	96	89–106	6	10
N-formyl-175-L	Grape	0.010	96	83–113	12	5
m/z Q1/Q3 774.3/203.0		1.0	91	86–98	6	5
		0.010-1.0	94	83–113	9	10
XDE-175-J	Orange	0.010	102	92–114	9	5
m/z Q1/Q3 748.7/142.1		1.0	97	93–100	3	5
		0.010-1.0	99	92–114	7	10
XDE-175-L	Orange	0.010	94	86–104	7	5
m/z Q1/Q3 760.7/142.1		1.0	92	81–99	9	5
		0.010-1.0	93	81–104	7	10
<i>N</i> -demethyl-175-J	Orange	0.010	96	89–112	10	5

Analyte	Matrix	Fortification	Recov	ery Rate (%)	RSD	
Anaryte	Maurix	Level (mg/kg)	mean	range	(%)	n
m/z Q1/Q3 734.7/127.9		1.0	93	88–96	4	5
		0.010-1.0	95	88–112	7	10
<i>N</i> -demethyl-175-L	Orange	0.010	91	83–113	14	5
m/z Q1/Q3 746.8/127.9		1.0	89	79–95	8	5
		0.010–1.0	90	79–113	11	10
N-formyl-175-J	Orange	0.010	102	95–105	4	5
m/z Q1/Q3 762.3/156.1		1.0	95	92–98	3	5
		0.010-1.0	99	92–105	5	10
N-formyl-175-L	Orange	0.010	96	90–107	7	5
m/z Q1/Q3 774.3/155.9		1.0	102	97–105	3	5
		0.010-1.0	99	90–107	6	10
XDE-175-J	Orange	0.010	95	87–104	8	5
m/z Q1/Q3 748.7/98.2		1.0	93	89–96	3	5
		0.010-1.0	94	87–104	6	10
XDE-175-L	Orange	0.010	93	86–101	6	5
m/z Q1/Q3 760.7/98.2		1.0	90	79–97	9	5
		0.010-1.0	92	79–101	8	10
N-demethyl-175-J	Orange	0.010	89	82–100	8	5
m/z Q1/Q3 734.7/84.2		1.0	90	85–92	4	5
		0.010-1.0	90	82–100	6	10
N-demethyl-175-L	Orange	0.010	92	84–106	10	5
m/z Q1/Q3 746.8/84.2		1.0	87	79–92	8	5
		0.010-1.0	89	79–106	9	10
N-formyl-175-J	Orange	0.010	102	98–108	5	5
m/z Q1/Q3 762.3/203.0		1.0	95	91–98	3	5
		0.010-1.0	99	91–108	5	10
N-formyl-175-L	Orange	0.010	101	92–104	6	5
m/z Q1/Q3 774.3/203.0		1.0	101	98–104	3	5
		0.010-1.0	101	92–104	4	10

The FDA PAM I multi-residue methods were found to be unsuitable for the determination of spinetoram and its metabolites in plant and animal matrices for enforcement purposes.

The DFG S19 multi-residue method was only successfully validated for the determination of spinetoram and it metabolites in apples, grapes and oranges when using matrix-matched calibration standards. It was not successful for the determination of the analytes in wheat grain and for oilseed rape seed due to matrix effects. The addition of GPC clean-up was not helpful in achieving successful results, most likely due to the fact that the large molecules elute early with the oily biomolecules. It was therefore predicted that this method would not work for determination of spinetoram and its metabolites in animal matrices.

Analytical Methods for Determination of Residues in Foods and Feeding stuffs of Plant Origin Used in Supervised Trials

Methods GRM 05.03 and GRM 05.04 have been developed for the determination of residues of XDE-175-J and XDE-175-L and their metabolites N-demethyl-175-J and -L, N-formyl-175-J and -L in plant matrices using HPLC with positive-ion electron-spray (ESI) tandem mass spectrometry (LC-MS/MS).

Method GRM 05.03

In method GRM 05.03 developed by Hastings and Wendelburg (2005), residues of spinetoram and its metabolites are extracted from agricultural commodities by shaking with an acetonitrile/water (80:20, v/v) solution. An aliquot of the extraction solvent is removed and a stable isotope internal standard solution containing XDE-175 and metabolites is added. XDE-175 and its metabolites are analysed without sample clean-up by liquid chromatography with positive-ion electron-spray ionization (ESI) tandem mass spectrometry (LC-MS/MS).

The method was validated over the concentration range of 0.01-1.0 mg/kg for all crops (dry crops, wet crops, oily crops and acidic crops), except lettuce which was validated over the concentration range of 0.01–10 mg/kg. The validated limit of quantification (LOQ) for all crops was 0.01 mg/kg (Hastings, 2005, Report 041021). Recoveries of spinetoram and its metabolites in a number of crops fortified at various levels are summarized in Table 13. Average recovery ranged between 82% and 106%, with relative standard deviations of \leq 15% for analytes in all crops, except for N-formyl-175-L in oily crops, where the relative standard deviation was 27%.

Table 13 Procedural recovery for spinetoram and its metabolites in various plant matrices using Method GRM 05.03

A last -	Matrice	Fortification	Recover	ry Rate (%)	RSD	
Analyte	Matrix	Level (mg/kg)	mean	range	(%)	n
XDE-175-J	Wet Crop ^a	0.01	99	89–107	5.1	18
		1.0	96	88-101	3.7	16
		10	93	89–99	4.3	6
		0.01-10	97	88–107	4.9	40
XDE-175-L	Wet Crop	0.01	106	97–116	4.4	18
		1.0	102	97–107	3.0	16
		10	93	90–96	2.1	6
		0.01-10	103	90–116	5.4	40
N-demethyl-175-J	Wet Crop	0.01	99	89–116	7.1	18
		1.0	102	98–109	3.1	16
		10	92	87–97	3.6	6
		0.01-10	99	87–116	6.4	40
N-demethyl-175-L	Wet Crop	0.01	98	89–114	6.6	18
		1.0	100	96–103	2.1	16
		10	93	90–95	1.8	6
		0.01-10	98	89–114	5.2	40
N-formyl-175-J	Wet Crop	0.01	90	79–106	8.1	18
		1.0	94	87–100	4.6	16
		10	88	82–96	5.9	6
		0.01-10	91	79–106	6.8	40
N-formyl-175-L	Wet Crop	0.01	89	76–101	6.9	18
		1.0	92	86–100	4.0	16
		10	89	87–92	2.6	6
		0.01-10	90	76–101	5.4	40
XDE-175-J	Dry Crop ^b	0.01	90	83–103	5.6	20
		1.0	91	84–100	4.5	20
		0.01-1.0	91	83–103	5.0	40
XDE-175-L	Dry Crop	0.01	104	97–114	4.5	20
		1.0	102	96–109	2.9	20

A 1.	24.	Fortification	Recover	y Rate (%)	RSD	
Analyte	Matrix	Level (mg/kg)	mean	range	(%)	n
		0.01-1.0	103	96–114	3.9	40
N-demethyl-175-J	Dry Crop	0.01	102	94–109	3.6	20
		1.0	103	97–110	3.1	20
		0.01-1.0	103	94–110	3.3	40
N-demethyl-175-L	Dry Crop	0.01	103	94–109	4.1	20
		1.0	104	98–113	3.5	20
		0.01-1.0	104	94–113	3.8	40
N-formyl-175-J	Dry Crop	0.01	99	77–115	9.1	20
		1.0	100	82–111	7.0	20
		0.01-1.0	99	77–115	8.0	40
N-formyl-175-L	Dry Crop	0.01	91	71–109	11.0	20
		1.0	96	81–109	8.5	20
		0.01-1.0	93	71–109	10.1	40
XDE-175-J	Acidic Crop ^c	0.01	99	89–114	7.2	17
		1.0	94	86–100	4.0	18
		0.01-1.0	96	86–114	6.4	35
XDE-175-L	Acidic Crop	0.01	107	97–116	5.4	17
		1.0	102	95–107	3.1	18
		0.01-1.0	104	95–116	5.2	35
N-demethyl-175-J	Acidic Crop	0.01	102	95–115	5.1	17
·		1.0	100	96–106	2.8	18
		0.01-1.0	101	95–115	4.2	35
N-demethyl-175-L	Acidic Crop	0.01	102	93–112	5.7	17
·	1	1.0	101	96–106	2.8	18
		0.01-1.0	101	93–112	4.4	35
N-formyl-175-J	Acidic Crop	0.01	94	79–112	10.5	17
		1.0	94	80–106	7.2	18
		0.01-1.0	94	79–112	8.8	35
N-formyl-175-L	Acidic Crop	0.01	93	73–107	10.2	17
		1.0	94	86–100	3.9	18
		0.01-1.0	93	73–107	7.5	35
XDE-175-J	Oily Crop d	0.01	94	88–102	5.3	10
		1.0	95	87–102	4.7	10
		0.01-1.0	95	87–102	4.9	20
XDE-175-L	Oily Crop	0.01	96	90–102	4.5	10
		1.0	100	95–107	4.3	10
		0.01-1.0	98	90–107	4.8	20
N-demethyl-175-J	Oily Crop	0.01	96	92–101	3.2	10
·		1.0	102	95–110	5.1	10
		0.01-1.0	99	92–110	5.1	20
N-demethyl-175-L	Oily Crop	0.01	89	80–99	6.0	10
,	J - 1	1.0	94	84–101	5.2	10
		0.01–1.0	92	80–101	6.3	20
N-formyl-175-J	Oily Crop	0.01	89	71–110	15.0	10

Analyte	Matrix	Fortification	Recover	RSD	n	
Allaryte	Wattix	Level (mg/kg)	mean	range	(%)	11
		1.0	96	85–105	6.8	10
		0.01-1.0	92	71–110	11.7	20
N-formyl-175-L	Oily Crop	0.01	80	48–113	27.5	10
		1.0	83	48–108	25.1	10
		0.01-1.0	82	48–113	25.7	20

^a "Wet crop" includes: broccoli, cabbage, grape, green beans (succulent), leek, lettuce, onion, sweet peppers, and tomato.

An independent validation was carried out at PTRL Europe by analysts having no previous experience with the residue method (Richter, 2005; Report 050051). The limit of quantification was established at 0.01 mg/kg for spinetoram and its metabolites in lettuce (wet crop) and in whole oranges (acidic crop). The final extracts were analysed for residues of spinetoram and its metabolites by LC-MS/MS. Table 14 summarizes the validation data for spinetoram and its metabolites in lettuce and oranges. Average recoveries ranged between 82% and 110%, with relative standard deviations of \leq 14%. Method GRM 05.03 was demonstrated to be applicable for use in the determination of spinetoram and its metabolites residues in these matrices.

Table 14 Procedural recovery obtained by an independent laboratory validation of Method GRM 05.03

Analyte	Matrix	Fortification	Recove	ry Rate (%)	RSD	n
Allaryte	Iviauix	Level (mg/kg)	mean	range	(%)	11
XDE-175-J	T	0.01	92	89–95	3	5
	Lettuce (Wet Crop)	10	83	80–86	3	5
	(wet crop)	0.01-10	88	80–95	6	10
XDE-175-L	Lettuce	0.01	82	77–86	5	5
	(Wet Crop)	10	85	83–90	4	5
	(wet crop)	0.01-10	84	77–90	5	10
N-demethyl-175-J	T ()	0.01	89	84–92	3	5
	Lettuce (Wet Crop)	10	92	85–98	6	5
		0.01-10	90	84–98	5	10
N-demethyl-175-L	T	0.01	87	71–105	14	5
	Lettuce (Wet Crop)	10	87	75–101	14	5
	(wet crop)	0.01-10	87	71–105	13	10
N-formyl-175-J	T	0.01	100	93–111	8	5
	Lettuce (Wet Crop)	10	83	76–97	10	5
	(wet crop)	0.01-10	91	76–111	13	10
N-formyl-175-L	T	0.01	99	89–112	10	5
	Lettuce (Wet Crop)	10	82	77–88	5	5
	(Wet Crop)	0.01-10	91	77–112	12	10

^b "Dry crop" includes: barley grain, forage and straw, grass forage, maize grain forage and stover, and wheat grain, forage and straw.

^c "Acidic crop" includes: apple, cherry (without seed), lemon, orange (whole fruit, peel and pulp), peach (without seed), pear and plum (without seed).

^d "Oily crop" canola seed, cotton seed, olive (without seed), olive oil and soya bean.

A 1 4	Mari	Fortification	Recove	ry Rate (%)	RSD	
Analyte	Matrix	Level (mg/kg)	mean	range	(%)	n
XDE-175-J		0.01	95	87–100	5	5
	Whole Orange (Acidic Crop)	0.10	92	89–98	4	5
	(ricidic Crop)	0.01-0.10	94	87–100	5	10
XDE-175-L		0.01	85	76–92	8	5
	Whole Orange (Acidic Crop)	0.10	93	89–98	5	5
	(Acidic Crop)	0.01-0.10	89	76–98	8	10
N-demethyl-175-J		0.01	89	87–93	3	5
	Whole Orange (Acidic Crop)	0.10	87	84–91	4	5
	(ricidic Crop)	0.01-0.10	88	84–93	4	40
N-demethyl-175-L		0.01	90	81–105	11	5
	Whole Orange (Acidic Crop)	0.10	90	73–100	12	5
	(Acidic Crop)	0.01-0.10	90	73–105	11	10
N-formyl-175-J		0.01	91	83–104	11	5
	Whole Orange (Acidic Crop)	0.10	89	71–95	11	5
	(Acidic Crop)	0.01-0.10	90	71–104	10	10
N-formyl-175-L		0.01	98	96–104	3	5
	Whole Orange (Acidic Crop)	0.10	95	87–103	6	5
	(Acidic Crop)	0.01-0.10	97	87–104	5	10
XDE-175-J		0.01	90	81–102	9	5
	Lettuce	10	90	87–94	2	5
	(Wet Crop)	0.01–10	90	81–102	6	10
XDE-175-L		0.01	96	92–103	7	5
	Lettuce (Wet Crop)	10	86	84–90	2	5
		0.01–10	91	84–103	8	10
N-demethyl-175-J	_	0.01	89	82–93	5	5
	Lettuce (Wet Crop)	10	89	83–99	7	5
	(wet Clop)	0.01–10	89	82- 99	6	10
N-demethyl-175-L	-	0.01	83	72–92	9	5
	Lettuce (Wet Crop)	10	87	77–95	8	5
	(wet Clop)	0.01-10	85	72–95	8	10
N-formyl-175-J	-	0.01	88	82–96	7	5
	Lettuce (Wet Crop)	10	98	92–110	10	5
	(wet Clop)	0.01-10	93	82–110	10	10
N-formyl-175-L	_	0.01	93	70–104	15	5
	Lettuce (Wet Crop)	10	89	74–109	14	5
	(wet crop)	0.01-10	91	70–109	14	10
XDE-175-J		0.01	89	82–100	8	5
	Whole Orange (Acidic Crop)	0.10	96	91–100	4	5
	(Acture Crop)	0.01-0.10	93	82–100	7	10
XDE-175-L		0.01	91	87–96	4	5
	Whole Orange (Acidic Crop)	0.10	92	84–100	6	5
	(reduce crop)	0.01-0.10	91	84–100	5	10

Analyte	Matrix	Fortification	Recover	ry Rate (%)	RSD (%)	n
Anaryte		Level (mg/kg)	mean	range		11
N-demethyl-175-J	W/h - l - O	0.01	89	84–93	4	5
	Whole Orange (Acidic Crop)	0.10	72	70–76	4	5
	17	0.01-0.10	80	70–93	11	40
N-demethyl-175-L	Whole Orange (Acidic Crop)	0.01	94	87–107	8	5
		0.10	106	100- 109	3	5
		0.01-0.10	100	87–109	9	10
N-formyl-175-J		0.01	100	88–119	13	5
	Whole Orange (Acidic Crop)	0.10	110	101 -118	7	5
	(Figure Grop)	0.01-0.10	105	88–119	11	10
N-formyl-175-L		0.01	100	88–108	7	5
	Whole Orange (Acidic Crop)	0.10	110	103- 118	6	5
	(Figure Grop)	0.01-0.10	105	88–118	8	10

Method GRM 05.04

Method GRM 05.04 was developed for the determination of residues of XDE-175-J and XDE-175-L and their corresponding metabolites, N-demethyl-175-J and -L, and N-formyl-175-J and -L in plant matrices by on-line solid phase extraction and HPLC with positive-ion electron-spray ionization (ESI) tandem mass spectrometry (LC-MS/MS) (Hastings and Wendelburg, 2005). The method was proposed as a data-gathering method.

Residues of spinetoram and its metabolites are extracted from agricultural commodities by shaking with an acetonitrile/water (80:20, v/v). An aliquot of the extraction solvent is removed and a stable isotope internal standard solution containing XDE-175 and metabolites is added. The solution is purified by on-line solid phase extraction (SPE) using a C18 cartridge. The extract is loaded onto the SPE cartridge with an acetonitrile/methanol/water (15:15:70, v/v) containing ammonium acetate. The SPE cartridge is washed with an acetonitrile/methanol/water (25:25:50, v/v) containing ammonium acetate and eluted onto the analytical C18 column with a gradient elution technique using the HPLC mobile phase. Spinetoram and its metabolites are analysed by liquid chromatography with positive-ion electron-spray ionization (ESI) tandem mass spectrometry (LC-MS/MS).

The method was validated over the concentration range of 0.01-.0 mg/kg for dry crops, oily crops and acidic crops, and over the concentration range of 0.01-10 mg/kg for wet crops. The validated limit of quantification (LOQ) for all crops was 0.01 mg/kg (Hastings, 2005, Report 041021). Recoveries of spinetoram and its metabolites in a number of crops fortified at various levels are summarized in Table 15. Average recovery ranged between 88% and 111%, with relative standard deviations of $\leq 20\%$ for all analytes and crops tested.

Table 15 Procedural recovery for spinetoram and its metabolites in various plant matrices using Method GRM 05.04 (Hastings, 2005)

Analyte	Matrix	Fortification Level (mg/kg)	Recover	y Rate (%)	RSD (%)	n
	iviauix		mean	range		11
XDE-175-J	Wet Crop ^a	0.01	103	95–118	5.6	18
		1.0	99	92–103	3.5	16
		10	92	90–94	1.3	6
		0.01-10	100	90–118	5.7	40
XDE-175-L	Wet Crop	0.01	104	96–114	5.3	18
		1.0	101	95–107	3.0	16
		10	92	89–98	3.2	6

A 1.	76.1	Fortification	Recover	ry Rate (%)	RSD	
Analyte	Matrix	Level (mg/kg)	mean	range	(%)	n
		0.01–10	101	89–114	6.0	40
N-demethyl-175-J	Wet Crop	0.01	100	94–117	5.8	18
		1.0	100	94–105	2.8	16
		10	94	89–101	5.5	6
		0.01–10	99	89–117	5.3	40
N-demethyl-175-L	Wet Crop	0.01	102	92–120	6.6	18
		1.0	101	95–104	2.3	16
		10	91	87–99	5.4	6
		0.01-10	100	87–120	6.3	40
N-formyl-175-J	Wet Crop	0.01	93	86–106	7.2	18
		1.0	101	91–116	6.9	16
		10	91	84–101	7.6	6
		0.01–10	96	84–116	8.3	40
N-formyl-175-L	Wet Crop	0.01	100	79–119	11.2	18
•		1.0	99	91–111	4.9	16
		10	90	81–99	7.8	6
		0.01–10	98	79–119	9.3	40
XDE-175-J	Dry Crop ^b	0.01	88	81–105	6.0	20
		1.0	88	81–98	5.6	20
		0.01-1.0	88	81–105	5.7	40
XDE-175-L	Dry Crop	0.01	102	94–114	5.5	20
		1.0	102	92–114	6.2	20
		0.01-1.0	102	92–114	5.8	40
N-demethyl-175-J	Dry Crop	0.01	102	92–114	5.1	20
,		1.0	102	95–114	3.8	20
		0.01-1.0	102	92–114	4.4	40
N-demethyl-175-L	Dry Crop	0.01	105	99–114	4.3	20
		1.0	104	96–110	3.6	20
		0.01-1.0	105	96–114	4.0	40
N-formyl-175-J	Dry Crop	0.01	105	77–131	12.5	20
		1.0	110	97–121	5.3	20
		0.01-1.0	108	77–131	9.7	40
N-formyl-175-L	Dry Crop	0.01	95	78–119	12.1	20
		1.0	93	77–117	12.5	20
		0.01-1.0	94	77–119	12.2	40
XDE-175-J	Acidic Crop ^c	0.01	102	93–115	6.0	17
		1.0	101	94–105	3.3	18
		0.01-1.0	102	93–115	4.8	35
XDE-175-L	Acidic Crop	0.01	106	98–114	4.6	17
		1.0	100	92–104	3.0	18
		0.01-1.0	103	92–114	5.0	35
N-demethyl-175-J	Acidic Crop	0.01	104	96–114	5.6	17
		1.0	102	97–107	3.1	18
		0.01-1.0	103	96–114	4.5	35

Amalista	Matrix	Fortification	Recover	ry Rate (%)	RSD	
Analyte	Matrix	Level (mg/kg)	mean	range	(%)	n
N-demethyl-175-L	Acidic Crop	0.01	100	87–113	7.8	17
		1.0	101	96–107	2.9	18
		0.01-1.0	100	87–113	5.8	35
N-formyl-175-J	Acidic Crop	0.01	97	76–118	12.4	17
		1.0	103	94–118	6.0	18
		0.01-1.0	100	76–118	9.8	35
N-formyl-175-L	Acidic Crop	0.01	94	86–107	7.4	17
		1.0	97	91–107	4.2	18
		0.01-1.0	95	86–107	6.1	35
XDE-175-J	Oily Crop ^d	0.01	94	84–105	7.4	10
		1.0	96	89–102	4.9	10
		0.01-1.0	95	84–105	6.2	20
XDE-175-L	Oily Crop	0.01	99	90–110	6.3	10
		1.0	101	95–107	4.7	10
		0.01-1.0	100	90–110	5.5	20
N-demethyl-175-J	Oily Crop	0.01	97	86–105	6.2	10
		1.0	103	93–111	5.1	10
		0.01-1.0	100	86–111	6.3	20
N-demethyl-175-L	Oily Crop	0.01	90	83–101	6.4	10
		1.0	98	92–102	3.4	10
		0.01-1.0	94	83–102	6.2	20
N-formyl-175-J	Oily Crop	0.01	103	82–120	11.2	10
		1.0	115	102–140	9.8	10
		0.01-1.0	109	82–140	11.7	20
N-formyl-175-L	Oily Crop	0.01	105	63–138	19.7	10
		1.0	111	76–139	15.4	10
		0.01-1.0	108	63–139	17.3	20

^a "Wet crop" includes: broccoli, cabbage, grape, green beans (succulent), leek, lettuce, onion, sweet peppers, and tomato.

Methods GRM 05.03 and GRM 05.04 are considered to be suitable for the determination of residues of spinetoram and its metabolites in commodities of plant origin. The methods were fully validated for a range of crops and crop types, with a limit of quantification (LOQ) of 0.01 mg/kg for all crops. Method GRM 05.03 has been designated by EPA as an appropriate enforcement method for analysis of residues of spinetoram and its metabolites in plant matrices.

Analytical Methods for Determination of Residues in Foods and Feedstuffs of Animal Origin Used in Supervised Trials

Methods GRM 05.15 and GRM 06.08 were developed for the determination of residues of spinetoram and its metabolites in bovine and poultry tissues, milk, cream, and eggs, using HPLC with positive ion electron-spray (ESI) tandem mass spectrometry (LC-MS/MS).

b "Dry crop" includes: barley grain, forage and straw, grass forage, maize grain forage and stover, and wheat grain, forage and straw.

^c "Acidic crop" includes: apple, cherry (without seed), lemon, orange (whole fruit, peel and pulp), peach (without seed), pear and plum (without seed).

^d "Oily crop" canola seed, cotton seed, olive (without seed), olive oil and soya bean.

Method GRM 05.15

Method GRM 05.15 was developed for the quantitative determination of XDE-175-J and XDE-175-L and their metabolites N-demethyl-175-J, N-demethyl-175-L, 3'-O-deethyl-175-J and 3'-O-deethyl-175-L in bovine tissues (muscle, kidney, liver and fat), milk (whole and skim) and cream, and poultry tissues (muscle, liver and fat) and poultry eggs (Shackelford and Hastings, 2005).

Residues of spinetoram and its metabolites are extracted from the samples of animal origin by homogenizing and shaking with an acetonitrile/water solution (80:20, v/v). The samples are filtered through a 0.45 μ m membrane GHP filter. A stable isotope internal standard solution containing XDE-175-J, XDE-175-L, and metabolites N-demethyl-175-J and N-demethyl-175-L is added to each sample. Spinetoram and its metabolites are analysed without sample cleanup by liquid chromatography with positive-ion electron-spray ionization (ESI) tandem mass spectrometry (LC-MS/MS).

The method was validated over the concentration range of 0.01-15 mg/kg for bovine and poultry muscle and bovine kidney; 0.01-50 mg/kg for bovine and poultry liver, bovine milk and cream and poultry eggs; and 0.01-150 mg/kg for bovine and poultry fat (Shackelford, 2005; Report 051022). The validated limit of quantification (LOQ) for all of the animal matrices was 0.01 mg/kg. Recoveries of spinetoram and its metabolites in animal matrices fortified at various levels are summarized in Table 16. Average recovery ranged between 87% and 119%, with relative standard deviations of $\leq 18.2\%$ for all analytes and matrices tested.

Table 16 Procedural recovery for spinetoram and its metabolites in various animal matrices, using Method GRM 05.15 (Shackelford, 2005)

A 1.	34.	Fortification	Recove	ry Rate (%)	RSD	
Analyte	Matrix	Level (mg/kg)	mean	range	(%)	n
XDE-175-J	J Muscle	0.010	108	101–115	4.3	12
	(bovine &	1.0	98	93–102	3.2	10
	poultry)	15	100	93–106	4.6	10
		0.010–15	103	93–115	6.0	32
XDE-175-L	Muscle	0.010	100	92–114	6.5	12
	(bovine &	1.0	92	89–95	2.1	10
	poultry)	15	97	92–102	3.7	10
		0.010–15	97	89–114	5.7	32
<i>N</i> -demethyl-175-J	Muscle (bovine &	0.010	111	106–117	2.7	12
		1.0	103	99–108	3.1	10
	poultry)	15	105	96–112	5.6	10
		0.010–15	106	96–117	5.0	32
<i>N</i> -demethyl-175-L	Muscle	0.010	101	92–110	6.2	12
	(bovine &	1.0	100	95–104	2.4	10
	poultry)	15	108	96–116	7.4	10
		0.010–15	103	92–116	6.6	32
3'- <i>O</i> -deethyl-175-J	Muscle	0.010	113	103–118	4.9	12
	(bovine &	1.0	93	90–99	3.4	10
	poultry)	15	102	96–109	3.9	10
		0.010–15	103	90–118	9.0	32
3'-O-deethyl-175-L	Muscle	0.010	119	115–125	2.0	12
	(bovine &	1.0	91	87–93	2.2	10
	poultry)	15	104	97–109	4.2	10
		0.010–15	105	87–125	11.3	32

	M ()	Fortification	Recove	ery Rate (%)	RSD	
Analyte	Matrix	Level (mg/kg)	mean	range	(%)	n
XDE-175-J	T :	0.010	102	84–117	9.6	12
	Liver (bovine &	1.0	100	91–108	6.2	10
	poultry)	15	103	100–109	3.1	10
		50	99	84–111	11.7	10
		0.010-50	101	84–117	8.2	42
XDE-175-L	τ.	0.010	104	83–119	11.5	12
	Liver (bovine &	1.0	93	87–99	4.7	10
	poultry)	15	98	93–101	2.5	10
		50	93	81–103	9.9	10
		0.010-50	97	81–119	9.3	42
N-demethyl-175-J	T :	0.010	108	99–118	4.3	12
	Liver (bovine &	1.0	97	92–102	3.2	10
	poultry)	15	102	97–114	5.4	10
		50	97	80–112	14.6	10
		0.010-50	101	80–118	8.9	42
N-demethyl-175-L	Liver	0.010	103	95–115	6.7	12
	(bovine &	1.0	104	93–119	8.2	10
	poultry)	15	103	97–116	6.9	10
		50	98	80–113	15.9	10
		0.010-50	102	80–119	9.7	42
3'-O-deethyl-175-J	Liver	0.010	112	101–120	5.4	12
	(bovine &	1.0	103	93–118	8.6	10
	poultry)	15	106	95–116	6.7	10
		50	100	86–114	11.0	10
		0.010-50	106	86–120	8.7	42
3'-O-deethyl-175-L	Liver	0.010	110	104–116	4.4	12
	(bovine &	1.0	93	85–100	5.7	10
	poultry)	15	102	91–115	8.1	10
		50	101	85–116	11.4	10
		0.010-50	102	85–116	9.6	42
XDE-175-J	Kidney	0.010	101	91–112	7.1	6
	(bovine)	1.0	90	88–92	2.1	5
		15	97	95–100	2.3	5
		0.010–15	96	88–112	6.4	16
XDE-175-L	Kidney	0.010	103	97–109	4.6	6
	(bovine)	1.0	87	84–93	4.2	5
		15	91	87–96	4.6	5
		0.010–15	94	84–109	8.4	16
<i>N</i> -demethyl-175-J	Kidney	0.010	111	103–116	4.4	6
	(bovine)	1.0	105	100–112	5.3	5
		15	110	99–119	8.3	5
		0.010–15	109	99–119	6.2	16

	3.5	Fortification	Recove	ry Rate (%)	RSD	
Analyte	Matrix	Level (mg/kg)	mean	range	(%)	n
<i>N</i> -demethyl-175-L	Kidney	0.010	108	102–116	4.8	6
	(bovine)	1.0	104	101–110	3.9	5
		15	112	102–119	7.3	5
		0.010–15	108	101–119	6.0	16
3'-O-deethyl-175-J	Kidney	0.010	107	104–112	3.1	6
	(bovine)	1.0	93	90–96	3.2	5
		15	100	94–104	4.7	5
		0.010–15	100	90–112	7.0	16
3'-O-deethyl-175-L	Kidney	0.010	106	90–119	11.9	6
	(bovine)	1.0	88	84–92	3.6	5
		15.0	99	94–102	3.7	5
		0.010-15.0	98	84–119	11.3	16
XDE-175-J	Fat	0.01	105	87–119	8.2	14
	(bovine &	1.0	101	92–109	5.2	10
	poultry)	15	105	102–110	2.3	10
		150	95	89–101	3.9	10
		0.010-150	102	87–119	6.8	44
XDE-175-L	Fat	0.01	100	88–114	7.2	14
	(bovine &	1.0	92	88–96	2.9	10
	poultry)	15	98	94–101	1.7	10
		150	93	88–97	3.2	10
		0.010-150	96	88–114	5.8	44
<i>N</i> -demethyl-175-J	Fat	0.01	110	102–117	3.6	14
	(bovine &	1.0	101	95–109	4.5	10
	poultry)	15	100	94–111	6.0	10
		150	99	95–104	2.8	10
		0.010–150	103	94–117	6.0	44
<i>N</i> -demethyl-175-L	Fat	0.01	102	89–118	8.6	14
	(bovine &	1.0	103	100–108	2.5	10
	poultry)	15	102	93–118	7.5	10
		150	101	97–106	2.8	10
		0.010-150	102	89–118	6.1	44
3'- <i>O</i> -deethyl-175-J	Fat	0.01	109	91–118	8.0	14
	(bovine &	1.0	94	86–100	4.5	10
	poultry)	15	103	99–110	2.8	10
		150	94	89–100	3.4	10
		0.010–150	101	86–118	8.6	44
3'-O-deethyl-175-L	Fat	0.01	111	102–119	5.4	14
	(bovine &	1.0	93	89–97	2.6	10
	poultry)	15	105	100–113	3.1	10
		150	94	89–99	3.6	10
		0.010–150	102	89–119	8.8	44

	3.6	Fortification	Recove	ery Rate (%)	RSD	
Analyte	Matrix	Level (mg/kg)	mean	range	(%)	n
XDE-175-J	Milk	0.010	100	92–109	6.2	12
		1.0	95	86–104	6.5	10
		15	99	88–107	6.1	10
		50	93	82–110	13.7	10
		0.010-50	97	82–110	8.7	42
XDE-175-L	Milk	0.010	103	93–115	5.9	12
		1.0	96	92–103	3.5	10
		15	95	86–99	4.2	10
		50	88	79–102	11.6	10
		0.010-50	96	79–115	8.7	42
N-demethyl-175-J	Milk	0.010	114	105–119	3.9	12
		1.0	99	95–107	3.7	10
		15	103	95–109	4.7	10
		50	94	80–115	16.9	10
		0.010-50	103	80–119	10.9	42
N-demethyl-175-L	Milk	0.010	108	96–117	6.5	12
		1.0	102	98–109	3.8	10
		15	107	99–114	5.6	10
		50	95	80–116	17.3	10
		0.010-50	103	80–117	10.3	42
3'-O-deethyl-175-J	Milk	0.010	107	96–114	6.4	12
		1.0	93	86–99	4.5	10
		15	98	83–107	7.6	10
		50	93	83–108	11.6	10
		0.010-50	98	83–114	9.6	42
3'-O-deethyl-175-L	Milk	0.010	110	98–115	4.6	12
		1.0	92	86–102	5.0	10
		15	99	84–106	6.9	10
		50	92	81–105	11.0	10
		0.010-50	99	81–115	10.1	42
XDE-175-J	Egg	0.010	100	92–113	8.8	6
		1.0	96	93–102	4.2	4
		15	100	98–102	1.5	5
		50	96	82–106	12.0	5
		0.010-50	99	82–113	7.6	20
XDE-175-L	Egg	0.010	98	95–103	3.0	6
		1.0	92	88–95	3.4	4
		15	96	94–97	1.0	5
		50	90	78–97	10.1	5
		0.010–50	94	78–103	6.1	20
<i>N</i> -demethyl-175-J	Egg	0.010	108	97–116	6.6	6
		1.0	100	99–103	1.9	4
		15	103	97–113	7.1	5
		50	102	79–116	16.6	5

Analyte	Matrix	Fortification	Recove	ry Rate (%)	RSD	n
Analyte	Maurx	Level (mg/kg)	mean	range	(%)	11
		0.010-50	104	79–116	9.4	20
N-demethyl-175-L	Egg	0.010	95	83–108	11.7	6
		1.0	104	102–105	1.1	4
		15	106	100–115	7.5	5
		50	103	78–119	18.2	5
		0.010-50	101	78–119	11.7	20
3'-O-deethyl-175-J	Egg	0.010	104	96–111	5.7	6
		1.0	91	90–92	1.1	4
		15	99	97–102	2.6	5
		50	92	78–101	11.3	5
		0.010-50	97	78–111	8.2	20
3'-O-deethyl-175-L	Egg	0.010	108	100–117	5.8	6
		1.0	90	89–92	1.3	4
		15	98	94–102	3.7	5
		50	93	78–102	11.6	5
		0.010-50	98	78–117	9.6	20

An independent validation was carried out at PTRL Europe by analysts having no previous experience with the residue method (Richter, 2005, Report 050049). Bovine milk (high water content) and poultry muscle were used as the representative matrices. Control bovine milk samples and control poultry muscle samples that were fortified at 0.01 mg/kg (at LOQ) and 15 mg/kg (1500× the LOQ) were analysed by liquid chromatography with positive-ion electron-spray ionization (ESI) tandem mass spectrometry (LC-MS/MS).

Table 17summarizes the validation data for spinetoram and its metabolites in bovine and poultry tissues. Average recovery ranged between 85% and 109%, with relative standard deviations of $\leq 12\%$.

Table 17 Procedural recovery obtained by an independent laboratory validation of Method GRM 05.15

Analyte	Matrix	Fortification	Recove	ry Rate (%)	RSD	n
Anaryte	Width	Level (mg/kg)	mean	range	(%)	11
XDE-175-J	Bovine Milk	0.010	103	97–108	4	5
		15	91	87–99	5	5
		0.010–15	97	87–108	8	10
XDE-175-L	Bovine Milk	0.010	105	99–109	4	5
		15	89	84–98	6	5
		0.010–15	97	84–109	10	10
N-demethyl-175-J	Bovine Milk	0.010	93	89–102	6	5
		15	89	82–94	7	5
		0.010–15	91	82–102	6	10
N-demethyl-175-L	Bovine Milk	0.010	85	72–97	12	5
		15	93	81–107	11	5
3'-O-deethyl-175-J	Bovine Milk	0.010	109	91–118	10	5
		15	100	90–105	6	5
		0.010–15	105	90–118	9	10

Analyte	Matrix	Fortification	Recove	ry Rate (%)	RSD	
Anaryte	Matrix	Level (mg/kg)	mean	range	(%)	n
3'-O-deethyl-175-L	Bovine Milk	0.010	108	100–115	6	5
		15	92	86–99	5	5
		0.010–15	100	86–115	10	10
XDE-175-J	Poultry Muscle	0.010	105	94–117	8	5
		15	101	98–105	3	5
		0.010–15	103	94–117	6	10
XDE-175-L	Poultry Muscle	0.010	103	97–113	7	5
		15	101	96–105	3	5
		0.010–15	102	96–113	5	10
N-demethyl-175-J	Poultry Muscle	0.010	107	102–117	6	5
		15	94	90–98	3	5
		0.010–15	101	90–117	8	10
N-demethyl-175-J	Poultry Muscle	0.010	98	88–112	10	5
		15	98	87–109	10	5
		0.010–15	98	87–112	9	10
3'-O-deethyl-175-J	Poultry Muscle	0.010	98	91–119	12	5
		15	93	90–98	5	5
		0.010–15	95	90–119	9	10
3'-O-deethyl-175-J	Poultry Muscle	0.010	109	100–113	5	5
		15	101	94–107	5	5
		0.010–15	105	94–113	6	10

Method GRM 06.08

Method GRM 06.08 supersedes method GRM 05.15. Initially method GRM 05.15 was developed to include the determination of metabolites N-demethyl-175-J, N-demethyl-175-L, 3'-O-deethyl-175-J and 3'-O-deethyl-175-L. Method GRM 06.08 was developed to only include the determination of N-demethyl-175-J and N-formyl-175-J (Shackelford, *et. al.*, 2007).

Residues of spinetoram and its metabolites are extracted from the animal matrices by homogenizing and by shaking with an acetonitrile/water solution (80:20, v/v). A stable isotope internal standard solution containing XDE-175-J, XDE-175-L, and metabolites N-demethyl-175-J and N-demethyl-175-L is added to each sample. Samples are analysed by direct injection of the extraction solution without further cleanup. Determination of spinetoram and its metabolites is by liquid chromatography with positive ion electron-spray ionization (ESI) tandem mass spectrometry (LC-MS/MS).

Method GRM 06.08 is identical to method GRM 05.15, except in slight differences in the metabolites determined. Method GRM 06.08 was validated for the quantitative determination of XDE-175-J and XDE-175-L and the major metabolites of XDE-175-J, i.e., N-demethyl-175-J and N-formyl-175-J, in bovine tissues (muscle, kidney, liver and fat), milk (whole and skim) and cream, and poultry tissues (muscle, liver and fat) and poultry eggs (Shackelford, 2007, Report 061023). With the exception of metabolite N-formyl-175-J, the method was validated over the concentration range of 0.01–15 mg/kg in bovine and poultry muscle and bovine kidney; 0.01–50 mg/kg for bovine and poultry liver, bovine milk and cream and poultry eggs; and 0.01-150 mg/kg for bovine and poultry fat.

For N-formyl-175-J, the method was validated over the concentration range of 0.01-0.10 mg/kg for all bovine matrices and over the concentration range of 0.01-5.0 mg/kg for all poultry matrices. The validated limit of quantification (LOQ) for all of the animal matrices was

0.01 mg/kg. Recoveries of spinetoram and its metabolites in bovine and poultry tissues are summarized Table 18. Average recovery ranged between 83% and 118%, with relative standard deviations of $\leq 17\%$ for all analytes and matrices tested.

Table 18 Procedural recovery for spinetoram and its metabolites in various animal matrices, using Method 06.08 (Shackelford, 2007)

Analyte	Matrix	Fortification	Reco	overy Rate (%)	RSD	n
rmaryte	Maura	Level (mg/kg)	mean	range	(%)	11
XDE-175-J	Muscle	0.010	99	73–117	13.9	22
	(bovine &	0.10	90	85–94	3.2	10
	poultry)	1.0	98	93–102	3.4	10
		15	100	93–106	4.7	10
		0.010–15	97	73–117	10.2	52
XDE-175-L	Muscle	0.010	90	68–114	13.3	22
	(bovine &	0.10	86	82–89	2.3	10
	poultry)	1.0	92	89–95	2.2	10
		15	97	92–102	3.7	10
		0.010–15	91	68–114	9.6	52
<i>N</i> -demethyl-175-J	Muscle	0.010	106	90–117	6.6	22
	(bovine &	0.10	111	107–117	2.8	10
	poultry)	1.0	103	99–108	3.1	10
		15	105	96–112	5.6	10
		0.010–15	106	90–117	5.7	52
N-formyl-175-J	Muscle	0.010	89	77–109	9.7	24
	(bovine &	0.10	88	79–107	7.9	20
	poultry)	5.0	105	100–110	3.2	6
		0.010-5.0	90	77–110	10.2	50
XDE-175-J	Liver	0.010	103	84–117	9.6	12
	(bovine &	1.0	100	91–108	6.2	10
	poultry)	15	103	100–109	3.0	10
		50	99	84–111	11.8	10
		0.010-50	101	84–117	8.3	42
XDE-175-L	Liver	0.010	104	83–119	11.5	12
	(bovine &	1.0	94	87–99	4.7	10
	poultry)	15	98	93–101	2.6	10
		50	93	81–103	10.0	10
		0.010-50	98	81–119	9.3	42
<i>N</i> -demethyl-175-J	Liver	0.010	108	99–118	4.4	12
	(bovine &	1.0	97	92–102	3.3	10
	poultry)	15	102	97–114	5.5	10
		50	98	80–112	14.8	10
		0.010-50	101	80–118	9.0	42
N-formyl-175-J	Liver	0.010	89	72–107	13.1	14
	(bovine &	0.10	91	80–102	7.8	10
	poultry)	5.0	107	103–114	3.7	6
		0.010-5.0	93	72–114	12.4	30

Amalasta	Matria	Fortification	Reco	overy Rate (%)	RSD	
Analyte	Matrix	Level (mg/kg)	mean	range	(%)	n
XDE-175-J	Kidney	0.010	101	91–112	7.0	6
	(bovine)	1.0	91	88–92	1.8	5
		15.0	97	95–100	2.2	5
		0.010-15.0	96	88-112	6.3	16
XDE-175-L	Kidney	0.010	103	97–109	4.6	6
	(bovine)	1.0	87	84–93	4.3	5
		15	91	87–96	4.7	5
		0.010–15	95	84–109	8.5	16
N-demethyl-175-J	Kidney	0.010	111	103–116	4.4	6
	(bovine)	1.0	105	100–112	5.4	5
		15	110	99–119	8.5	5
		0.010–15	109	99–119	6.2	16
N-formyl-175-J	Kidney	0.010	83	75–103	12.6	8
	(bovine)	0.10	86	80–91	5.3	4
		0.010-0.10	84	75–103	10.4	12
XDE-175-J	Fat	0.01	105	87–119	8.0	14
	(bovine &	1.0	101	92–109	5.1	10
	poultry)	15	105	102-110	2.3	10
		150	95	89–101	3.9	10
		0.010-150	102	87–119	6.7	44
XDE-175-L	Fat	0.01	100	88–114	7.1	14
	(bovine &	1.0	92	88–96	2.9	10
	poultry)	15	98	94–101	1.9	10
		150	93	88–97	3.1	10
		0.010-150	96	88–114	5.8	44
<i>N</i> -demethyl-175-J	Fat	0.01	110	102–117	3.7	14
	(bovine &	1.0	101	95–109	4.6	10
	poultry)	15	101	94–111	6.0	10
		150	99	95–104	2.9	10
		0.010-150	103	94–117	6.1	44
N-formyl-175-J	Fat	0.01	94	79–122	9.3	42
	(bovine &	0.10	96	84–109	6.5	24
	poultry)	5.0	113	105–121	5.4	6
		0.010-5.0	96	79–122	9.6	72
XDE-175-J	Milk and Cream	0.010	100	92–109	6.1	12
		1.0	95	86–104	6.6	10
		15	99	88–107	6.2	10
		50	93	82–110	13.6	10
		0.010-50	97	82–110	8.7	42
XDE-175-L	Milk and Cream	0.010	103	93–115	5.9	12
		1.0	96	92–103	3.4	10
		15	95	86–99	4.3	10
		50	88	79–102	11.6	10
		0.010–50	96	79–115	8.6	42

Analyte	Matrix	Fortification	Reco	overy Rate (%)	RSD	n
Anaryte	Wiatrix	Level (mg/kg)	mean	range	(%)	11
N-demethyl-175-J	Milk and Cream	0.010	114	105–119	3.9	12
		1.0	99	95–107	3.7	10
		15	103	95–109	4.7	10
		50	94	80–115	17.0	10
		0.010-50	103	80–119	11.0	42
N-formyl-175-J	Milk and Cream	0.010	88	67–100	7.8	38
		0.10	92	81–104	6.4	18
		1.0	91	87–94	NA	2
		0.010-1.0	89	67–104	7.5	58
XDE-175-J	Egg	0.010	101	92–113	8.8	6
		1.0	96	93–102	4.2	4
		15	100	98–102	1.7	5
		50	96	82–106	12.0	5
		0.010-50	99	82–113	7.6	20
XDE-175-L	Egg	0.010	98	95–103	3.1	6
		1.0	92	88–95	3.4	4
		15	96	94–97	1.3	5
		50	89	78–97	10.2	5
		0.010-50	94	78–103	6.3	20
<i>N</i> -demethyl-175-J	Egg	0.010	108	97–116	6.5	6
		1.0	101	99–103	1.9	4
		15	104	97–113	7.1	5
		50	102	79–116	16.6	5
		0.010-50	104	79–116	9.3	20
N-formyl-175-J	Egg	0.010	93	74–111	15.5	6
		0.10	92	88–96	4.1	6
		5.0	106	98–111	5.2	6
		0.010–5.0	97	74–111	11.1	18

Methods GRM 05.15 and GRM 06.08 are considered to be suitable for the determination of spinetoram and its metabolites in commodities of animal origin. The methods were fully validated for a range of animal matrices, with a limit of quantification (LOQ) of 0.01 mg/kg for all matrices. Method GRM 06.08 has been designated by EPA as an appropriate enforcement method for analysis of residues of spinetoram and its metabolites in animal matrices.

Stability of residues in stored analytical samples

The Meeting received information on frozen storage stability of residues of spinetoram and its metabolites in wheat, soya bean, orange, lettuce and sugar beet samples.

Commodities of plant origin

A storage stability study was conducted with XDE-175 and metabolites in agricultural commodities to determine the stability of the residues in homogenized samples while stored frozen (Wendelburg, 2006; Report 050027.01). Wheat grain, soya bean seed, orange whole fruit, lettuce leaf, and sugar beet root were selected to represent each of five crop groupings (dry, oily, acidic, wet, and root crops) to obtain information on a wide variety of crops. Samples taken from trial studies (report 040063 and 040051) were used.

The stability of the compounds, XDE-175-J, XDE-175-L, N-demethyl-175-J, N-demethyl-175-L, N-formyl-175-J, and N-formyl-175-L, were investigated for twelve months. All samples were weighed into individual high density polyethylene (HDPE) containers, fortified at 0.10 mg/kg and stored in a temperature-monitored freezer set at approximately –20 °C. Samples were analysed at 0, 63, 90, 231, and 372 days of frozen storage after fortification, by LC-MS/MS following method GRM 05.04.

The efficiency of the analytical method was determined at the time of analysis of each sampling event by creating two concurrent recovery samples and analysing them according to the method above. Concurrent recovery results confirmed the suitability of the method for determination of spinetoram and its metabolites in the agricultural commodities selected.

Results of the storage stability tests are summarized in Table 19. The results show that spinetoram and its metabolites are stable for at least 12 months in agricultural commodities, except lettuce, under frozen storage at approximately $-20\,^{\circ}$ C. In lettuce, remaining XDE-175-L and N-dimethyl-175-L were 60 and 65% respectively (unadjusted for procedural recovery) on 372 days after initiation of the study.

Table 19 Stability of spinetoram and specific metabolites in agricultural commodities at fortification level of 0.10 mg/kg following storage at –20 °C (Wendelburg, 2006)

Commodity	Storage interval (days)	% Remaining ^a	Concurrent recovery (%)
	XDE-	175-J	
Wheat Grain	0	104	103
	63	119	119
	90	95	110
	231	102	102
	372	99	109
Soya bean Seed	0	105	108
	63	97	95
	90	98	110
	231	103	96
	372	95	92
Orange Fruit	0	101	102
	63	99	104
	90	95	106
	231	100	98
	372	98	97
Lettuce Leaf	0	90	96
	63	91	89
	90	95	91
	231	85	87
	372	87	96
Sugar Beet Root	0	100	98
	63	105	103
	90	96	106
	231	98	101
	372	87	99

	Storage		
Commodity	interval	% Remaining ^a	Concurrent recovery (%)
	(days)		
	XDE-	-175-L	
Wheat Grain	0	107	104
	63	98	100
	90	108	111
	231	95	103
	372	91	98
Soya bean Seed	0	101	108
	63	100	85
	90	104	108
	231	92	99
	372	84	89
Orange Fruit	0	98	96
	63	88	94
	90	102	104
	231	92	96
	372	86	88
Lettuce Leaf	0	88	89
	63	87	76
	90	105	105
	231	73	77
	372	60	81
Sugar Beet Root	0	100	102
	63	88	91
	90	100	105
	231	100	93
	372	111	102
	N-demet	hyl-175-J	
Wheat Grain	0	106	116
	63	96	95
	90	103	114
	231	97	101
	372	89	101
Soya bean Seed	0	100	106
	63	97	89
	90	96	104
	231	92	93
	372	86	87
Orange Fruit	0	99	99
	63	96	95
	90	94	100
	231	91	95
	372	83	84

	Storage		
Commodity	interval	% Remaining ^a	Concurrent recovery (%)
	(days)		
Lettuce Leaf	0	94	109
	63	90	98
	90	91	102
	231	84	102
	372	80	95
Sugar Beet Root	0	98	96
	63	97	93
	90	89	102
	231	88	91
	372	83	90
	N-demet	hyl-175-L	
Wheat Grain	0	103	105
	63	85	90
	90	98	106
	231	90	98
	372	80	94
Soya bean Seed	0	101	107
	63	89	95
	90	96	102
	231	87	93
	372	85	88
Orange Fruit	0	100	102
	63	88	93
	90	88	97
	231	91	95
	372	84	87
Lettuce Leaf	0	93	104
	63	91	95
	90	90	99
	231	77	93
	372	65	89
Sugar Beet Root	0	100	102
	63	88	94
	90	87	102
	231	88	95
	372	80	95
	N-form	yl-175-J	
Wheat Grain	0	133	141
	63	83	82
	90	90	105
	231	83	88
	372	78	85

	Storage		
Commodity	interval	% Remaining ^a	Concurrent recovery (%)
	(days)		
Soya bean Seed	0	106	108
	63	102	87
	90	111	127
	231	91	95
	372	96	92
Orange Fruit	0	91	94
	63	93	90
	90	86	90
	231	90	88
	372	79	75
Lettuce Leaf	0	98	101
	63	105	110
	90	89	94
	231	90	85
	372	86	95
Sugar Beet Root	0	92	90
	63	86	88
	90	91	99
	231	77	102
	372	79	85
	N-form	yl-175-L	
Wheat Grain	0	99	106
	63	94	94
	90	112	117
	231	97	87
	372	79	96
Soya bean Seed	0	116	108
	63	84	88
	90	129	145
	231	104	108
	372	98	90
Orange Fruit	0	89	85
	63	78	96
	90	94	101
	231	90	90
	372	84	75
Lettuce Leaf	0	80	99
	63	75	75
	90	102	102
	231	86	84
	372	86	91

Commodity	Storage interval (days)	% Remaining ^a	Concurrent recovery (%)
Sugar Beet Root	0	82	83
	63	89	79
	90	93	101
	231	93	104
	372	77	87

^a Mean of three subsamples.

Commodities of animal origin

Samples of animal tissues, milk, and eggs from the metabolism and feeding studies were analysed within 20 days of sample collection in the supervised trials. Therefore, storage stability tests were not conducted.

USE PATTERN

The Meeting received approved labels from Canada and the USA.

Information on registered formulations, applications methods and dosage rates of spinetoram for uses on the crops for which supervised trial data were provided is summarized in Table 20. Unless otherwise noted, each of the following GAPs are for field use and all applications are foliar applications. Other than the crops for which supervised trial data were available to the Meeting, spinetoram is approved for use on asparagus, banana, beet greens, Brassica vegetables, bulb vegetables, bush berries, caneberries, cereals, corn (field and sweet), cotton, cranberry, cucurbits, fig, grape, herbs, legume vegetables, mint, peanuts, pulses, soya bean, strawberry, tropical fruits and turnip greens in these countries.

Table 20 Registered uses of spinetoram

Crop	Country	Formul	ation		Application				PHI
		g ai/L or g ai/kg	type	Method	Rate g ai/ha	Rate g ai/hL	Min. interval days	Max. no./ g ai/ season	days
Apricot	Canada	250	WG	Foliar	53–105		7	3	14
Apricot	USA	250	WG	Foliar	53-123		7	4 / (490) ^a	14
Cherry	Canada	250	WG	Foliar	53–105		7	3	7
Cherry	USA	250	WG	Foliar	53–123		3	4 / (490) ^a	7
Citrus fruits	USA	250	WG	Foliar	53-105		7	3 / (210) ^a	1
Fruiting vegetables	Canada	250	WG	Foliar	35–50		5	3	1
Fruiting vegetables	USA	120	SC	Foliar	44–88		4	6 / (298) ^a	1
Leaf of root, tuber & legume vegetables	USA	120	SC	Foliar	44–88		7	6 / (298) ^a	3
Leafy vegetables	Canada	250	WG	Foliar	35–50		5	3	1
Leafy vegetables	USA	120	SC	Foliar	44–88		4	6 / (298) ^a	1
Nectarine	Canada	250	WG	Foliar	53-105		7	3	14
Nectarine	USA	250	WG	Foliar	53–123		7	4 / (490) ^a	1
Peach	Canada	250	WG	Foliar	53-105		7	3	14

Crop	Country	Formul	ation			Application	n		PHI
		g ai/L or g ai/kg	type	Method	Rate g ai/ha	Rate g ai/hL	Min. interval days	Max. no./ g ai/ season	days
Peach	USA	250	WG	Foliar	53–123		7	4 / (490) ^a	14
Plums, prunes	Canada	250	WG	Foliar	53-105		7	3	7
Plums, prunes	USA	250	WG	Foliar	53–123		3	4 / (490) ^a	7
Pome fruits	Canada	250	WG	Foliar	53-105		7	3	7
Pome fruits	New Zealand	250	WG	Foliar	50	2.5	14	4	7
Pome fruits	USA	250	WG	Foliar	79–123		7	4 / (490) ^a	7
Root vegetables (carrot, horse radish, radish, rutabaga, turnip)	Canada	250	WG	Foliar	35–50		5	3	3
Root vegetables (potato, sugar beet, etc)	USA	120	SC	Foliar	53–70		7	4 / (281) ^a	7
Root vegetables (radish, turnip)	USA	120	SC	Foliar	53–70		4	3 / (210) ^a	3
Tree nuts	USA	250	WG	Foliar	26–123		7	4 / (490) ^a	14

^a Total application per season or year.

RESIDUES RESULTING FROM SUPERVISED TRIALS ON CROPS

The Meeting received information on supervised field trials of spinetoram on the following crops:

Commodity	Group	Table No.
Oranges	Citrus	Table 21
Apple	Pome fruits	Table 22
Apple	Pome fruits	Table 23
Apple	Pome fruits	Table 24
Pear	Pome fruits	Table 25
Cherry	Stone fruit	Table 26
Peach	Stone fruit	Table 27
Nectarine	Stone fruit	Table 28
Apricot	Stone fruit	Table 29
Tomatoes	Fruiting vegetables, Other than Cucurbits	Table 30
Leafy lettuce	Leafy vegetables	Table 31
Sugarbeet	Root and tuber vegetables	Table 32
Almonds	Tree nuts	Table 33
Pecan nut	Tree nuts	Table 34
Sugar beet leaves and tops	Animal feedstuffs	Table 35
Almond hulls	Animal feedstuffs	Table 36

Trials were generally well documented with full laboratory and field reports. Laboratory reports included method validation, with batch recoveries at residue levels similar to those occurring in samples from the supervised trials. Dates of analyses or duration of residue sample storage were also provided. In general, data on procedural recoveries were within the acceptable range 70-120%, with relative standard deviation of < 20%. In addition, frozen storage periods from harvest to analysis of all samples were within those confirmed to be stable through separate storage stability determinations.

Field reports provide data on the dates of spray applications, methods used and sampling dates. Although trials included control plots, but no control data are recorded in the tables below unless residues in control samples exceeded the LOQ.

The residue concentrations are reported for XDE-175-J, XDE-175-L, N-demethyl-175-J and N-formyl-175-J. They are unadjusted for procedural recovery. Where residues were below the limit of detection (0.003 mg/kg in US and Canadian trials and 0.05 mg/kg in trials in Australia and New Zealand), they are expressed as "ND". Where they are below the limit of quantification (0.01 mg/kg) and at or above the limit of detection, they are expressed as "< 0.01".

Total residues for estimation of maximum residue levels were calculated by summing up the concentrations of XDE-175-J and XDE-175-L. XDE-175-J was the primary residue reflecting higher ratio in spinetoram formulations. The method for calculation of the total residues is illustrated below. In the calculation, "ND" is treated in the same manner as "< 0.01".

XDE-175-J	XDE-175-L	Total
	mg/kg	
< 0.01	< 0.01	< 0.01
0.05	< 0.01	0.05
0.06	0.02	0.08

Total residues for estimation of STMRs were calculated by summing up the concentrations of XDE-175-J, XDE-175-L, *N*-demethyl-175-J and *N*-formyl-175-J. In most trials, XDE-175-J was the primary residue at shorter PHIs while, in many cases, *N*-formyl-175-J was found at higher concentrations than XDE-175-J at longer PHIs. On the other hand, XDE-175-L, with its concentration being one third of that of XDE-175-J, was a minor component in the four compounds. *N*-demethyl-J was in most cases at lower concentrations than XDE-175-J and in a number of trials was not determined. The method for calculation of the total residues taking the above into account is illustrated below. In the calculation, "ND" is treated in the same manner as "< 0.01".

XDE-175-J	XDE-175-L	N-demethyl-175-J	N-formyl-175-J	Total
		mg/kg		
< 0.01	< 0.01	< 0.01	< 0.01	< 0.02
0.05	< 0.01	< 0.01	< 0.01	0.06
< 0.01	< 0.01	< 0.01	0.05	0.06
0.05	< 0.01	< 0.01	0.05	0.10
0.06	0.02	0.02	0.06	0.16

In all the US trials, duplicate samples were taken and analysed. In accordance with the 2007 JMPR's decision, the higher value of the two was used for estimating maximum residue level.

Total values of XDE-175-J and XDE-175-L residues from the trials conducted according to the GAP have been underlined and used for the estimation of maximum residue levels. Corresponding total values of XDE-175-J, XDE-175-L, N-demethyl-175-J and N-formyl-175-J residues were used for estimation of STMRs and they are double underlined.

Orange

Twelve supervised trials on oranges were conducted in commercial citrus growing areas in the US in 2004 and 2007 (Dolder and Wendelberg, 2005; Report 040063; McKellar, 2008; Report ARAP 07D-001).

Trials were conducted in six sites, each site consisting of a control plot and 2 treated plots. Trial plots had a minimum of 6 orange trees, with samples taken from the 4 inner trees. In each site, one plot was treated using a low spray volume (700 L/ha) and the other, a high spray volume (approximately 3300 L/ha). Each treated plot received three applications of a suspension concentrate (SC) formulation containing 100 g/L of spinetoram at the nominal rate of 70 g ai/ha or a seasonal total of 210 g ai/ha. The applications were made at 4-day intervals and mature fruits were collected at the PHI of 1 day. In one of each of the low volume and high volume spray trials, samples were also taken on days 0, 3, 7, and 14 after the last application.

Approximately 4.5 kg of mature oranges were collected by hand. At each sampling, a single composite sample was taken from the control plot while duplicate composite samples were taken independently from each treated plot. Samples were frozen within 4 h of collection, placed in plastic bags, labelled, shipped frozen, and kept in temperature-monitored freezers at -20 °C until analysis.

Spinetoram and the metabolites were analysed using the method GRM 05.04 with an LOQ of 0.01 mg/kg and LOD of 0.003 mg/kg. Samples from the 2008 trials were analysed using method GRM 05.03, with the same LOQ and LOD values as GRM 05.04.

Table 21-1 Residues of spinetoram from supervised trials on orange in the USA (for estimation of maximum residue level)

	Form	A	pplication		Total/	PHI,	R	esidue, mg/l	ζg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	Total ^a	No.
GAP, USA Citrus fruits	SC or WG		53– 105	3	210	1				
Foliar ap	plication	using low	spray volu	me (~70	00 L/ha)					
Deleon Springs, FL, 2004	SC	10	70–72	3	213	1	0.030	< 0.01	0.030	040063
(Valencia)							0.028	< 0.01	0.028	
Mount Dora, FL, 2004	SC	11	71–72	3	214	1	0.011	ND	0.011	040063
(Valencia)							0.022	< 0.01	0.022	
Raymondville,	SC	10	70–72	3	213	1	< 0.01	ND	< 0.01	040063
TX, 2004 (N-33)							< 0.01	ND	< 0.01	
Richgrove, CA,	SC	9	70–71	3	211	1	< 0.01	ND	< 0.01	040063
2004 (Olinda)							< 0.01	ND	< 0.01	
Porterville,CA,	SC	9	70	3	210	0	0.011	ND	0.011	040063
2004 (Cutter Valencia)							0.014	< 0.01	0.014	
(Cutter varenera)						1	0.012	< 0.01	0.012	
							0.010	ND	0.010	
						3	< 0.01	ND	< 0.01	
							0.011	ND	0.011	
						7	< 0.01	ND	< 0.01	
							< 0.01	ND	< 0.01	
						14	< 0.01	ND	< 0.01	
							< 0.01	ND	< 0.01	
FL, 2007	WG	11	69- 72	3	212	1	0.028	< 0.01	0.028	ARAP
(Not specified)							0.025	ND	0.025	07D-001
Foliar app	lication u	sing high	spray volu	me (~33	00 L/ha)					
Deleon Springs,	SC	2	70-71	3	212	1	0.015	< 0.01	0.015	040063

	Form	A	pplication		Total/	PHI,	R	esidue, mg/l	κg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	Total ^a	No.
GAP, USA Citrus fruits	SC or WG		53– 105	3	210	1				
FL, 2004 (Valencia)							0.014	ND	0.014	
Mount Dora, FL, 2004	SC	2	70-71	3	212	1	0.017	< 0.01	0.017	040063
(Valencia)							0.018	< 0.01	0.018	
Raymondville, TX, 2004	SC	2	70-71	3	211	1	< 0.01	ND	< 0.01	040063
(N-33)							ND	ND	< 0.01	
Richgrove, CA, 2004	SC	2	70-71	3	211	1	0.021	< 0.01	0.021	040063
(Olinda)							0.020	< 0.01	0.020	
Porterville, CA,	SC	2	70	3	210	0	0.021	< 0.01	0.021	040063
2004 (Cutter Valencia)							0.012	< 0.01	0.012	
						1	0.010	ND	0.010	
							0.012	ND	0.012	
						3	< 0.01	ND	< 0.01	
							ND	ND	< 0.01	
						7	< 0.01	ND	< 0.01	
							< 0.01	ND	< 0.01	
						14	< 0.01	ND	< 0.01	
							< 0.01	ND	< 0.01	
FL, USA, 2007	WG	2	69- 70	3	209	1	0.021	< 0.01	0.021	ARAP
(Not specified)							0.024	< 0.01	0.024	07D-001

 $^{^{}a}$ Total residues = XDE-175-J + XDE-175-L

Table 21-2 Residues of spinetoram and metabolites from supervised trials on orange in the USA (for estimation of STMR) $\,$

Location, year		App	plication	1	Total/			Res	sidue, mg	/kg		
(Variety)	Form	g ai/hL	g ai/ha	No	season, g ai/ha	PHI, days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total a	Report No.
GAP, USA Citrus fruits	SC or WG		53- 105	3	210	1						
			Folia	appli	cation usi	ng low	spray vol	ume (~70	00 L/ha)			
Deleon			70				0.030	< 0.01	0.011	0.016	0.057	
Springs, FL, 2004 (Valencia)	SC	10	70- 72	3	213	1	0.028	< 0.01	0.014	0.024	0.066	040063
Mount Dora,	90	11	71-	3	21.4	1	0.011	ND	< 0.01	< 0.01	0.021	0.400.62
FL, 2004 (Valencia)	SC	11	72	3	214	1	0.022	< 0.01	0.012	0.017	0.051	040063
Raymondville,	9.0	10	70-	2	212	1	< 0.01	ND	< 0.01	0.012	0.022	0.400.62
TX, 2004 (N-33)	SC	10	72	3	213	1	< 0.01	ND	< 0.01	0.011	0.021	040063
Richgrove,	SC	9	70-	3	211	1	< 0.01	ND	ND	ND	< 0.02	040063

Location, year		App	plication	n	Total/			Res	sidue, mg	/kg		
(Variety)	Form	g ai/hL	g ai/ha	No	season, g ai/ha	PHI, days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total	Report No.
GAP, USA Citrus fruits	SC or WG		53- 105	3	210	1						
CA, 2004 (Olinda)			71				< 0.01	ND	ND	< 0.01	< 0.02	
Porterville,CA,							0.011	ND	< 0.01	0.014	0.025	
2004 (Cutter Valencia)	SC	9	70	3	210	0	0.014	< 0.01	< 0.01	0.012	0.026	040063
						1	0.012	< 0.01	ND	0.015	0.027	
						1	0.010	ND	< 0.01	0.020	0.030	
						3	< 0.01	ND	ND	< 0.01	< 0.02	
							0.011	ND	< 0.01	0.022	0.033	
						7	< 0.01	ND	ND	0.015	0.025	
							< 0.01	ND	ND	< 0.01	< 0.02	
						14	< 0.01	ND	ND	0.013	0.023	
							< 0.01	ND	ND	0.015	0.025	
FL, 2007 (Not	WG	11	69-	3	212	1	0.028	< 0.01	0.013	< 0.01	0.051	ARAP
specified)	W G	11	72	3	212	1	0.025	ND	< 0.01	ND	0.035	07D-001
Foliar ap	plication	using h	igh spr	ay vol	ume (~330	00 L/ha)					
Deleon							0.015	< 0.01	< 0.01	0.026	0.041	
Springs, FL, 2004 (Valencia)	SC	2	70- 71	3	212	1	0.014	ND	< 0.01	0.024	0.038	040063
Mount Dora,		_	70-	_		_	0.017	< 0.01	< 0.01	0.022	0.039	
FL, 2004 (Valencia)	SC	2	71	3	212	1	0.018	< 0.01	< 0.01	0.021	0.039	040063
Raymondville,	9.0	_	70-		211		< 0.01	ND	< 0.01	< 0.01	< 0.02	0.400.62
TX, 2004 (N-33)	SC	2	71	3	211	1	ND	ND	ND	< 0.01	< 0.02	040063
Richgrove,			70-				0.021	< 0.01	< 0.01	0.032	0.053	
CA, 2004 (Olinda)	SC	2	71	3	211	1	0.020	< 0.01	< 0.01	0.049	0.069	040063
Porterville,	SC	2	70	3	210	_	0.021	< 0.01	< 0.01	0.038	0.059	
CA, 2004						0	0.012	< 0.01	< 0.01	0.026	0.038	
(Cutter Valencia)							0.010	ND	< 0.01	0.035	0.045	0.400.60
ŕ						1	0.012	ND	< 0.01	0.035	0.047	040063
						2	< 0.01	ND	ND	0.019	0.029	
						3	ND	ND	ND	< 0.01	< 0.02	
						7	< 0.01	ND	ND	0.026	0.036	
						7	< 0.01	ND	ND	0.026	0.036	
						14	< 0.01	ND	ND	0.023	0.033	
						14	< 0.01	ND	ND	0.020	0.03	
FL, USA,							0.021	< 0.01	0.010	< 0.01	0.041	
2007 (Not specified)	WG	2	69- 70	3	209	1	0.024	< 0.01	0.012	< 0.01	0.046	ARAP 07D- 001

 $^{^{}a}$ Total residues = XDE-175-J + XDE-175-L + ND-J + NF-J

Pome Fruits

Supervised trials on apples were conducted in Australia, Canada, New Zealand, and the USA during 2004-2007. Trials on pears were also conducted in Australia and New Zealand during 2005/2006 season.

Supervised trials on apple in the USA

Supervised trials on apples were conducted in major apple growing areas in the USA in 2004 and 2007 (Dolder and Wendelburg, 2005; Report 040063; McKellar, 2008; Report ARAP 07D-001).

Twelve trials were carried out in six different sites, with ten trials conducted in 2004 and two conducted in 2007. Each site consisted of one control plot and two treated plots. Each treated plot had a minimum of 6 apple trees, with samples taken from the four inner trees. In each site, one plot was treated using low spray volume application (~700 L/ha) and the other, a high volume spray (~3300 L/ha). Five applications of a formulation (100 g/L SC or 250 g/kg WG) were made to each treated plot using tractor-mounted airblast sprayers at a nominal rate of 100 g ai/ha or a total seasonal rate of 500 g ai/ha. Applications were made at intervals of about 28 days and mature fruits were harvested at the PHI of 7 days. In one of each of the low volume and high volume spray trials, samples were also taken on days 0, 1, 3, 7, and 14 after the last application.

Approximately 2.6 kg of apples were harvested by hand on each sampling date. A single composite sample was taken from the control plot while duplicate composite samples were taken independently from each treated plot. Samples were frozen within 4 h of collection, placed in plastic bags, labelled, shipped frozen, and kept in temperature-monitored freezers at -20 °C until analysis.

Spinetoram and the metabolites were analysed using the method GRM 05.04 with an LOQ and LOD of 0.01 mg/kg and 0.003 mg/kg, respectively.

Table 22-1 Residues of spinetoram from supervised trials on apples in the USA (for estimation of maximum residue level)

	Form	A	pplication		Total/	PHI	Ro	esidue, mg/l	κg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	day s	XDE- 175-J	XDE- 175-L	Total ^a	No.
GAP, USA Pome fruits	SC or WG		79-123	4	490	7				
		Fol	iar applicat	ion usin	g low spray	volume	(~700 L/ha	1)		
North Rose, NY, 2004	SC	14	100	5	500	7	< 0.01	ND	< 0.01	040063
(Rome)							< 0.01	ND	< 0.01	
Blairsville, GA, 2004	SC	14	99-103	5	505	7	0.011	ND	0.011	040063
(Red Rome)							0.013	ND	0.013	
Paynesville, MN, 2004	SC	15	101- 102	5	508	7	ND	ND	< 0.01	040063
(Honey Crisp)			102				ND	ND	< 0.01	
Orosi, CA, 2004	SC	13	97-101	5	494	7	ND	ND	< 0.01	040063
(Granny Smith)							< 0.01	ND	< 0.01	
Ephrata, WA,	SC	14	100	5	500	0	0.048	0.014	0.062	040063
2004 (Red							0.044	0.013	0.057	
Delicious)						1	0.019	< 0.01	0.019	
							0.027	< 0.01	0.027	
						3	0.014	ND	0.014	

	Form	A	pplication		Total/	PHI	Re	esidue, mg/l	kg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	day s	XDE- 175-J	XDE- 175-L	Total ^a	No.
GAP, USA Pome fruits	SC or WG		79-123	4	490	7				
							0.019	< 0.01	0.019	
						7	0.013	ND	0.013	
							< 0.01	ND	< 0.01	
						14	< 0.01	ND	< 0.01	
							< 0.01	ND	< 0.01	
MI, 2007 (Not specified)	WG	16	99- 101	5	501	7	0.028	ND	0.028	ARAP 07D-001
(r tot specifica)			101				0.023	ND	0.023	072 001
		Folia	ar applicati	on using	g high spray	volume	(~3300 L/h	a)		
North Rose, NY, 2004	SC	4	98-100	5	498	7	< 0.01	ND	< 0.01	040063
(Rome)							< 0.01	ND	< 0.01	
Blairsville, GA, 2004	SC	3	100- 103	5	509	7	0.012	ND	0.012	040063
(Red Rome)			103				0.010	ND	0.010	
Paynesville, MN, 2004	SC	3	101- 102	5	506	7	ND	ND	< 0.01	040063
(Honey Crisp)			102				ND	ND	< 0.01	
Orosi, CA, 2004 (Granny	SC	3	98-100	5	498	7	< 0.01	ND	< 0.01	040063
Smith)							< 0.01	ND	< 0.01	
Ephrata, WA,	SC	3	99-100	5	498	0	0.041	0.011	0.052	040063
2004 (Red							0.045	0.012	0.057	
Delicious)						1	0.015	ND	0.015	
							0.012	ND	0.012	
						3	0.013	ND	0.013	
							0.011	ND	0.011	
						7	< 0.01	ND	< 0.01	
							< 0.01	ND	< 0.01	
						14	< 0.01	ND	< 0.01	
							< 0.01	ND	< 0.01	
MI, 2007	WG	3	96- 101	5	498	7	0.020	ND	0.020	ARAP 07D-001
(Not specified)			101				0.020	ND	0.020	U/D-001

^a Total residue = XDE-175-J + XDE-175-L.

Table 22-2 Residues of spinetoram and metabolites from supervised trials on apples in the USA (for estimation of STMR)

	Form	Ap	plication	1	Total/	PHI,		Res	sidue, mg/	'kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total ^a	No.
GAP, USA Pome fruits	SC or WG		79- 123	4	490	7						

	Form	Ap	plication	ı	Total/	PHI,		Re	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total	No.
GAP, USA Pome fruits	SC or WG		79- 123	4	490	7						
Folia	r applicat	ion using	g low spi	ay vol	ume (~700	L/ha)						
North Rose, NY, 2004	SC	14	100	5	500	7	< 0.01	ND ND	ND ND	ND < 0.01	< 0.02 < 0.02	040063
(Rome) Blairsville,	SC	14	99-	5	505	7	0.011	ND	< 0.01	ND	0.021	040063
GA, 2004 (Red Rome)	SC	14	103	3	303	/	0.011	ND ND	< 0.01	< 0.01	0.021	040003
Paynesville,	SC	15	101-	5	508	7	ND	ND	ND	ND	< 0.02	040063
MN, 2004 (Honey Crisp)			102				ND	ND	ND	ND	< 0.02	
Orosi, CA, 2004	SC	13	97- 101	5	494	7	ND	ND	ND	ND	< 0.02	040063
(Granny Smith)							< 0.01	ND	ND	ND	< 0.02	
Ephrata, WA,	SC	14	100	5	500	0	0.048	0.014	< 0.01	< 0.01	0.072	040063
2004 (Red							0.044	0.013	< 0.01	< 0.01	0.067	
Delicious)						1	0.019	< 0.01	< 0.01	0.010	0.029	
							0.027	< 0.01	0.012	0.014	0.053	
						3	0.014	ND	< 0.01	< 0.01	0.024	
							0.019	< 0.01	< 0.01	0.013	0.032	-
						7	0.013	ND	0.010	0.013	0.036	-
							< 0.01	ND	< 0.01	< 0.01	< 0.02	
						14	< 0.01	ND	< 0.01	0.010	0.020	-
							< 0.01	ND	< 0.01	< 0.01	< 0.02	
MI, 2007 (Not specified)	WG	16	99- 101	5	501	7	0.028	ND ND	< 0.01	0.010	0.038	ARAP 07D- 001
Foliar	application	on using	high spr	av voli	ıme (~330	0 L/ha)						
North Rose,	SC	4	98-	5	498	7	< 0.01	ND	ND	< 0.01	< 0.02	040063
NY, 2004 (Rome)			100				< 0.01	ND	< 0.01	0.012	0.022	
Blairsville,	SC	3	100-	5	509	7	0.012	ND	< 0.01	< 0.01	0.022	040063
GA, 2004 (Red Rome)			103				0.010	ND	< 0.01	< 0.01	0.020	
Paynesville, MN, 2004	SC	3	101-	5	506	7	ND	ND	ND	ND	< 0.02	040063
MN, 2004 (Honey Crisp)		_	102				ND	ND	ND	ND	< 0.02	
Orosi, CA, 2004	SC	3	98- 100	5	498	7	< 0.01	ND	ND	< 0.01	< 0.02	040063
(Granny Smith)							< 0.01	ND	ND	< 0.01	< 0.02	
Ephrata, WA, 2004	SC	3	99- 100	5	498	0	0.041	0.011	< 0.01	< 0.01	0.062	040063
(Red			100				0.045	0.012	< 0.01	< 0.01	0.067	
						1	0.015	ND	< 0.01	0.017	0.032	

	Form	Ap	plication	1	Total/	PHI,		Re	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total a	No.
GAP, USA Pome fruits	SC or WG		79- 123	4	490	7						
Delicious)							0.012	ND	< 0.01	0.016	0.028	
						3	0.013	ND	< 0.01	0.017	0.030	
							0.011	ND	< 0.01	0.018	0.029	
						7	< 0.01	ND	ND	0.012	0.022	
							< 0.01	ND	< 0.01	0.016	0.026	
						14	< 0.01	ND	ND	0.015	0.025	
							< 0.01	ND	ND	0.010	0.020	
MI, 2007 (Not	WG	3	96- 101	5	498	7	0.020	ND	< 0.01	0.017	0.037	ARAP 07D-
specified)		_	_			_	0.020	ND	< 0.01	0.016	0.036	001

^a Total residue = XDE-175-J + XDE-175-L + ND-J + NF-J.

Supervised trials on apple in Canada

Six supervised trials were conducted on apples in Canada in 2005 (Dolder and Schelle, 2006; Report 050041). Two additional trials included in the study were conducted in Idaho, USA.

A total of eight trials were carried out in four different sites, four trials in two sites in Ontario, two trials in Quebec, and two trials in Idaho. Each site consisted of one control plot and two treated plots. Three applications of a water dispersible granule formulation (WG) containing 250 g /kg spinetoram were made at two different rates, one plot receiving a seasonal rate of 240 g ai/ha and the other, 315 g ai/ha. Applications were made at 6–8-day intervals, using airblast sprayers with 5–7 nozzles, with spray volumes ranging from 916 to 993 L/ha. Mature apples were harvested 5–7 days after the last application.

A minimum of 24 fruits, weighing approximately 2.8 kg were harvested by hand. A single composite sample was taken from the control plot while duplicate composite samples were taken independently from each treated plot. Samples were frozen within 4 h of collection, placed in plastic bags, labelled, shipped frozen, and kept in temperature-monitored freezers at -20 °C until analysis.

Spinetoram and the metabolites were analysed using the method GRM 05.04 with an LOQ and LOD of 0.01 mg/kg and 0.003 mg/kg, respectively.

Table 23-1 Residues of spinetoram from supervised trials on apple in Canada (for estimation of maximum residue level)

	Form	Aj	pplication		Total/	PHI	Re	esidue, mg/k	ιg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	day s	XDE- 175-J	XDE- 175-L	Total ^a	No.
GAP, Canada	WG		53-105	3		7				
St. George, Ontario, 2005	WG	8	79- 81	3	239	6	< 0.01	ND	< 0.01	050041
(Red Delicious)							< 0.01	ND	< 0.01	

	Form	Aj	pplication		Total/	PHI	Re	esidue, mg/l	ιg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	day s	XDE- 175-J	XDE- 175-L	Total ^a	No.
St. George, Ontario, 2005 (Red Delicious)	WG	11	103- 106	3	314	6	0.017	< 0.01	0.017	050041
St.George,	WG	8	76- 80	3	233	6	0.010	ND	0.010	050041
Ontario, 2005 (IdaRed)							0.010	ND	0.010	
St.George,	WG	11	99-	3	309	5	0.011	ND	0.011	050041
Ontario, 2005 (IdaRed)			108				0.015	ND	0.015	
S-Paul d'Abbotsford,	WG	9	77- 80	3	237	7	< 0.01	ND	< 0.01	050041
Quebec, 2005 (McIntosh)							< 0.01	ND	< 0.01	
S-Paul d'Abbotsford,	WG	11	101- 107	3	314	7	< 0.01	ND	< 0.01	050041
Quebec, 2005 (McIntosh)							< 0.01	ND	< 0.01	
Rockland, Idaho, USA,	WG	8	78- 80	3	236	7	0.018	< 0.01	0.018	050041
2005 (McIntosh)							0.020	< 0.01	0.020	
Rockland, Idaho, USA,	WG	11	101- 103	3	306	7	0.028	< 0.01	0.028	050041
2005 (McIntosh)							0.019	< 0.01	0.019	

^a Total residue = XDE-175-J + XDE-175-L.

Table 23-2. Residues of spinetoram and metabolites from supervised trials on apple in Canada (for estimation of STMR)

	Form	Ap	plication	n	Total/	PHI,		Re	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total	No.
GAP, Canada	WG		53- 105	3		7						
St. George, Ontario,	WG	8	79- 81	3	239	6	< 0.01	ND	< 0.01	ND	< 0.02	050041
2005 (Red Delicious)							< 0.01	ND	< 0.01	< 0.01	< 0.02	
St. George, Ontario, 2005	WG	11	103- 106	3	314	6	0.017	< 0.01	0.011	< 0.01	0.038	050041
(Red Delicious)							0.012	ND	< 0.01	< 0.01	0.022	
St.George,	WG	8	76-	3	233	6	0.010	ND	< 0.01	< 0.01	0.020	050041
Ontario, 2005 (IdaRed)			80				0.010	ND	< 0.01	ND	0.020	
St.George,	WG	11	99-	3	309	5	0.011	ND	< 0.01	< 0.01	0.021	050041
Ontario, 2005 (IdaRed)			108				0.015	ND	< 0.01	< 0.01	0.025	

	Form	Ap	plication	1	Total/	PHI,		Re	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total a	No.
GAP, Canada	WG		53- 105	3		7						
S-Paul d'Abbotsford,	WG	9	77- 80	3	237	7	< 0.01	ND	< 0.01	ND	< 0.02	050041
Quebec, 2005 (McIntosh)							< 0.01	ND	< 0.01	ND	< 0.02	
S-Paul d'Abbotsford,	WG	11	101- 107	3	314	7	< 0.01	ND	< 0.01	ND	< 0.02	050041
Quebec, 2005 (McIntosh)							< 0.01	ND	< 0.01	ND	< 0.02	
Rockland, Idaho, USA,	WG	8	78- 80	3	236	7	0.018	< 0.01	< 0.01	< 0.01	0.028	050041
2005 (McIntosh)							0.020	< 0.01	< 0.01	< 0.01	0.030	
Rockland, Idaho, USA,	WG	11	101- 103	3	306	7	0.028	< 0.01	< 0.01	< 0.01	0.038	050041
2005 (McIntosh)							0.019	< 0.01	< 0.01	< 0.01	0.029	

^a Total residue = XDE-175-J + XDE-175-L + ND-J + NF-J.

Supervised trials on apple in Australia

A total of 20 supervised field trials on apples and eight on pears were conducted at six sites in Australia during 2004/2005 and 2005/2006 season (Cowles, 2006; Reports 040114 and 050062).

In trials in the 2004/2005 season at one site in Australia, apple trees were treated with one or four applications of a SC formulation of spinetoram, using two treatment regimes. The first treatment schedule consisted of four applications of spinetoram formulation at 63, 49, 35 and 21 days before harvest. The second treatment schedule consisted of one application at 21 days before harvest. Treatments were made as high volume applications at rates of 2.5 g/hL, 3.75 g/hL, 5 g/hL and 7.5 g/hL of spinetoram, respectively.

During the 2005/2006 season, trials were conducted in five sites in Australia where apple plots, consisting of 1 to 6 trees per plot ranging from early flowering to full size maturing fruit, were treated with 4 or 7 applications of a WG formulation of spinetoram (250 g ai/L). Two treatment regimens were used. The first treatment schedule consisted of three applications at 5-day intervals over flowering followed by four applications at 14-day intervals commencing 63 days before harvest so that the last application was 21 days before harvest. The results show that these early season treatments had no cumulative effect on residues resulting from the 4 applications at 14-day interval sprays used later in the season. The second treatment schedule consisted of four applications at 14-day intervals commencing 63 days before commercial harvest so that the last application is 21 days before harvest. Treatments were made as foliar spray applications at rates of 5 g ai/hL and 7.5 g ai/hL of spinetoram.

Twenty-five whole apples or pears, weighing approximately 2–5 kg, were collected from each field at 0, 7, 14, 21, 28 and 35 days after the last application. Samples were collected by hand, packed in plastic bags, labelled, and transported frozen to the laboratory.

Spinetoram and the metabolites were analysed using the method GRM 05.03, with an LOQ 0.01 mg/kg. The LOD was determined to be 0.005 mg/kg.

Supervised trials on apple and pear in New Zealand

A total of 20 supervised field trials on apples and 8 on pears were carried out at 6 sites in New Zealand during the 2004/05 and 2005/06 seasons (Cowles, 2006; Reports 040114 and 050074). All the trials were decline trials.

Each field site consisted of one untreated and four treated plots, each with four trees (except for the trial on apple in 2004/2005, which had two trees). Each treated plot received one of four treatment rates of 2.5, 3.75, 5, and 7.5 g ai/hL (SC 100 or 120 g ai/L spinetoram) applied at intervals of 14–21 days. For the 2004/2005 trials, two application regimes were used; one using 4 applications at 60–63, 49, 36, and 21 days before harvest, and the other using a single application 21 days before harvest. The 2005/2006 trials used four applications. Mature apples were harvested 5–7 days after the last application. Applications were by hand-gun to run-off (1500–1800 L water/ha).

About 3–4 kg (12 fruits) of whole mature fruits were harvested by hand on 0, 1, 3, 7, 14, 28, and 35 or 42 days after the last application. Samples were placed in plastic bags, labelled, frozen and transported frozen to the laboratory, where they were kept in freezers at -18 °C until analysis.

Spinetoram and the metabolites were analysed using the method GRM 05.03, previously validated with an LOQ 0.01 mg/kg. The LOD was 0.005 mg/kg.

Table 24-1 Residues of spinetoram from supervised trials on apple in Australia and New Zealand (for estimation of maximum residue level)

	Form	A	pplication		Total/	PHI	Re	esidue, mg/l	kg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	day s	XDE- 175-J	XDE- 175-L	Total ^a	No.
GAP, New Zealand Pome fruits	WG	2.5		4		7				
AU	STRALIA	N TRIALS	S, 2004/200	5 SEAS	SON					
Shepparton	SC	2.5	37.5	4	150	0	0.051	< 0.01	0.051	040114
East, VIC, 2005						3	0.021	ND	0.021	
(Sundowner)						7	< 0.01	ND	< 0.01	
040114-01						14	< 0.01	ND	< 0.01	
						21	< 0.01	ND	< 0.01	
						28	ND	ND	< 0.01	
						35	ND	ND	< 0.01	
Shepparton	SC	3.75	56–57	4	225	0	0.120	0.023	0.143	040114
East, VIC, 2005						3	0.050	< 0.01	0.05	
(Sundowner)						8	0.020	ND	0.02	
040114-01						14	0.011	ND	0.011	
						21	< 0.01	ND	< 0.01	
						28	< 0.01	ND	< 0.01	
						36	ND	ND	< 0.01	
Shepparton	SC	5	75	4	300	0	0.120	0.026	0.146	040114
East, VIC, 2005						3	0.094	0.012	0.106	
(Sundowner)						8	0.042	ND	0.042	
040114-01						14	0.016	ND	0.016	
						21	< 0.01	ND	< 0.01	
						28	< 0.01	ND	< 0.01	
						36	< 0.01	ND	< 0.01	

	Form	Aj	pplication		Total/	PHI	Ro	esidue, mg/l	ζg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	, day	XDE- 175-J	XDE- 175-L	Total ^a	No.
						s	1 / J-J	173-L		
Shepparton	SC	7.5	112-	4	450	0	0.186	0.039	0.225	040114
East, VIC, 2005			113			3	0.101	0.017	0.118	
(Sundowner)						8	0.049	ND	0.049	
040114-01						14	0.024	ND	0.024	
						21	< 0.01	ND	< 0.01	
						28	0.024	ND	0.024	
						36	< 0.01	ND	< 0.01	
Shepparton	SC	2.5	37.5	1	37.5	0	0.030	< 0.01	0.03	040114
East, VIC, 2005						3	0.024	ND	0.024	
(Sundowner)						8	< 0.01	ND	< 0.01	
040114-01						14	ND	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
Shepparton	SC	3.75	56	1	56	0	0.053	0.012	0.065	040114
East, VIC, 2005						3	0.026	ND	0.026	
(Sundowner)						8	< 0.01	ND	< 0.01	
040114-01						14	ND	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
Shepparton	SC	5	75	1	75	0	0.095	0.022	0.117	040114
East, VIC, 2005						3	0.040	< 0.01	0.04	
(Sundowner)						8	0.017	ND	0.017	
040114-01						14	0.010	ND	0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
Shepparton	SC	7.5	112.5	1	112.5	0	0.123	0.031	0.154	040114
East, VIC, 2005						3	0.076	0.014	0.09	
(Sundowner)						8	0.014	ND	0.014	
040114-01						14	0.034	ND	0.034	
						21	< 0.01	ND	< 0.01	
						28	< 0.01	ND	< 0.01	
						36	ND	ND	< 0.01	
AUSTR	ALIAN T	RIALS, 20	005/2006 S	EASON	ſ					
Ardmona VIC,	SC	5	62	4	248	0	0.04	< 0.01	0.04	050062
2006 (Granny Smith)						7	< 0.01	ND	< 0.01	
050062-01						14	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						35	ND	ND	< 0.01	

	Form	Aj	pplication		Total/	PHI	Re	esidue, mg/	kg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	day s	XDE- 175-J	XDE- 175-L	Total ^a	No.
Ardmona VIC,	SC	7.5	93	4	372	0	0.06	0.02	0.08	050062
2006 (Granny Smith)						7	0.01	ND	0.01	
050062-01						14	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						35	ND	ND	< 0.01	
Poziers, QLD,	WG	5	186–	4	805	0	0.05	0.01	0.06	050062
2005-06 (Summer Del)			229			7	< 0.01	ND	< 0.01	
$050062-02^2$						28	ND	ND	< 0.01	
						35	ND	ND	< 0.01	
Poziers, QLD,	WG	7.5	278–	4	1207	0	0.04	0.02	0.06	050062
2005-06 (Summer Del)			344			7	0.03	ND	0.03	
$050062-02^2$						28	0.01	ND	0.01	
						35	ND	ND	< 0.01	
Spreyton,	WG	5	128–	4	551	0	0.09	0.01	0.10	050062
Tasmania, 2006 (Fuji)			149			7	0.03	ND	0.03	
050062-03						14	0.02	ND	0.02	
						28	< 0.01	ND	< 0.01	
						35	ND	ND	< 0.01	
Spreyton,	WG	7.5	188-	4	797	0	0.11	0.02	0.13	050062
Tasmania, 2006 (Fuji)			207			7	0.04	ND	0.04	
050062-03						14	0.02	ND	0.02	
						28	< 0.01	ND	< 0.01	
						35	ND	ND	< 0.01	
Ardmona VIC,	SC	5	62	7	434	0	0.06	0.02	0.08	050062
2006 (Granny Smith)						7	< 0.01	ND	< 0.01	
050062-01						14	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						35	ND	ND	< 0.01	
Ardmona VIC,	SC	7.5	93	7	651	0	0.04	0.01	0.05	050062
2006 (Granny Smith)						7	0.01	ND	0.01	
050062-01						14	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						35	ND	ND	< 0.01	
Poziers, QLD	WG	5	65-	7	848	0	ND	ND	< 0.01	050062
2005-06 b (Summer Del) 050062-02			229			35	ND	ND	< 0.01	
Poziers, QLD,	WG	7.5	98–	7	1549	0	ND	ND	< 0.01	050062
2005-06 b (Summer Del) 050062-02			344			35	ND	ND	< 0.01	

	Form	A	pplication		Total/	PHI	Re	esidue, mg/l	kg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	, day s	XDE- 175-J	XDE- 175-L	Total ^a	No.
Spreyton,	WG	5	70-	7	785	0	0.05	< 0.01	0.05	050062
Tasmania, 2006 (Fuji)			149			7	0.03	ND	0.03	
APPLE						14	0.02	ND	0.02	
050062-03						28	ND	ND	< 0.01	
						35	ND	ND	< 0.01	
Spreyton,	WG	7.5	101-	7	1017	0	0.11	0.02	0.13	050062
Tasmania, 2006 (Fuji)			207			7	0.03	ND	0.03	
050062-03						14	0.01	ND	0.01	
						28	0.01	ND	0.01	
						35	ND	ND	< 0.01	
NEW ZE	EALAND	TRIALS, 2	2004/2005	SEASO	N	•				
Hort Research,	SC	2.5	52–62	4	233	0	0.032	< 0.01	0.032	040114
2005 (Fuji)						3	< 0.01	ND	< 0.01	
040114-02						8	ND	ND	< 0.01	
						14	ND	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
Hort Research,	SC	3.75	69–83	4	307	0	0.050	0.016	0.066	040114
2005 (Fuji)						3	< 0.01	ND	< 0.01	
040114-02						8	< 0.01	ND	< 0.01	
						14	ND	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
Hort Research,	SC	5	122-	4	498	0	0.068	0.020	0.088	040114
2005 (Fuji)			130			3	0.012	ND	0.012	
040114-02						8	< 0.01	ND	< 0.01	
						14	ND	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
Hort Research,	SC	7.5	159–	4	681	0	0.075	0.023	0.098	040114
2005 (Fuji)			180			3	0.025	< 0.01	0.025	
040114-02						8	< 0.01	ND	< 0.01	
						14	< 0.01	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	

	Form	A	pplication		Total/	PHI	Re	esidue, mg/l	kg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	day s	XDE- 175-J	XDE- 175-L	Total ^a	No.
Hort Research,	SC	2.5	62	1	62	0	0.021	< 0.01	0.021	040114
2005 (Fuji)						3	ND	ND	< 0.01	
040114-02						8	ND	ND	< 0.01	
						14	ND	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
Hort Research,	SC	3.75	81	1	81	0	0.046	0.015	0.061	040114
2005 (Fuji)						3	0.015	ND	0.015	
040114-02						8	< 0.01	ND	< 0.01	
						14	ND	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
Hort Research,	SC	5	104	1	104	0	0.086	0.027	0.113	040114
2005 (Fuji)						3	0.016	ND	0.016	
040114-02						8	< 0.01	ND	< 0.01	
						14	ND	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
Hort Research,	SC	7.5	147	1	147	0	0.127	0.041	0.168	040114
2005 (Fuji)						3	0.026	< 0.01	0.026	
040114-02						8	0.015	ND	0.015	
						14	ND	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
NEW ZE	EALAND	TRIALS, 2	2005/2006	SEASO	N	,				
Twyford,	SC	2.5	38-39	4	154	0	0.015	ND	0.015	050074
Hawkes Bay, 2006						3	0.012	ND	0.012	
(Braeburn)						7	< 0.01	ND	< 0.01	
050074-01						14	ND	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
						42	ND	ND	< 0.01	

	Form	Al	pplication		Total/	PHI	Ro	esidue, mg/l	cg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	day s	XDE- 175-J	XDE- 175-L	Total ^a	No.
Twyford,	SC	3.72	55- 58	4	226	0	0.035	< 0.01	0.035	050074
Hawkes Bay, 2006						3	0.031	< 0.01	0.031	
(Braeburn) 050074-01						7	0.013	ND	0.013	
030074-01						14	0.011	ND	0.011	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
						42	ND	ND	< 0.01	
Twyford,	SC	5	75- 79	4	307	0	0.037	< 0.01	0.037	050074
Hawkes Bay, 2006						3	0.038	< 0.01	0.038	
(Braeburn) 050074-01						7	0.016	ND	0.016	
030074-01						14	0.011	ND	0.011	
						21	< 0.01	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
						42	ND	ND	< 0.01	
Twyford,	SC	7.6	112-	4	461	0	0.068	0.018	0.086	050074
Hawkes Bay, 2006			118			3	0.058	0.014	0.072	
(Braeburn) 050074-01						7	0.028	0.008	0.036	
030074-01						14	0.016	ND	0.016	
						21	< 0.01	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
						42	ND	ND	< 0.01	
Havelock	SC	2.5	36- 38	4	150	0	0.020	< 0.01	0.02	050074
North, Hawkes Bay, 2006						3	< 0.01	ND	< 0.01	
(Royal Gala) 050074-02						7	< 0.01	ND	< 0.01	
030074-02						14	ND	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
						42	ND	ND	< 0.01	

	Form	Aj	pplication		Total/	PHI	Re	esidue, mg/l	κg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	day s	XDE- 175-J	XDE- 175-L	Total ^a	No.
Havelock	SC	3.7	54- 56	4	222	0	0.023	< 0.01	0.023	050074
North, Hawkes Bay, 2006						3	< 0.01	ND	< 0.01	
(Royal Gala)						7	ND	ND	< 0.01	
050074-02						14	ND	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
						42	ND	ND	< 0.01	
Havelock	SC	5	73- 76	4	300	0	0.045	0.012	0.057	050074
North, Hawkes Bay, 2006						3	< 0.01	ND	< 0.01	
(Royal Gala)						7	< 0.01	ND	< 0.01	
050074-02						14	< 0.01	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
						42	ND	ND	< 0.01	
Havelock	SC	7.6	109-	4	450	0	0.074	0.019	0.093	050074
North, Hawkes Bay, 2006			115			3	0.015	ND	0.015	
(Royal Gala) 050074-02						7	< 0.01	ND	< 0.01	
030074-02						14	< 0.01	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
						42	ND	ND	< 0.01	
Riwaka,	SC	2.5	38- 40	4	154	0	0.017	ND	0.017	050074
Nelson, 2006 (Royal Gala)						3	ND	ND	< 0.01	
050074-04						7	ND	ND	< 0.01	
						14	ND	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
						42	ND	ND	< 0.01	
Riwaka,	SC	3.72	56- 59	4	228	0	0.016	ND	0.016	050074
Nelson, 2006 (Royal Gala)						3	< 0.01	ND	< 0.01	
050074-04						7	ND	ND	< 0.01	
						14	ND	ND	< 0.01	
						21	ND ND	ND	< 0.01	
						28 36	ND ND	ND ND	< 0.01 < 0.01	
						42	ND ND	ND	< 0.01	
		<u> </u>		<u> </u>		72	ND	ND	< 0.01	

	Form	Aj	pplication		Total/	PHI	Ro	esidue, mg/l	ζg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	day s	XDE- 175-J	XDE- 175-L	Total ^a	No.
Riwaka,	SC	5	76- 79	4	308	0	0.025	< 0.01	0.025	050074
Nelson, 2006 (Royal Gala)						3	ND	ND	< 0.01	
050074-04						7	< 0.01	ND	< 0.01	
						14	ND	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
						42	ND	ND	< 0.01	
Riwaka,	SC	7.6	113-	4	461	0	0.060	0.018	0.078	050074
Nelson, 2006 (Royal Gala)			119			3	0.012	ND	0.012	
050074-04						7	< 0.01	ND	< 0.01	
						14	< 0.01	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
						42	ND	ND	< 0.01	

^a Total residue = XDE-175-J + XDE-175-L.

Table 24-2 Residues of spinetoram and metabolites from supervised trials on apple in Australia and New Zealand (for estimation of STMR).

	Form	Ap	plication	1	Total/	PHI,		Re	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total a	No.
GAP, New Zealand Pome fruits	WG	2.5		4		7						
A	USTRA	LIAN TR	RIALS, 2	004/20	005 SEASC	N						
Shepparton	SC	2.5	37.5	4	150	0	0.051	< 0.01	-	ND	0.061	040114
East, VIC, 2005						3	0.021	ND	-	ND	0.031	
(Sundowner)						7	< 0.01	ND	-	ND	< 0.02	
040114-01						14	< 0.01	ND	-	ND	< 0.02	
						21	< 0.01	ND	-	ND	< 0.02	
						28	ND	ND	-	ND	< 0.02	
						35	ND	ND	-	ND	< 0.02	
Shepparton	SC	3.75	56-	4	225	0	0.120	0.023	-	< 0.01	0.153	040114
East, VIC, 2005			57			3	0.050	< 0.01	-	< 0.01	0.060	
(Sundowner)						8	0.020	ND	-	ND	0.030	
040114-01						14	0.011	ND	-	ND	0.021	
						21	< 0.01	ND	-	ND	< 0.02	
						28	< 0.01	ND	-	ND	< 0.02	

^b Samples on Days 0, 7, and 28 are from replacement trials in Cottonvale, QLD, referred to as 050062-06 in the study.

	Form	Ap	plication	l	Total/	PHI,		Re	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total	No.
						36	ND	ND	-	ND	< 0.02	
Shepparton	SC	5	75	4	300	0	0.120	0.026	-	< 0.01	0.156	040114
East, VIC, 2005						3	0.094	0.012	-	< 0.01	0.116	-
(Sundowner)						8	0.042	ND	-	< 0.01	0.052	
040114-01						14	0.016	ND	-	ND	0.026	
						21	< 0.01	ND	-	ND	< 0.02	
						28	< 0.01	ND	-	ND	< 0.02	
						36	< 0.01	ND	-	ND	< 0.02	
Shepparton	SC	7.5	112-	4	450	0	0.186	0.039	-	0.011	0.236	040114
East, VIC, 2005			113			3	0.101	0.017	-	0.012	0.130	
(Sundowner)						8	0.049	ND	-	< 0.01	0.059	
040114-01						14	0.024	ND	-	< 0.01	0.034	
						21	< 0.01	ND	-	ND	< 0.02	
						28	0.024	ND	-	< 0.01	0.034	
						36	< 0.01	ND	-	ND	< 0.02	
Shepparton	SC	2.5	37.5	1	37.5	0	0.030	< 0.01	-	ND	0.04	040114
East, VIC, 2005						3	0.024	ND	-	ND	0.034	-
(Sundowner)						8	< 0.01	ND	-	ND	< 0.02	
040114-01						14	ND	ND	-	ND	< 0.02	-
						21	ND	ND	-	ND	< 0.02	-
						28	ND	ND	-	ND	< 0.02	-
						36	ND	ND	-	ND	< 0.02	
Shepparton East, VIC,	SC	3.75	56	1	56	0	0.053	0.012	-	ND	0.075	040114
2005						3	0.026	ND	-	ND	0.036	-
(Sundowner) 040114-01						8	< 0.01	ND	-	ND	< 0.02	1
040114-01						14	ND	ND	-	ND	< 0.02	-
						21	ND	ND	-	ND	< 0.02	-
						28	ND	ND	-	ND	< 0.02	-
Chamantan	CC	_	75	1	75	36	ND 0.005	ND	-	ND	< 0.02	040114
Shepparton East, VIC,	SC	5	/3	1	75	0	0.095	0.022	-	ND	0.127	040114
2005						8	0.040	< 0.01 ND	-	ND ND	0.050	
(Sundowner) 040114-01						14	0.017	ND	-	ND	0.027	-
						21	ND	ND ND		ND	< 0.020	
						28	ND	ND	-	ND	< 0.02	-
						36	ND	ND	-	ND	< 0.02	-
Shepparton	SC	7.5	112.5	1	112.5	0	0.123	0.031	-	ND	0.164	040114
East, VIC,	50	1.5	112.3	1	112.3	3	0.123	0.031	-	< 0.01	0.104	0.70114
2005						8	0.076	ND	-	ND	0.100	-
(Sundowner) 040114-01						14	0.014	ND	-	ND	0.024	-
						21	< 0.034	ND	-	ND	< 0.02	1
						28	< 0.01	ND	-	ND	< 0.02	-
						36	ND	ND	-	ND	< 0.02	1
L	L	<u> </u>		<u> </u>		50	עויו	עויו		עא	< 0.0∠	

	Form	Ap	plication	l	Total/	PHI,		Re	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total a	No.
AUSTRA	LIAN TR	IALS, 2	005/2006	SEAS	SON	I						
Ardmona	SC	5	62	4	248	0	0.04	< 0.01	ND	ND	0.05	050062
VIC, 2006 (Granny						7	< 0.01	ND	ND	ND	< 0.02	
Smith)						14	ND	ND	ND	ND	< 0.02	
050062-01						28	ND	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
Ardmona	SC	7.5	93	4	372	0	0.06	0.02	ND	ND	0.09	050062
VIC, 2006 (Granny						7	0.01	ND	< 0.01	ND	0.02	
Smith)						14	ND	ND	ND	ND	< 0.02	
050062-01						28	ND	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
Poziers,	WG	5	186-	4	805	0	0.05	0.01	< 0.01	ND	0.07	050062
QLD, 2005- 06			229			7	< 0.01	ND	0.01	< 0.01	0.03	
(Summer						28	ND	ND	< 0.01	< 0.01	< 0.02	
Del) 050062-02 ^b						35	ND	ND	ND	ND	< 0.02	
Poziers,	WG	7.5	278-	4	1207	0	0.04	0.02	< 0.01	< 0.01	0.07	050062
QLD, 2005- 06			344			7	0.03	ND	0.03	0.01	0.07	
(Summer						28	0.01	ND	0.02	0.01	0.04	
Del) 050062-02 ^b						35	ND	ND	< 0.01	< 0.01	< 0.02	
Spreyton,	WG	5	128-	4	551	0	0.09	0.01	< 0.01	< 0.01	0.11	050062
Tasmania, 2006			149			7	0.03	ND	< 0.01	< 0.01	0.04	
(Fuji)						14	0.02	ND	ND	ND	0.03	
050062-03						28	< 0.01	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
Spreyton,	WG	7.5	188-	4	797	0	0.11	0.02	< 0.01	< 0.01	0.14	050062
Tasmania, 2006			207			7	0.04	ND	0.010	< 0.01	0.06	
(Fuji)						14	0.02	ND	ND	ND	0.03	
050062-03						28	< 0.01	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
Ardmona	SC	5	62	7	434	0	0.06	0.02	< 0.01	ND	0.09	050062
VIC, 2006 (Granny						7	< 0.01	ND	ND	ND	< 0.02	
Smith)						14	ND	ND	ND	ND	< 0.02	
050062-01						28	ND	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
Ardmona	SC	7.5	93	7	651	0	0.04	0.01	ND	ND	0.06	050062
VIC, 2006 (Granny						7	0.01	ND	< 0.01	ND	0.02	
Smith)						14	ND	ND	ND	ND	< 0.02	
050062-01						28	ND	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	

	Form	Aŗ	plication	1	Total/	PHI,		Re	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total	No.
Poziers,	WG	5	65-	7	848	0	ND	ND	ND	< 0.01	< 0.02	050062
QLD 2005-06 (Summer Del) 050062-02			229			35	ND	ND	ND	ND	< 0.02	
Poziers, QLD, 2005-	WG	7.5	98– 344	7	1549	0	ND	ND	ND	< 0.01	< 0.02	050062
06 (Summer Del) 050062-02			311			35	ND	ND	ND	< 0.01	< 0.02	
Spreyton,	WG	5	70–	7	785	0	0.05	< 0.01	ND	ND	0.06	050062
Tasmania, 2006			149			7	0.03	ND	< 0.01	ND	0.04	
(Fuji)						14	0.02	ND	ND	ND	0.03	
APPLE 050062-03						28	ND	ND	ND	ND	< 0.02	
030002-03						35	ND	ND	ND	ND	< 0.02	
Spreyton,	WG	7.5	101-	7	1017	0	0.11	0.02	0.01	< 0.01	0.15	050062
Tasmania, 2006			207			7	0.03	ND	< 0.01	< 0.01	0.04	
(Fuji)						14	0.01	ND	ND	ND	0.02	
050062-03						28	0.01	ND	ND	ND	0.02	
						35	ND	ND	ND	ND	< 0.02	
NEW ZEA	LAND T	RIALS,	2004/200	5 SEA	SON							
Hort	SC	2.5	52-	4	233	0	0.032	< 0.01	-	< 0.01	0.042	040114
Research, 2005			62			3	< 0.01	ND	-	< 0.01	< 0.02	
(Fuji)						8	ND	ND	-	ND	< 0.02	
040114-02						14	ND	ND	-	ND	< 0.02	
						21	ND	ND	-	ND	< 0.02	
						28	ND	ND	-	ND	< 0.02	
						36	ND	ND	-	ND	< 0.02	
Hort Research,	SC	3.75	69– 83	4	307	0	0.050	0.016	-	< 0.01	0.076	040114
2005			0.5			3	< 0.01	ND	-	< 0.01	< 0.02	
(Fuji) 040114-02						8	< 0.01	ND	-	< 0.01	< 0.02	
040114-02						14	ND	ND	-	< 0.01	< 0.02	
						21	ND	ND	-	ND	< 0.02	
						28	ND	ND	-	ND	< 0.02	
***	0.0		122		400	36	ND	ND	-	ND	< 0.02	0.4011.4
Hort Research,	SC	5	122– 130	4	498	0	0.068	0.020	-	< 0.01	0.098	040114
2005						3	0.012	ND	-	< 0.01	0.022	
(Fuji) 040114-02						8	< 0.01	ND	-	< 0.01	< 0.02	
0.511102						14	ND	ND	-	< 0.01	< 0.02	
						21	ND	ND	-	ND	< 0.02	
						28	ND	ND	-	ND	< 0.02	
						36	ND	ND	-	ND	< 0.02	

Decision Season		Form	Ap	plication	l	Total/	PHI,		Re	sidue, mg	/kg		Report
Research, 2005 (Fuji) O40114-02 Part	year		g ai/hL	g ai/ha	No		days			ND-J	NF-J		No.
2005 (Fuji) 040114-02		SC	7.5		4	681	0	0.075	0.023	-	< 0.01	0.108	040114
GFuji) 040114-02 Horizontal Price Horizonta				180			3	0.025	< 0.01	-	< 0.01	0.035	
Hort Research, 2005 (Fuji) 040114-02 ND SC SC SC SC SC SC SC S	(Fuji)						8	< 0.01	ND	-	< 0.01	< 0.02	
Hort Research, 2005 (Fuji) 040114-02	040114-02						14	< 0.01	ND	-	< 0.01	< 0.02	
Hort Research, 2005 (Fuji) O40114-02 Part P							21	ND	ND	-	ND	< 0.02	
Hort Research, 2005 (Fuji) O40114-02 Hort Research, 2005 (Fuji) O40114							28	ND	ND	-	ND	< 0.02	
Research 2005							36	ND	ND	-	ND	< 0.02	
2005 (Fuji) 040114-02 140 14		SC	2.5	62	1	62	0	0.021	< 0.01	-	ND	0.031	040114
Hort Research, 2005 (Fuji) 040114-02 SC 7.5								ND	ND	-	ND	< 0.02	
Hort SC S 104 1 104 100 10	(Fuji)						8	ND	ND	-	< 0.01	< 0.02	
Hort Research, 2005 (Fuji) O40114-02 Hort Research, 2005 (Fuji) O40114	040114-02							ND		-	ND		
Hort Research, 2005 (Fuji) O40114-02 Hort Research, 2005 (Fuji) O40114							21	ND		-	ND		
Hort Research, 2005 (Fuji) O40114-02 Hort Research, 2005 (Fuji) O40114										-			
Research, 2005										-			
2005 (Fuji) 040114-02 Hort SC 5 104 1 104 10 104 14 10 10		SC	3.75	81	1	81	0	0.046	0.015	-	ND	0.071	040114
14	2005						3	0.015	ND	-	< 0.01	0.025	
Hort Research, 2005 (Fuji) O40114-02 Hort Research, 2005 (Fuji) O40114							8	< 0.01	ND	-	ND	< 0.02	
Hort SC 5 104 1 104 0 0.086 0.027 - ND < 0.02	040114-02						14	ND	ND	-	ND	< 0.02	
Hort SC 5 104 1 104 0 0.086 0.027 - ND 0.123 040114							21	ND	ND	-	ND	< 0.02	
Hort Research, 2005 (Fuji) 040114-02 Hort Research, 2005 (Fujii)							28	ND	ND	-	ND	< 0.02	
Research, 2005 (Fuji) 040114-02							36	ND	ND	-	ND	< 0.02	
Solution Solution		SC	5	104	1	104	0	0.086	0.027	-	ND	0.123	040114
Research, 2005 (Fuji) 040114-02							3	0.016	ND	-	< 0.01	0.026	
Hort SC 7.5 147 1 147 0 0.127 0.041 - ND 0.02	(Fuji)						8	< 0.01	ND	-	< 0.01	< 0.02	
Hort Research, 2005 (Fuji) 040114-02 NEW ZEALAND TRIALS, 2005/2006 SEASON Twyford, Hawkes Bay, 2006 (Braeburn) 050074-01 NEW ZEALAND TRIALS, 2005/2006 SEASON SC 3.5 3.6 3.0 1.5	040114-02						14	ND	ND	-	ND	< 0.02	
Hort SC 7.5 147 1 147 0 0.127 0.041 - ND 0.178 040114							21	ND	ND	-	ND	< 0.02	
Hort Research, 2005 2005 (Fuji) 040114-02 SC 7.5 147 1 147 0 0.127 0.041 - ND 0.178 040114							28	ND	ND	-	ND	< 0.02	
Research, 2005 (Fuji) 040114-02							36	ND	ND	-	ND	< 0.02	
2005 (Fuji) 040114-02		SC	7.5	147	1	147				-			040114
14 ND ND - ND < 0.02 21 ND ND - ND < 0.02 28 ND ND - ND < 0.02 28 ND ND - ND < 0.02 36 ND ND - ND < 0.02 37 ND < 0.01 ND - ND < 0.02 37 ND < 0.01 ND - ND < 0.02 37 ND < 0.02 37 ND < 0.02 37 ND < 0.01 37 ND < 0.02 37 ND <										-	< 0.01		
ND ND ND ND ND ND ND ND										-			
28 ND ND - ND < 0.02	040114-02									-			
36 ND ND - ND < 0.02													
NEW ZEALAND TRIALS, 2005/2006 SEASON													
Twyford, Hawkes Bay, 2006 (Braeburn) 050074-01 SC 2.5 38- 4 154 0 0.015 ND - ND 0.025 050074					<u> </u>	2017	36	ND	ND	-	ND	< 0.02	
Hawkes Bay, 2006 (Braeburn) 39 3 0.012 ND - ND 0.022		1					0	0.015	ND	_	ND	0.025	050074
2006 (Braeburn) 050074-01 7 < 0.01 ND - ND < 0.02 14 ND ND - < 0.01 < 0.02	Hawkes Bay,	30	2.3		-	134							030074
050074-01 14 ND ND - < 0.01 < 0.02	2006												
17 10 10 1001 0.02							-						
28 ND ND - ND < 0.02													

	Form	Ap	plication	1	Total/	PHI,		Re	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total	No.
						36	ND	ND	-	ND	< 0.02	
						42	ND	ND	-	ND	< 0.02	
Twyford,	SC	5	75-	4	307	0	0.037	< 0.01	-	< 0.01	0.047	050074
Hawkes Bay, 2006			79			3	0.038	< 0.01	-	< 0.01	0.048	
(Braeburn)						7	0.016	ND	-	0.010	0.026	
050074-01						14	0.011	ND	-	< 0.01	0.021	
						21	< 0.01	ND	-	< 0.01	< 0.02	
						28	ND	ND	-	ND	< 0.02	
						36	ND	ND	-	ND	< 0.02	
						42	ND	ND	-	0.008	0.018	
Twyford, Hawkes Bay,	SC	3.72	55– 58	4	226	0	0.035	< 0.01	-	< 0.01	0.045	050074
2006			30			3	0.031	< 0.01	-	ND	0.041	
(Braeburn) 050074-01						7	0.013	ND	-	< 0.01	0.023	
030074-01						14	0.011	ND	-	< 0.01	0.021	
						21	ND	ND	-	ND	< 0.02	
						28	ND	ND	-	ND	< 0.02	
						36	ND	ND	-	< 0.01	< 0.02	
						42	ND	ND	-	ND	< 0.02	
Twyford,	SC	7.6	112-	4	461	0	0.068	0.018	-	< 0.01	0.096	050074
Hawkes Bay, 2006			118			3	0.058	0.014	-	0.013	0.085	
(Braeburn)						7	0.028	0.008	-	0.012	0.048	
050074-01						14	0.016	ND	-	0.011	0.027	
						21	< 0.01	ND	-	< 0.01	< 0.02	
						28	ND	ND	-	< 0.01	< 0.02	
						36	ND	ND	-	ND	< 0.02	
						42	ND	ND	-	ND	< 0.02	
Havelock	SC	2.5	36-	4	150	0	0.020	< 0.01	-	ND	0.030	050074
North, Hawkes Bay,			38			3	< 0.01	ND	-	< 0.01	< 0.02	
2006						7	< 0.01	ND	-	< 0.01	< 0.02	
(Royal Gala) 050074-02						14	ND	ND	-	ND	< 0.02	
000071.02						21	ND	ND	-	ND	< 0.02	
						28	ND	ND	-	ND	< 0.02	
						36	ND	ND	-	ND	< 0.02	
						42	ND	ND	_	ND	< 0.02	
Havelock	SC	3.7	54-	4	222	0	0.023	< 0.01	-	< 0.01	0.033	050074
North,			56			3	< 0.01	ND	-	< 0.01	< 0.02	
Hawkes Bay, 2006						7	ND	ND	_	< 0.01	< 0.02	
(Royal Gala) 050074-02						14	ND	ND	_	< 0.01	< 0.02	
030074-02						21	ND	ND	_	ND	< 0.02	
						28	ND	ND		ND	< 0.02	
						28	מא	מא	-	מא	< 0.02	

	Form	Ap	plication	1	Total/	PHI,		Re	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total	No.
						36	ND	ND	-	ND	< 0.02	
						42	ND	ND	-	ND	< 0.02	
Havelock	SC	5	73–	4	300	0	0.045	0.012	-	< 0.01	0.067	050074
North, Hawkes Bay,			76			3	< 0.01	ND	-	< 0.01	< 0.02	
2006						7	< 0.01	ND	-	< 0.01	< 0.02	
(Royal Gala) 050074-02						14	< 0.01	ND	-	< 0.01	< 0.02	
						21	ND	ND	-	ND	< 0.02	
						28	ND	ND	-	ND	< 0.02	
						36	ND	ND	-	ND	< 0.02	
						42	ND	ND	-	ND	< 0.02	
Havelock North,	SC	7.6	109– 115	4	450	0	0.074	0.019	-	0.012	0.105	050074
Hawkes Bay,			113			3	0.015	ND	-	< 0.01	0.025	
2006 (Royal Gala)						7	< 0.01	ND	-	< 0.01	< 0.02	
050074-02						14	< 0.01	ND	-	< 0.01	< 0.02	
						21	ND	ND	-	< 0.01	< 0.02	
						28	ND	ND	-	< 0.01	< 0.02	
						36	ND	ND	-	< 0.01	< 0.02	
						42	ND	ND	-	ND	< 0.02	
Riwaka,	SC	2.5	38-	4	154	0	0.017	ND	-	< 0.01	0.027	050074
Nelson, 2006 (Royal Gala)			40			3	ND	ND	-	ND	< 0.02	
050074-04						7	ND	ND	-	ND	< 0.02	
						14	ND	ND	-	ND	< 0.02	
						21	ND	ND	-	ND	< 0.02	
						28	ND	ND	-	ND	< 0.02	
						36	ND	ND	-	ND	< 0.02	
						42	ND	ND	-	ND	< 0.02	
Riwaka,	SC	3.72	56– 59	4	228	0	0.016	ND	-	< 0.01	0.026	050074
Nelson, 2006 (Royal Gala)			39			3	< 0.01	ND	-	ND	< 0.02	
050074-04						7	ND	ND	-	ND	< 0.02	
						14	ND	ND	-	ND	< 0.02	
						21	ND	ND	-	ND	< 0.02	
						28	ND	ND	-	ND	< 0.02	
						36	ND	ND	-	ND	< 0.02	
						42	ND	ND	-	ND	< 0.02	
Riwaka,	SC	5	76-	4	308	0	0.025	< 0.01	-	ND	0.035	050074
Nelson, 2006 (Royal Gala)			79			3	ND	ND	-	ND	< 0.02	
050074-04						7	< 0.01	ND	-	< 0.01	< 0.02	
						14	ND	ND	-	ND	< 0.02	
						21	ND	ND	-	ND	< 0.02	

	Form	Ap	plication	l	Total/	PHI,		Re	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total a	No.
						28	ND	ND	-	ND	< 0.02	
						36	ND	ND	-	ND	< 0.02	
						42	ND	ND	-	ND	< 0.02	
Riwaka,	SC	7.6	113-	4	461	0	0.060	0.018	-	< 0.01	0.088	050074
Nelson, 2006 (Royal Gala)			119			3	0.012	ND	-	< 0.01	0.022	
050074-04						7	< 0.01	ND	-	< 0.01	< 0.02	
						14	< 0.01	ND	-	< 0.01	< 0.02	
						21	ND	ND	-	ND	< 0.02	
						28	ND	ND	-	ND	< 0.02	
						36	ND	ND	-	ND	< 0.02	
						42	ND	ND	-	ND	< 0.02	

^a Total residue = XDE-175-J + XDE-175-L + ND-J + NF-J.

Table 25-1 Residues of spinetoram from supervised trials on pear in Australia and New Zealand (for estimation of maximum residue level)

	Form	A	pplication		Total/	PHI,	R	esidue, mg/l	κg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	Total ^a	No.
GAP, New Zealand Pome fruits	WG	2.5		4	200	7				
AUST	RALIAN	TRIALS, 2	005/2006 S	EASON						
Ardmona VIC,	WG	5	74	4	296	0	0.05	0.01	0.06	050062
2006 (Packham)						7	0.01	ND	0.01	
050062-04						14	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						35	ND	ND	< 0.01	
Ardmona VIC,	WG	7.5	111	4	444	0	0.11	0.02	0.13	050062
2006 (Packham)						7	0.03	ND	0.03	
050062-04						14	0.02	ND	0.02	
						28	< 0.01	ND	< 0.01	
						35	ND	ND	< 0.01	
Paracombe,	WG	5	97–105	4	404	0	0.02	ND	0.02	050062
SA, 2006 (Packham)						7	ND	ND	< 0.01	
050062-05						14	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						35	ND	ND	< 0.01	
Paracombe,	WG	7.5	145-	4	606	0	0.03	ND	0.03	050062
SA, 2006 (Packham)			158			7	< 0.01	ND	< 0.01	
050062-05						14	< 0.01	ND	< 0.01	
						28	ND	ND	< 0.01	

^b Samples on Days 0, 7, and 28 are from replacement trials in Cottonvale, QLD, referred to as 050062-06 in the study.

	Form	A	pplication		Total/	PHI,	R	esidue, mg/l	cg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	Total ^a	No.
					1	35	ND	ND	< 0.01	
Ardmona VIC,	WG	5	73–74	7	517	0	0.04	< 0.01	0.04	050062
2006 (Packham)						7	0.02	ND	0.02	
050062-04						14	0.01	ND	0.01	
						28	ND	ND	< 0.01	
						35	ND	ND	< 0.01	
Ardmona VIC, 2006	WG	7.5	102– 111	7	768	0	0.09	0.02	0.11	050062
(Packham)			111			7	0.02	ND	0.02	
050062-04						14	< 0.01	ND	< 0.01	
						28	ND	ND	< 0.01	
						35	ND	ND	< 0.01	
Paracombe, SA, 2006	WG	5	91–105	7	677	0	0.01	ND	0.01	050062
(Packham)						7	ND	ND	< 0.01	
050062-05						14	ND	ND	< 0.01	
						35	ND ND	ND ND	< 0.01	
Paracombe,	WG	7.5	136–	7	1014	0	0.02	ND	0.02	050062
SA, 2006	,,,,	7.0	158	,	1011	7	ND	ND	< 0.01	030002
(Packham) 050062-05						14	ND	ND	< 0.01	
00002 00						28	ND	ND	< 0.01	
						35	ND	ND	< 0.01	
NEW Z	EALAND	TRIALS,	1 2005/2006 S	I SEASO!	<u> </u> N	33	112	T,D	(0.01	
Hastings,	SC	2.5	46–47	4	187	0	0.011	ND	0.011	050074
Hawkes Bay, 2006						3	< 0.01	ND	< 0.01	
(Packham)						7	ND	ND	< 0.01	
050074-03						14	ND	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
						42	ND	ND	< 0.01	
Hastings,	SC	3.72	68–70	4	275	0	0.021	< 0.01	0.021	050074
Hawkes Bay, 2006						3	ND	ND	< 0.01	
(Packham) 050074-03						7	ND	ND	< 0.01	
030074-03						14	ND	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
						42	ND	ND	< 0.01	

	Form	A	pplication		Total/	PHI,	Re	esidue, mg/l	κg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	Total ^a	No.
Hastings,	SC	5	92–95	4	373	0	0.025	< 0.01	0.025	050074
Hawkes Bay, 2006						3	0.016	ND	0.016	
(Packham)						7	< 0.01	ND	< 0.01	
050074-03						14	< 0.01	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
						42	ND	ND	< 0.01	
Hastings,	SC	7.6	139–	4	561	0	0.037	0.011	0.048	050074
Hawkes Bay, 2006			142			3	0.027	ND	0.027	
(Packham)						7	ND	ND	< 0.01	
050074-03						14	< 0.01	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						36	ND	ND	< 0.01	
						42	ND	ND	< 0.01	
Upper	SC	2.5	38–41	4	154	0	0.032	ND	0.032	050074
Moutere, Nelson, 2006						3	0.026	ND	0.026	
(Taylor's						7	0.015	ND	0.015	
Gold) 050074-05						14	0.015	ND	0.015	
						21	< 0.01	ND	< 0.01	
						28	< 0.01	ND	< 0.01	
						36	ND	ND	< 0.01	
						42	ND	ND	< 0.01	
Upper	SC	3.72	56–60	4	233	0	0.043	< 0.01	0.043	050074
Moutere, Nelson, 2006						3	0.053	< 0.01	0.053	
(Taylor's						7	0.041	ND	0.041	
Gold) 050074-05						14	0.023	ND	0.023	
						21	0.018	ND	0.018	
						28	0.017	ND	0.017	
						36	0.012	ND	0.012	
						42	< 0.01	ND	< 0.01	
Upper	SC	5	76–81	4	315	0	0.064	< 0.01	0.064	050074
Moutere, Nelson, 2006						3	0.062	0.011	0.073	
(Taylor's						7	0.035	ND	0.035	
Gold) 050074-05						14	0.041	< 0.01	0.041	
						21	0.027	ND	0.027	
						28	0.019	ND	0.019	
						36	0.016	ND ND	0.016	
			<u> </u>			42	< 0.01	ND	< 0.01	

	Form	Aj	pplication		Total/	PHI,	Re	esidue, mg/l	ζg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	Total ^a	No.
Upper	SC	7.6	115-	4	472	0	0.096	0.017	0.113	050074
Moutere, Nelson, 2006			121			3	0.066	0.010	0.076	
(Taylor's Gold)						7	0.056	< 0.01	0.056	
050074-05						14	0.037	ND	0.037	
						21	0.049	< 0.01	0.049	
						28	0.027	ND	0.027	
						36	0.013	ND	0.013	
						42	0.010	ND	0.010	

^a Total residue = XDE-175-J + XDE-175-L.

Table 25-2 Residues of spinetoram and metabolites from supervised trials on pear in Australia and New Zealand (for estimation of STMR)

	Form	Ap	plication	1	Total/	PHI,		Re	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J a	NF-J	Total b	No.
GAP, New Zealand Pome fruit	WG	2.5		4	200	7						
AUSTRAI	JAN TR	IALS, 20	005/2006	SEAS	SON							
Ardmona	WG	5	74	4	296	0	0.05	0.01	ND	ND	0.07	050062
VIC, 2006 (Packham)						7	0.01	ND	ND	ND	0.02	
050062-04						14	ND	ND	ND	ND	< 0.02	
						28	ND	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
Ardmona	WG	7.5	111	4	444	0	0.11	0.02	< 0.01	ND	0.14	050062
VIC, 2006 (Packham)						7	0.03	ND	< 0.01	ND	0.04	
050062-04						14	0.02	ND	< 0.01	ND	0.03	
						28	< 0.01	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
Paracombe,	WG	5	97–	4	404	0	0.02	ND	ND	ND	0.03	050062
SA, 2006 (Packham)			105			7	ND	ND	ND	ND	< 0.02	
050062-05						14	ND	ND	ND	ND	< 0.02	
						28	ND	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
Paracombe,	WG	7.5	145-	4	606	0	0.03	ND	ND	ND	0.04	050062
SA, 2006 (Packham)			158			7	< 0.01	ND	ND	ND	< 0.02	
050062-05						14	< 0.01	ND	ND	ND	< 0.02	
						28	ND	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	

	Form				Total/	PHI,		Re	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J a	NF-J	Total b	No.
Ardmona	WG	5	73-	7	517	0	0.04	< 0.01	ND	ND	0.05	050062
VIC, 2006 (Packham)			74			7	0.02	ND	ND	ND	0.03	
050062-04						14	0.01	ND	ND	ND	0.02	
						28	ND	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
Ardmona	WG	7.5	102-	7	768	0	0.09	0.02	ND	< 0.01	0.12	050062
VIC, 2006 (Packham)			111			7	0.02	ND	ND	< 0.01	0.03	
050062-04						14	< 0.01	ND	ND	ND	< 0.02	
						28	ND	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
Paracombe, SA, 2006	WG	5	91– 105	7	677	0	0.01	ND	ND	ND	0.02	050062
(Packham)			103			7	ND	ND	ND	ND	< 0.02	
050062-05						14	ND	ND	ND	ND	< 0.02	
						28	ND	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
Paracombe,	WG	7.5	136– 158	7	1014	0	0.02	ND	ND	ND	0.03	050062
SA, 2006 (Packham)			138			7	ND	ND	ND	ND	< 0.02	
050062-05						14	ND	ND	ND	ND	< 0.02	
						28	ND	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
NEW ZEAI	LAND TI	RIALS, 2	2005/200	6 SEA	SON	•	•	•	•		•	•
Hastings,	SC	2.5	46–	4	187	0	0.011	ND	-	ND	0.021	050074
Hawkes Bay, 2006			47			3	< 0.01	ND	-	ND	< 0.02	
(Packham)						7	ND	ND	-	ND	< 0.02	
050074-03						14	ND	ND	-	ND	< 0.02	
						21	ND	ND	-	ND	< 0.02	
						28	ND	ND	-	ND	< 0.02	
						36	ND	ND	-	ND	< 0.02	
						42	ND	ND	_	ND	< 0.02	
Hastings,	SC	3.72	68-	4	275	0	0.021	< 0.01	-	ND	0.031	050074
Hawkes Bay,			70			3	ND	ND	_	ND	< 0.02	
2006 (Packham)						7	ND	ND	_	ND	< 0.02	
050074-03						14	ND	ND	-	ND	< 0.02	-
						21						-
							ND	ND	-	ND	< 0.02	-
						28	ND	ND	-	ND	< 0.02	-
						36	ND	ND	-	ND	< 0.02	
						42	ND	ND	-	ND	< 0.02	

	Form	Ap	plication	1	Total/	PHI,		Re	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J a	NF-J	Total b	No.
Hastings,	SC	5	92-	4	373	0	0.025	< 0.01	-	ND	0.035	050074
Hawkes Bay, 2006			95			3	0.016	ND	-	ND	0.026	
(Packham) 050074-03						7	< 0.01	ND	-	ND	< 0.02	
030074-03						14	< 0.01	ND	-	< 0.01	< 0.02	
						21	ND	ND	-	ND	< 0.02	
						28	ND	ND	-	ND	< 0.02	
						36	ND	ND	-	ND	< 0.02	
						42	ND	ND	-	ND	< 0.02	
Hastings,	SC	7.6	139-	4	561	0	0.037	0.011	-	ND	0.058	050074
Hawkes Bay, 2006			142			3	0.027	ND	-	< 0.01	0.037	
(Packham) 050074-03						7	ND	ND	-	ND	< 0.02	
030074-03						14	< 0.01	ND	-	ND	< 0.02	
						21	ND	ND	-	ND	< 0.02	
						28	ND	ND	-	ND	< 0.02	
						36	ND	ND	-	ND	< 0.02	
						42	ND	ND	-	ND	< 0.02	
Upper Moutere,	SC	2.5	38– 41	4	154	0	0.032	ND	-	ND	0.042	050074
Nelson, 2006			41			3	0.026	ND	-	ND	0.036	
(Taylor's Gold)						7	0.015	ND	-	ND	0.025	
050074-05						14	0.015	ND	-	ND	0.025	
						21	< 0.01	ND	-	ND	< 0.02	
						28	< 0.01	ND	-	ND	< 0.02	
						36	ND	ND	-	ND	< 0.02	
						42	ND	ND	-	ND	< 0.02	
Upper Moutere,	SC	3.72	56– 60	4	233	0	0.043	< 0.01	-	ND	0.053	050074
Nelson, 2006						3	0.053	< 0.01	-	ND	0.063	
(Taylor's Gold)						7	0.041	ND	-	ND	0.051	
050074-05						21	0.023	ND ND	-	ND ND	0.033	
						28	0.017	ND	-	ND	0.028	
						36	0.017	ND	_	ND	0.027	
						42	< 0.01	ND	-	ND	< 0.02	
Upper	SC	5	76–	4	315	0	0.064	< 0.01	-	ND	0.074	050074
Moutere, Nelson, 2006			81			3	0.062	0.011	-	ND	0.083	
(Taylor's						7	0.035	ND	-	ND	0.045	
Gold) 050074-05						14	0.041	< 0.01	-	ND	0.051	
						21	0.027	ND	-	ND	0.037	
						28	0.019	ND	-	ND	0.029	
						36	0.016	ND	-	ND	0.026	
						42	< 0.01	ND	-	ND	< 0.02	

	Form	Ap	plication	n	Total/	PHI,		Re	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J a	NF-J	Total b	No.
Upper	SC	7.6	115-	4	472	0	0.096	0.017	-	ND	0.123	050074
Moutere, Nelson, 2006			121			3	0.066	0.010	-	ND	0.086	
(Taylor's Gold)						7	0.056	< 0.01	-	ND	0.066	
050074-05						14	0.037	ND	-	ND	0.047	
						21	0.049	< 0.01	-	ND	0.059	
						28	0.027	ND	-	ND	0.037	
						36	0.013	ND	-	ND	0.023	
						42	0.010	ND	-	ND	0.020	

^a Not determined.

Stone Fruits

Supervised trials on cherry, peach, apricot and nectarine in Australia

Trials on cherries (8), peaches (7), nectarines (4) and apricots (4) were conducted during the 2005 to 2006 season in Australia (Cowles, 2006; Report 050063).

Trials were conducted in six sites (two sites for cherries, two for peaches, and one each for apricots and nectarines), each site consisting of one untreated and four treated plots. Each plot, which had 1 to 4 trees ranging from flowering to 7 cm diameter fruits, was treated with 4 or 7 applications of a WG formulation of spinetoram containing 250 g ai/kg. Two treatment regimes were used. The first treatment schedule consisted of three applications at 5-day intervals over flowering followed by four applications at 14-day intervals commencing 63 days before harvest so that the last applications at 14-day intervals commencing 63 days before harvest so that the last application was 21 days before harvest. Treatments were made as foliar spray applications at rates of 5 g ai/hL or 7.5 g ai/hL of spinetoram.

Approximately 2 kg each of whole peaches, apricots, and nectarines and 0.5 kg of cherries were collected 0, 7, 14 or 15, 21 or 22, 28 and 35 days after the last application. Samples were placed in plastic bags, labelled, and frozen at -20°C until analysed in June 2006 to August 2006.

Samples were analysed for residues of spinetoram and the metabolites using the method GRM 05.03. The LOQ for the method was 0.01 mg/kg. The LOD was 0.005 mg/kg. Recoveries were within the acceptable range of 70-120% and relative standard deviation < 20%, with the exception of apricots. The mean recovery of XDE-175-J from apricots fortified at the LOQ (66%) was lower than 70%.

Supervised trials on cherries, peach and apricot in New Zealand

Trials on cherries (12), peach (8), and apricot (8) were conducted during the 2005 to 2006 season in New Zealand (Cowles, 2006; Report 050075).

Trials were conducted in seven sites (three sites for cherries, two for peaches, and two for apricots), each site consisting of one untreated and four treated plots. Each peach and apricot site had 4 trees while the cherry plots had 2 trees each. Four applications of a SC formulation of spinetoram containing 120 g ai/L were made to each plot using high volume spray applications at rates of 2.5 g ai/hL, 3.7 g ai/hL, 5 g ai/hL, or 7.6 g ai/ha. Treatments were made at intervals of 13–15 days to the stone fruit plots containing fruits ranging from 1.1 cm to 5 cm in diameter at the first application to 1.2 cm to 6.5 cm in diameter at the fourth application.

^b Total residue = XDE-175-J + XDE-175-L + ND-J + NF-J

Approximately 2 kg each of whole peaches and apricots, and 1 kg of cherries were collected by hand 0, 1, 3, 7, 14, 21, and 28 days after the last application. Samples were placed in plastic bags, labelled, and frozen at -20 °C until analysed.

Spinetoram and its metabolites were analysed using method GRM 05.03 with an LOQ of 0.01 mg/kg. The LOD was determined to be 0.005 mg/kg.

Table 26-1 Residues of spinetoram from supervised trials on cherry in Australia and New Zealand (for estimation of maximum residue level.

	Form	A	pplication		Total/	PHI,	Re	esidue, mg/l	kg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	Total ^a	No.
	AUS'	TRALIAN	TRIALS	•	•				•	
Sheffield Rd,	WG	5	54–92	4	271	0	0.05	0.01	0.06	050063
Spreyton TAS 2005						7	< 0.01	ND	< 0.01	
(Van)						14	< 0.01	ND	< 0.01	
050063-01						21	< 0.01	ND	< 0.01	
						28	< 0.01	ND	< 0.01	
Sheffield Rd,	WG	7.5	93-125	4	432	0	0.09	0.03	0.12	050063
Spreyton TAS 2005						7	0.02	ND	0.02	
(Van)						14	ND	ND	< 0.01	
050063-01						21	< 0.01	ND	< 0.01	
						28	< 0.01	ND	< 0.01	
Sheffield Rd,	WG	5	54–92	7	471	0	0.14	0.04	0.18	050063
Spreyton TAS 2005						7	0.02	ND	0.02	
(Van)						14	0.01	ND	0.01	
050063-01						21	ND	ND	< 0.01	
						28	< 0.01	ND	< 0.01	
Sheffield Rd,	WG	7.5	93–125	7	758	0	0.13	0.04	0.17	050063
Spreyton TAS 2005						7	0.02	ND	0.02	
(Van)						14	< 0.01	ND	< 0.01	
050063-01						21	< 0.01	ND	< 0.01	
						28	< 0.01	ND	< 0.01	
Wilgro	WG	5	70–90	4	330	7	< 0.01	ND	< 0.01	050063
Orchards, Tumut Road,						14	ND	ND	< 0.01	
Batlow, NSW,						21	ND	ND	< 0.01	
2005 (Stella)						28	ND	ND	< 0.01	
050063-02						35	ND	ND	< 0.01	
Wilgro	WG	7.5	105-	4	495	7	0.01	ND	0.01	050063
Orchards,			135			14	< 0.01	ND	< 0.01	
Tumut Road, Batlow, NSW,						21	ND	ND	< 0.01	
2005						28	ND	ND	< 0.01	
(Stella) 050063-02						35	ND	ND	< 0.01	
Wilgro	WG	5	60–90	7	560	7	< 0.01	ND	< 0.01	050063
Orchards,						14	ND	ND	< 0.01	
Tumut Road, Batlow, NSW						21	ND	ND	< 0.01	
2005						28	ND	ND	< 0.01	
(Stella) 050063-02						35	ND	ND	< 0.01	

	Form	A	pplication		Total/	PHI,	Re	esidue, mg/l	kg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	Total ^a	No.
Wilgro	WG	7.5	90–135	7	694	7	< 0.01	ND	< 0.01	050063
Orchards, Tumut Road,						14	< 0.01	ND	< 0.01	
Batlow, NSW,						21	ND	ND	< 0.01	
2005						28	ND	ND	< 0.01	
(Stella) 050063-02						35	ND	ND	< 0.01	
	NEW 2	ZEALAND	TRIALS	•		•				
Tukituki,	SC	2.5	38–40	4	157	0	0.019	< 0.01	0.019	050075
Hawke's Bay, 2005						1	0.018	ND	0.018	
(Stella)						3	< 0.01	ND	< 0.01	
050075-01						7	< 0.01	ND	< 0.01	
						14	ND	ND	< 0.01	
Tukituki,	SC	3.7	57–58	4	231	0	0.041	0.011	0.052	050075
Hawke's Bay, 2005						1	0.025	< 0.01	0.025	
(Stella)						3	0.018	ND	0.018	
050075-01						7	< 0.01	ND	< 0.01	
						14	ND	ND	< 0.01	
						21	ND	ND	< 0.01	
Tukituki,	SC	5	78–79	4	313	0	0.069	0.019	0.088	050075
Hawke's Bay, 2005						1	0.048	0.013	0.061	
(Stella)						3	0.036	< 0.01	0.036	
050075-01						7	0.018	ND	0.018	
						14	< 0.01	ND	< 0.01	
						21	< 0.01	ND	< 0.01	
Tukituki,	SC	7.6	116-	4	469	0	0.136	0.037	0.173	050075
Hawke's Bay, 2005			119			1	0.092	0.024	0.116	
(Stella)						3	0.029	< 0.01	0.029	
050075-01						7	0.045	< 0.01	0.045	
						14	0.028	ND	0.028	
						21	< 0.01	ND	< 0.01	
Alexandra,	SC	2.5	35–41	4	156	0	0.030	< 0.01	0.030	050075
Central Otago, 2006						1	0.028	< 0.01	0.028	
(Lapins)						3	0.019	ND	0.019	
050075-04						7	0.019	ND	0.019	
						14	ND	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
Alexandra, Central Otago,	SC	3.7	52–61	4	231	0	0.060	0.015	0.075	050075
2006						1	0.032	< 0.01	0.032	
(Lapins)						3	0.019	< 0.01	0.019	
050075-04						7	0.019	ND	0.019	
						14	< 0.01	ND	< 0.01	
						21	< 0.01	ND	< 0.01	
						28	< 0.01	ND	< 0.01	

	Form	A	pplication		Total/	PHI,	R	esidue, mg/l	кg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	Total ^a	No.
Alexandra,	SC	5	70–82	4	311	0	0.102	0.027	0.129	050075
Central Otago, 2006						1	0.064	0.016	0.080	
(Lapins)						3	0.045	0.012	0.057	
050075-04						7	0.054	0.013	0.067	
						14	0.015	ND	0.015	
						21	0.013	ND	0.013	
						28	< 0.01	ND	< 0.01	
Alexandra,	SC	7.6	105-	4	467	0	0.139	0.032	0.171	050075
Central Otago,			123			1	0.100	0.026	0.126	
2006 (Lapins)						3	0.044	0.011	0.055	
050075-04						7	0.059	0.013	0.072	
						14	0.021	< 0.01	0.021	
						21	0.014	ND	0.014	
						28	< 0.01	ND	< 0.01	
Earnscleugh,	SC	2.5	41–49	4	187	0	0.040	0.012	0.052	050075
Central Otago 2005						1	0.039	0.011	0.050	
(Stella)						3	0.025	< 0.01	0.025	
050075-05						7	0.024	< 0.01	0.024	
						14	0.009	ND	0.009	
						21	0.010	ND	0.01	
						28	ND	ND	< 0.01	
Earnscleugh,	SC	3.7	60–73	4	275	0	0.065	0.021	0.086	050075
Central Otago 2005						1	0.055	0.019	0.074	
(Stella)						3	0.029	< 0.01	0.029	
050075-05						7	0.023	< 0.01	0.023	
						14	< 0.01	ND	< 0.01	
						21	< 0.01	ND	< 0.01	
						28	< 0.01	ND	< 0.01	
Earnscleugh,	SC	5	81–99	4	374	0	0.075	0.023	0.098	050075
Central Otago 2005						1	0.089	0.028	0.117	
(Stella)						3	0.047	0.011	0.058	
050075-05						7	0.025	< 0.01	0.025	
						14	0.024	< 0.01	0.024	
						21	< 0.01	ND	< 0.01	
						28	ND	ND	< 0.01	
Earnscleugh,	SC	7.6	122-	4	560	0	0.103	0.035	0.138	050075
Central Otago 2005			148			1	0.053	0.017	0.070	
(Stella)						3	0.059	0.017	0.076	
050075-05						7	0.052	0.015	0.067	
030073-03						14	< 0.01	< 0.01	< 0.01	
						21	0.013	ND	0.013	
						28	0.010	ND	0.010	

^a Total residue = XDE-175-J + XDE-175-L.

 $Table\ 26\ Residues\ of\ spinetoram\ and\ metabolites\ from\ supervised\ trials\ on\ cherry\ in\ Australia\ and\ New\ Zealand\ (for\ estimation\ of\ STMR)$

	Form	Ap	plication	1	Total/	PHI,		Re	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total	No.
					AUSTRA	ALIAN '	TRIALS					
Sheffield Rd,	WG	5	54–	4	271	0	0.05	0.01	ND	< 0.01	0.07	050063
Spreyton TAS 2005			92			7	< 0.01	ND	ND	< 0.01	< 0.02	
(Van)						14	< 0.01	ND	ND	< 0.01	< 0.02	
050063-01						21	< 0.01	ND	ND	< 0.01	< 0.02	
						28	< 0.01	ND	ND	0.010	0.020	
Sheffield Rd,	WG	7.5	93-	4	432	0	0.09	0.03	ND	0.02	0.14	050063
Spreyton TAS 2005			125			7	0.02	ND	< 0.01	0.03	0.05	
(Van)						14	ND	ND	ND	< 0.01	< 0.02	
050063-01						21	< 0.01	ND	ND	< 0.01	< 0.02	
						28	< 0.01	ND	ND	< 0.01	< 0.02	
Sheffield Rd,	WG	5	54-	7	471	0	0.14	0.04	< 0.01	< 0.01	0.19	050063
Spreyton TAS 2005			92			7	0.02	ND	ND	0.01	0.03	
(Van)						14	0.01	ND	ND	0.01	0.02	
050063-01						21	ND	ND	ND	ND	< 0.02	
						28	< 0.01	ND	ND	ND	< 0.02	
Sheffield Rd,	WG	7.5	93-	7	758	0	0.13	0.04	< 0.01	0.02	0.19	050063
Spreyton TAS 2005			125			7	0.02	ND	ND	0.02	0.04	
(Van)						14	< 0.01	ND	ND	< 0.01	< 0.02	
050063-01						21	< 0.01	ND	ND	< 0.01	< 0.02	
						28	< 0.01	ND	ND	< 0.01	< 0.02	
Wilgro	WG	5	70–	4	330	7	< 0.01	ND	ND	ND	< 0.02	050063
Orchards, Tumut Road,			90			14	ND	ND	ND	ND	< 0.02	
Batlow, NSW,						21	ND	ND	ND	ND	< 0.02	
2005 (Stella)						28	ND	ND	ND	ND	< 0.02	
050063-02						35	ND	ND	ND	ND	< 0.02	
Wilgro	WG	7.5	105-	4	495	7	0.01	ND	ND	ND	0.02	050063
Orchards, Tumut Road,			135			14	< 0.01	ND	ND	ND	< 0.02	
Batlow, NSW,						21	ND	ND	ND	ND	< 0.02	
2005 (Stella)						28	ND	ND	ND	ND	< 0.02	
050063-02						35	ND	ND	ND	ND	< 0.02	
Wilgro	WG	5	60-	7	560	7	< 0.01	ND	ND	ND	< 0.02	050063
Orchards, Tumut Road,			90			14	ND	ND	ND	ND	< 0.02	
Batlow, NSW						21	ND	ND	ND	ND	< 0.02	
2005 (Stella)						28	ND	ND	ND	ND	< 0.02	
050063-02						35	ND	ND	ND	ND	< 0.02	
Wilgro	WG	7.5	90–	7	694	7	< 0.01	ND	< 0.01	0.01	0.02	050063
Orchards, Tumut Road,			135			14	< 0.01	ND	ND	ND	< 0.02	
Batlow, NSW,						21	ND	ND	ND	ND	< 0.02	
2005 (Stalla)						28	ND	ND	ND	ND	< 0.02	
(Stella) 050063-02						35	ND	ND	ND	ND	< 0.02	

	Form	Ap	plication	1	Total/	PHI,		Re	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total	No.
				l .	NEW ZEA	ALAND	TRIALS					
Tukituki,	SC	2.5	38-	4	157	0	0.019	< 0.01	-	< 0.01	0.029	050075
Hawke's Bay, 2005			40			1	0.018	ND	-	0.012	0.030	
(Stella)						3	< 0.01	ND	-	0.013	0.023	1
050075-01						7	< 0.01	ND	-	< 0.01	< 0.02	-
						14	ND	ND	-	< 0.01	< 0.02	
Tukituki,	SC	3.7	57-	4	231	0	0.041	0.011	-	0.013	0.065	050075
Hawke's Bay, 2005			58			1	0.025	< 0.01	-	0.014	0.039	
(Stella)						3	0.018	ND	-	0.018	0.036	
050075-01						7	< 0.01	ND	-	0.013	0.023	
						14	ND	ND	-	< 0.01	< 0.02	
						21	ND	ND	-	ND	< 0.02	
Tukituki,	SC	5	78–	4	313	0	0.069	0.019	-	0.024	0.112	050075
Hawke's Bay, 2005			79			1	0.048	0.013	-	0.019	0.080	
(Stella)						3	0.036	< 0.01	-	0.026	0.062	
050075-01						7	0.018	ND	-	0.028	0.046	
						14	< 0.01	ND	-	0.019	0.029	
						21	< 0.01	ND	-	< 0.01	< 0.02	
Tukituki,	SC	7.6	116-	4	469	0	0.136	0.037	-	0.027	0.200	050075
Hawke's Bay, 2005			119			1	0.092	0.024	-	0.020	0.136	
(Stella)						3	0.029	< 0.01	-	0.021	0.050	
050075-01						7	0.045	< 0.01	-	0.027	0.072	
						14	0.028	ND	-	0.032	0.060	
						21	< 0.01	ND	-	0.010	0.020	
Alexandra, Central	SC	2.5	35– 41	4	156	0	0.030	< 0.01	-	0.015	0.045	050075
Otago, 2006			41			1	0.028	< 0.01	-	0.018	0.046	-
(Lapins)						3	0.019	ND	-	0.025	0.044	-
050075-04						7	0.019	ND	-	0.022	0.041	
						14	ND	ND	-	< 0.01	< 0.02	-
						21	ND	ND	-	< 0.01	< 0.02	-
						28	ND	ND	-	< 0.01	< 0.02	
Alexandra, Central	SC	3.7	52– 61	4	231	0	0.060	0.015	-	0.024	0.099	050075
Otago, 2006			01			1	0.032	< 0.01	-	0.019	0.051	1
(Lapins) 050075-04						3	0.019	< 0.01	-	0.020	0.039	
030073-04						7	0.019	ND	-	0.018	0.037	
						14	< 0.01	ND	-	0.012	0.022	_
						21	< 0.01	ND	-	< 0.01	< 0.02	_
., .	~-				0.11	28	< 0.01	ND	-	< 0.01	< 0.02	0500==
Alexandra, Central	SC	5	70– 82	4	311	0	0.102	0.027	-	0.029	0.158	050075
Otago, 2006			02			1	0.064	0.016	-	0.036	0.116	_
(Lapins) 050075-04						3	0.045	0.012	-	0.032	0.089	-
050075-04						7	0.054	0.013	-	0.037	0.104	-
						14	0.015	ND	-	0.016	0.031	

	Form	Ap	plication	1	Total/	PHI,		Re	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total	No.
						21	0.013	ND	-	0.017	0.030	
						28	< 0.01	ND	-	0.013	0.023	
Alexandra,	SC	7.6	105-	4	467	0	0.139	0.032	-	0.033	0.204	050075
Central Otago, 2006			123			1	0.100	0.026	-	0.027	0.153	
(Lapins)						3	0.044	0.011	-	0.017	0.072	
050075-04						7	0.059	0.013	-	0.037	0.109	
						14	0.021	< 0.01	-	0.026	0.047	
						21	0.014	ND	-	0.018	0.032	
						28	< 0.01	ND	-	0.016	0.026	
Earnscleugh,	SC	2.5	41-	4	187	0	0.040	0.012	-	0.018	0.070	050075
Central Otago 2005			49			1	0.039	0.011	-	0.020	0.070	
(Stella)						3	0.025	< 0.01	-	0.016	0.041	
050075-05						7	0.024	< 0.01	-	0.014	0.038	
						14	0.009	ND	-	0.015	0.024	
						21	0.010	ND	-	0.016	0.026	
						28	ND	ND	-	ND	< 0.02	
Earnscleugh,	SC	3.7	60-	4	275	0	0.065	0.021	-	0.024	0.110	050075
Central Otago 2005			73			1	0.055	0.019	-	0.026	0.100	
(Stella)						3	0.029	< 0.01	-	0.019	0.048	
050075-05						7	0.023	< 0.01	-	0.034	0.057	
						14	< 0.01	ND	-	0.016	0.026	
						21	< 0.01	ND	-	0.019	0.029	
						28	< 0.01	ND	-	< 0.01	< 0.02	
Earnscleugh,	SC	5	81-	4	374	0	0.075	0.023	-	0.022	0.120	050075
Central Otago 2005			99			1	0.089	0.028	-	0.041	0.158	
(Stella)						3	0.047	0.011	-	0.020	0.078	
050075-05						7	0.025	< 0.01	-	0.023	0.048	
						14	0.024	< 0.01	-	0.023	0.047	
						21	< 0.01	ND	-	0.015	0.025	
						28	ND	ND	-	ND	< 0.02	
Earnscleugh,	SC	7.6	122-	4	560	0	0.103	0.035	-	0.027	0.165	050075
Central Otago 2005			148			1	0.053	0.017	-	0.024	0.094]
(Stella)						3	0.059	0.017	-	0.027	0.103]
						7	0.052	0.015	-	0.024	0.091	1
050075-05						14	< 0.01	< 0.01	-	0.015	0.025]
						21	0.013	ND	-	0.024	0.037]
						28	0.010	ND	-	0.015	0.025]

^a Total residue = XDE-175-J + XDE-175-L + ND-J + NF-J.

Spinetoram Spinetoram

Table 27-1 Residues of spinetoram from supervised trials on peach in Australia and New Zealand (for estimation of maximum residue level)

	Form	A	pplication		Total/	PHI,	Re	esidue, mg/l	kg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	Total ^a	No.
	A	USTRALL	AN TRIALS	S	•		•			
McIssacs Rd,	SC	5	62	4	248	0	0.05	0.01	0.06	050063
Ardmona VIC						7	0.02	< 0.01	0.02	
2005 (Zea Lady)						14	< 0.01	ND	< 0.01	
050063-03						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						35	ND	ND	< 0.01	
McIssacs Rd,	SC	7.5	93	4	372	0	0.03	< 0.01	0.03	050063
Ardmona VIC						7	0.02	ND	0.02	
2005 (Zea Lady)						14	< 0.01	ND	< 0.01	
050063-03						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						35	ND	ND	< 0.01	
McIssacs Rd,	SC	5	54–62	7	415	0	0.03	< 0.01	0.03	050063
Ardmona VIC						7	0.01	ND	0.01	
2005 (Zea Lady)						14	ND	ND	< 0.01	
050063-03						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
McIssacs Rd,	SC	7.5	86–93	7	637	0	0.05	0.01	0.06	050063
Ardmona VIC 2005						7	0.01	ND	0.01	
(Zea Lady)						15	ND	ND	< 0.01	
050063-03						22	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
						35	ND	ND	< 0.01	
Neale Road	SC	5	81–91	4	360	0	0.11	0.03	0.14	050063
Upper Hermitage SA						7	0.02	ND	0.02	
2005/2006						15	< 0.01	ND	< 0.01	
(Tasty ee) 050063-04						22	< 0.01	ND	< 0.01	
030003-04						28	ND	ND	< 0.01	
						35	ND	ND	< 0.01	
Neale Road	WG	7.5	121-	4	538	0	0.12	0.03	0.15	050063
Upper Hermitage SA			136			7	0.03	ND	0.03	
2005/2006						15	0.01	ND	0.01	
(Tasty ee)						22	0.01	ND	0.01	
050063-04						28	< 0.01	ND	< 0.01	
						35	ND	ND	< 0.01	
Neale Road	WG	5	91–136	7	729	0	0.07	0.02	0.09	050063
Upper Hermitage SA						7	0.02	ND	0.02	
2005/2006						15	< 0.01	ND	< 0.01	
(Tasty ee) 050063-04						22	< 0.01	ND	< 0.01	
020002*U T						28	ND	ND	< 0.01	
						35	ND	ND	< 0.01	

	Form	A	pplication		Total/	PHI,	R	esidue, mg/l	kg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	Total ^a	No.
		NEW Z	EALAND '	TRIALS						
Havelock	SC	2.5	46	4	184	0	0.020	ND	0.020	050075
North, Hawke's Bay, 2006						1	0.010	ND	0.010	
(Golden						3	0.011	ND	0.011	
Queen) 050075-02						7	< 0.01	ND	< 0.01	
030073-02						14	ND	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
Havelock	SC	3.7	67–68	4	269	0	0.038	< 0.01	0.038	050075
North, Hawke's Bay, 2006						1	0.018	ND	0.018	
(Golden						3	0.020	ND	0.020	
Queen) 050075-02						7	< 0.01	ND	< 0.01	
030073-02						14	< 0.01	ND	< 0.01	
						21	< 0.01	ND	< 0.01	
						28	ND	ND	< 0.01	
Havelock	SC	5	91–92	4	365	0	0.049	0.012	0.061	050075
North, Hawke's Bay, 2006						1	0.032	< 0.01	0.032	
(Golden						3	0.026	< 0.01	0.026	
Queen)						7	0.024	< 0.01	0.024	
050075-02						14	< 0.01	ND	< 0.01	
						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
Havelock	SC	7.6	137-	4	549	0	0.058	0.016	0.074	050075
North, Hawke's Bay, 2006			138			1	0.041	< 0.01	0.041	
(Golden						3	0.036	< 0.01	0.036	
Queen) 050075-02						7	0.039	< 0.01	0.039	
030073-02						14	0.023	ND	0.023	
						21	0.012	ND	0.012	
						28	0.014	ND	0.014	
Earnscleugh,	SC	2.5	49-	4	199	0	0.060	0.014	0.074	050075
Central Otago 2006			52			1	0.094	0.024	0.118	
(Southern Ice)						3	0.032	< 0.01	0.032	
050075-06						7	0.013	ND	0.013	
						14	< 0.01	ND	< 0.01	
						21	0.019	ND	0.019	
						28	0.027	ND	0.027	
Earnscleugh,	SC	3.7	72–76	4	293	0	0.078	0.019	0.097	050075
Central Otago 2006						1	0.053	0.012	0.065	
(Southern Ice)						3	0.052	0.010	0.062	
050075-06						7	0.030	< 0.01	0.030	
						14	< 0.01	ND	< 0.01]
						21	0.014	ND	0.014	1
						28	< 0.01	ND	< 0.01	1
Earnscleugh,	SC	5	98–103	4	397	0	0.154	0.032	0.186	050075

	Form	A	pplication		Total/	PHI,	R	esidue, mg/k	cg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	Total ^a	No.
Central Otago 2006						1	0.053	0.012	0.065	
(Southern Ice)						3	0.071	0.013	0.084	
050075-06						7	0.039	< 0.01	0.039	
						14	0.015	ND	0.015	
						21	0.014	ND	0.014	
						28	0.020	ND	0.020	
Earnscleugh,	SC	7.6	147-	4	596	0	0.106	0.027	0.133	050075
Central Otago 2006			155			1	0.111	0.029	0.140	
(Southern Ice)						3	0.088	0.014	0.102	
050075-06						7	0.027	< 0.01	0.027	
						14	0.019	ND	0.019	
						21	0.020	ND	0.020	
						28	0.034	ND	0.034	

^a Total residue = XDE-175-J + XDE-175-L.

Table 27-2 Residues of spinetoram and metabolites from supervised trials on peach in Australia and New Zealand (for estimation of STMR)

	Form	Ap	plication	ì	Total/	PHI,		Re	sidue, mg/	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total a	No.
	AUS	TRALIA	N TRIA	LS	<u>I</u>			<u> </u>			<u> </u>	
McIssacs Rd,	SC	5	62	4	248	0	0.05	0.01	ND	ND	0.07	050063
Ardmona VIC 2005						7	0.02	< 0.01	ND	ND	0.03	
(Zea Lady)						14	< 0.01	ND	ND	ND	< 0.02	
050063-03						21	ND	ND	ND	ND	< 0.02	
						28	ND	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
McIssacs Rd,	SC	7.5	93	4	372	0	0.03	< 0.01	ND	ND	0.04	050063
Ardmona VIC 2005						7	0.02	ND	ND	ND	0.03	
(Zea Lady)						14	< 0.01	ND	ND	ND	< 0.02	
050063-03						21	ND	ND	ND	ND	< 0.02	
						28	ND	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
McIssacs Rd,	SC	5	54–	7	415	0	0.03	< 0.01	ND	ND	0.04	050063
Ardmona VIC 2005			62			7	0.01	ND	ND	ND	0.02	
(Zea Lady)						14	ND	ND	ND	ND	< 0.02	
050063-03						21	ND	ND	ND	ND	< 0.02	
						28	ND	ND	ND	ND	< 0.02	

	Form	Ap	plication	1	Total/	PHI,		Re	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total	No.
McIssacs Rd,	SC	7.5	86–	7	637	0	0.05	0.01	< 0.01	< 0.01	0.07	050063
Ardmona VIC 2005			93			7	0.01	ND	ND	ND	0.02	
(Zea Lady)						15	ND	ND	ND	ND	< 0.02	
050063-03						22	ND	ND	ND	ND	< 0.02	
						28	ND	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
Neale Road	SC	5	81-	4	360	0	0.11	0.03	< 0.01	< 0.01	0.15	050063
Upper Hermitage			91			7	0.02	ND	ND	< 0.01	0.03	
SA						15	< 0.01	ND	ND	ND	< 0.02	
2005/2006 (Tasty ee)						22	< 0.01	ND	ND	ND	< 0.02	
050063-04						28	ND	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
Neale Road	WG	7.5	121– 136	4	538	0	0.12	0.03	0.01	0.02	0.18	050063
Upper Hermitage			130			7	0.03	ND	ND	0.01	0.04	
SA 2005/2006						15	0.01	ND	ND	< 0.01	0.02	
2005/2006 (Tasty ee)						22	0.01	ND	ND	< 0.01	0.02	
050063-04						28	< 0.01	ND	ND	< 0.01	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
Neale Road Upper	WG	5	91– 136	7	729	0	0.07	0.02	< 0.01	< 0.01	0.010	050063
Hermitage			130			7	0.02	ND	ND	0.01	0.03	
SA 2005/2006						15	< 0.01	ND	ND	ND	< 0.02	
(Tasty ee)						22	< 0.01	ND	ND	ND	< 0.02	
050063-04						28	ND	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
	N	IEW ZE	ALAND	TRIA	LS							
Havelock	SC	2.5	46	4	184	0	0.020	ND	-	0.011	0.031	050075
North, Hawke's Bay,						1	0.010	ND	-	< 0.01	0.020	
2006						3	0.011	ND	-	0.011	0.022	
(Golden Queen)						7	< 0.01	ND	-	< 0.01	< 0.02	
050075-02						14	ND	ND	-	ND	< 0.02	
						21	ND	ND	-	ND	< 0.02	
						28	ND	ND	-	ND	< 0.02	
Havelock	SC	3.7	67–	4	269	0	0.038	< 0.01	-	0.013	0.051	050075
North, Hawke's Bay,			68			1	0.018	ND	-	0.011	0.029	
2006						3	0.020	ND	-	0.016	0.036	
(Golden Queen)						7	< 0.01	ND	-	< 0.01	< 0.02	
050075-02						14	< 0.01	ND	-	< 0.01	< 0.02	
						21	< 0.01	ND	-	< 0.01	< 0.02	
						28	ND	ND	-	< 0.01	< 0.02	

	Form	Ap	plication	1	Total/	PHI,		Re	sidue, mg	g/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total a	No.
Havelock	SC	5	91-	4	365	0	0.049	0.012	-	0.017	0.078	050075
North, Hawke's Bay,			92			1	0.032	< 0.01	-	0.013	0.045	
2006						3	0.026	< 0.01	-	0.015	0.041	
(Golden Queen)						7	0.024	< 0.01	-	0.015	0.039	
050075-02						14	< 0.01	ND	-	0.011	0.021	
						21	ND	ND	-	ND	< 0.02	
						28	ND	ND	-	< 0.01	< 0.02	
Havelock	SC	7.6	137-	4	549	0	0.058	0.016	-	0.018	0.092	050075
North, Hawke's Bay,			138			1	0.041	< 0.01	-	0.016	0.057	
2006						3	0.036	< 0.01	-	0.027	0.063	
(Golden Queen)						7	0.039	< 0.01	-	0.025	0.064	
050075-02						14	0.023	ND	-	0.017	0.040	
						21	0.012	ND	-	0.017	0.029	
						28	0.014	ND	-	0.020	0.034	
Earnscleugh,	SC	2.5	49-	4	199	0	0.060	0.014	-	0.022	0.096	050075
Central Otago			52			1	0.094	0.024	-	0.038	0.156	
2006						3	0.032	< 0.01	-	0.023	0.055	
(Southern Ice)						7	0.013	ND	-	< 0.01	0.023	
050075-06						14	< 0.01	ND	-	< 0.01	< 0.02	
						21	0.019	ND	-	< 0.01	0.029	
						28	0.027	ND	-	0.010	0.037	
Earnscleugh,	SC	3.7	72-	4	293	0	0.078	0.019	-	0.013	0.11	050075
Central Otago			76			1	0.053	0.012	-	0.020	0.085	
2006						3	0.052	0.010	-	0.025	0.087	
(Southern Ice)						7	0.030	< 0.01	-	0.017	0.047	
050075-06						14	< 0.01	ND	-	< 0.01	< 0.02	
						21	0.014	ND	-	0.011	0.025	
						28	< 0.01	ND	-	< 0.01	< 0.02	
Earnscleugh,	SC	5	98–	4	397	0	0.154	0.032	-	0.041	0.227	050075
Central Otago			103			1	0.053	0.012	-	0.026	0.091	
2006						3	0.071	0.013	-	0.033	0.117	
(Southern Ice)						7	0.039	< 0.01	-	0.036	0.075	
050075-06						14	0.015	ND	-	0.014	0.029	
						21	0.014	ND	-	0.017	0.031	
						28	0.020	ND	-	0.020	0.04	
Earnscleugh, Central	SC	7.6	147-	4	596	0	0.106	0.027	-	0.017	0.150	050075
Otago			155			1	0.111	0.029	-	0.035	0.175	
2006						3	0.088	0.014	-	0.041	0.143	
(Southern Ice)						7	0.027	< 0.01	-	0.019	0.046	
050075-06						14	0.019	ND	-	0.014	0.033	
						21	0.020	ND	-	0.012	0.032	
						28	0.034	ND	-	0.021	0.055	

 $^{^{}a}$ Total residue = XDE-175-J + XDE-175-L + ND-J + NF-J.

Table 28-1 Residues of spinetoram from supervised trials on nectarine in Australia (for estimation of maximum residue level)

	Form	A	pplication		Total/	PHI,	Ro	esidue, mg/l	κg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	Total ^a	No.
NEC	TARINE	S–AUSTR	ALIAN TR	IALS						
McIssacs Rd,	WG	5	62	4	248	0	0.01	ND	0.02	050063
Ardmona VIC 2005						7	ND	ND	< 0.02	
(Fire Pearl)						14	ND	ND	< 0.02	
050063-06						21	ND	ND	< 0.02	
						28	ND	ND	< 0.02	
						35	ND	ND	< 0.02	
McIssacs Rd,	WG	7.5	93	4	372	0	0.02	ND	0.03	050063
Ardmona VIC 2005						7	< 0.01	ND	< 0.02	
(Fire Pearl)						14	ND	ND	< 0.02	
050063-06						21	ND	ND	< 0.02	
						28	ND	ND	< 0.02	
						35	ND	ND	< 0.02	
McIssacs Rd,	WG	5	59–62	7	427	0	0.02	< 0.01	0.03	050063
Ardmona VIC 2005						7	< 0.01	ND	< 0.02	
(Fire Pearl)						14	ND	ND	< 0.02	
050063-06						21	ND	ND	< 0.02	
						28	ND	ND	< 0.02	
McIssacs Rd,	WG	7.5	77–93	7	629	0	0.01	ND	0.03	050063
Ardmona VIC 2005						7	< 0.01	ND	< 0.02	
(Fire Pearl) 050063-06						14	ND	ND	< 0.02	
030003-00						21	ND	ND	< 0.02	
						28	ND	ND	< 0.02	
						35	ND	ND	< 0.02	

^a Total residue = XDE-175-J + XDE-175-L.

Table 28-2 Residues of spinetoram and metabolites from supervised trials on nectarine in Australia (for estimation of STMR)

	Form	Ap	plication	1	Total/	PHI,		Res	idue, mg/	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total a	No.
NECTA	RINES-A	AUSTRA	LIAN T	RIALS	S							
McIssacs Rd,	WG	5	62	4	248	0	0.01	ND	ND	ND	0.02	050063
Ardmona VIC 2005						7	ND	ND	ND	ND	< 0.02	
(Fire Pearl)						14	ND	ND	ND	ND	< 0.02	
050063-06						21	ND	ND	ND	ND	< 0.02	
						28	ND	ND	ND	ND	< 0.02	

	Form	Ap	plication	l	Total/	PHI,		Res	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total a	No.
NECTA	RINES-A	AUSTRA	LIAN T	RIALS	S							
						35	ND	ND	ND	ND	< 0.02	
McIssacs Rd,	WG	7.5	93	4	372	0	0.02	ND	ND	ND	0.03	050063
Ardmona VIC 2005						7	< 0.01	ND	ND	ND	< 0.02	
(Fire Pearl)						14	ND	ND	ND	ND	< 0.02	
050063-06						21	ND	ND	ND	ND	< 0.02	
						28	ND	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
McIssacs Rd,	WG	5	59-	7	427	0	0.02	< 0.01	ND	ND	0.03	050063
Ardmona VIC 2005			62			7	< 0.01	ND	ND	ND	< 0.02	
(Fire Pearl)						14	ND	ND	ND	ND	< 0.02	
050063-06						21	ND	ND	ND	ND	< 0.02	
						28	ND	ND	ND	ND	< 0.02	
McIssacs Rd,	WG	7.5	77-	7	629	0	0.01	ND	ND	ND	0.03	050063
Ardmona VIC 2005			93			7	< 0.01	ND	ND	ND	< 0.02	
(Fire Pearl)						14	ND	ND	ND	ND	< 0.02	
050063-06						21	ND	ND	ND	ND	< 0.02	
						28	ND	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	

 $^{^{}a}$ Total residue = XDE-175-J + XDE-175-L + ND-J + NF-J.

Table 29-1 Residues of spinetoram from supervised trials on apricot in Australia and New Zealand (for estimation of maximum residue level)

	Form	A	pplication		Total/	PHI,	Residue, mg/kg			Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE-175-J	XDE- 175-L	Total ^a	No.
	AU	STRALL	AN TRIAL	S						
McIssacs Rd,	WG	5	51	4	204	0	0.08	0.02	0.10	050063
Ardmona VIC 2005						7	0.02	ND	0.02	
(Francesco) 050063-05						14	< 0.01	< 0.01	< 0.01	
030003-03						21	< 0.01	ND	< 0.01	
						28	ND	ND	< 0.01	
						35	ND	ND	< 0.01	
McIssacs Rd,	WG	7.5	76	4	304	0	0.08	0.02	0.10	050063
Ardmona VIC 2005						7	0.02	ND	0.02	
(Francesco) 050063-05						14	0.01	ND	0.01	
030003-03						21	< 0.01	ND	< 0.01	
						28	< 0.01	ND	< 0.01	
						35	ND	ND	< 0.01	

	Form	A	pplication		Total/	PHI,	Residue, mg/kg			Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE-175-J	XDE- 175-L	Total ^a	No.
McIssacs Rd,	WG	5	51–52	7	358	0	0.07	0.02	0.09	050063
Ardmona VIC 2005						7	0.02	ND	0.02	
(Francesco)						14	0.01	ND	0.01	
050063-05						21	ND	ND	< 0.01	
						28	ND	ND	< 0.01	
M-I D-I	WC	7.5	76–79	7	526	35	ND	ND 0.04	< 0.01	050062
McIssacs Rd, Ardmona VIC	WG	7.5	/0-/9	7	536	0	0.16	0.04	0.20	050063
2005						7	0.02	ND	0.02	
(Francesco) 050063-05						14	0.02	ND	0.02	
020000 02						21	0.01	ND	0.01	
						28	< 0.01	ND	< 0.01	
						35	< 0.01	ND	< 0.01	
	NEV	V ZEALA	ND TRIA	LS						
Bay View,	SC	2.5	31–33	4	129	0	0.038	< 0.01	0.038	050075
Hawke's Bay,						1	0.048	0.012	0.06	
2005 (Castlebright)						3	0.031	< 0.01	0.031	
050075-03						7	0.016	ND	0.016	
						14	0.010	ND	0.01	
Bay View,	SC	3.7	46–48	4	189	0	0.057	0.015	0.072	050075
Hawke's Bay,						1	0.048	0.012	0.06	
2005 (Castlebright)						3	0.032	< 0.01	0.032	
050075-03						7	0.041	< 0.01	0.041	
						14	0.014	ND	0.014	
Bay View,	SC	5	62–65	4	256	0	0.080	0.019	0.099	050075
Hawke's Bay,						1	0.091	0.022	0.113	
2005 (Castlebright)						3	0.063	0.015	0.078	
050075-03						7	0.055	0.012	0.067	
						14	0.017	ND	0.017	
Bay View,	SC	7.6	93–98	4	385	0	0.185	0.043	0.228	050075
Hawke's Bay, 2005						1	0.208	0.052	0.26	
(Castlebright)						3	0.153	0.033	0.186	
050075-03						7	0.080	0.017	0.097	
Б 1 1	n.c.	2.5	26.40	4	150	14	0.030	< 0.01	0.03	050075
Earnscleugh, Central Otago	SC	2.5	36–40	4	150	1	0.035	0.055	0.09	050075
2006						3	0.039	ND	0.051	
(Sundrop) 050075-07						7	0.018	ND	0.018	
000010 01						14	< 0.01	ND	< 0.01	
Earnscleugh,	SC	3.7	53–59	4	221	0	0.126	0.039	0.165	050075
Central Otago						1	0.124	0.037	0.161	
2006 (Sundrop)						3	0.063	0.018	0.081	
050075-07						7	0.041	0.011	0.052	
			I		1	14	0.022	< 0.01	0.022	

	Form	A	pplication		Total/	PHI,	Residue, mg/kg			Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE-175-J	XDE- 175-L	Total ^a	No.
Earnscleugh,	SC	5	72–79	4	298	0	0.101	0.031	0.132	050075
Central Otago 2006						1	0.080	0.023	0.103	
(Sundrop)						3	0.081	0.025	0.106	
050075-07						7	0.064	0.017	0.081	
						14	0.024	ND	0.024	
Earnscleugh,	SC	7.6	107-	4	447	0	0.222	0.067	0.289	050075
Central Otago 2006			119			1	0.213	0.062	0.275	
(Sundrop) 050075-07						3	0.139	0.038	0.177	
0300/3-0/						7	0.084	0.022	0.106	
						14	0.034	< 0.01	0.034	

^a Total residue = XDE-175-J + XDE-175-L.

Table 29-2 Residues of spinetoram and metabolites from supervised trials on apricot in Australia and New Zealand (for estimation of STMR)

	Form	Ap	plication	n	Total/	PHI,		Res	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total ^a	No.
	AUSTR	ALIAN	TRIAL	S								
McIssacs Rd,	WG	5	51	4	204	0	0.08	0.02	0.01	< 0.01	0.12	050063
Ardmona VIC 2005						7	0.02	ND	0.01	< 0.01	0.04	
(Francesco) 050063-05						14	< 0.01	< 0.01	ND	ND	< 0.02	
030003-03						21	< 0.01	ND	ND	ND	< 0.02	
						28	ND	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
McIssacs Rd,	WG	7.5	76	4	304	0	0.08	0.02	0.02	< 0.01	0.13	050063
Ardmona VIC 2005						7	0.02	ND	< 0.01	< 0.01	0.03	
(Francesco) 050063-05						14	0.01	ND	< 0.01	< 0.01	0.02	
030003-03						21	< 0.01	ND	ND	ND	< 0.02	
						28	< 0.01	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	
McIssacs Rd,	WG	5	51-	7	358	0	0.07	0.02	0.01	< 0.01	0.11	050063
Ardmona VIC 2005			52			7	0.02	ND	0.01	0.010	0.04	
(Francesco) 050063-05						14	0.01	ND	< 0.01	< 0.01	0.02	
030003-03						21	ND	ND	ND	ND	< 0.02	
						28	ND	ND	ND	ND	< 0.02	
						35	ND	ND	ND	ND	< 0.02	

	Form	Ap	plication	n	Total/	PHI,		Re	sidue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total ^a	No.
McIssacs Rd,	WG	7.5	76– 79	7	536	0	0.16	0.04	0.02	0.01	0.23	050063
Ardmona VIC 2005			79			7	0.02	ND	0.010	< 0.01	0.04	
(Francesco)						14	0.02	ND	0.010	< 0.01	0.04	
050063-05						21	0.01	ND	< 0.01	< 0.01	0.02	
						28	< 0.01	ND	< 0.01	< 0.01	< 0.02	
						35	< 0.01	ND	ND	ND	< 0.02	
N	EW ZE	ALAND	TRIAI	LS								-
Bay View, Hawke's	SC	2.5	31-	4	129	0	0.038	< 0.01	-	0.018	0.056	050075
Bay, 2005 (Castlebright)			33			1	0.048	0.012	-	0.023	0.083	
050075-03						3	0.031	< 0.01	-	0.023	0.054	
						7	0.016	ND	-	0.015	0.031	
						14	0.010	ND	-	0.013	0.023	
Bay View, Hawke's	SC	3.7	46-	4	189	0	0.057	0.015	-	0.020	0.092	050075
Bay, 2005 (Castlebright)			48			1	0.048	0.012	-	0.021	0.081	
050075-03						3	0.032	< 0.01	-	0.021	0.053	
						7	0.041	< 0.01	-	0.028	0.069	
						14	0.014	ND	-	0.017	0.031	
Bay View, Hawke's	SC	5	62-	4	256	0	0.080	0.019	-	0.026	0.125	050075
Bay, 2005 (Castlebright)			65			1	0.091	0.022	-	0.036	0.149	
050075-03						3	0.063	0.015	-	0.032	0.11	
						7	0.055	0.012	-	0.042	0.109	
						14	0.017	ND	-	0.020	0.037	
Bay View, Hawke's	SC	7.6	93–	4	385	0	0.185	0.043	-	0.045	0.273	050075
Bay, 2005 (Castlebright)			98			1	0.208	0.052	-	0.058	0.318	
050075-03						3	0.153	0.033	-	0.065	0.251	
						7	0.080	0.017	-	0.055	0.152	
						14	0.030	< 0.01	-	0.030	0.06	
Earnscleugh, Central	SC	2.5	36– 40	4	150	0	0.035	0.055	-	0.010	0.1	050075
Otago 2006 (Sundrop)			40			1	0.039	0.012	-	0.013	0.064	
050075-07						3	0.018	ND	-	< 0.01	0.028	
						7	0.019	ND	-	< 0.01	0.029	
						14	< 0.01	ND	-	0.004	0.014	
Earnscleugh, Central Otago 2006	SC	3.7	53– 59	4	221	0	0.126	0.039	-	0.024	0.189	050075
(Sundrop)			37			1	0.124	0.037	-	0.036	0.197	
050075-07						3	0.063	0.018	-	0.028	0.109	_
						7	0.041	0.011	-	0.023	0.075	4
Б. 1.16.1	0.0		70	,	200	14	0.022	< 0.01	-	0.016	0.038	050055
Earnscleugh, Central Otago 2006	SC	5	72– 79	4	298	0	0.101	0.031	-	0.025	0.157	050075
(Sundrop)						1	0.080	0.023	-	0.019	0.122	1
050075-07						3	0.081	0.025	-	0.031	0.137	4
						7	0.064	0.017	-	0.034	0.115	4
						14	0.024	ND	-	0.021	0.045	

	Form	Ap	plicatio	n	Total/ PHI, Residue, mg/kg							Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total ^a	No.
Earnscleugh,Central	SC	7.6	107-	4	447	0	0.222	0.067	-	0.038	0.327	050075
Otago 2006 (Sundrop)			119			1	0.213	0.062	-	0.046	0.321	
050075-07						3	0.139	0.038	-	0.038	0.215	
						7	0.084	0.022	-	0.036	0.142	
						14	0.034	< 0.01	-	0.017	0.051	

^a Total residue = XDE-175-J + XDE-175-L + ND-J + NF-J.

Fruiting Vegetables other than Cucurbits

Supervised trials on tomato in the USA

Six supervised trials on tomatoes were conducted in major tomato growing areas in the USA in 2004 and 2007 (Dolder and Wendelburg, 2005; Report 040063; McKellar, 2008; Report ARAP 07D-001).

Trials were conducted in six sites, each site consisting of a control plot and a treated plot. Each treated plot received five applications of a suspension concentrate (SC) formulation containing either 100 g/L or 120 g/L of spinetoram. The initial application was made at a nominal rate of 20 g ai/ha, followed by four additional applications at the higher rate of 70 g ai/ha for a seasonal maximum of approximately 300 g ai/ha. The applications were made as low volume spray applications at 4-day intervals. Mature tomatoes were collected at the PHI of 1 day. In one trial, samples were also taken on days 0, 1, 3, 7, and 14 days after the last treatment.

Approximately 2 kg of mature whole tomato fruits were collected by hand. At each sampling, a single composite sample was taken from the control plot while duplicate, composite samples were taken independently from each treated plot. Samples were frozen within 4 hours of collection, placed in plastic bags, labelled, shipped frozen, and kept in temperature-monitored freezers at -20 °C until analysis.

Spinetoram and its metabolites were analysed using the method GRM 05.04 with an LOQ and LOD of 0.01 mg/kg and 0.003 mg/kg, respectively. Samples from the 2007 trials were analysed using method GRM 05.03 with the same LOQ and LOD as GRM 05.04.

Table 30-1 Residues of spinetoram from supervised trials on tomato in the USA (for estimation of maximum residue level)

	Form	A_1	pplication		Total/	PHI,	Re	esidue, mg/l	κg	Report	
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	Total ^a	No.	
GAP, USA Fruiting vegetables	SC		44-88	6	298	1					
North Rose, NY, 2004 (Floradade)	SC	5, initial, + 21, 4 appl.	21, initial, + 70- 71, 4 appl.	5	304	1	0.010	ND ND	0.010	040063	
Lake Park, GA, 2004 (Sonoma Roam)	SC	5, initial, + 17- 19, 4 appl.	20, initial, + 71- 72, 4 appl.	5	306	1	ND ND	ND ND	< 0.01	040063	

	Form	A	pplication		Total/	PHI,	R	esidue, mg/l	κg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	Total ^a	No.
Jennings, FL,	SC	5,	20,	5	308	1	< 0.01	ND	< 0.01	040063
2004 (Florida)		initial, + 17- 19, 4 appl.	initial, + 70- 76, 4 appl.				ND	ND	< 0.01	
Paynesville,	SC	7,	20,	5	301	1	0.025	< 0.01	0.025	040063
MN, 2004 (Olinda)		initial, +24, 4 appl.	initial, + 70- 71, 4 appl.				0.022	< 0.01	0.022	
Porterville,	SC	22-25	70-71	4	281	0	0.043	0.015	0.058	040063
CA, 2004 (Early Girl)							0.020	< 0.01	0.02	
(Larry Giri)						1	0.015	< 0.01	0.015	
							0.012	< 0.01	0.012	
						3	< 0.01	ND	< 0.01	
							< 0.01	ND	< 0.01	
						7	ND	ND	< 0.01	
							< 0.01	ND	< 0.01	
						14	ND	ND	< 0.01	
							ND	ND	< 0.01	
GA, 2007	SC	5,	21,	5	295	1	0.022	< 0.01	0.022	040063
(Not specified)		initial, + 14- 16, 4 appl.	initial, + 68- 70, 4 appl.				0.024	< 0.01	0.024	

^a Total residue = XDE-175-J + XDE-175-L.

Table 30-2 Residues of spinetoram and metabolites from supervised trials on tomato in the USA (for estimation of STMR) $\,$

	Eam	Ap	plication		Total/	DIII		Re	sidue, mg	/kg		ъ.
Location, year (Variety)	Form	g ai/hL	g ai/ha	No	season, g ai/ha	PHI, days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total ^a	Report No.
GAP, USA Fruiting vegetables	SC		44-88	6	298	1						
North Rose,		5,	21,				0.010	ND	< 0.01	0.010	0.020	
NY, 2004 (Floradade)	SC	initial, + 21, 4 appl.	initial, + 70- 71, 4 appl.	5	304	1	< 0.01	ND	< 0.01	< 0.01	< 0.02	040063
Lake Park,		5,	20,				ND	ND	ND	< 0.01	< 0.02	
GA, 2004 (Sonoma Roam)	SC	initial, + 17- 19, 4 appl.	initial, + 71- 72, 4 appl.	5	306	1	ND	ND	ND	< 0.01	< 0.02	040063
		5,	20,				< 0.01	ND	ND	< 0.01	< 0.02	
Jennings, FL, 2004 (Florida)	SC	initial, + 17- 19, 4 appl.	initial, + 70- 76, 4 appl.	5	308	1	ND	ND	ND	< 0.01	< 0.02	040063

	Г	Ap	plication		Total/	PHI.		Res	sidue, mg	/kg		
Location, year (Variety)	Form	g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total ^a	Report No.
D:11-		7,	20,				0.025	< 0.01	< 0.01	< 0.01	0.035	
Paynesville, MN, 2004 (Olinda)	SC	initial, +24, 4 appl.	initial, + 70- 71, 4 appl.	5	301	1	0.022	< 0.01	< 0.01	< 0.01	0.032	040063
						0	0.043	0.015	< 0.01	0.011	0.069	
						U	0.020	< 0.01	< 0.01	< 0.01	0.030	
						1	0.015	< 0.01	< 0.01	< 0.01	0.025	
						1	0.012	< 0.01	< 0.01	< 0.01	0.022	35 040063 369 30 225 22 040063 02 02 02 02 02 02 02 02 02 02 02 02 02 0
Porterville, CA, 2004	CC.	22.25	70-71	4	281	3	< 0.01	ND	ND	< 0.01	01 0.022 01 < 0.02 040063	040062
(Early Girl)	SC	22-25	/0-/1	4	281	3	< 0.01	ND	< 0.01	< 0.01	< 0.02	040063
OIII)						7	ND	ND	ND	< 0.01	< 0.02	
						,	< 0.01	ND	< 0.01	< 0.01	< 0.02	
						14	ND	ND	ND	ND	< 0.02	
						14	ND	ND	ND	< 0.01	< 0.02	
CA 2007		5,	21,				0.022	< 0.01	ND	< 0.01	0.032	
GA, 2007 (Not specified)	SC	initial, + 14- 16, 4 appl.	initial, + 68- 70, 4 appl.	5	295	1	0.024	< 0.01	ND	< 0.01	0.034	040063

^a Total residue = XDE-175-J + XDE-175-L + ND-J + NF-J.

Leafy Vegetables

Supervised trials on lettuce in the US

Six supervised trials on leaf lettuce were conducted in major lettuce growing areas in the US in 2004 and 2007 (Dolder and Wendelburg, 2005; Report 040063; McKellar, 2008; Report ARAP 07D-001).

Trials were conducted in six sites, each site consisting of a control plot and a treated plot. Each treated plot received five applications of a suspension concentrate (SC) formulation containing either 100 g/L or 120 g/L of spinetoram. The initial application was made at a nominal rate of 20 g ai/ha, followed by four additional applications at the higher rate of 70 g ai/ha for a seasonal total of approximately 300 g ai/ha. The applications were made as low volume spray applications at 4-day intervals. Mature lettuce samples were collected at the PHI of 1 day. In one trial, samples were also taken on days 0, 1, 3, 7, and 14 after the last application.

Approximately 1 kg of mature lettuce leaves was collected by hand. At each sampling, a single composite sample was taken from the control plot while duplicate, composite samples were taken independently from each treated plot. Samples were frozen within 4 h of collection, placed in plastic bags, labelled, shipped frozen, and kept in temperature-monitored freezers at -20 °C until analysis.

Spinetoram and its metabolites were analysed using the method GRM 05.04 with an LOQ and limit of detection LOD of 0.01 mg/kg and 0.003 mg/kg, respectively. Samples from the 2007 trials were analysed using method GRM 05.03 with the same LOQ and LOD values as GRM 05.04.

Table 31-1 Residues of spinetoram from supervised trials on leaf lettuce in the USA (for estimation of maximum residue level)

Location, year	For m	A	pplication		Total/ season,	PHI ,	Residue , mg/kg			Report No.
(Variety)		g ai/hL	g ai/ha	No	g ai/ha	day s	XDE- 175-J	XDE- 175-L	Total ^a	
GAP, USA Leafy vegetables	WG or SC		44–88	6	298	1				
North Rose, NY, 2004 (Black Seeded	WG	6 initial and 22, 4 appl.	21 initial and 71-	5	309	1	0.226	0.057	0.283	040063
Simpson)			73, 4 appl.				0.247	0.067	0.314	
Chula, GA, 2004 (Tehama)	WG	7 initial and 23, 4 appl.	21 initial and 70-	5	305	1	0.120	0.010	0.130	040063
			71, 4 appl.				0.142	0.012	0.154	
Jennings, FL, 2004 (Tehama)	WG	6 initial and 21- 22, 4	20 initial and 70-	5	305	1	0.222	0.060	0.282	040063
		appl.	74, 4 appl.				0.265	0.076	0.341	
King City, CA, 2004 (Sunbelt)	WG	7 initial and 22- 24, 4	21 initial and 67-	5	299	1	0.153	0.043	0.196	040063
(Suissis)		appl.	71, 4 appl.				0.245	0.075	0.320	
Porterville,	WG	6 initial	20	5	300	0	0.97	0.31	1.28	040063
CA, 2004 (Shining Star)		and 19- 20, 4	initial and 70,				1.42	0.44	1.86	
(Similing Star)		appl.	4 appl.			1	0.42	0.12	0.54	040063
							0.43	0.12	0.55	
						3	0.11	0.03	0.14	040063
							0.12	0.03	0.15	
						7	0.05	0.01	0.06	040063
							0.07	0.02	0.09	
						14	0.02	< 0.01	0.02	040063
							0.02	< 0.01	0.02	
GA, 2007 (Not specified)	WG	4 initial and 15,	21 initial	5	305	1	5.62	1.40	7.02	040063
		4 appl.	and 70, 4 appl.				6.26	1.54	7.80	

^a Total residue = XDE-175-J + XDE-175-L.

Table 31-2 Residues of spinetoram and metabolites from supervised trials on leaf lettuce in the USA (for estimation of STMR)

	Form	Ap	plication		Total/	PHI,		Res	idue, mg/	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total a	No.
GAP, USA Leafy vegetables	WG or SC		44– 88	6	298	1						
North Rose, NY, 2004 (Black Seeded Simpson)	WG	6 initial and 22, 4 appl.	21 initial and 71- 73, 4 appl.	5	309	1	0.226	0.057	0.183	0.094	0.560	040063
Chula, GA, 2004 (Tehama)	WG	7 initial and 23, 4 appl.	21 initial and 70-71, 4 appl.	5	305	1	0.120	0.010	0.056	0.077	0.263	040063
Jennings, FL, 2004 (Tehama)	WG	6 initial and 21-22, 4 appl.	20 initial and 70- 74, 4 appl.	5	305	1	0.222	0.060	0.141	0.700	1.123	040063
King City, CA, 2004 (Sunbelt)	WG	7 initial and 22-24, 4 appl.	21 initial and 67- 71, 4 appl.	5	299	1	0.153	0.043	0.076	0.056	0.328	040063
Porterville, CA, 2004	WG	6 initial	20 initial and	5	300	0	0.97 1.42	0.31 0.44	0.31 0.42	0.19 0.22	1.78 2.50	040063
(Shining Star)		and 19-20, 4 appl.	70, 4 appl.			1	0.42	0.12 0.12	0.32 0.35	0.25 0.26	1.11 1.16	040063
						3	0.11 0.12	0.03	0.13 0.16	0.10 0.12	0.37 0.43	040063
						7	0.05 0.07	0.01	0.08	0.08	0.22 0.27	040063
						14	0.02	< 0.01	0.02	0.02	0.06	040063
GA, 2007 (Not specified)	WG	4 initial and 15, 4 appl.	21 initial and 70, 4 appl.	5	305	1	5.62 6.26	1.40	0.66	0.94	8.62 9.55	040063

^a Total residue = XDE-175-J + XDE-175-L + ND-J + NF-J.

Root and Tuber Vegetables

Supervised trials sugar beet in the USA

Six supervised trials on sugar beet were conducted in major sugar beet growing areas in the US in 2004 and 2007 (Dolder and Wendelburg, 2005; Report 040063; McKellar, 2008; Report ARAP 07D-001).

Trials were conducted in six sites, each site consisting of a control plot and a treated plot. Each treated plot received four applications of a suspension concentrate (SC) formulation containing either 100 g/L or 120 g/L of spinetoram at a nominal rate of 70 g ai/ha for a seasonal maximum of 280 g ai/ha. The applications were made as low volume spray applications at 4-day intervals. Mature roots and tops were collected 3 days after the last application. In one trial, samples were also taken on days 0, 1, 3, 7, and 14 days after the last treatment.

Approximately 2.3 kg of mature roots and 2.8 kg of tops were collected by hand. At each sampling, a single composite sample was taken from the control plot while duplicate, composite samples were taken independently from each treated plot. Due to the large size of the sugar beet, the root portion was quartered longitudinally and only one quarter from each of 12 beets were retained as a sample. Samples were frozen within 4 h of collection, placed in plastic bags, labelled, shipped frozen, and kept in temperature-monitored freezers at –20 °C until analysis.

Spinetoram and its metabolites were analysed using the method GRM 05.04 with an LOQ and LOD of 0.01 mg/kg and 0.003 mg/kg, respectively. Samples from the 2007 trials were analysed using method GRM 05.03 with the same LOQ and LOD values as GRM 05.04.

Table 32-1 Residues of spinetoram from supervised trials on sugar beet in the USA (for estimation of maximum residue level)

Location, year	Form	A	pplication		Total/	PHI,	R	esidue, mg/l	kg	Report
(Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	Total ^a	No.
GAP, US, Tuberous vegetables	SC		53-70	4	281	7				
	SUG	AR BEET	ROOTS							
Olivia, MN, 2004	SC	24–25	70–71	4	282	3	ND	ND	< 0.01	040063
(Beta)							ND	ND	< 0.01	
Edgeley, ND, 2004	SC	24–25	70–71	4	281	3	ND	ND	< 0.01	040063
(Beta)							ND	ND	< 0.01	
Levelland, TX, 2004	SC	19	70–72	4	284	3	ND	ND	< 0.01	040063
(Wrangler)							ND	ND	< 0.01	
Porterville, CA, 2004	SC	19	70–71	4	283	3	ND	ND	< 0.01	040063
(Beta 4030R)							ND	ND	< 0.01	
Ephrata, WA,	SC	21	70–71	4	283	0	< 0.01	ND	< 0.01	040063
2004 (Blazer)							< 0.01	ND	< 0.01	
						1	ND	ND	< 0.01	
							ND	ND	< 0.01	
						3	< 0.01	ND	< 0.01	
							ND	ND	< 0.01	
						7	ND	ND	< 0.01	
							ND	ND	< 0.01	

Location, year	Form	Aj	pplication		Total/	PHI,	Re	esidue, mg/k	cg .	Report
(Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	Total ^a	No.
						14	ND	ND	< 0.01	
							ND	ND	< 0.01	
GA, 2007	SC	21-22	69-72	4	283	3	< 0.01	ND	< 0.01	ARAP
(Not specified)							< 0.01	ND	< 0.01	07D-001

^a Total residue = XDE-175-J + XDE-175-L.

Table 32-2 Residues of spinetoram and metabolites from supervised trials on sugar beet in the USA (for estimation of STMR)

Location,	Form	Aŗ	plication	l	Total/	PHI,		Re	sidue, mg	/kg		Report
year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175- L	ND-J	NF-J	Total ^a	No.
GAP, US, Root vegetables	SC		53-70	4	281	7						
	SUGAI	R BEET	ROOTS									
Olivia, MN, 2004	SC	24– 25	70– 71	4	282	3	ND	ND	ND	ND	< 0.02	040063
(Beta)							ND	ND	ND	ND	< 0.02	
Edgeley, ND, 2004	SC	24– 25	70– 71	4	281	3	ND	ND	ND	ND	< 0.02	040063
(Beta)		23	, 1				ND	ND	ND	ND	< 0.02	
Levelland, TX, 2004	SC	19	70– 72	4	284	3	ND	ND	ND	< 0.01	< 0.02	040063
(Wrangler)			, 2				ND	ND	ND	ND	< 0.02	
Porterville, CA, 2004	SC	19	70– 71	4	283	3	ND	ND	< 0.01	< 0.01	< 0.02	040063
(Beta 4030R)							ND	ND	ND	ND	< 0.02	
Ephrata, WA,	SC	21	70–	4	283	0	< 0.01	ND	< 0.01	< 0.01	< 0.02	040063
2004 (Blazer)			71				< 0.01	ND	< 0.01	< 0.01	< 0.02	
(Blazer)						1	ND	ND	ND	ND	< 0.02	
							ND	ND	ND	< 0.01	< 0.02	
						3	< 0.01	ND	< 0.01	< 0.01	< 0.02	
							ND	ND	ND	ND	< 0.02	
						7	ND	ND	ND	< 0.01	< 0.02	
							ND	ND	ND	ND	< 0.02	
						14	ND	ND	< 0.01	< 0.01	< 0.02	
							ND	ND	< 0.01	ND	< 0.02	
GA, 2007 (Not	SC	21- 22	69-72	4	283	3	< 0.01	ND	< 0.01	ND	< 0.02	ARAP 07D-
specified)							< 0.01	ND	< 0.01	ND	< 0.02	001

^a Total residue = XDE-175-J + XDE-175-L + ND-J + NF-J.

Tree Nuts

Supervised trials on almond and pecan in the USA

Six supervised trials on almond and six on pecan were conducted in the USA in 2007 (Dolder and Schelle, 2008; Report 070069).

Trials on almond were conducted in six sites in California, while trials on pecan were conducted in six different States in the USA. Each site consisted of a control plot and a treated plot, with at least six trees in each plot. Each treated plot received four applications of WG formulation containing either 250 g/kg of spinetoram at a nominal rate of 123 g ai/ha for a seasonal maximum of 500g ai/ha. The applications were made at intervals of 7 days, using airblast sprayers with four to seven nozzles. Spray volumes ranged from 889 to 1074 L/ha. Whole nuts were collected 7 days after the last application. In decline trials (one each for almond and pecan), samples were taken on days 0, 1, 3, 7 and 14 days after the last treatment.

For each commodity at each site, a single, composite sample was taken from the control plot while duplicate, composite samples were taken independently from the treated plot. At each sampling the nuts were knocked or shaken from the trees. Trees at the ends of plots were not sampled. The whole nuts were collected and, within approximately 24 hours, were cracked using a nut cracker and the nutmeat (and hulls in the case of almonds) was collected by hand or with a sheller. A minimum of 1 kg was collected for each sample. Samples were placed in sample bags, labelled and frozen within approximately 24 h of sampling and remained frozen up through shipment overnight to the analytical laboratory. All samples were received frozen and were stored in temperature-monitored freezers at approximately $-20~^{\circ}$ C.

Spinetoram and its metabolites were analysed using the method GRM 05.04 with an LOQ and LOD of 0.01 mg/kg and 0.003 mg/kg, respectively.

Table 33-1 Residues of spinetoram from supervised trials on almonds in the USA (for estimation of maximum residue level)

	Form	A _l	pplication		Total/	PHI,	Residue, mg/kg			Report
Location, year (Variety)		g ai/hL	g ai/ha	No	year g ai/ha	days	XDE-175-J	XDE- 175-L	Total ^a	No.
GAP, USA, Tree Nuts	WG		26– 123	4	490	14				
Terra Bella, CA, 2007 070069CA1	WG	13	117– 144	4	532	7	< 0.01	ND	< 0.01	070069
(Carmel)							< 0.01	ND	< 0.01	
Wasco, CA, 2007	WG	13	117– 130	4	501	7	< 0.01	ND	< 0.01	070069
070069CA2 (Price)							< 0.01	ND	< 0.01	
Earlimart, CA, 2007	WG	13	117– 133	4	491	7	ND	ND	< 0.01	070069
070069CA3 (Fritz)							ND	ND	< 0.01	
Sultana, CA, 2007	WG	13	123– 124	4	494	7	< 0.01	ND	< 0.01	070069
070069CA4 (Carmel)							< 0.01	ND	< 0.01	
Sanger, CA, 2007	WG	13	110– 124	4	480	7	ND	ND	< 0.01	070069
070069CA5 (Neplus)							ND	ND	< 0.01	

	Form	Al	plication		Total/	PHI,	Residue, mg/kg			Report
Location, year (Variety)		g ai/hL	g ai/ha	No	year g ai/ha	days	XDE-175-J	XDE- 175-L	Total ^a	No.
Sanger, CA,	WG	13	123-	4	495	0	0.011	ND	0.011	070069
2007 070069CA6			123				0.011	< 0.01	0.011	
(Nonpareil)						1	< 0.01	ND	< 0.01	
							0.011	ND	0.011	
						3	< 0.01	ND	< 0.01	
							0.011	ND	0.011	
						7	< 0.01	ND	< 0.01	070069
							< 0.01	ND	< 0.01	
						14	< 0.01	ND	< 0.01	070069
							< 0.01	ND	< 0.01	

^a Total residue = XDE-175-J + XDE-175-L.

Table 33-2 Residues of spinetoram and metabolites from supervised trials on almonds in the USA (for estimation of STMR)

	Form	A	pplication		Total/	PHI,		Resi	due, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	year g ai/ha	days	XDE- 175-J	XDE- 175-L	ND- J	NF-J	Total	No.
GAP, USA, Tree Nuts	WG		26– 123	4	490	14						
Terra Bella, CA, 2007 070069CA1	WG	13	117– 144	4	532	7	< 0.01	ND	ND	ND	< 0.02	070069
(Carmel)							< 0.01	ND	ND	ND	< 0.02	
Wasco, CA, 2007 070069CA2	WG	13	117– 130	4	501	7	< 0.01	ND	ND	ND	< 0.02	070069
(Price)							< 0.01	ND	ND	ND	< 0.02	
Earlimart, CA, 2007	WG	13	117– 133	4	491	7	ND	ND	ND	ND	< 0.02	070069
070069CA3 (Fritz)							ND	ND	ND	ND	< 0.02	
Sultana, CA, 2007 070069CA4	WG	13	123– 124	4	494	7	< 0.01	ND	ND	ND	< 0.02	070069
(Carmel)							< 0.01	ND	ND	ND	< 0.02	
Sanger, CA, 2007 070069CA5	WG	13	110– 124	4	480	7	ND	ND	ND	ND	< 0.02	070069
(Neplus)							ND	ND	ND	ND	< 0.02	
Sanger, CA,	WG	13	123-	4	495	0	0.011	ND	ND	ND	0.021	070069
2007 070069CA6			123				0.011	< 0.01	ND	ND	0.021	
(Nonpareil)						1	< 0.01	ND	ND	ND	< 0.02	
							0.011	ND	ND	ND	0.021	
						3	< 0.01	ND	ND	ND	< 0.02	
							0.011	ND	ND	ND	0.021	

	Form	A	pplication		Total/	PHI,		Resi	due, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	year g ai/ha	days	XDE- 175-J	XDE- 175-L	ND- J	NF-J	Total a	No.
						7	< 0.01	ND	ND	ND	< 0.02	070069
							< 0.01	ND	ND	ND	< 0.02	
						14	< 0.01	ND	ND	ND	< 0.02	070069
							< 0.01	ND	ND	ND	< 0.02	

^a Total residue = XDE-175-J + XDE-175-L + ND-J + NF-J.

Table 34-1 Residues of spinetoram from supervised trials on pecan in the USA (for estimation of maximum residue level)

	Form	A	pplication		Total/	PHI,	Ro	esidue, mg/l	ζg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	Total ^a	No.
GAP, USA, Tree Nuts	WG		26–123	4	490	14				
Scott, AR, 2007 070069AR	WG	13	123– 126	4	495	7	ND	ND	< 0.01	070069
(Stuart)							ND	ND	< 0.01	
Albany, GA, 2007 070069GA2	WG	13	122– 126	4	495	7	ND	ND	< 0.01	070069
(Sumner)							ND	ND	< 0.01	
Madill, OK, 2007 070069OK1	WG	13	123	4	492	7	0.011	ND	0.011	070069
(Graking)							0.010	ND	0.010	
Mannsville, OK, 2007 070069OK2	WG	13	122– 123	4	491	7	0.012	ND	0.012	070069
(Beckett)							0.010	ND	0.010	
Claytonville, TX, 2007 070069TX	WG	13	123– 124	4	493	7	0.011	ND	0.011	070069
(Beckett)							0.011	ND	0.011	
Chula, GA,	WG	13	123-	4	496	0	< 0.01	ND	< 0.01	070069
2007 070069GA1			125				0.011	ND	0.011	
(Sumner)						1	0.010	ND	0.010	
							0.011	ND	0.011	
						3	< 0.01	ND	< 0.01	
							< 0.01	ND	< 0.01	
						7	ND	ND	< 0.01	
							ND	ND	< 0.01	
						14	ND	ND	< 0.01	
							ND	ND	< 0.01	

^a Total residue = XDE-175-J + XDE-175-L.

Spinetoram Spinetoram

Table 34-2 Residues of spinetoram and metabolites from supervised trials on pecan in the USA (for estimation of STMR)

	Form	Aj	plication		Total/	PHI,		Res	idue, mg	/kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175- L	ND- J	NF-J	Total a	No.
GAP, USA, Tree Nuts	WG		26– 123	4	490	14						
Scott, AR, 2007 070069AR	WG	13	123– 126	4	495	7	ND	ND	ND	ND	< 0.02	070069
(Stuart)							ND	ND	ND	ND	< 0.02	
Albany, GA, 2007 070069GA2	WG	13	122– 126	4	495	7	ND	ND	ND	ND	< 0.02	070069
(Sumner)							ND	ND	ND	ND	< 0.02	
Madill, OK, 2007 070069OK1	WG	13	123	4	492	7	0.011	ND	ND	ND	0.021	070069
(Graking)							0.010	ND	ND	ND	0.020	
Mannsville, OK, 2007 070069OK2	WG	13	122- 123	4	491	7	0.012	ND	ND	ND	0.022	070069
(Beckett)							0.010	ND	ND	ND	0.020	
Claytonville, TX, 2007 070069TX	WG	13	123– 124	4	493	7	0.011	ND	ND	ND	0.021	070069
(Beckett)							0.011	ND	ND	ND	0.021	
Chula, GA,	WG	13	123-	4	496	0	< 0.01	ND	ND	ND	< 0.02	070069
2007 070069GA1			125				0.011	ND	ND	ND	0.021	
(Sumner)						1	0.010	ND	ND	ND	0.02	
							0.011	ND	ND	ND	0.021	
						3	< 0.01	ND	ND	ND	< 0.02	
							< 0.01	ND	ND	ND	< 0.02	
						7	ND	ND	ND	ND	< 0.02	
							ND	ND	ND	ND	< 0.02	
						14	ND	ND	ND	ND	< 0.02	
							ND	ND	ND	ND	< 0.02	

^a Total residue = XDE-175-J + XDE-175-L + ND-J + NF-J.

Animal Feedstuffs

Table 35-1 Residues of spinetoram in sugar beet leaves or tops from supervised trials on sugar beet in the USA (for estimation of maximum residue level)

Location, year	Form	Aŗ	plication		Total/	PHI,	Residue, mg/kg			Report
(Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE-175-J	XDE- 175-L	Total ^a	No.
GAP, US, Root vegetables	SC		53-70	4	281	7				
S	SUGAR I	ВЕЕТ ТОР	S/LEAVES	S						
Olivia, MN, 2004	SC	24–25	70–71	4	282	3	0.089	0.025	0.114	040063
(Beta) Edgeley, ND,	SC	24–25	70–71	4	281	3	0.153	0.047	0.200	040063
2004 (Beta)							0.097	0.029	0.126	
Levelland, TX, 2004	SC	19	70–72	4	284	3	0.073	0.013	0.086	040063
(Wrangler)							0.062	0.013	0.075	
Porterville, CA, 2004	SC	19	70–71	4	283	3	0.063	0.013	0.076	040063
(Beta 4030R)							0.081	0.018	0.099	
Ephrata, WA, 2004	SC	21	70–71	4	283	0	0.589	0.190	0.779	040063
(Blazer)							0.542	0.172	0.714	
						1	0.129	0.038	0.167	
							0.175	0.054	0.229	
						3	0.111	0.033	0.144	
							0.122	0.035	0.157	
						7	0.024	< 0.01	0.024	
							0.019	< 0.01	0.019	
						14	< 0.01	ND	< 0.01	
							0.011	ND	0.011	
Michigan, 2007	SC	21–22	69–72	4	283	3	0.085	0.016	0.101	ARAP 07D-001
(Not specified)							0.168	0.031	0.199	0.2 001

^a Total residue = XDE-175-J + XDE-175-L.

Table 35-2 Residues of spinetoram and metabolites in sugar beet leaves or tops from supervised trials on sugar beet in the USA (for estimation of STMR)

Location,	Form	Aı	pplication		Total/	PHI,		Resi	due, mg/	kg		Report
year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total ^a	No.
GAP, US, Root vegetables	SC		53-70	4	281	7						
SU	SUGAR BEET TOPS/LEAVES											
Olivia, MN,	SC	24–	70–71	4	282	3	0.089	0.025	0.052	0.069	0.235	040063

Spinetoram Spinetoram

Location,	Form	Aj	pplication		Total/	PHI,		Res	idue, mg/	'kg		Report
year (Variety)		g ai/hL	g ai/ha	No	season, g ai/ha	days	XDE- 175-J	XDE- 175-L	ND-J	NF-J	Total ^a	No.
2004 (Beta)		25					0.055	0.016	0.046	0.046	0.163	
Edgeley, ND, 2004	SC	24– 25	70–71	4	281	3	0.153	0.047	0.095	0.101	0.396	040063
(Beta)		23					0.097	0.029	0.072	0.067	0.265	
Levelland,	SC	19	70–72	4	284	3	0.073	0.013	0.101	0.167	0.354	040063
TX, 2004 (Wrangler)							0.062	0.013	0.090	0.141	0.306	
Porterville,	SC	19	70–71	4	283	3	0.063	0.013	0.175	0.231	0.482	040063
CA, 2004 (Beta 4030R)							0.081	0.018	0.165	0.204	0.468	
Ephrata, WA,	SC	21	70–71	4	283	0	0.589	0.190	0.249	0.144	1.172	040063
2004 (Blazer)							0.542	0.172	0.214	0.111	1.039	
						1	0.129	0.038	0.161	0.271	0.599	
							0.175	0.054	0.165	0.316	0.71	
						3	0.111	0.033	0.125	0.343	0.612	
							0.122	0.035	0.125	0.347	0.629	
						7	0.024	< 0.01	0.038	0.171	0.233	
							0.019	< 0.01	0.032	0.133	0.184	
						14	< 0.01	ND	0.016	0.048	0.074	
							0.011	ND	0.019	0.049	0.079	
Michigan, 2007	SC	21– 22	69–72	4	283	3	0.085	0.016	0.174	0.121	0.396	ARAP 07D-
(Not specified)		22					0.168	0.031	0.261	0.214	0.674	001

 $^{^{}a}$ Total residue = XDE-175-J + XDE-175-L + ND-J + NF-J.

 $Table\ 36\text{--}1\ Residues\ of\ spinetoram\ and\ metabolites\ in\ almond\ hulls\ from\ supervised\ trials\ on\ almond\ in\ the\ USA$

	Form	A	pplication		Total/	PHI,	R	esidue, mg/l	ζg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	year g ai/ha	days	XDE- 175-J	XDE- 175-L	Total ^a	No.
GAP, USA, Tree Nuts	WG		26–123	4	490	14				
Terra Bella, CA, 2007 070069CA1 (Carmel)	WG	13	117– 144	4	532	7	0.695	0.135	0.830	070069
Wasco, CA, 2007 070069CA2 (Price)	WG	13	117– 130	4	501	7	0.378	0.083	0.461	070069
Earlimart, CA, 2007 070069CA3 (Fritz)	WG	13	117– 133	4	491	7	0.705 0.358	0.146	0.851	070069

	Form	Aj	pplication		Total/	PHI,	R	esidue, mg/l	ζg	Report
Location, year (Variety)		g ai/hL	g ai/ha	No	year g ai/ha	days	XDE- 175-J	XDE- 175-L	Total ^a	No.
Sultana, CA, 2007	WG	13	123– 124	4	494	7	1.901	0.419	2.320	070069
070069CA4 (Carmel)							1.394	0.306	1.700	
Sanger, CA, 2007	WG	13	110– 124	4	480	7	0.737	0.137	0.874	070069
070069CA5 (Neplus)							0.702	0.129	0.831	
Sanger, CA,	WG	13	123-	4	495	0	1.326	0.246	1.572	070069
2007 070069CA6			123				1.366	0.281	1.647	
(Nonpareil)						1	1.182	0.216	1.398	070069
							1.270	0.234	1.504	
						3	0.875	0.173	1.048	070069
							0.883	0.171	1.054	
						7	0.858	0.190	1.048	070069
							0.883	0.202	1.085	
						14	0.525	0.107	0.632	070069
							0.616	0.134	0.750	

^a Total residue = XDE-175-J + XDE-175-L.

Table 36-2 Residues of spinetoram and metabolites in almond hulls from supervised trials on almond in the USA (for estimation of STMR)

	Form	A	pplication		Total/	PHI,		Res	idue, mg/	'kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	year g ai/ha	days	XDE- 175-J	XDE- 175- L	ND-J	NF-J	Total	No.
GAP, USA, Tree Nuts	WG		26– 123	4	490	14						
Terra Bella, CA, 2007 070069CA1 (Carmel)	WG	13	117– 144	4	532	7	0.695	0.135	0.128	0.308	1.27 0.88	070069
Wasco, CA, 2007 070069CA2 (Price)	WG	13	117– 130	4	501	7	0.378	0.083	0.040	0.051	0.55	070069
Earlimart, CA, 2007 070069CA3 (Fritz)	WG	13	117– 133	4	491	7	0.705	0.146	0.086	0.102	1.04 0.52	070069
Sultana, CA, 2007 070069CA4 (Carmel)	WG	13	123– 124	4	494	7	1.901	0.419	0.152	0.177 0.124	2.65	070069
Sanger, CA, 2007 070069CA5 (Neplus)	WG	13	110– 124	4	480	7	0.737	0.137	0.094	0.105 0.108	1.07	070069

Spinetoram Spinetoram

	Form	A	pplication		Total/	PHI,		Res	idue, mg/	kg		Report
Location, year (Variety)		g ai/hL	g ai/ha	No	year g ai/ha	days	XDE- 175-J	XDE- 175- L	ND-J	NF-J	Total	No.
Sanger, CA,	WG	13	123-	4	495	0	1.326	0.246	0.098	0.095	1.77	070069
2007 070069CA6			123				1.366	0.281	0.112	0.111	1.87	
(Nonpareil)						1	1.182	0.216	0.098	0.097	1.59	070069
							1.270	0.234	0.107	0.101	1.71	
						3	0.875	0.173	0.092	0.135	1.28	070069
							0.883	0.171	0.093	0.125	1.27	
						7	0.858	0.190	0.084	0.091	1.22	070069
							0.883	0.202	0.080	0.079	1.25	
						14	0.525	0.107	0.055	0.088	0.78	070069
							0.616	0.134	0.068	0.097	0.91	

^a Total residue = XDE-175-J + XDE-175-L + ND-J + NF-J.

FATE OF RESIDUES IN STORAGE AND PROCESSING

In storage

No information was provided to the Meeting as spinetoram is not intended for use in stored products.

In processing

The Meeting received information on processing of oranges to juice and oil, and apples to juice and puree (sauce).

Orange

An orange processing study was conducted in the USA in 2007 where orange samples were processed to dried pulp, juice, and oil with corresponding processing factors calculated (McKellar, 2008; Report ARAP 07D-001).

Orange trees were treated three times with a SC formulation containing 120 g/L spinetoram, at the exaggerated rate of 350 g ai/ha, equivalent to 5× the GAP rate. Applications were made at intervals of 4 days, using low spray volume application. Samples (approximately 200 kg) of mature orange fruits from the control and treated plots were harvested at the PHI of 1 day and shipped at ambient temperature, to the processing facility. The following processed commodities were prepared, following procedures that simulated commercial practices: orange peel, pulp, dried pulp, juice, and oil.

<u>Peel:</u> After inspection of the samples, an aliquot of the oranges were removed and peeled by hand to generate orange peel, from which a representative sample fraction was removed, packaged, labelled and placed in frozen storage.

Oil: The remaining oranges were washed for 5 minutes. Approximately 3 kg of oranges per batch were abraded for 1 minute. The scarified fruit was weighed and retained for juice processing. The collected oil-water emulsion was screened using a ca. 180 µm screen to separate any fragments from the oil-water emulsion. The separated fragments were set aside for later addition to the shredded peel. The first run oil-water emulsion was processed through the cream separator and IEC centrifuge to separate the oil. The free oil was removed and measured with a 50 mL volumetric pipette and frozen. The residual emulsion was then frozen overnight, thawed, centrifuged and removed with a 50 mL volumetric pipette and added to the oil collected the previous day. The entire collected combined sample of the oil recovered from both processing days was weighed and then packaged, labelled and placed in frozen storage.

<u>Juice and peel/pulp:</u> An aliquot of the scarified oranges were weighed and transferred to the juice extractor to recover the juice from the peel. The juice and peel/pulp recovered were weighed and a representative orange pulp sample fraction was removed, packaged, labelled and placed in frozen storage.

The collected juice was transferred to the pulper finisher and screened using ca. 1.19 mm screen to remove vesicular membranes, seeds, segment membranes and peel fragments from the juice. The collected rag and seeds were set aside for later addition to the shredded peel. A representative sample of the fresh filtered juice was removed, packaged, labelled and placed in frozen storage for the required sample fraction. The remaining juice was discarded.

<u>Dried pulp:</u> The peel from the juice extractor was shredded and the shredded peel was combined with the scarified flavedo from the scarification process and the rag and seeds from the juice finisher extraction process to generate wet peel. Lime was added to the wet peel and mixed for 17 minutes. The limed peel was pressed in a fruit press and the expressed liquid weighed, checked for pH and Brix and discarded. The wet peel/pulp was placed on the air drier and dried to below 10% moisture. The dried pulp was milled using a hammermill. A representative sample of the dried pomace was removed, packaged, labelled and placed in frozen storage. The remaining dried pomace was discarded.

All samples were stored up to 87 days before analysis, which is within the storage time of the stability study conducted separately.

Residues of spinetoram and its metabolites were determined in whole orange samples collected from plots treated at 5× the GAP, and each processed fraction prepared, using the method GRM 05.04, with an LOQ and LOD of 0.01 mg/kg and 0.003 mg/kg, respectively.

Residues in orange and processed fractions are summarized in Table 37. Residues are reported as the sum of residues of XDE-175-J, XDE-175-L, N-demethyl-175-J and N-formyl-175-J.

Processed	Spinetora	m residue	Spinetoram + t	wo metabolites	
Product	Residues mg/kg	Processing factor	Residues mg/kg	Processing factor	Reference
Orange (RAC)	0.201		0.280		McKellar, 2008
Peel	0.279	1.4	0.384	1.4	Report ARAP 07D-001
Pulp after juicing	0.061	0.30	0.091	0.33	
Dried pulp	0.478	2.4	0.478	2.3	
Juice	< 0.01	< 0.05	< 0.02	< 0.07	
Oil	28.8	143	32.23	115	

Table 37 Residues of spinetoram and its metabolites in orange and orange processed products

Apple

Two processing trials on apples were carried out in France in 2005 (Dolder and Schelle, 2006; Report No. 050028). Apple trees were treated with four applications of the WG formulation containing 250 g/kg spinetoram at the rate of 100 g ai/ha. Applications were made at approximately 7-day intervals with a calibrated sprayer simulating a commercial application. Water volumes used were approximately 800–1200 L/ha for apples. Samples were collected 7 days after the last application.

About 60 kg of mature apples were sent to the processing facility for processing into dry pomace, juice, sauce and canned apples. Processing procedures simulated commercial practices. The fruits were washed manually with tap water. The following processed samples were prepared:

Apple juice: The washed apples were crushed in an apple mill and thereafter pressed in a fruit pack press to get wet pomace and raw juice. A part of the wet pomace was dried overnight in an oven and the sample of dry pomace was weighed to determine the dry weight.

The raw juice was filtered and enzymes were added to the filtered juice. The time and temperature during enzyme treatment was recorded. The cleared juice was filtered once or twice and, after filtration, the clear juice was filled in glass bottles which were closed airtight and pasteurized at 81–89 °C and cooled down. The cooled clear juice samples were stored deep frozen until analysis.

Apple puree: Washed fruits were cut in small pieces before putting them into a stainless steel pot. After adding water to the apples (about 150–200 mL per kg of apples), the mixture was heated until the fruits were soft. The fruits were then passed through a food mill-sieve combination in order to remove the peel and cores. A part of the raw stewed sauce was filled in preserving glass bottles. The glass bottles were closed airtight and pasteurized at 80–89 °C for 30 minutes. After cooling down, the apple puree samples were stored deep frozen until analysis.

<u>Canned apples</u>: Washed fruits were peeled manually with a knife and the peel and cores weighed and removed. The peeled fruits were cut into halves or quarters and filled in suitable glass containers. A heated sugar-water solution was put into the container filled with fruits. The glass containers were then closed airtight and pasteurized for 30–35 minutes at 80–85 °C. After heating, the glass containers were opened and sample of canned fruits taken. After cooling down, the samples were stored deep frozen until analysis.

Residues of spinetoram and metabolites were determined using method GRM 05.04 with an LOQ of 0.01 mg/kg and an LOD of 0.003 mg/kg. The results are summarized in Table 37. Residues reported are the sum of residues of XDE-175-J, XDE-175-L, *N*-demethyl-175-J, and *N*-formyl-175-J.

Processed	Spinetora	m residue	Spinetoram +	two metabolites	
product	Residues mg/kg	Processing factor	Residues mg/kg	Processing factor	Reference
Apple (RAC)	0.020		0.041		Dolder and Schelle,
Juice	< 0.01	< 0.50	< 0.02	< 0.48	2008
Dry pomace	0.212	10.6	0.331	7.9	Report 050028
Puree (sauce)	0.012	0.60	0.022	0.53	
Canned apples	< 0.01	< 0.50	< 0.02	< 0.48	
Apple (RAC)	0.041		0.051		Dolder and Schelle,
Juice	< 0.01	< 0.24	< 0.02	< 0.39	2008
Dry pomace	0.173	5.6	0.210	4.1	Report 050028
Puree (sauce)	0.015	0.29	0.022	0.42	
Canned apples	< 0.01	< 0.24	< 0.02	< 0.39	
		Mean		Mean	
Juice		< 0.37		< 0.44	
Dry pomace		8.1		6.0	

0.47

< 0.44

Table 38 Residues of spinetoram and metabolites in apples and processed fractions

0.45

< 0.50

RESIDUES I ANIMAL COMMODITIES

Farm animal feeding studies

Lactating cows

Puree (sauce)

Canned apples

Six groups of lactating Holstein dairy cows (3–5 years old; body weight 474-656 kg at receipt) were dosed daily for 29 consecutive days via gelatine capsules containing spinetoram (Dolder and Schelle, 2007; Report 060052) (Table 39). Each group consisted of at least 3 cows. Four of the groups were dosed with test substance A, containing 26% XDE-175-J, 6% XDE-175-L, 30% *N*-formyl-175-J and

28% *N*-demethyl-175-J. One group was given test substance B containing only XDE-175-J and XDE-175-L (86% total). The remaining group was the control group which was dosed with empty gelatine capsules.

Table 39 Animal Dosing Group	Table 39	Animal	Dosing	Group
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	Ni-mb f		Dose, ppm in diet							
Dose Level	Number of Cows	Test Sub	stance A	Test Substance B						
	Cows	Total ^a	Spinetoram	Spinetoram						
Control	4	-	-	-						
0.3×	3	1.2	0.4	-						
1×	12	3.7	1.3	-						
3×	3	11.5	3.8	-						
10×	3	38.6	12.9	-						
10×	3	-		37.6						

^a Total dose= sum of XDE-175-J, XDE-175-L, N-demethyl-175-J, and N-formyl-175-J.

Milk was collected twice daily (AM and PM) via surge milking machines and placed into labelled collection containers. The milk from AM and PM milking of each treated animal was pooled in proportion to the amount of milk collected at each of the two daily milkings. Milk samples from the control animals were also pooled in this manner. Only milk from the same cow for a given study day's AM and PM milking was combined to form a pooled sample. The calculated amounts of PM and AM milk were combined to form the pooled 250 g sample. The average daily milk production in dose groups ranged between 26 and 29 kg during the dosing period.

Milk samples from Days 14 and 28 were separated into cream and skim milk from the three cows each in the control and $1\times$ dose groups that were not designated for use in the depletion phase and the three cows in each of the $10\times$ -A and $10\times$ -B dose groups.

Three cows from each group were sacrificed at the end of a 29-day dosing period in order to determine residue levels in tissues (muscle, fat, liver and kidney). One extra cow in the control group along with nine extra cows in the 1X dose group were used to evaluate residue depletion in milk and tissues during a withdrawal period following the completion of the dosing period. Of the nine cows in the 1x dose group, three cows were assigned to each of three sacrifice dates following completion of the dosing period. The three sacrifice dates in the depletion phase were 14, 28 and 56 days after completion of dosing. The one extra cow in the control group used in the withdrawal phase of the study was sacrificed at 56 days after completion of the dosing period. Samples of liver, kidney, muscle and fat (approximately 1 kg each, if available, of mesenterial, perirenal and subcutaneous) were collected from all animals, cubed, and frozen until analysis. Samples were analysed within 20 days of collection; therefore, no storage stability studies were conducted.

Milk and tissue samples were analysed for residues of XDE-175-J, XDE-175-L, *N*-formyl-175-J and *N*-demethyl-175-J by method GRM 6.08 with LOQ of 0.01 mg/kg for each of the analytes.

Residues found in milk samples collected during the dosing phase of the study are summarized in Table 40. No residue was detected in any of the control samples throughout the dosing phase. Residues in the 0.3× dose group were generally between the LOD (0.003 mg/kg) and LOQ (0.01 mg/kg) throughout the dosing period. The average spinetoram residues in milk for study days 10–28 for the 0.3×, 1×, 3×, 10×-A and 10×-B dose groups were < 0.01, 0.018, 0.072, 0.253, and 0.711 mg/kg, respectively. Residues found in milk from cows given the parent compounds only were about three times higher than those in milk from cows given a mixture of the parent, *N*-demethyl-175-J and *N*-formyl-175-J, reflecting the ratio of the parent in the test substances.

Spinetoram Spinetoram

Table 40 Mean spinetoram residues in milk during dosing phase

Dose		Residues of XDE-175-J and XDE-175-L in milk on specific dosing day (mg/kg)										
Group	-2	3	7	10	14	16	18	20	22	24	26	28
Control a	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
0.3×	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
1×	< 0.01	0.013	0.022	0.021	0.019	0.017	0.019	0.017	0.018	0.018	0.019	0.017
3×	< 0.01	0.049	0.065	0.076	0.077	0.071	0.070	0.067	0.076	0.087	0.076	0.080
10×-A	< 0.01	0.21	0.20	0.23	0.23	0.17	0.25	0.25	0.37	0.28	0.30	0.29
10×-B	< 0.01	0.059	0.69	0.70	0.70	0.69	0.72	0.71	0.73	1.01	0.94	0.87

Dose	Resid	Residues of XDE-J, XDE-L, N-demethyl-175-J and N-formy-175-J in milk on specific dosing day (mg/kg)										
Group	-2	3	7	10	14	16	18	20	22	24	26	28
Control ^a	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02
0.3×	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02
1×	< 0.02	0.023	0.032	0.031	0.027	0.027	0.029	0.027	0.028	0.028	0.029	0.027
3×	< 0.02	0.059	0.075	0.086	0.087	0.081	0.080	0.077	0.086	0.097	0.086	0.090
10×-A	< 0.02	0.22	0.22	0.26	0.26	0.19	0.28	0.26	0.40	0.30	0.32	0.32
10×-B	< 0.02	0.60	0.70	0.71	0.71	0.70	0.73	0.72	0.74	1.02	0.95	0.88

^a Includes cows designated for the depletion phase.

Samples from milk collected on days 14 and 28 were separated into skim milk and cream. Table 41 shows a summary of the residues. No residues were detected in skim milk from the $1\times$ dose group. In the $10\times$ -A and $10\times$ -B dose groups average total residues in skim milk ranged from just below the LOQ to 0.075mg/kg. In all of the dose groups on both study days residues in cream were much higher than the residues in skim milk. Mean total residues from the $1\times$ dose group in cream were 0.187 and 0.231 mg/kg. Mean total residues in the $10\times$ -A and $10\times$ -B groups ranged 0.64–1.95 and 3.11–5.84 mg/kg. These results indicate that residues of spinetoram in whole milk preferentially partition into cream. The median ratio of spinetoram residues in cream to those in whole milk is 6.6.

Table 41 Mean spinetoram residues in skim milk and cream

Б.	Residues of XDE-175-J and XDE-175-L on specific dosing day (mg/kg)								
Dose Group	Whole	Milk	Cre	am	Skim Milk				
Group	Day 14	Day 28	Day 14	Day 28	Day 14	Day 28			
1X	0.027	0.023	0.177	0.221	< 0.01	< 0.01			
10X-A	0.12	0.29	0.616	1.84	< 0.01	0.028			
10X-B	0.70	0.87	3.09	5.81	0.060	0.065			

Dana	Residues of XDE-J, XDE-L, N-demethyl-175-J and N-formy-175-J on specific dosing day (mg/kg)								
Dose Group	Whole	e Milk	Cre	am	Skim Milk				
Group	Day 14 Da		Day 14	Day 28	Day 14	Day 28			
1X	0.037	0.033	0.187	0.231	< 0.02	< 0.02			
10X-A	0.13	0.32	0.642	1.95	< 0.02	0.038			
10X-B	0.71	0.88	3.11	5.84	0.070	0.075			

Residues found in kidney, liver, muscle, and fat samples collected from animals sacrificed upon completion of the dosing phase are presented in Table 42. With one exception, no residues were detected in the control samples. One kidney control sample contained residues below the LOQ possibly due to contamination. All tissues from treated cows contained residues and they increased from the lowest to highest dose groups. Residue concentrations were lowest in the muscle followed by kidney, liver, and fat. Residues in fat were significantly higher than residues in the other tissues. Across all the dose levels the mesenterial and perirenal fat samples had similar residue concentrations which were roughly twice the concentrations found in the subcutaneous fat. As expected, the composite fat samples, which consisted of equal parts of the three fats collected, yielded residues very close to the average residue concentration found in the three fats. These results indicate that residues of spinetoram tend to accumulate in fatty tissue.

Table 42 Spinetoram residues in kidney, liver, muscle and fat following a dosing period of 29 days

ъ		Residue	es of XDE-J and	l XDE-L on speci	fic dosing day	(mg/kg)	
Dose Group	Kidney	Liver	Muscle	Subcutaneous Fat	Mesenterial Fat	Perirenal Fat	Composite Fat ¹
Control	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
0.3×	< 0.01	< 0.01	< 0.01	0.077	0.093	0.10	0.082
	< 0.01	< 0.01	< 0.01	0.063	0.10	0.11	0.11
	< 0.01	< 0.01	< 0.01	0.021	0.10	0.091	0.075
Maximum	< 0.01	< 0.01	< 0.01	0.077	0.10	0.11	0.11
Mean	< 0.01	< 0.01	< 0.01	0.054	0.10	0.10	0.090
1×	0.016	0.057	< 0.01	0.087	0.44	0.50	0.38
	0.040	0.052	0.043	0.52	0.59	0.69	0.55
	0.011	0.040	< 0.01	0.14	0.29	0.34	0.26
Maximum	0.040	0.057	0.043	0.52	0.59	0.69	0.55
Mean	0.022	0.050	0.021	0.25	0.44	0.51	0.39
3×	0.074	0.11	0.086	0.74	1.22	1.41	1.25
	0.068	0.11	0.019	0.57	0.83	0.64	0.39
	0.048	0.11	0.010	0.035	0.25	0.20	0.13
Maximum	0.074	0.11	0.086	0.74	1.22	1.41	1.25
Mean	0.063	0.11	0.038	0.45	0.76	0.75	0.59
10×-A	0.29	0.47	0.082	0.15	0.29	0.84	0.35
	0.25	0.41	0.13	1.14	3.54	3.07	2.68
	0.30	0.21	0.24	2.96	3.69	3.64	3.26
Maximum	0.30	0.47	0.24	2.96	3.69	3.64	3.26
Mean	0.28	0.36	0.15	1.42	2.51	2.52	2.10
10×B	1.74	1.29	0.53	8.69	11.23	16.52	15.29
	0.81	0.62	0.49	4.69	8.37	11.04	9.72
	1.01	2.39	0.41	2.32	9.22	14.77	8.85
Maximum	1.74	2.39	0.53	8.69	11.23	16.52	15.29
Mean	1.19	1.43	0.48	5.24	9.61	14.14	11.28

Dose	Residues o	of XDE-J, XDE	-L, N-demethyl-	-175-J and <i>N</i> -forr	ny-175-J on sp	ecific dosing da	ıy (mg/kg)
Group	Kidney	Liver	Muscle	Subcutaneous Fat	Mesenterial Fat	Perirenal Fat	Composite Fat ^a
Control	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02
	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02
	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02
0.3×	< 0.02	< 0.02	< 0.02	0.087	0.10	0.11	0.092
	< 0.02	0.010	< 0.02	0.073	0.11	0.12	0.12
	< 0.02	< 0.02	< 0.02	0.031	0.11	0.10	0.085
Maximum	< 0.02	< 0.02	< 0.02	0.087	0.11	0.12	0.12
Mean	< 0.02	< 0.02	< 0.02	0.064	0.10	0.11	0.10
1×	0.043	0.10	< 0.02	0.097	0.46	0.53	0.40
	0.075	0.095	0.053	0.54	0.61	0.72	0.57
	0.036	0.092	< 0.02	0.15	0.30	0.36	0.27
Maximum	0.075	0.10	0.053	0.54	0.61	0.72	0.57
Mean	0.052	0.097	0.031	0.26	0.46	0.54	0.41
3×	0.13	0.22	0.11	0.81	1.29	1.49	1.32
	0.12	0.19	0.029	0.60	0.86	0.66	0.40
	0.091	0.21	0.033	0.045	0.27	0.23	0.15
Maximum	0.13	0.22	0.11	0.81	1.29	1.49	1.33
Mean	0.12	0.20	0.057	0.48	0.81	0.79	0.63
10×-A	0.57	0.93	0.17	0.18	0.34	0.95	0.42
	0.44	0.73	0.18	1.22	3.72	3.23	2.80
	0.40	0.36	0.29	3.12	3.89	3.84	3.44
Maximum	0.57	0.93	0.29	3.12	3.89	3.84	3.44
Mean	0.47	0.68	0.21	1.51	2.65	2.67	2.22
10×-B	1.80	1.37	0.54	8.75	11.29	16.61	15.36
	0.84	0.66	0.50	4.71	8.41	11.08	9.76
	1.08	2.57	0.44	2.36	9.30	14.84	8.92
Maximum	1.80	2.57	0.54	8.75	11.29	16.61	15.36
Mean	1.24	1.54	0.50	5.28	9.67	14.18	11.34

^a Contained equal parts of subcutaneous, mesenterial, and perirenal fats.

Residues found in milk samples collected from cows during the depletion phase are shown in Table 43. With one exception, residues were not detectable in milk by the fourth day after the last dose was administered. Concentrations just above the LOD were detected in one cow through the ninth day after the final dose. No further residue was detected beyond that point. No residue was ever detected in milk from the cow in the control group.

Table 43 Residues of spinetoram in milk during the depletion phase

Davis	Re	Residues of XDE-J, XDE-L, N-demethyl-175-J and N-formy-175-J on days after final dose (mg/kg)										
Dose Group	0	2	4	6	9	13	20	27	34	41	48	55
Group	(29)	(31)	(33)	(35)	(38)	(42)	(49)	(56)	(63)	(70)	(77)	(84)
Control	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02
1X	0.036	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	-	-	-	-	-	-
	0.029	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	-	-	-	-	-	-
	0.020	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	-	-	-	-	-	-

D	Re	sidues of	XDE-J, X	KDE-L, <i>N</i>	-demethy	l-175-J ar	nd N-form	ıy-175-J o	n days af	ter final d	ose (mg/l	kg)
Dose Group	0	2	4	6	9	13	20	27	34	41	48	55
Group	(29)	(31)	(33)	(35)	(38)	(42)	(49)	(56)	(63)	(70)	(77)	(84)
	0.025	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	-	-	-	-
	0.024	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	-	-	-	-
	0.027	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	-	-	-	-
	0.024	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02
	0.029	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02
	0.026	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02
Mean	0.027	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02

Days in parentheses are days after the beginning of dosing period.

Residues found in kidney, liver, muscle, and fat samples collected during the depletion phase are presented in Table 44. Residue concentrations were lowest in the muscle followed by kidney, liver, and fat. On the day the last dose was given, the mesenterial and perirenal fat samples had similar residue concentrations which were roughly twice the concentrations found in the subcutaneous fat. However, in the fat samples taken 14 days after the last dose was given, the residue in the subcutaneous fat was higher than in the other two fat samples. As expected, the composite fat samples, which consisted of equal parts of the three fats collected, yielded residues very close to the average residue concentration found in the three fats. Residue decline continued through 28-days after the last dose. No residue was detected in kidney, liver or muscle from any cow by 28 days after the final dose or in fat 56 days following the final dose.

Table 44 Residues of spinetoram in tissues during the depletion phase

Days After	Study	Residues	of XDE-J, XD	E-L, <i>N</i> -demeth	nyl-175-J and a (mg/kg)	N-formy-175-J	on days after	final dose
Final Dose	Day				Sub.	Mes.	Per.	Comp.
		Kidney	Liver	Muscle	Fat	Fat	Fat	Fat
0	29	0.043	0.103	0.020	0.097	0.46	0.53	0.40
0	29	0.075	0.095	0.053	0.55	0.61	0.72	0.57
0	29	0.036	0.092	0.023	0.15	0.30	0.36	0.27
	Mean	0.052	0.097	0.032	0.26	0.46	0.54	0.41
14	43	< 0.02	< 0.02	< 0.02	0.26	0.23	0.14	0.22
14	43	< 0.02	< 0.02	< 0.02	0.051	< 0.02	0.021	< 0.02
14	43	< 0.02	< 0.02	< 0.02	0.10	0.055	0.026	0.063
	Mean	< 0.02	< 0.02	< 0.02	0.14	0.10	0.062	0.10
28	57	< 0.02	< 0.02	< 0.02	0.022	0.022	< 0.02	< 0.02
28	57	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02
28	57	< 0.02	< 0.02	< 0.02	0.044	0.024	0.023	0.037
	Mean	< 0.02	< 0.02	< 0.02	0.029	0.022	< 0.02	0.025S
56	85	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02
56	85	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02
56	85	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02
	Mean	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02
Control			_				_	
56	85	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02

Cattle, except those in the control group, were dosed with a total dose of 3.7 ppm in the diet.

Note: Residues are reported as sum of XDE-175-J, XDE-175-L, *N*-demethyl-175-J and *N*-formyl-175-J concentrations. All concentrations are reported in mg/kg.

Laying hens

The Meeting received no information on poultry feeding studies.

ADDITIONAL INFORMATION

The currently existing national residue definitions for plant and animal commodities are:

Australia: XDE-175-J and XDE-175-L;

Canada: XDE-175-J, XDE-175-L, N-demethyl-175-J and N-formyl-175-J (for risk assessment in poultry commodities, 3'-O-deethyl-175-J and L, and O-demethyl-175-L are added);

New Zealand: XDE-175-J and XDE-175-L; and

USA: XDE-175-J, XDE-175-L, N-demethyl-175-J and N-formyl-175-J;

APPRAISAL

Spinetoram, a multi-component tetracyclic macrolide in the class of spinosyn insecticides, consists of two components shown below, present approximately in a three to one ratio. It was identified as a priority new compound at the 39th Session of the CCPR in 2007 (ALINORM 07/30/24—Rev.1) for evaluation by the 2008 JMPR. The Meeting received information on physical and chemical properties, animal and plant metabolism, environmental fate, analytical methods, storage stability, use patterns, supervised trials, processing and farm animal feeding.

XDE-175-J (Major component)

XDE-175-L (Minor component)

In this appraisal, the following abbreviated names were used for metabolites:

N-demethyl-175-J

N-formyl-175-J

3'-O-deethyl-175-J

4'-O-demethyl-175-L

Animal metabolism

The Meeting received information on the fate of orally-dosed spinetoram in lactating goats and laying hens.

When either ¹⁴C-XDE-175-J or ¹⁴C-XDE-175-L, uniformly labelled with ¹⁴C in the macrolide ring, was administered orally at a dose equivalent to a dietary concentration of 10–11 ppm to a <u>lactating goat</u> once a day for five consecutive days, 51% or 78% of the administered dose of ¹⁴C-XDE-175-J or ¹⁴C-XDE-175-L, respectively, were recovered in faeces. Radioactivity recovered in urine was insignificant; less than 0.2% of the administered dose.

Radioactivity started to appear in milk within 24 hours after the first application but cumulative milk sample contained only about 0.3% of the total administered dose of 14 C-XDE-175-J or 0.2% of that of 14 C-XDE-175-L. The maximum total radioactive residues were 0.047 mg/kg in parent equivalents for 14 C-XDE-175-L and 0.039 mg/kg 14 C-XDE-175-L.

Total radioactive residues (TRR) in tissues after sacrifice (21 h after the last dose) showed the tendency to be higher in fatty tissues, with 0.235 mg/kg and 0.119 mg/kg in parent equivalents in fat (after the administration of ¹⁴C -XDE-175-J and ¹⁴C-XDE-175-L, respectively), and 0.116 mg/kg and 0.099 mg/kg in liver. TRR in kidney, muscle and milk were much lower.

The primary residue was XDE-175-J or XDE-175-L (42-84% of TRR) in all tissues and milk, except liver, indicating that minimal metabolism had occurred. In liver, XDE-175-J or XDE-175-L was the primary residue but at lower levels (30 or 26% of TRR) with N-demethyl-175-J or -L at very low levels (<2% of the TRR), and an unidentified metabolite. No residue components other than the unchanged parent compounds were identified in milk, kidney or fat. Radioactive residues in muscle

also consisted primarily of XDE-175-J or XDE-175-L and much lesser amounts of what seemed to be the same unidentified metabolite as found in liver. There were many minor metabolites detected but all were less than 10% of the TRR. Unextracted radioactivity was less than 15% of the TRR in all samples except liver in which it was around 25%.

When either ¹⁴C-XDE-175-J or ¹⁴C-XDE-175-L, (uniformly labelled with ¹⁴C in the macrolide ring), was administered orally at a dose equivalent to a dietary concentration of 10 ppm to a group of <u>laying hens</u> once a day for seven consecutive days, 93% or 91% of the administered dose of ¹⁴C-XDE-175-J or ¹⁴C-XDE-175-L, respectively, were recovered in excreta.

Eggs and tissues contained a low proportion of the administered dose (< 3%) for both ¹⁴C-XDE-175-J and ¹⁴C-XDE-175-L. TRR in eggs increased over the experimental period and reached a maximum of 0.20 and 0.49 mg/kg for ¹⁴C-XDE-175-J and ¹⁴C-XDE-175-L, respectively, on day 7. TRR in the tissues were highest in abdominal fat (1.04 mg/kg for XDE-175-J, and 2.46 mg/kg for XDE-175-L), followed by skin with subcutaneous fat, liver, eggs and muscle. There is a tendency for radioactivity to be found at higher levels in tissues with higher fat content.

Unchanged XDE-175-J or XDE-175-L remained as the primary residue in the egg (58–49%) and tissues (45–70% of TRR) other than liver (13–12%). 3'-O-deethyl-175-J was detected in abdominal fat (1.8% of TRR) and in liver (18%) and O-demethyl-175-J was present in all tissues (3.2–6.5% of TRR) while 3'-O-deethyl-175-L and O-demethyl-175-L were found in all tissues and eggs (5.2–13% and 13–20% of TRR respectively). Unextracted radioactivity was less than 7% of the TRR in all samples except muscle and eggs in which it was 12–20%.

Limited metabolism of spinetoram was observed in ruminants and hens as the unchanged parent compound was the primary residue component in milk and all ruminant tissues as well as eggs and all avian tissues except liver for both XDE-175-J and XDE-175-L. Metabolism of spinetoram appears to be primarily through demethylation of the N-dimethyl moiety on the forosamine sugar to give the *N*-demethyl metabolite (goat) and dealkylation of the rhamnose sugar to give the *O*-deethyl and/or *O*-demethyl (two possible isomers) metabolites (hen).

Plant metabolism

The Meeting received information on the fate of spinetoram after foliar applications of ¹⁴C-XDE-175-J or ¹⁴C-XDE-175-L, uniformly labelled with ¹⁴C in the macrolide ring, on apple, lettuce and turnip representing the fruits, leafy crops and root crops respectively.

In all three crops tested, applied parent compounds decreased over test period. Among the two spinetoram components, XDE-175-L tended to be metabolized faster than XDE-175-J.

Apple

When a branch of <u>apple</u> tree with immature fruits was treated with single foliar application of either ¹⁴C-XDE-175-J or ¹⁴C-XDE-175-L at the rate of 1.8 kg ai/ha (4.8×) or 1.1 kg ai/ha (8.9×), respectively, apple fruits harvested seven days (PHI in US GAP) after the application contained 1.2 mg/kg or 0.36 mg/kg of radioactive residues. Washing of fruits (0–30 DAT) with acetonitrile and then dichloromethane removed 63–97% of TRR.

In the case of XDE-175-J application, in apples taken seven DAT, the parent compound was the major residue at 43% of the TRR with N-demethyl-175-J at 9.5% and N-formyl-175-J at 5.1% of the TRR. After the treatment with XDE-175-L, the parent compound was extensively degraded and metabolized and only 1.3% of the TRR remained as the parent compound with 1.0% of the TRR as N-demethyl-175-L and another 1.0% as N-formyl-175-L.

In the washed fruits obtained from the 14 C-XDE-175-J treated branch, peels contained 2–11% of TRR while pulp contained less than 1% (\leq 0.007 mg/kg). In washed fruits from 14 C-XDE-175-L treated branch, peel contained 6–33% of TRR while pulp contained less than 4%.

Unextractable radioactive residues were < 10% of TRR in or on all samples after extraction with a mixture of acetonitrile and water (80:20, v/v). Several minor metabolites were also detected in

the treated apples and leaves, each at $\leq 7.5\%$ of TRR. A multi-component mixture of extensively degraded compounds represented up to 39–77% of TRR but each component was less than 1% of TRR.

Comparison of radioactivity in treated fruits and shielded fruits indicates that translocation was negligible.

Lettuce

Red <u>leaf lettuce</u> was treated with either single or multiple foliar spray applications of ¹⁴C-XDE-175-J or ¹⁴C-XDE-175-J. For ¹⁴C-XDE-175-J, single applications were made at rates equivalent to 0.90 kg ai/ha while the same amount of the test compound was sprayed in three separate applications with the equal rate at weekly intervals. For ¹⁴C-XDE-175-L, plants were treated in a similar fashion but at a rate equivalent to 0.30 kg ai/ha. For both compounds, the applied amounts approximately correspond to four times the maximum seasonal rate on the label and reflect the ratio between XDE-175-J and XDE-175-L in spinetoram formulations.

Washing leaves (0-7 DAT) with dichloromethane and then ace tonitrile removed 76–96% of TRR.

The lettuce samples taken one day (PHI in US GAP) after the single application of XDE-175-J or XDE-175-L contained 34 mg/kg in parent equivalents or 7.6 mg/kg of radioactive residues, respectively. For treatment with XDE-175-J, the parent was 31%, N-demethyl-175-J was 20% and N-formyl-175-J was 11% of the TRR. For treatment with XDE-175-L, the parent was 12%, N-demethyl-175-L was 7.2% and N-formyl-175-L was 4.0% of the TRR. With the multiple applications, both TRR and percentage of these three compounds tended to be lower than with single applications of the same total rate for both parent compounds. Only 0.2–5.2% of TRR remained unextractable in all treated lettuce samples after extraction with acetonitrile/water (75:25, v/v).

Several minor metabolites were observed in the 14C-XDE-175-J and 14C-XDE-175-L treated lettuce at \leq 6% of TRR. A multi-component mixture of extensively degraded compounds represented up to 13–78% of TRR, each component at less than 3% of TRR.

Turnip

<u>Turnips</u> were treated with either a single or multiple foliar applications of ¹⁴C-XDE-175-J or ¹⁴C-XDE-175-L in the same manner as in the lettuce study. In turnip roots harvested three days after the application, TRR was quite low at 0.12 mg/kg for XDE-175-J treatment and 0.031 mg/kg for XDE-175-L treatment.

In ¹⁴C-XDE-175-J treated turnip tops, 59–91% of TRR were surface residues found in the dichloromethane and acetonitrile washings. In ¹⁴C-XDE-175-L treated turnip tops, surface residues were 39–80% of TRR.

Major components identified at three DAT were the parent compounds (XDE-175-J and -L), *N*-dimethyl-175-J and *N*-formyl-175-J in roots and tops but all less than 25% of the TRR.

Several minor metabolites assumed to be structurally similar to the parent compound were also observed in treated turnip roots and tops, each less than 4% of TRR. A multi-component mixture of extensively degraded compounds represented 10–74% of TRR; each compound at less than 1% TRR.

Metabolism of spinetoram was observed to be similar in the three crops studied—apple, lettuce and turnip—indicating that a common metabolism is expected for not only fruits, leafy vegetables and root vegetables but also other plants. It appears that three metabolic pathways are responsible for the breakdown of spinetoram in plants. The first one involves changes to the N-demethyl moiety on the forosamine sugar to give the N-demethyl and N-formyl metabolites. N-formyl metabolites were found only in plants. The second involves cleavage of the macrolide ring system at one or more positions, ultimately resulting in a complex residue mixture consisting of numerous components. The third, (applicable only to XDE-175-J), involves changes to the rhamnose sugar

producing the 3-O-deethyl and C9-pseudoaglycone-175-J metabolites. All the metabolites occurring as a result of changes in forosamine and rhamnose are further degraded via the second pathway.

Environmental fate in soil

The Meeting reviewed information on aerobic soil metabolism, aqueous photolysis and hydrolysis, and rotational crop study, as spinetoram was intended for protection of root vegetables.

Aerobic soil metabolism

Aerobic soil metabolism studies were conducted using ¹⁴C-XDE-175-J or ¹⁴C-XDE-175-L, uniformly labelled with ¹⁴C in the macrolide ring applied to various soils and incubated under aerobic conditions at 25, 20 or 10 °C. Under aerobic conditions, spinetoram applied to soil was degraded relatively rapidly. In all soils tested, XDE-175-L was degraded faster than XDE-175-J. After one year of incubation at 25 °C, 1.2–2.8% and 0.3–2.9% of applied XDE-175-J and XDE-175-L respectively, remained as the parent in US soils tested. In European soils (except the loamy sand), after 127 days of incubation at 20 °C, 2.0–4.9% and 1.4–5.0% of applied XDE-175-J and XDE-175-L respectively remained as the parent. Carbon dioxide was evolved slowly from all soils and accounted for 5.0–35% and 9.5–32% of the applied XDE-175-J and XDE-175-L respectively after one year at 25 °C, and 0.8–1.1% and 1.2–3.2% of the applied XDE-175-J and XDE-175-L respectively after 127 days at 20 °C.

Major degradation products, N-demethyl-175-J and N-demethyl-175-L were formed and then degraded during the study periods. As minor products (at or less than 10% of the applied dose), N-demethyl-N-nitroso-175-J, N-demethyl-N-nitroso-175-L, N-succinyl-175-J and N-succinyl-175-L were also formed and degraded. Many other degradates were formed but at very low concentrations.

While extractable radioactivity decreased, non-extractable radioactivity steadily increased to reach 22–29% and 32–37% of the applied XDE-175-J and XDE-175-L respectively after one year at 25 $^{\circ}$ C; and 5–15% and 11–24% of the applied XDE-175-J and XDE-175-L respectively after 127 days at 20 and 10 $^{\circ}$ C.

Aqueous photolysis

Under xenon light (simulating 40° N latitude summer sunlight) in aqueous buffer solution at pH 7 at 25 °C, XDE-175-J and XDE-175-L degraded rapidly with DT₅₀ of 0.5 days and 0.3 days respectively. Numerous (more than 70) minor degradates were observed after irradiation of XDE-175-J and -L. N-demethyl-175-L was observed as a major photodegradation product of XDE-175-L.

At test termination, greater than 90% of the applied amount remained as parent in the dark controls indicating that negligible transformation of the parent compounds occurred in the dark. No degradates were observed in the dark controls.

Aqueous hydrolysis

In sterile aqueous buffer solutions at pH 5 and 7, no degradation was observed for both XDE-J and XDE-175-L for 30 days at 20 °C. At pH 9, a degradate of XDE-175-J was observed but the concentration of XDE-175-J did not decrease below 89% and therefore, XDE-175-J can be regarded to be relatively stable, also at pH 9. After 30 days at pH 9, XDE-175-L decreased from 92% to 82% with N-demethyl-175-L as the major degrade at 12% at the end of the testing period. No minor degradates were detected. DT_{50} of XDE-175-L at pH 9 was calculated to be 154 days.

Residues in succeeding crops

In an outdoor confined rotation study, radish, lettuce and wheat were planted at 30, 120 and 365 days after the application of ¹⁴C-XDE-175-J or ¹⁴C-XDE-175-L at rate of 405 or 135 g ai/ha respectively to soil corresponding to the maximum seasonal rate and reflecting the ratio of these two active ingredients in spinetoram formulations.

TRRs were very low for all samples at all plant back intervals with the maximum at 0.085 mg/kg in parent equivalents. Unextractable residues in crops were less than 0.019 mg/kg.

Extraction of residues indicated that no greater than 0.065, 0.004 and 0.007 mg/kg were found in the neutral organic phases, acidic organic phases, and in the extracted aqueous phase, respectively. In any immature or mature sample, no single component exceeded 0.025 mg/kg or 0.007 mg/kg, respectively. At 120 DAT and 365 DAT, no radioactive residues were associated with any of XDE-175-L, N-demethyl-175-L or N-formyl-175-L. Of the XDE-175-J treated 120 DAT and 365 DAT crop samples, radish (immature tops and mature tops), lettuce (immature), and wheat forage, hay, and straw contained TRR greater than 0.010 mg/kg, but the concentrations were too low for identification. The lower residues in 30 DAT samples were characterized, but could not be identified.

The levels of radioactivity taken up from soil treated with [14C]spinetoram into the three succeeding crops (radish, lettuce, and wheat) planted 30, 120, or 365 days after treatment, were below 0.085 mg/kg spinetoram equivalents. Since such low radioactive residues were found in analysed fractions of these rotational crop samples, spinetoram is unlikely to be taken up readily by succeeding crops.

Methods of analysis

Analytical methods for determination of residues of spinetoram and its metabolites were developed for a wide range of matrices of plant and animal origin. In general, these methods employ extraction of spinetoram and its metabolites with a mixture of acetonitrile and water (80:20, v/v), addition of a stable isotope internal standard solution containing XDE-175 and metabolites, and then, without any clean-up or with solid phase clean-up using a C18 cartridge, analysis with HPLC with positive-ion electrospray tandem mass spectrometry (LC-MS/MS). Three methods, two for plant matrices and one for animal matrices, are capable of determining XDE-175-J, XDE-175-L, N-demethyl-175-J and -L, One method for animal matrices, however, determines XDE-175-J, XDE-175-L, N-demethyl-175-J and -L, and 3'-O-deethyl-175-J and -L.

The methods for plant matrices were validated for each analyte at 0.01–1.0 mg/kg, and in the case of lettuce at 0.01–10 mg/kg. Mean recovery was in a range of 82–111%. The validated limit of quantification was 0.01 mg/kg for all matrices.

The methods for animal matrices were validated for each analyte at 0.01–15 mg/kg in bovine muscle and kidney; 0.01–0.10 mg/kg in poultry muscle; 0.01–50 mg/kg in liver, milk and cream, and eggs; and 0.01–150 mg/kg in fat. Mean recovery ranged between 83 and 119%. The validated limit of quantification was 0.01 mg/kg for all matrices.

The existing multi-residue enforcement methods, FDA PAM I screen methods, were found to be unsuitable for the determination of spinetoram and its metabolites in plant and animal matrices. The DFG S19 multi-residue method was validated successfully only for the determination of spinetoram and its N-demethyl and N-formyl metabolites in apples, grapes and oranges.

Stability of pesticide residues in stored analytical samples

Stability of spinetoram and its N-demethyl and N-formyl metabolites (each at a fortification level of 0.10 ppm) in homogenized orange, lettuce, sugar beet, soya bean and wheat grain stored in deep freezer at -20 °C was investigated over 12 months. No significant decrease of spinetoram was observed in all samples, except lettuce, during the test period. In lettuce, remaining XDE-175-L and N-dimethyl-175-L were 60 and 65% respectively (unadjusted for procedural recovery) at 372 days after initiation of the study.

The Meeting concluded that at $-20\,^{\circ}\text{C}$, spinetoram and its N-demethyl and N-formyl metabolites were stable for 12 months in orange, sugar beet, soya bean and wheat. In lettuce, XDE-175-J, the major component, and its N-demethyl and N-formyl metabolites were also stable for 12 months but XDE-175-L and N-dimethyl-175-L were stable only up to eight months.

As samples of animal tissues, milk and eggs from the metabolism and feeding studies were analysed within 20 days of sample collection in supervised trials, no information was provided to the Meeting on storage stability of spinetoram in animal commodities.

Residue definition

Spinetoram consists of two closely related active ingredients, XDE-175-J and XDE-175-L, present approximately in a three to one ratio.

In apple, lettuce and turnip receiving 4× to 9× the rate of either of the two active ingredients, major metabolites were the parent compounds (XDE-175-J and XDE-175-L), N-demethyl-175-J and N-formyl-175-J. In most cases, PHI XDE-175-J was the primary component of residues. N-demethyl-175-L and N-formyl-175-L were also detected but no more than 7.2% and 4.0% respectively of TRR on one DAT or thereafter.

In goats and hens, metabolism of spinetoram was limited. The parent compounds remained as major components in milk and all ruminant tissues as well as eggs, and all avian tissues except liver, in which 3'-O-deethyl metabolites were detected at similar levels as the parent, but less than 20%.

Sufficiently validated LC-MS/MS methods were available for determining the parent compounds and their N-demethyl and N-formyl metabolites in a wide range of plant commodities and animal tissues, milk and eggs.

Based on the above findings, the Meeting considered that the two parent compounds, XDE-175-J and XDE-175-L, were suitable residues for enforcement. However, as N-demethyl-175-J is a major metabolite in both plants and animals and covered by the ADI, and N-formyl-175-J, a major metabolite in plants, is also found in crops after application of spinetoram, the Meeting decided to include these two metabolites as well as the two spinetoram components in the residue definition for estimation of dietary intake.

XDE-175-J and XDE-175-L have logPow of 4.09 and 4.49 respectively at pH 7 at 20 °C, implying that spinetoram may be fat-soluble. In animal metabolism studies, residue concentrations were found to be higher in tissues with higher fat content. In addition, an animal feeding study with lactating cows indicates that spinetoram residue concentrations in milk fat were 4.4–9.5 times higher than those in whole milk and those in composite fat were 14–24 times higher than in muscle. The Meeting agreed that spinetoram residue is fat-soluble.

The Meeting recommended the following residue definition for plant and animal commodities:

- Definition of the residue (for compliance with the MRL): Spinetoram.
- Definition of the residue (for estimation of dietary intake): Spinetoram and N-demethyl and N-formyl metabolites of the major spinetoram component.
- The residue is fat-soluble.
- Note: Spinetoram consists of two related components.

Results of supervised residue trials on crops

The Meeting received supervised trial data for spinetoram on orange, pome fruits, stone fruits, leaf lettuce, tomato, sugar beet and tree nuts.

For all analytes and matrices, the LOQ was 0.01 mg/kg. The LOD was reported to be 0.003 mg/kg for trials conducted in the USA and 0.005 mg/kg for trials conducted in Australia.

Citrus fruits

Twelve supervised trials were conducted on <u>oranges</u> in the USA.

Six trials conducted using low spray volume applications (approximately 700 L/ha) were in accordance with US GAP for citrus fruits (maximum rate of 103 g ai/ha, three applications, maximum

seasonal rate of 210 g ai/ha, PHI one day). Spinetoram residues from these trials in rank order were: < 0.01 (2), 0.012, 0.022, 0.028 and 0.03 mg/kg.

Corresponding total residues of spinetoram and the two metabolites in rank order were: < 0.02, 0.022, 0.03, 0.052, 0.052 and 0.066 mg/kg.

Six other trials conducted using high spray volume applications (approximately 3300 L/ha) were in accordance with US GAP. Residues from these trials in rank order were: < 0.01, 0.012, 0.015, 0.018, 0.021 and 0.02 mg/kg.

Corresponding total residues of spinetoram and the two metabolites in rank order were: < 0.02, 0.039, 0.041, 0.046, 0.047 and 0.069 mg/kg.

The Meeting considered that these two sets of trials conducted in the same locations could not be regarded as independent from each other and decided to use one data set for the estimation of maximum residue level. Taking into consideration the results of the two data sets being mutually supportive, the Meeting estimated a maximum residue level based on spinetoram residues and an STMR based on the total residues of spinetoram and the two metabolites for oranges at 0.07 and 0.0435 mg/kg.

Pome fruits

Numerous supervised trials were conducted on <u>apple</u> in Australia (20), Canada (8), New Zealand (20) and the USA (12).

Six trials conducted in the USA using low spray volume applications (approximately 700 L/ha) were in accordance with US GAP for pome fruits (maximum rate of 123 g ai/ha, five applications, maximum seasonal rate of 500 g ai/ha, PHI seven days). Spinetoram residues from these trials in rank order were: < 0.01 (3), 0.01, 0.013 and 0.028 mg/kg.

Corresponding total residues of spinetoram and the two metabolites in rank order were < 0.02(3), 0.023, 0.036 and 0.038 mg/kg.

Six other trials conducted in the USA using high spray volume applications (approximately 3300 L/ha) were in accordance with US GAP. Spinetoram residues from these trials in rank order were < 0.01 (4), 0.012 and 0.02 mg/kg.

Corresponding total residues of spinetoram and the two metabolites in rank order were < 0.02(2), 0.022, 0.022, 0.026 and 0.037 mg/kg.

Supervised trials were conducted in four locations in Canada and in the USA in accordance with Canadian GAP for pome fruits (maximum rate of $103 \, \mathrm{g}$ ai/ha, three applications, maximum seasonal rate of $315 \, \mathrm{g}$ ai/ha, PHI seven days). Since two plots were in each location, only higher residues were selected for each location. Spinetoram residues from these trials in rank order were < 0.01, 0.015, 0.017 and $0.028 \, \mathrm{mg/kg}$. These trials were also in compliance with US GAP. Corresponding total residues of spinetoram and the two metabolites in rank order were < 0.02, 0.025, $0.038 \, \mathrm{and} \, 0.038 \, \mathrm{mg/kg}$.

Two trials conducted in Australia and five trials in New Zealand were according to GAP in New Zealand for pome fruits (maximum 2.5 g ai/hL, minimum 50 g ai/ha, four applications and PHI seven days). Spinetoram residues from these trials in rank order were < 0.01 (7) mg/kg. Corresponding total residues of spinetoram and the two metabolites in rank order were < 0.02 (7) mg/kg.

Although application rates were different (PHI in all related GAPs is seven days), results of trials matching three different GAPs were mutually supportive. In ten trials conducted in the USA and Canada following US GAP, which would lead to the highest residues, spinetoram residues were in rank order: < 0.01 (4), 0.01 0.013 0.015 0.017 and 0.028 (2) mg/kg. Corresponding total residues of spinetoram and the two metabolites in rank order were: <0.02 (3), 0.023, 0.025, 0.036, 0.038 (3) mg/kg.

Eight trials were conducted on <u>pear</u> in Australia and eight other in New Zealand. No trials in Australia matched GAP of New Zealand for pome fruits. Spinetoram residues from two trials conducted in New Zealand in accordance with GAP of New Zealand in rank order were < 0.01 and 0.02 mg/kg. Corresponding total residues of spinetoram and the two metabolites in rank order were < 0.02 and 0.03 mg/kg.

Since the results from trials on apple and pear were similar, the Meeting estimated a maximum residue level based on spinetoram residues and an STMR based on the total residues of spinetoram and the two metabolites, for pome fruits on a basis of apple trials, at 0.05 and 0.025 mg/kg respectively.

Stone fruits

A large number of supervised trials were conducted on cherry, peach and apricot in Australia and New Zealand. A few trials on nectarines were also conducted in Australia.

However, since proposed GAP in Australia for stone fruits has not been approved, no maximum residue level could be estimated.

Tomato

Six supervised trials conducted in the USA were according to US GAP for fruiting vegetables (maximum rate of 88 g ai/ha, six applications, maximum seasonal rate of 298 g ai/ha and PHI one day). Residues from these trials in rank order were < 0.01 (2), 0.01, 0.0156, 0.024 and 0.025 mg/kg.

Corresponding total residues of spinetoram and the two metabolites in rank order were < 0.02(3), 0.02, 0.025, 0.034 and 0.035 mg/kg.

The Meeting estimated a maximum residue level based on spinetoram residues and an STMR based on the total residue of spinetoram and the two metabolites in tomato at 0.06 and 0.02 mg/kg respectively.

Lettuce

Six supervised trials were conducted on leaf lettuce in the USA in accordance with US GAP for leafy vegetables (maximum rate of 88 g ai/ha, six applications, maximum seasonal rate of 298 g ai/ha and PHI one day). Residues in rank order were 0.15, 0.31, 0.32, 0.34, 0.55 and 7.80 mg/kg.

Corresponding total residues of spinetoram and the two metabolites in rank order were 0.28, 0.56, 0.64, 1.16, 1.35 and 9.55 mg/kg.

The residue values of 7.80 and 9.55 mg/kg were very high compared to the rest of results. The study report indicates that the trial was conducted in the same manner as the other trials and there was no indication for this trial being invalid. Using all the residue values, the Meeting estimated a maximum residue level based on spinetoram residues and an STMR for spinetoram in lettuce at 10 and 0.895 mg/kg.

As foliar applications on leaf lettuce were expected to result in higher residues than those on head lettuce, the Meeting agreed that these maximum residue level and STMR are applicable also to head lettuce.

Sugar beet

Six supervised trials were conducted in the USA. In one trial according to US GAP (maximum rate of 70 g ai/ha, four applications, maximum seasonal rate of 281 g ai/ha and PHI seven days) for tuberous vegetables, e.g., potato and sugar beet, the residue was < 0.01 mg/kg. In the other five trials, root samples were taken three days after the last application, earlier than the required PHI of seven days in GAP. Residues in these five trials were < 0.01 mg/kg (5).

Corresponding total residues of spinetoram and the two metabolites in rank order were < 0.02 (6) mg/kg.

The Meeting estimated a maximum residue level based on spinetoram residues and an STMR for spinetoram in sugar beet at 0.01 (*) and 0.02 mg/kg respectively.

Tree nuts

Six supervised trials were conducted on <u>almonds</u> in the USA. One trial was in accordance with US GAP for tree nuts (maximum rate of $123 \, g$ ai/ha, four applications, maximum seasonal rate of 490 g ai/ha and PHI 14 days), and residues were < 0.01 mg/kg. In the other five trials, nut samples were taken seven days after the last application rather than the required PHI of 14 days in GAP. Residues in these five trials were < 0.01 mg/kg (5).

Corresponding total residues of spinetoram and the two metabolites in rank order were < 0.02 (6) mg/kg.

Six supervised trials were also conducted on $\underline{\text{pecan}}$ in the USA. In one trial according to US GAP for tree nuts, residues were < 0.01 mg/kg. In the other five trials, nut samples were taken seven days after the last application rather than the required PHI of 14 days in GAP. Residues in these five trials were < 0.01 mg/kg (2) and 0.01 (3) mg/kg. A decline study indicates that it is likely that residues would be less than 0.01 mg/kg if samples were taken on the required PHI of 14 days and in general residues were not expected to occur in edible portions of tree nuts due to negligible translocation of spinetoram.

From the results of trials on almonds and pecan, the Meeting estimated a maximum residue level based on spinetoram residues and an STMR based on the total residue of spinetoram and the two metabolites in tree nuts at 0.01 and 0.02 mg/kg.

Sugar beet leaves or tops

Among six supervised trials conducted in the USA, only one was in compliance with US GAP and residues were 0.024 mg/kg. In other trials, samples were collected only three days after the last application instead of the required PHI of seven days and they contained finite level of residues. However, these trials were in compliance with US GAP for leaf of root and tuberous vegetables for forage (maximum rate of 80 g ai/ha, four applications, maximum seasonal rate of 281 g ai/ha and PHI of three days). Residues from these trials were 0.086, 0.099, 0.11, 0.16 and 0.20 (2) mg/kg.

The Meeting estimated an STMR and a highest residue in sugar beet leaves or tops based on spinetoram residues for calculation of animal burden at 0.135 and 0.20 mg/kg.

Almond hulls

Among six supervised trials conducted in the USA, only one trial was in accordance with US GAP and residues are 0.75 mg/kg. In other trials, samples were collected seven days after the last application instead of the required PHI of 14 days and they contained finite level of residues. The Meeting concluded that it was not possible to estimate a maximum residue level for spinetoram in almond hulls from the results of these trials.

Fate of residues during processing

The Meeting received information on processing of oranges to juice and oil, and apples to juice and puree (sauce).

Processing factors were calculated for oranges (juice, peel and pulp after juicing, dried pulp and oil) and for apples (juice, dry pomace and puree (sauce)).

Processed Orange	Process	ing factor	STMR/STMR-P	
Product	Spinetoram residues	Spinetoram+2 metabolites	(mg/kg)	
Orange	-	-	0.045	
Juice	< 0.05	< 0.07	0.003	
Dried pulp	2.4	2.3	0.105	
Apple	-	-	0.025	
Juice	< 0.37	< 0.44	0.011	
Dry pomace	8.1	6.0	0.15	

Processed Orange	Processir	ng factor	STMR/STMR-P	
Product	Spinetoram residues	Spinetoram+2 metabolites	(mg/kg)	
Puree (sauce)	0.45	0.47	0.012	

For the purpose of calculating animal dietary burden for estimating maximum residue levels for commodities of animal origin, STMR-P for citrus pulp, dry and apple pomace, dry were calculated based on spinetoram residues to be 0.048 and 0.081 mg/kg.

Farm animal dietary burden

Dry apple pomace, dry citrus pulp and sugar leaves or tops may be fed to dairy cattle and beef cattle but not as a major ingredient. The dietary burdens were calculated from the highest residue and STMR of sugar beet leaves or tops, and the STMRs of apple pomace, dry, using the OECD feedstuffs tables (Annex 6 of the 2006 Report of the JMPR).

Summary of livestock dietary burdens (ppm of dry matter diet)

	US-Canada		EU		Australia	
	max	mean	max	mean	max	mean
Beef cattle	0.018	0.018	0.192 ^a	0.135 ^b	0.018	0.018
Dairy cattle	0.0089	0.0089	0.270°	0.185^{d}	0.0089	0.0089

^a Suitable for estimating maximum residue levels for meat and edible offal.

Residues in milk and cattle tissues

Lactating dairy cows were dosed daily for 29 consecutive days via gelatin capsules containing a mixture of spinetoram and N-demethyl and N-formyl metabolites of XDE-175-J (1.2–38.6 ppm in diet) or spinetoram only (37.6 ppm).

Residues in the milk in 1.2 ppm (equivalent to 0.4 mg XDE-175-J and -L) dose group were generally between the LOD (0.003 mg/kg) and LOQ (0.01 mg/kg) throughout the dosing period.

No or low concentration residues were detected in skim milk; even in 11.5 and 38.6 ppm (total) doses groups, mean total residues of the four compounds in skim milk ranged from just below the LOQ to 0.075 mg/kg. In all of the dose groups on day 14 and 28, total residues in cream were much higher than the residues in skim milk at 0.187 and 0.237 mg/kg in the 1.2 ppm doses groups. The mean total residues in cream from the 11.5 and 38.6 ppm doses groups ranged from 0.64 to 5.84 mg/kg. The average ratio of residues in cream to those in whole milk is 6:6.

All tissues from treated cows contained residues and they increased from the lowest to highest dose groups. Residue concentrations were lowest in the muscle followed by kidney, liver, and fat. Residues in fat were significantly higher than residues in the other tissues. These results indicate that residues of spinetoram tend to accumulate in fatty tissue.

With one exception, residues were not detectable in milk by the fourth day after the last dose was administered. Concentrations just above the LOD were detected in one cow through day nine after the final dose. No further residue was detected beyond that point.

Residues in tissues continuously declined through 28-day depletion period after the last dose. No residue was detected in kidney, liver or muscle from any cow by 28 days after the final dose or in fat 56 days following the final dose.

^b Suitable for estimating STMRs for meat and edible offal.

^c Suitable for estimating a maximum residue level for milk and fat.

^d Suitable for estimating an STMR for milk and fat.

The dietary burdens for beef and dairy cattle are both lower than the lowest feeding level (1.2 ppm, equivalent to 0.4 ppm of spinetoram only) in the feeding study. Therefore the MRL and STMR were estimated using the residue concentrations in milk and tissues at the lowest feeding level and the dietary burden. The calculated residues in cattle tissues and milk are summarized below.

Dietary burden	Spinetoram residu	ies, mg/kg			
mg/kg Feeding level [mg/kg]	Milk	Muscle	Liver	Kidney	Fat
MRL	highest	highest	highest	highest	highest
0.270/0.192	< 0.00675	< 0.00480	< 0.00480	< 0.00480	0.0743
[0.4]	[< 0.01]	[< 0.01]	[< 0.01]	[< 0.01]	[0.11]
	Spinetoram, N-de	methyl-175-J and Λ	-formyl-175-J, mg	/kg	
STMR	mean	mean	mean	mean	mean
0.185/0.135	< 0.00925	< 0.00625	< 0.00625	< 0.00625	0.0463
[0.4]	[< 0.02]	[< 0.02]	[< 0.02]	[< 0.02]	[0.10]

The Meeting estimated a maximum residue level for spinetoram in edible offal (mammalian) and whole milk at 0.01(*) mg/kg and in mammalian fats at 0.2 mg/kg. STMRs were estimated to be 0.00925 mg/kg for whole milk, 0.00625 mg/kg for meat (muscle) and edible offal (mammalian), and 0.046 mg/kg for mammalian fats.

The Meeting estimated a maximum residue level for spinetoram and an STMR for spinetoram and the two metabolites in milk fat, using the median ratio between residues in cream and whole milk of 6:6 and assuming that cream contains 50% fat, at 0.1 mg/kg and 0.12 mg/kg respectively.

Poultry

No data were provided on poultry feeding study. Since there was no treated commodities that can be fed to hens, the Meeting considered that it was unnecessary to estimate maximum residue levels for poultry tissues or eggs.

RECOMMENDATIONS

On the basis of the data from supervised trials, the Meeting concluded that the residue levels listed below are suitable for establishing maximum residue limits and for IEDI assessment. Spinetoram is considered to be fat soluble.

Definition of the residue (for compliance with the MRL): Spinetoram.

Definition of the residue (for estimation of dietary intake): *Spinetoram and N-demethyl and N-formyl metabolites of the major spinetoram component.*

The residue is fat-soluble.

Note: Spinetoram consists of two related components.

CCN	Commodity	Recommended	d MRL, mg/kg	STMR or STMR-P
CCIV	Commodity	New	Previous	(mg/kg)
MO 0105	Edible offal, mammalian	0.01*	-	0.00625
VL 0482	Lettuce, Head	10	-	0.895
VL 0483	Lettuce, Leaf	10		0.895
MM 0095	Meat (from mammals other than	0.2 (fat)	-	0.00625 (muscle)
	marine mammals)			0.042 (fat)
FM 0183	Milk fats	0.1	-	0.12
ML 0106	Milks	0.01*	-	0.00925

CCN	Commodity	Recommend	ed MRL, mg/kg	STMR or STMR-P
CCN	Commodity	New	Previous	(mg/kg)
FC 0004	Oranges, Sweet, Sour	0.07	-	0.0435
JF 0004	Orange juice			0.003
FP 0009	Pome fruits	0.05		0.025
JF 0226	Apple juice			0.011
	Apple puree or sauce			0.012
VR 0596	Sugar beet	0.01*	-	0.02
VO 0448	Tomato	0.06	-	0.02
TN 0085	Tree nuts	0.01	-	0.02
AB 0001	Citrus pulp, dry			0.048 ^a
AB 0226	Apple pomace, dry			0.081 ^a
AV 0596	Sugar beet leaves or tops			0.135 ^a

^a For dietary burden calculation. based on spinetoram residues only.

DIETARY RISK ASSESSMENT

Long-term intake

The International Estimated Dietary Intakes (IEDIs) of spinetoram were calculated for the 13 GEMS/Food cluster diets using STMRs estimated by the current Meeting (see Annex 3 of the 2008 Report of the JMPR). The ADI is 0–0.05 mg/kg bw and the calculated IEDIs were 0–1% of the maximum ADI. The Meeting concluded that the long-term intake of residues of spinetoram resulting from the uses considered by the current JMPR is unlikely to present a public health concern.

Short-term intake

The 2008 JMPR decided that an ARfD is unnecessary. The Meeting therefore concluded that the short-term intake of residues of spinetoram is unlikely to present a public health concern.

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