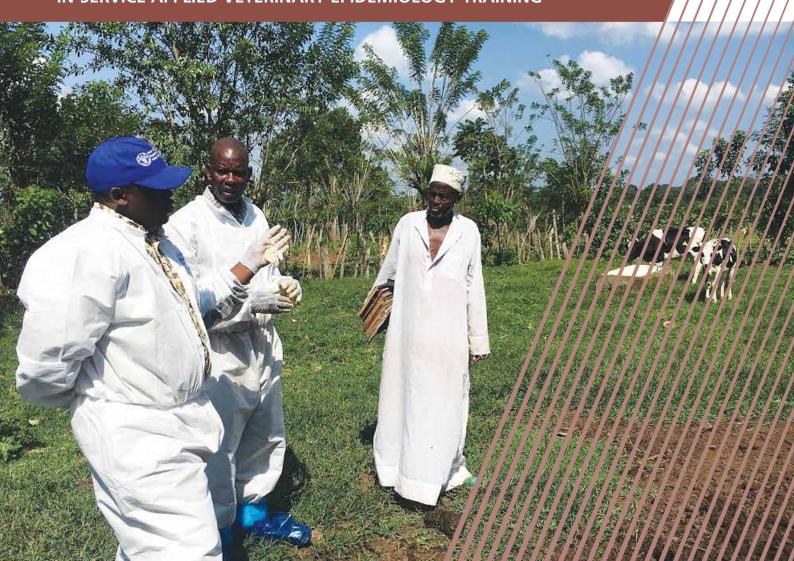




IN-SERVICE APPLIED VETERINARY EPIDEMIOLOGY TRAINING



Published by:

Food and Agriculture Organization of the United Nations and

Texas A&M AgriLife – Institute for Infectious Animal Diseases

Rome, 2021

Required citation:

FAO and AGRILIFE. 2021. Frontline ISAVET Curriculum Participant Manual. Rome.

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Acronyms/Abbreviations

AI Avian Influenza

A-P-T Animal, Place and Time ASF African Swine Fever

AU-IBAR African Union InterAfrican Bureau for African Resources

BSE Bovine Spongiform Encephalopathy

BT Blue Tongue

CAHWs Community Animal Health Workers
CBPP Contagious Bovine Pleuropneumonia

CCA Critical Control Areas
CFR Case Fatality Rate

CQI Continuous Quality Improvement CRD Chronic Respiratory Disease

CSF Classical Swine Fever

DAR Directorate of Animal Resources
DTR Disease Transmission Risk
DVO Local area Veterinary Officer

ECF East Coast Fever

EID Emerging Infectious Disease

ELISA Enzyme Linked Immunisorbent Assay

FAO Food and Agriculture Organization of the United Nations FETPV Field Epidemiology Training Program for Veterinarians

FMD Foot and Mouth Disease

FN False Negative
FP False Positive

GEMP Good Emergency Management Practices

GIS Geographic Information Systems HPAI Highly Pathogenic Avian Influenza

IFA Immunofluorescent Antibody

IR Incidence Rate

ISAVET In Service Applied Veterinary Epidemiology Training

KAP Knowledge, Attitude and Practices

LBM Live Bird Market

LPAI Low Pathogenic Avian Influenza

MAIFF Ministry of Agriculture, Animal Industry and Fisheries

MS Microsoft

NARO National Agricultural Research Organisation

NCD New Castle Disease

NGOs Non-governmental Organisations

No. Number
OB Outbreak
OH One Health

OIE World Organisation for Animal Health

PAR Population at Risk

PCR Polymerase Chain Reaction

PM Post-Mortem

PPE Personal Protective Equipment
PPR Peste des Petits Ruminants

RT-PCR Reverse Transcriptase Polymerase Chain Reaction

RTPs Risk Transmission Pathways

RVF Rift Valley Fever

RVFv Rift Valley Fever Virus SCC Somatic Cell Count

SOP Standard Operating Procedure

SSRT Surveillance Rapid Response Teams
TAD Transboundary Animal Disease

TB Tuberculosis

USDA United States Department of Agriculture

UVA Uganda Veterinary Association
VFE Veterinary Field Epidemiology
WHO World Health Organization

ZDCO Zoonotic Disease Coordination Office

Acknowledgements

These guidelines were written on behalf of the Food and Agriculture Organization of the United Nations (FAO) under the overall guidance and responsibility of J. Lubroth, Chief Veterinary Officer, FAO and the Senior Animal Health Officer, Yilma Makonnen.

The competencies and skills that serve as the framework for Frontline ISAVET were developed by a global technical working group organised by FAO in July 2018. The lessons, exercises, case studies, field activities and standard operating procedures were developed by the following authors and contributors:

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Publication of this document was made possible by financial support from FAO and the United States Agency for International Development (USAID), Global Health Security Agenda (GHSA).

About this Manual

This manual serves as the primary reference for Frontline ISAVET participant trainees to utilise the Frontline ISAVET course curriculum and is a companion manual to the Frontline ISAVET Trainer Manual. It includes the 36 lessons and exercises of the Frontline ISAVET course curriculum based on two pilot trainings in Uganda and Senegal between October and December, 2018 as well as extensive post-pilot internal and external reviews.

Target Audience

The intended audience for this Manual is the Frontline ISAVET participants who will be trainees during the four-week didactic and applied Frontline ISAVET course. Participants include field level veterinarians and paraveterinary professionals enrolled in the course.

Development of the Frontline ISAVET Curriculum

The curriculum foundation work for Frontline ISAVET was developed by a global technical working group organised by FAO in July 2018. The FAO technical working group includes animal health, public health and wildlife health professionals under a One Health approach. This group developed a competency-skills matrix for the frontline veterinary field epidemiology level which was used to develop specific lesson learning objectives.

Specific Objectives of the Frontline ISAVET Curriculum Participant Manual

The specific objectives of the Frontline ISAVET Curriculum Participant Manual are as follows:

- 1. Provide the structure, format and content of the Frontline ISAVET curriculum lessons, exercises and case studies.
 - a. This includes specific learning objectives for each lessons.
 - b. English or French language versions are available.
- Demonstrate clear linkage between lesson learning objectives and expected outputs of exercises and case studies.
- 3. Provide supportive curriculum resource materials include MS Excel training videos, standard operating procedures and relevant references.
- 4. Provide a flexible base upon which to include relevant examples based on the regional and national context.
- 5. Emphasise the flexibility of classroom exercises designed to accommodate veterinary and paraveterinary trainees that may have different levels of computer experience.

Content Development and Review

The content of the Frontline ISAVET curriculum was developed according to the following steps:

- 1. The specific learning objectives were developed by TAMUS, IIAD based on the FAO technical working group competency-skills matrix.
- 2. Lesson outlines were developed which were completed by contributors previously listed.
- 3. Lessons were adjusted in real-time during the pilot trainings as part of a continuous quality improvement process.

Following the Uganda and Senegal pilot trainings, the Frontline ISAVET Curriculum was re-aligned and further developed as follows:

- 1. English internal review by FAO IIAD epidemiologists.2. English external review by experienced field epidemiologists from animal health (Thailand), public health (USA) and wildlife health (Uganda).
- 2. French internal review by FAO epidemiologists from west and central Africa.
- 3. Final internal review by FAO IIAD epidemiologists.
- 4.

Frontline ISAVET Curriculum and Exercise Matrix Overview

The Frontline ISAVET course is comprised of 4 weeks of classroom training that includes one week of field application. The classroom portion of the training covers 8 domains, 14 competencies and 51 skills that were developed by an FAO technical working group. The Frontline ISAVET curriculum is the common framework to adapt the Frontline ISAVET curriculum for country programmes in the future. The Frontline ISAVET Curriculum Matrix is provided in Table 1.

For National Programmes wanting access to the training Power Point Slides, please send a request to ISAVET@fao.org

 Table 1. Frontline ISAVET Curriculum Matrix

| LESSON NUMBER | LESSON TITLE | EXERCISE |
|---------------|-----------------------------------|------------------------|
| | Course Introduction | Group introductions, |
| | and Orientation | sharing and discussion |
| | PRE-TEST | NA |
| 1 | Introduction to | Exercise 1: |
| | Animal Health | |
| | Surveillance | |
| 2 | Data for Information | Exercise 2: |
| _ | and Response | Exercise 3: |
| 3 | Defining and | Exercise 4: |
| | counting disease | Exercise 5: |
| 4 | cases Data Quality | Exercise 6: |
| 4 | Principles | Exercise 6. |
| 5.1 | Describing and Acting | Exercise 7: |
| 3.1 | upon Animal Disease | Zacreise 71 |
| | and Health Data: | |
| | Central tendency | |
| 5.2 | Describing and Acting | Exercise 8: |
| | upon Animal Disease | |
| | and Health Data: | |
| | Disease Occurrence | |
| | and Impact | |
| 5.3 | Describing and Acting | Exercise 9: |
| | upon Animal Disease | |
| | and Health Data: | |
| | Descriptive analysis | |
| | by animal-place-time | |
| 6 | Display data for | Exercise 10: |
| | decision making | Exercise 11: |
| CASE STUDY 4 | 200 | Exercise 12: |
| CASE STUDY 1: | PPR | Case study 1: |
| 7 | Data Interpretation | Exercise 13: |
| | and Reporting to | Exercise 14: |
| | Improve Situational Awareness and | |
| | Decision Making | |
| 8 | Elements of a | Exercise 15.1: |
| | surveillance report | |
| 9 | Making | Exercise 15.2: |
| | recommendations for | |
| | animal disease | |
| | prevention and | |
| | control | |

| LESSON NUMBER | LESSON TITLE | EXERCISE |
|---------------|--|----------------|
| 10 | Sharing surveillance information in a network for animal disease prevention and control | Exercise 16: |
| 11 | Assessing Surveillance in Your Local Area to Improve Response to Animal Disease and Health Events | Exercise 17: |
| 12 | Animal field investigations | Exercise 18: |
| 13 | Investigation strategies for early prevention and control of animal disease transmission | Exercise 19: |
| 14 | Establish Two-Way Linking Between Laboratory Disease Diagnosis and Field Investigation | Exercise 20: |
| 15 | Biosafety and biosecurity for animal disease investigations | Exercise 21: |
| 16.1 | Apply the steps of a animal health outbreak investigation for animal-specific and zoonotic diseases | Exercise 22: |
| 16.2 | Apply the steps of a public health outbreak investigation for zoonotic diseases | |
| CASE STUDY 2 | Al Tabletop Exercise | Case Study 2: |
| 17.1 | Managing Outbreak Investigation Data: Collect Data and create a line listing | Exercise 23.1: |
| 17.2 | Managing Outbreak Investigation Data | Exercise 23.2: |
| 18 | Follow up investigations and special studies | Exercise 24: |

| LESSON NUMBER | LESSON TITLE | EXERCISE |
|---------------|--|----------------------------------|
| 19 | Surveillance situation assessment for prevention and control | Exercise 25: |
| 20 | Display outbreak investigation findings and make relevant recommendations for prevention and control | Exercise 26: |
| 21 | One Health Panel Discussion: Multi- disciplinary Outbreak Investigation | Panel Discussion and Q&A session |
| 22 | Preparing MS PowerPoint Presentations | Exercise 27: |
| 23 | Guidelines for Outbreak Investigation Reports | Exercise 28: |
| 24 | Communicating disease transmission risk to diverse audiences | Exercise 29: |
| 25 | Stakeholder Risk Communication Before, During and Following an Animal Disease Event | Exercise 30: |
| 26 | How to Plan and Prepare the Frontline ISAVET Field Study Proposal and Report | Exercise 31: |
| CASE STUDY 3 | FACE - Ethical Decision Making Framework | Case Study 3 |
| 28 | Professionalism in the Practice of Veterinary Field Epidemiology | Exercise 32: |
| 29 | Emergency Preparedness for Animal Health Emergencies at the Local Level: Part 1 | Exercise 33: |
| 30 | Emergency Preparedness for Animal Health Emergencies at the District Level: Part 2 | Exercise 34: |

| LESSON NUMBER | LESSON TITLE | EXERCISE |
|---------------|--|--------------------------------|
| 31 | Disease preparedness and response at the district level | Exercise 35: |
| 32 | Characteristics of a functional disease prevention and control program at the district level | Exercise 36: |
| 33 | Local Area Profile | Question and Answer Session |
| 34 | (Disease name) | Question and Answer Session |
| 35 | The Role of Wildlife for Priority Disease(s) | Question and Answer Session |
| 36 | Preparing for Field Work | Question and Answer Session |

Section I: Week 1 – Epidemiological Surveillance

Introduction

| Estimated Lesson and Exercise Time | 1 hour |
|------------------------------------|-------------------|
| Instructor Materials | Introduction.pptx |
| | |
| Participant Materials | Introduction.PDF |

INSTRUCTOR COMMENTS: LESSON ACRONYMS AND ABBREVIATIONS

| In Service Applied Veterinary Epidemiology Training (ISAVET) Introduction | | |
|---|---|--------|
| INSTITUTE FOR INFECTIOUS ANIMAL DISEASES | Food and Agriculture Organization of the United Nations | l ———— |

Outline

- ${\color{red} \bullet \, Welcome}$
- Introduction of Trainees
- About the Frontline ISAVET Programme
- About the Frontline ISAVET Course

Introduction of Trainees

- · Please introduce yourself, where you live and work
- · Why are you attending this course?
- · What do you want to gain from the Frontline ISAVET training?



3

Importance of Frontline Veterinarians and Veterinary Paraprofessionals

Frontline Veterinarians and Veterinary Paraprofessionals:

- Have strong ties to the community level where disease events occur
- · Are at the leading edge of an animal disease outbreak
- · Have the best access to field data
- · Have high quality field data for analysis of risk factors for disease prevention and control
- epidemiological information for decision-making and action



Reference: Frontline ISAVET, 2018

What is Frontline ISAVET?

- Based on the field epidemiology training programme (FETP) public health training
- model in 73 countries
 One Health focus: animal-human-wildlife interface
- $\cdot \ \ Focuses on applied veterinary-specific competencies$ and skills
- The field epidemiology training programme for veterinarians (FETPV) was initiated in 2008 in Thailand and has since developed in China and Indonesia $\,$



Levels of ISAVET Programme

· ISAVET is the new name of the programme with Three levels (Frontline, Intermediate and

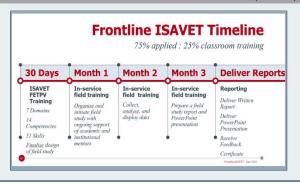
Overview of Frontline ISAVET

- Frontline ISAVET includes 14 countries in West, Central and East Africa:
- Burkina Faso, Cameroon, Côte d'Ivoire, Democratic Republic of Congo (DRC), Ethiopia, Ghana, Guinea, Kenya, Liberia, Mali, Sierra Leone, Senegal, Tanzania and Uganda
- Frontline ISAVET Programme training occurs in 5 locations:
 - Pilot Phase: Uganda and Senegal



6

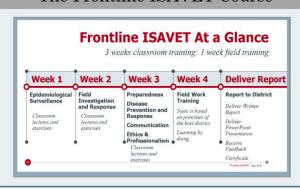
Overview of Frontline ISAVET (cont.)



7

8

The Frontline ISAVET Course



Requirements · Attend all course modules/lectures · Sit for all quizzes, pre and post tests · Complete all homework & assignments · Active participation in case studies and other learning exercises Completion of field project · Presentation of your project findings to your peers and stakeholders 9 Frontline ISAVET Programme Requirements TRAINEE WORK REQUIREMENTS FOR CERTIFICATE IN FRONTLINE ISAVET PROGRAMME Required field products for Frontline ISAVET in 12 weeks of field activities: 1. Conduct Two Data Exercises: COMPULSORY Weekly surveillance reports and systematic disease monitoring; $2. \ \ \, \textbf{Data Quality Audits at the animal health of fice level} \, (summarize \, findings \, through \, a$ $SWOT\ Analysis\ or\ Problem\ Analysis\ using\ a\ Fishbone\ Diagram\ to\ produce\ a\ report):$ COMPULSORY 3. Conduct One Brief Field Study (maximum of 10 pages in length): COMPULSORY • Field or Outbreak investigation; • Survey or KAP study; · Secondary data analysis; ${}^{\bullet}$ Other, including value chain mapping and risk pathway analysis. 10

How Can You Apply Your Knowledge and Skills?

- · Introduction to animal health surveillance
- · Using data to inform follow up action
- · Defining and counting disease cases
- · Apply data quality principles
- · Describing and acting upon animal disease and health data
- · Collecting high quality data
- $\boldsymbol{\cdot}$ How data analysis can provide information and knowledge for decision-making
- · Data interpretation and reporting to improve situational awareness and decision-making
- · Develop weekly of surveillance reports
- $\boldsymbol{\cdot}$ Making recommendations for animal disease prevention and control
- · Sharing surveillance information in a network for animal disease prevention and control
- · Assessing surveillance in your district to improve response to animal disease events

| ISAVE | T Contributing Universities | | |
|--------------|-------------------------------|-----|--|
| Partners | TEXAS A&M TEXAS A&M GRILIFE | | |
| | A CRILITE | | |
| | XXX | | |
| | Section 2 | | |
| Contributors | | | |
| | MAKERIER UNIVERSITY | 9.0 | |
| | | 20 | |

Lesson 1 - Introduction to Animal Health Surveillance

| Estimated Lesson and Exercise Time | 1 hour 30 minutes |
|------------------------------------|--|
| Participant Materials | ISAVET Lesson 1 Introduction to Animal Health Surveillance Participant Guide.PDF |
| | Computer and Microsoft Word |
| | Pen or Pencil |

In Service Applied Veterinary Epidemiology Training (ISAVET)

Lesson 1: Introduction to Animal Health Surveillance

| SSTITUTI FOR. A INFECTIOUS ANIMAL DIMENTS | Food and Agricultur Organization of the United Nations |
|--|--|

Learning Objectives

At the end of this lesson, you will be able to:

- 1. Define surveillance, survey and monitoring and surveillance system;
- $2. \;\;$ Describe the objectives and components of an animal health surveillance system;
- 3. Describe the flow of surveillance data and how it is shared; and
- 4. Explain how surveillance data is used.

What is a Survey?

- A survey is an investigation in which information is systematically collected using samples from a defined subpopulation group within a defined time period (Salman, 2003)
 Example: One elephant group in a park
- Survey sample must be representative of the entire population "A sample is a selected subset of a population". (Dictionary of Epidemiology)



Reference: Google images

3

What is Monitoring?

- The ongoing collection of data related to various disease factors
- NO action is implied
- · Examples:
 - Insect vector monitoring
 - Wild bird populations
- Carcass quality
- · Can you think of other examples?



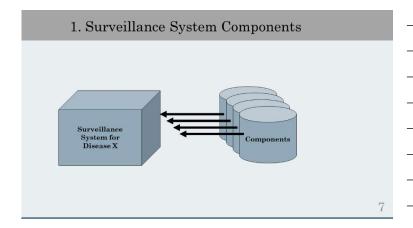
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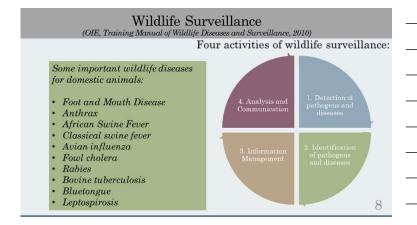
What is Surveillance?

- Surveillance is the <u>systematic</u>, <u>ongoing</u> collection, collation and analysis of data and the <u>timely dissemination</u> of information to <u>those who need to know so</u> <u>that action can be taken</u>. (OIE Terrestrial Animal Health Code)
- · Desired outcomes of animal disease surveillance:



What is a Surveillance System? • A surveillance system is described by: • A method of surveillance that may involve one or more component activities that generates information on the health or disease status of animal populations." (Source: OIE) • What is a 'system'? 1. Identified parts/components of the system 2. Coordinated objectives and methods 3. Inter-connections using mechanisms and networks





Examples of Surveillance System Components – Bovine TB

- · Slaughter inspection of bovine TB-susceptible species
- · Tuberculin testing
- · Passive reporting of suspect cases

9

Examples of Surveillance System Components – African Swine Fever

- Sampling and testing of sick /dead wild boar/domestic pig
- Sampling and testing hunted boar/domestic pig
- Sampling and testing suspect holdings
- Sampling and testing meat and animal product
- · Testing home slaughter
- Inspection of personal baggage
- Sampling, testing, quarantine of imported pigs

10

Food Safety: Surveillance Along The Food Chain Agricultural inputs Hazard or Contan in the Food Chain Food processing Food consumption Reference: FAO

2. Coordinated Objectives

A country disease surveillance system will require clear objectives that will be adapted for each country and each disease situation.

Some types of objectives may include:

- · Early detection of livestock diseases
- · Enabling early reaction to such diseases
- · Correct identification of resource needs in the field.
- · Provision of strategic decision-making support
- · Measurement of surveillance system performance

Source: FAO Manual on Livestock Disease Surveillance and Information Systems

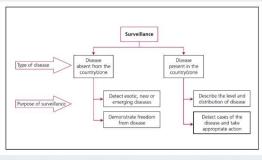
12

What components does surveillance Target?

- 1. Early reporting
- 2. Early detection
 - a. Targeted surveillance at poultry aggregation sites using value chains
 - b. Rapid screening tests
 - c. Collaboration and communication with poultry producers and marketers
- 3.Rapid response
- 4.Containment
- $5. \\ \\ Continual improvement of the surveillance system in terms of sensitivity and timeliness$

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Surveillance Objectives Will Depend on Whether the Disease is Present or Absent



Reference: Manual of Basic Animal Disease Surveillance, AU-IBAR, 2012

Example: Zoonotic Disease Surveillance

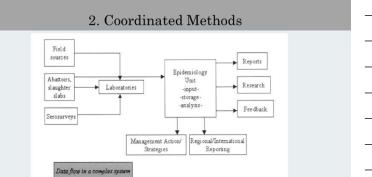
Definition of a zoonosis:

A disease that can be transmitted between animals and humans.

The objective of surveillance for zoonotic disease is to detect the pathogen as early as possible to prevent further animal and human exposure as shown in the graph.



15



(FAO Manual on Livestock Disease Surveillance and Information Systems)

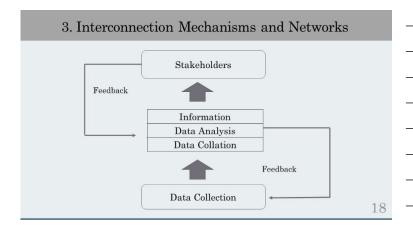
16

Exercise 1: Epidemiological Surveillance

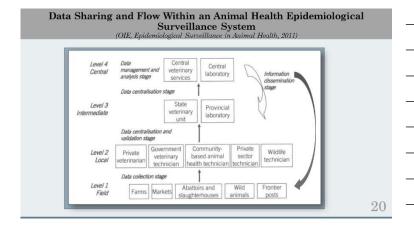
- This exercise will take 60 minutes take 20 minutes to draw a flow diagram to be followed by 4 group reports of 10 minutes each.
- Form yourselves into groups based on country.
- 3. Draw a diagram on a flip chart that describes the surveillance system with reference to animal health, wildlife, zoonotic diseases, food safety and antimicrobial resistance in your local area, province and country. Include the following elements in your diagram:

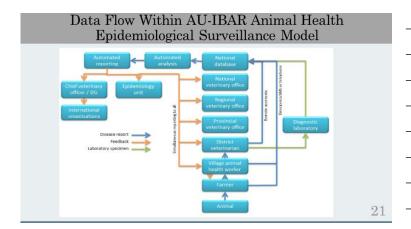
 Label country stakeholders at each administration level from field to national levels.

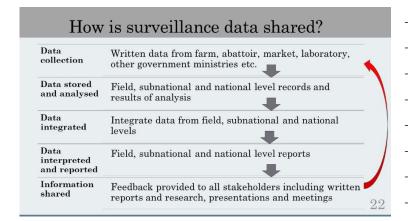
 - Information inputs including direction, origin and destinations
 Information outputs including direction, origin and destinations
- One group will describe their surveillance system related to food safety linking local area, province and country.









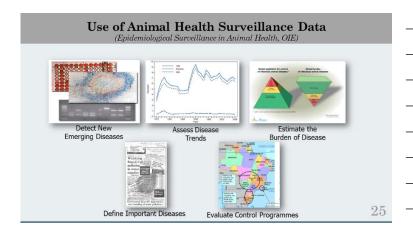


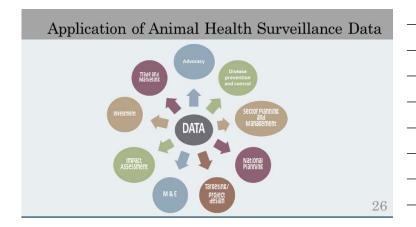
Think, Pair and Share!

Why do we need animal health information?

Need For Animal Health Information

- · Identify what diseases exist (in a country, area or farm)
- $\boldsymbol{\cdot}$ Determine frequency and distribution of diseases
- \bullet Determine the importance or "burden" of different diseases
- Set priorities for the use of resources for disease control
- Plan, implement and evaluate disease control programmes
- · Respond to disease outbreaks
- ${}^{\raisebox{1pt}{\text{\circle*{1.5}}}}$ Meet reporting requirements of international organizations
- \cdot Demonstrate disease status to trading partners





In Summary...

Animal Health Surveillance:

- Includes public and private sectors and the communities.
- Is a system that includes disease detection, data collection, analysis, interpretation and sharing at all levels so that action is taken on the best available evidence.
- · Occurs at all levels from local to international levels.
- Is used to assess disease status, trends, and animal disease programs to take action for animal disease prevention and control.

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In Summary: Flow of Surveillance Data 1. Detect Disease 2. Report Disease and Collect Data Communication and Coordination 4. Take Action 3. Analyse and Interpret Data



Exercise 1 - Epidemiological Surveillance

Description of Exercise:

Develop a flow diagram that describes the surveillance system in your local area, province, and country with reference to animal health, wildlife, zoonotic diseases, food safety and antimicrobial resistance. Should you have any questions about the exercise, please ask a trainer for clarification before, during, and after the exercise.

Time: 30 minutes

Organisation of Group Work:

- 1. Form yourselves into groups based on country.
- 2. Draw a diagram on a flip chart that describes the surveillance system related to food safety in your local area, province and country.

At least one group will describe their surveillance system linking local area, province and country with reference to animal health, wildlife, zoonotic diseases, food safety and antimicrobial resistance.

Include the following elements in your diagram:

Label country stakeholders at each administration level from field to national levels

- Information inputs including direction, origin and destinations
- Information outputs including direction, origin and destinations

Materials, Data or Information:

1. Preferably, information is collected and displayed using MS Word or MS PowerPoint Flip chart and markers if computers are not available

Lesson 2 – Data for Information and Response

| Estimated Lesson and Exercise Time | 1 hour 30 minutes |
|------------------------------------|---|
| Participant Materials | Frontline ISAVET Lesson 2 Data for Information and Response Participant Guide.doc |
| | Computer and Microsoft Word |
| | Pen or Pencil |

| Lesson 2: Data for Information and Response | |
|---|--|
| In Service Applied Veterinary Epidemiology Training (ISAVET) | |
| | |

Learning Objectives

At the end of this lesson you will be able to:

- 1. Explain the terms population, unit of interest, case definition;
- $_{\rm 2.}$ Explain the surveillance approaches for data collection , active and passive surveillance.

What is Epidemiology? "Study of distribution and determinants of health related states or events in specified populations, and the application of this to the control of health problems" Dictionary of Epidemiology · Surveillance is used to collect epidemiological data · Epidemiologic Approach at the population level is necessary to achieve surveillance 3 ...Distribution... · What = Clinical signs in affected animals • Who = Type of animals that are affected Descriptive Epidemiology · Where = Place where the disease is found • When = Date when clinical signs appear • Why/How = Cause / Risk factors 4 ...Determinants of Disease... · What · Who · Where • When • Why = Possible risk factors **Analytic Epidemiology** • How = Modes of transmission

... Health- and Disease-Related States or Events... · Laboratory diagnosis • FMD laboratory confirmation • Indicators of disease · Mortality, production • Syndromes or signs drops · Risk factors for disease · Animal movement, lack of · Ancillary data biosecurity · Animal census data 6 What is a Population? · A collection of individual elements (i.e. units) that share common traits, such - Attributes (species, age, sex, breed, production, etc) - Place - Time Cattle Sheep Goats Milk Meat Dual purpose Breeder Example of populations with corresponding subpopulation Meat producing Eggs producing Poultry Wildlife (Carnivores) 7

Populations: A Collection of Elements

- 1. All cattle in a country
- 2. All animals susceptible to PPR
- 3. Three chickens in a cage on a poultry farm
- 4. All fish in the Atlantic ocean
- 5. All buffaloes in a national park
- 6. All pigs in a village on 1 January 2009

Unit of Interest

- · Unit of interest:
 - Refers to what we actually identify and count when we conduct surveillance $\,$
 - Requires that we are able to identify each element of a population
 - · Identification numbers or names are required for each unit (e.g. animal, herd and farm)
 - Need to specify at what level we are measuring
- · Examples of animal health units of interest:
 - Individual animal or person
 - $^{\circ}$ Herd or flock
- $^{\circ}$ Village, local area, region, etc.



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What Do We Count?

| Local Area | Total Cattle | Beef Cattle | Milking Dairy Cows | Sheep | Swine | Broilers | Egg Layers | TOTAL |
|------------|--------------|-------------|-----------------------|--------|--------|----------|------------|---------|
| A | 18,000 | 8,000 | 500 | 4,224 | 4,581 | | 1,556 | 28361 |
| В | 15,000 | 10,000 | | 6,336 | 120 | | 133 | 21,589 |
| С | 12,000 | 1,000 | 3,300 | 71 | 27 | 150 | 229 | 12,477 |
| D | 60,000 | 16,000 | 17,900 | 6,722 | 2,362 | | 764 | 69,848 |
| E | 55,000 | 20,000 | 16,200 | 3,601 | 1,561 | | 1,552 | 61,714 |
| F | 7,000 | 4,000 | | 1,607 | 1,128 | | 6,133 | 15,868 |
| G | 44,000 | 25,000 | | 4,138 | 913 | | 459 | 49,510 |
| н | 32,000 | 9,000 | 10,200 | 11,146 | | | 358 | 43,504 |
| 1 | 18,000 | 10,000 | | 9,418 | 2,408 | 510 | 4,451 | 34,787 |
| J | 67,000 | 46,000 | | 7,055 | 143 | | 359 | 74,557 |
| TOTAL | 328,000 | 149,000 | 48,100 | 54,318 | 13,243 | 660 | 15,994 | 412,215 |

10

What is Case Definition?

A set of standard criteria for deciding whether a unit of interest has a disease

When Do We Use Case Definitions? · For surveillance · For reportable diseases · For outbreak investigations · For epidemiological studies - For disease eradication programs Case definitions will vary for a given disease depending on the purpose for the collection of data 12 Case Definition Objectives 1. To create systematic method to classify units of interest in a population with respect to a disease Link the field and the laboratory diagnostic components in defining a 'case' through "two-way linking" 13

Components of a Case Definition

- · Clinical criteria

 - The characteristic symptoms of the disease Are not intended for use for individual clinical diagnosis
- · Production criteria
- Early, subclinical signals due to production drops, feed and water intake
- · Epidemiologic criteria
- Defined epidemiological links
- · Laboratory criteria
 - Specific test must be named (i.e., PCR, culture)
 - * Positive and negative results must be defined
- · Case classification criteria
- Suspect, probable, confirmed

Rift Valley Fever (RVF) in Kenya: 2015 - 2016

| Type | Definition | |
|-------------------------------|---|---|
| Suspect Case | A suspect herd is one reporting abortion (clinical signs) in any of the livestock from herds in an area experiencing heavy rainfall and flooding (ancillary data) | Probable RVF Cases Suspects RVF Cases |
| Probable Case | A probable RVF herd is defined as a herd reporting abortions and/or hemorrhagic signs (clinical signs), mortalities (production indicator) in the young ones, in any of the livestock in the herd in an area experiencing heavy rainfall and flooding (ancillary data) | 1 - 2 - 2 - 3 - 5 - 5 - 5 - 5 - 5 - 5 - 5 - 5 - 5 |
| Confirmed Positive Case | A confirmed RVF herd is defined as a herd where an animal tested positive to RVF by RVF IgM | Reference: Oyas et al 2018 |

Exercise 2: Develop Case Definitions

 $\label{lowing-information-in-loss} In pairs, please answer the following information and share in a plenary. This exercise should take 10 minutes total – 6 for developing case definition and 4 for sharing in plenary$

- Develop suspect, probable and confirmed case definitions for one of the following diseases in animals
 Rabies in wild carnivores
 Food and Mouth Disease in Goats and sheep
 Anthrax in cattle
 Highly Pathogenic Influenza in birds

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Types of Data Collected in Surveillance

- · Identifying information
- · Demographic information
- · Clinical information
- · Exposure / risk factor information
- Reporter information (name of laboratory/clinic)

Detecting Disease "Events" - Passive Reporting - Effective and efficient - Rumor tracking - Reports from owners, veterinarians or allied industry - Laboratory - Active Reporting - Existing disease surveillance programmes - Zero-reporting (reporting negative results especially during an outbreak))

Methods for Gathering Surveillance Data

Passive surveillance

- Incentive to report**
- Awareness of what to report
- · How to report
- Access to supportive infrastructure (laboratory)

Active surveillance

- Resource intensive (manpower)
- Probability based sampling
- Potentially less subject to bias
- \cdot Targeted
- · Expensive!

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Exercise 3: Data Reporting Active and Passive Surveillance

- This exercise should take 20 minutes.
- · Work in small groups of four persons per group.
 - 1. Review the disease scenario provided; and
- Answer the questions provided to describe how you would collect data both actively and passively.

In Summary...

- To achieve surveillance we use the epidemiologic approach targeting populations as opposed to individual animals.
- A population is a collection of individual elements (i.e. units) that share common traits. When conducting surveillance we distinguish between target, study and sample populations.
- * A case definition is essential to enable us to count positive cases of disease or disease related events.
- ${\boldsymbol{\cdot}}$ Surveillance data can be gathered actively or passively.

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Partners Contributors Contributors Contributors

Exercise 2 - Creating Suspect, Probable, and Confirmed Case Definitions

Description of Exercise:

Develop suspect, probable and confirmed case definitions for one of the following diseases in animals

Rabies in wild carnivores

Foot and Mouth Disease in Goats and sheep

Anthrax in cattle

Highly Pathogenic Influenza in birds;

Time: 25 minutes

Organisation of Group Work:

For each group, create the suspect, probable and confirmed case definitions in MS Word or MS PowerPoint and be prepared to share your list.

Exercise Components and Structure:

For each group, create the suspect, probable and confirmed case definitions in MS Word or MS PowerPoint and be prepared to share your list. for one of the following diseases in animals:

Rabies in wild carnivores

Foot and Mouth Disease in Goats and sheep

Anthrax in cattle

Highly Pathogenic Influenza in birds;

Materials, Data or Information:

1. MS Word or MS PowerPoint

Expected Outputs and Deliverables of Each Participant:

- 1. Provide examples of active and passive surveillance.
- 2. Show how surveillance data flows including two-way linking between field and laboratory data.

| Disease | Suspect | Probable | Confirmed |
|---|---------|----------|-----------|
| Rabies in wild carnivores | | | |
| Foot and Mouth Disease in Goats and Sheep | | | |
| Anthrax in cattle | | | |
| Highly Pathogenic Influenza in birds | | | |

Exercise 3 - Use of Active and Passive Surveillance for Brucella Mellitensis in Sheep and Goats

Description of Exercise:

Review the scenario provided and answer the questions related to use of active and passive surveillance. Should you have any questions, please ask a trainer for clarification before during, and after the exercise.

Work in pairs to answer the questions for the data report forms.

Exercise Components and Structure:

- 1. This exercise should take 20 minutes.
- 2. Work in small groups of four persons per group. Be prepared to report back to the class to share your group responses.
- 3. Review the disease scenario provided; and
- 4. Answer the questions provided to describe how you would collect data both actively and passively.

Materials, Data or Information:

- 1. Scenario for Exercise 3.
- 2. Provide responses in MS Word or MS PowerPoint

Expected Outputs and Deliverables of Each Participant:

- 1. Provide examples of active and passive surveillance.
- 2. Show how surveillance data flows including two-way linking between field and laboratory data.

Scenario

Background (Reference: Franc et al. BMC Public Health (2018) 18:125 DOI 10.1186/s12889-017-5016-y):

In 2013, a study. was conducted to estimate the economic impact of brucellosis in the developing nations of Africa and South/ Southeast Asia. In all, 259 studies on brucellosis from these regions were analyzed and encompassed observations from 500,000 animals, 30,000 people, and 600 food samples. The data revealed an average prevalence range of 0–88.8% in sheep and goats, 0–68.8% in cattle, 0.4–20% in camels, and 0–12.9% in other species (pigs and dogs).

In the case of brucellosis, visible losses include livestock abortion, reduced milk production, lost draught power, reduced weight gain from chronic infections and ill-thrift, premature death or culling of unproductive stock, veterinary costs associated with treating clinically ill animals and diminished animal welfare. In endemic areas, Brucella spp. can cause a significant reduction in herd productivity that compromises food security and the livelihood of farmers who depend on the sale or trade of surplus meat, dairy, and offspring from their animals.

Scenario:

You receive a call from the local health department concerning a rise in the number of human cases of B. mellitensis who went to the local hospital in your local area that are confirmed by the national laboratory based on positive bacterial culture.. Twice as many females as males are affected and there has been a significant increase in human abortions has occurred during the past 3 months. Brucellosis is a reportable animal health disease and is a high priority disease in your country. Your veterinary supervisor in the national government has asked you to become a member of a special task force that has been created by the Ministry of Health and the Ministry of Agriculture to investigate the presence of B. mellitensis in your local area.

Please answer the following questions:

- 1. Is the discovery of B. mellitensis in humans from the local area hospital an example of active or passive surveillance? Please explain your answer.
- 2. Your supervisor would like you to conduct ongoing surveillance of the local abattoirs to identify sheep and goats that react positively to the rose bengal test. Is the surveillance passive or active? Briefly explain your answer.
- 3. The local health department has decided to conduct as survey of the abattoir workers and sheep and goat farmers in your local area. They would like to collaborate with you and have asked you to collect information about abortion history and chronic illness in some selected sheep and goat herds.
 - a. Is the farm survey active or passive? Briefly explain your answer.
 - b. Some farmers choose to contact you following the survey to report cases of of abortion in their sheep and goats of their own free will. Is this voluntary reporting by farmers active or passive? Briefly explain your answer.

Lesson 3 – Defining and Counting Disease Cases

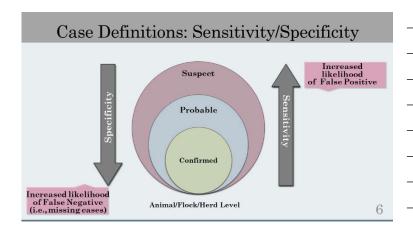
| 8 | 8 |
|------------------------------------|--|
| Estimated Lesson and Exercise Time | 2 hours |
| Participant Materials | Frontline ISAVET Lesson 3 Defining and |
| | Counting Disease Cases Participant |
| | Guide.PDF |
| | Microsoft Word and Excel |
| | Computer |
| Handout Materials for Exercises | Frontline ISAVET_PivotTable |
| | Handout.PDF |
| | Frontline ISAVET_PivotTable.xls |
| | |

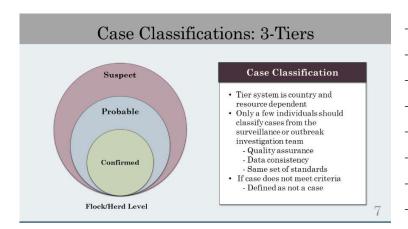
| In-Service Applied Veterinary Epidemiology Training (ISAVET) Lesson 3: Defining and Counting Disease Cases | | |
|--|---|--|
| INTERIOR DE LOS DELOS DE LOS DELOS DE LOS DELOS DE LOS DELOS DE | Food and Agriculture Organization of the United Nations | |

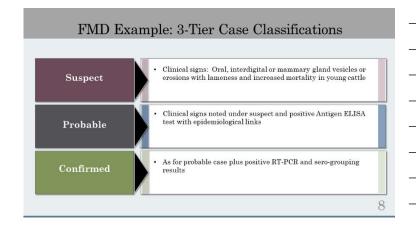
Case Definitions A set of standard criteria for deciding whether a unit of interest has a disease Uses of Case Definition? 1. Reportable diseases 2. Disease control/eradication programmes 3. Outbreak investigation

Case Definitions will vary depending on the purpose for the collection of data Surveillance Surveillance Standard across a country or area Revised as more is learned about the disease Revised as laboratory tests improve Comparisons can be made across animal (herd/flock), place, and time Outbreak Investigation Different for every outbreak Contains specific time and place constraints Revised as more is learned about the outbreak Comparisons might be possible with surveillance data

| Clinical | Characteristic clinical signs, progression and duration of the disease |
|----------------|---|
| | Used at herd/flock level not at the individual animal level |
| Epidemiologic | Determined epidemiological exposures and linkages |
| Laboratory | • Specific test must be named (e.g., PCR ELISA) • Positive and negative results must be defined |
| 3-Tier Case | • Suspect |
| Classification | Probable Confirmed |







Exercise 4: Case Definitions

- · This exercise should take 30 minutes.
- Divide into four groups of roughly equal size. Select one spokesperson from your groups to report on outcomes.
- · Develop a 3 tiered case-definition.
- Provide examples of a suspect, probable and confirmed case definition for one of the following diseases
- · Rabies
- \cdot Anthrax
- * Foot and Mouth Disease
- Please provide examples of wildlife where applicable

9

Case Counts

- Represents a number of suspect, probable or confirmed cases
- · Can be calculated from a line listing
- It may be affected by underreporting and how certain or inclusive is the case definition is that is being applied.

10

What is a Line List?

- A list of each possible case in a field investigation, with detailed information presented in a consistent manner
 - Provides information on possible cases in an ordered and uniform fashion (species, gender, date of onset, etc.)
- · Used to evaluate if each suspect case meets the case definition

| Herd ID | Species | Gender | Onset Date | Current Status | Case Category | Location | Epidemiologica l Linkage |
|------------|---------|--------|---------------|-------------------|------------------|------------|-----------------------------|
| 1 | Caprine | Male | 7/2/2018 | Clinical signs | Probable | District A | No vaccination |
| 2 | Caprine | Male | 7/2/2018 | Clinical signs | Suspect | District A | Vaccination |
| 3 | Ovine | Male | 7/3/2018 | Clinical signs | Confirmed | District B | No vaccination |
| 4 | Caprine | Male | 7/402018 | Dead | Confirmed | District C | No vaccination |

Creating and Managing a Line Listing

- Creation Paper (hard copy) Computer (electronically)
- Information received from field questionnaires
 Animal (Herd/Flock/Clinical signs/Species/Production class)
 Time (Date of onset of clinical signs and mortality/date of visit)
 Place (Geographic coordinates)
- $\boldsymbol{\cdot}$ Should include the components of the developed case definition of the outbreak
- · Provides a quick assessment of different aspects of an outbreak
- Refer to Excel YouTube Video # 3.1: Creating a Line Listing by Hand
- · Refer to Excel YouTube Video #3.2: Creating a Line List in MS Excel

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Line Listing

| Herd ID | | | Onset Date | Current Status | Case Category | | Epidemiological Linkage |
|------------|---------|------|---------------|-------------------|------------------|--------------|----------------------------|
| 1 | Caprine | Male | 7/2/2018 | Clinical signs | Probable | Local area A | No vaccination |
| 2 | Caprine | Male | 7/2/2018 | Clinical signs | Suspect | Local area A | Vaccination |
| 3 | Ovine | Male | 7/3/2018 | Clinical signs | Confirmed | Local area B | No vaccination |
| 4 | Caprine | Male | 7/402018 | Dead | Confirmed | Local area C | No vaccination |

Herd ID – Unique identifier assigned to each species for a investigation
Species – Species under investigation
Gender – Male or female
Onset date – Date of clinical signs onset
Current status – No clinical signs, clinical signs, or dead
Case category – Confirmed, probable, or suspect
Epidemiology linkage – Exposure or connections with other cases

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Line Listing (Continued)

| Herd ID | | | Fever of Unknown Origin | | |
|---------|---------|----------|-------------------------------|----------|----------|
| 1 | Caprine | Negative | Negative | Negative | Positive |
| 2 | Caprine | Negative | Positive | Negative | Positive |
| 3 | Ovine | Positive | Negative | Positive | Positive |
| 4 | Caprine | Negative | Positive | Positive | Positive |

Herd ID – Unique identifier assigned to each herd for an investigation
 Clinical Signs – Varies by disease

Line Listing (Continued)

Reporting Province or Local area: Local areas A-C $\,\,\,\,\,$ Date of Initial Report:_7/3/2018

| Herd ID | | Testing Requested | Results (Screening) | Results (Confirmatory) |
|---------|--------------------|----------------------|------------------------|---------------------------|
| 1 | Blood, Serum | Serology | ****** | ****** |
| 2 | Blood, Serum | Serology | Negative | |
| 3 | Blood, Serum, Swab | Serology | Positive | Positive |
| 4 | Blood, Serum, Swab | Serology | Positive | Positive |

- Herd ID Unique identifier assigned to each herd for an investigation
 Specimens collected Examples include blood, urine, serum, swab etc.
 Testing requested Examples include: culture, antigen detection, serology, PCR Results (screening) Findings of field laboratory testing
 Results (confirmatory) Finding of diagnostic laboratory testing

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MS Excel PivotTable: A Tool to Review Your Count Data

- · MS Excel tool that can calculate, summarise and analyse data in your MS Excel worksheet
- · Allows for a 2x2 cross tabular table in MS Excel
- · Can develop multiple graphical displays from this tool
- · Allows you to see comparisons, patterns, and trends in your data
- · Provide information for case counts
- · Allows you to identify data errors and/or missing data
- Refer to Excel YouTube Video # 7: Developing a Pivot Table

Developing a PivotTable from a Line Listing

- Step 1: Click the Insert Tab
- Step 2: Select the data
- Step 3: Click PivotTable Icon
- · Step 4: Select new worksheet. Click Ok.
- Step 5: Drag and drop variables to pivot table squares



- Filter: Stratifies data (looks at counts by each category of variable)
 Rows: Sets variable up in the exposure status of 2X2 table
 Columns: Set variable up in the outcome status of 2X2 table
 Values: Sets variable up as a count (will need to be changed from "sum" to "Count"

Demonstration: Developing a PivotTable From a Line Listing Foot-and-Mouth Disease MS Excel Demonstration of Developing a PivotTable Microsoft Excel Worksheet From our line listing, lets determine how many cases are classified as suspect, probable, and confirmed in each Country using the PivotTableTool 18 Exercise 5: Line Listing · This exercise should take 45 minutes. · Divide into groups of three participants. · Develop a line listing from the data provided in L3_Ex5_1.xlsx as follows: · Create and enter by hand first $^{\circ}$ Create and enter using MS Excel. 19 Underreporting of Disease · Notifiable/reportable diseases are usually based on passive reporting - Result: only a fraction of cases overall are reported · What are the causes of under-reporting?

Underreporting of Disease

- · Underreporting occurs due to:
 - Lack of knowledge of reporting requirements
 - Lack of awareness of responsibility to report or which diseases must be reported
 - Lack of awareness of how or to who to report to
- Lack of access to the field
- Assumption that another group (i.e., the laboratory) will report

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Underreporting of Disease

- Underreporting can occur due to a negative attitude towards reporting
 - Take to much time
 - Lengthy and complex report or procedure
 - Lack of incentive
 - Lack of feedback to information sources
 - Distrust in the government
- Concern than report might result in an economic issue
- Disagreement with need to report
- Lack of access to remote areas

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In Summary...

- Case definitions identify what is and is not a case and what we count
- · Case definitions include four common criteria
- The type of case definition needed is dependent upon the objectives at the time and can have the draw back of having too many or too many false positives
- ${\cdot}\, A$ line listing provides an up to date "pulse" of an outbreak situation for case classifications.
- PivotTables are a powerful tool to calculate, summarise, and analyse data that lets you see comparisons, patterns, and trends in your data
- · Always consider factors which can cause underreporting!

| ISAVE | T Contributing Universities | | |
|--------------|---------------------------------|----|--|
| Partners | TEXAS ASM TEXAS ASM TEXAS ASM | | |
| | TIGRILIFE | | |
| | John V. Dr. day. | | |
| Contributors | MAKERBE UNIVERSITY | | |
| | MAKERER UNIVERSITY | 24 | |

Exercise 4 – Case Definition

Description of Exercise:

Based on the information provided in the handout, use the following exercise to develop a case definition for brucellosis or theileriosis (east coast fever). Use the example provided in the case definition lecture as a reference for how a case definition should be structured and what information should be included at each level (i.e., 3-tier case definition). Should you have any questions over the exercise, please ask a trainer for clarification before during, and after the exercise.

Allotted Time: 45 minutes

Exercise Components and Structure:

- 1. Develop the following descriptions for your case definition.
 - a. Case classification (3-tiered case definition)
- 2. After 45 minutes, each spokesperson will present their findings.

Organization of Group Work:

- 1. Divide into four groups. Select one spokesperson from your group to report outcomes to the entire group.
 - a. Group A Conducting routine surveillance for brucellosis in your country.
 - b. Group B Conducting outbreak investigation for brucellosis in your local area.
 - c. Group C Conducting routine surveillance for east coast fever in your country.
 - d. Group D Conducting outbreak investigation for east coast fever in your local area.

Materials, Data or Information:

- 1. Internet access
- 2. Computer with Microsoft Word
- 3. Paper and pen

Expected Outputs and Deliverables of Each Participant:

1. Provide examples of a suspect, probable, and confirmed case definition.

Group A Conducting Routine Surveillance for Brucellosis in Your Country

| | 1. Case Classification (3-Tiered Case Definition): Suspect: |
|----|--|
| | Probable: |
| | Confirmed: |
| Co | Group B Inducting Outbreak Investigation for Brucellosis in Your Local area |
| 1. | Case Classification (3-Tiered Case Definition): Suspect: |
| | Probable: |
| | Confirmed: |

Group C Conducting Routine Surveillance for East Coast Fever in Your Country

| 1. | Case Classification (3-Tiered Case Definition): Suspect: |
|---------|--|
| | Probable: |
| | Confirmed: |
| Conduct | Group D ing an Outbreak Investigation for East Coast Fever in Your Local area |
| 1. | Case Classification (3-Tiered Case Definition): Suspect: |
| | Probable: |
| | Confirmed: |
| | |

Exercise 5 – Create a Line Listing in MS Excel Using the Field and Laboratory Data Provided

Description of Exercise:

Based on the information provided in the handouts, use the following exercise create a line listing in an electronic format (MS Excel) and paper-based format using the field case reports and laboratory information for a specific disease. Use the example provided in the case definition lecture as a reference for how a line listing should be structured and what information should be included. Should you have any questions over the exercise, please ask a trainer for clarification before during, and after the exercise.

Allotted Time: 30 minutes

Exercise Components and Structure:

- 1. Determine the variables that should be included in a line listing from field and laboratory case reports.
- 2. Create a line listing.
- 3. Use a PivotTable in MS Excel to obtain values of count data from variables in a line listing.

Organisation of Group Work:

- 1. Each group has 45 minutes to complete the work.
- 2. Divide into groups of three participants.
 - a. Participants can choose from two case report types to develop their line listing.
 - b. The line listing should be developed at the herd level.
 - c. Each group will develop a paper-based and electronic format of their line listing.

Materials, Data or Information:

- 1. Computer with Microsoft Word and Microsoft Excel
- 2. Participant Exercise ISAVET_Exercise_5_Line_Listing_ECF_Participant
- 3. Paper and pen

Expected Outputs and Deliverables of Each Participant:

- 1. Develop a line listing from field and laboratory data for a disease.
- 2. Be able to count and sort data using a pivot table in an electronic format.

Field Case Report and Screening Test Results: East Coast Fever

Review the following three field case reports from local area A.

| Field Case | Report for Country X | |
|--|---|-----------------------------------|
| Case Report Number: 1 Date of Visit: 9/2/2018 | Field Inspector: B. Mbasa | Region: Central Sub-Local area: A |
| Premise ID: 1 Herd ID: 1 Age: Not recorded Farm Production Type: Commercial beef with a hobby-scale goat | GPS Coordinates: Longitude: 22.324761 Latitude: 63.452188 | |

| Species | Total Number at Risk | Number Sick | Number Dead | Age (M) = Mature (Y) = Young | Breed | Sex (F) = Female (M) = Male | |
|------------------------|----------------------------|----------------|----------------|-----------------------------------|------------|---|--|
| Dairy Cattle | | | | | | | |
| Beef Cattle | 15 | 13 | 3 | M | Angus | F | |
| Ducks | | | | | | | |
| Poultry | | | | | | | |
| Sheep | | | | | | | |
| Goats | 4 | 0 | 0 | Y | Boer | F | |
| Swine | | | | | | | |
| Herd/Flock Le Signs | vel Clinical | Yes | No | N | lotes: | | |
| Anorexia | | ✓ | | Clinical signs of | nly seen i | n beef | |
| Loss of body | condition | ✓ | | cattle. No other species affected | | | |
| Lacrimation | | ✓ | | on farm. | | | |
| Corneal opac | ity | ✓ | | | | | |
| Nasal discha | rge | ✓ | | | | | |
| Dyspnoea | | ✓ | | | | | |
| Diarrhoea | | ✓ | | | | | |

Case History

1. What diseases are present in the local area?

Trypanosomiasis, East Coast Fever, Anaplasmosis, Babesiosis, and Brucellosis

2. What were the dates for the following?

• Symptoms first observed: 8/31/2018

• First death: 9/2/2018

• Laboratory submission: 9/2/2018

| Field Case | Field Case Report for Country X | | | | | | | | | | | |
|---|---|-----------------------------------|--|--|--|--|--|--|--|--|--|--|
| Case Report Number: 2 Date of Visit: 9/3/2018 | Field Inspector: C. Raseasala | Region: Central Sub-Local area: A | | | | | | | | | | |
| Premise ID: 2 Herd ID: 2 Age: Not recorded Farm Production Type: Hobby- Scale | GPS Coordinates: Longitude: 23.224578 Latitude: 74.312217 | | | | | | | | | | | |

| Number at Risk | Number Sick | Number Dead | Age (M) = Mature (Y) = Young | Breed | Sex (F) = Female (M) = Male | |
|-------------------|---------------------------------|---|--|--|--|--|
| | | | | | | |
| 4 | 2 | 0 | M | Angus | F | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| vel Clinical | Yes | No | Notes: | | | |
| | | | | | | |
| | ✓ | | | | | |
| ondition | ✓ | | | | | |
| | | | | | | |
| ty | | | | | | |
| ge | ✓ | | | | | |
| | ✓ | | | | | |
| | ✓ | | | | | |
| | Risk 4 vel Clinical ondition | Risk 4 2 vel Clinical Yes ondition ty ge ✓ | Risk 4 2 0 vel Clinical Yes No ondition ty ge ✓ | Risk Mature (Y) = Young 4 2 0 M vel Clinical Yes No No ondition ✓ V ty ✓ ✓ ✓ ✓ ✓ | Risk Mature (Y) = Young 4 2 0 M Angus vel Clinical Yes No Notes: ondition ✓ Ondition ✓ | |

Case History

3. What diseases are present in the local area?

Trypanosomiasis, East Coast Fever, Anaplasmosis, Babesiosis, and Brucellosis

4. What were the dates for the following?

• Symptoms first observed: 9/1/2018

• First death: Not applicable

• Laboratory submission: 9/3/2018

| Field Case | Field Case Report for Country X | | | | | | | | | | | |
|---|---|-----------------------------------|--|--|--|--|--|--|--|--|--|--|
| Case Report Number: 3 Date of Visit: 9/3/2018 | Field Inspector: R . Mutowembwa | Region: Central Sub-Local area: A | | | | | | | | | | |
| Village ID: 3 Herd ID: 3 Age: Not recorded Farm Production Type: Hobby- Scale | GPS Coordinates: Longitude: 99.115678 Latitude: 88.257322 | | | | | | | | | | | |

| Species | Total Number at Risk | Number Sick | Number Dead | Age (M) = Mature (Y) = Young | Breed | Sex (F) = Female (M) = Male | |
|----------------|----------------------------|----------------|----------------|--|----------------|---|--|
| Dairy Cattle | 3 | 2 | 1 | M | Holstein | F | |
| Beef Cattle | | | | | | | |
| Ducks | | | | | | | |
| Poultry | | | | | | | |
| Sheep | | | | | | | |
| Goats | 15 | 10 | 0 | M | LaMancha | F | |
| Swine | 2 | 0 | 0 | Y | Duroc | F | |
| Herd/Flock L | evel | Yes | No | Notes: | | | |
| Clinical Signs | S | | | | | | |
| Anorexia | | ✓ | | Late term abo | rtions seen ii | n goats. | |
| Loss of body | y condition | ✓ | | Clinical signs | in all species | s, but | |
| Lacrimation | า | | | swine. | | | |
| Corneal opa | ncity | | | | | | |
| Nasal disch | arge | | | | | | |
| Dyspnoea | | | | | | | |
| Diarrhoea | | | | | | | |

Case History

5. What diseases are present in the local area?

Trypanasomiasis, East Coast Fever, Anaplasmosis, Babesiosis, and Brucellosis

6. What were the dates for the following?

• Symptoms first observed: 9/1/2018

• First death: 9/3/2018

• Laboratory submission: 9/3/2018

1. What categories of information should be collected from each of the case reports and transferred to a line listing? Identify specific variables for each category.

| Category | Variable(s) |
|----------|-------------|
| | |
| | |
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| | |
| | |
| | |
| | |
| | |
| | |
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| | |

2. Create a line listing from the three field case reports using MS Word.

| | Animal (Herd/Flock) | | | | | | | | | | | | | | | | |
|------------|---------------------|---------|--------------------|-------------------------|-------------|-------------|-----|-------|-----|-------|---------|-------|-----------|-----------------|--------------------------|-----------------|----------------|
| | | | | | | | | | | | | Cli | nical S | igns | | | |
| Premise ID | Herd ID | Species | Production Type | Total No. At Risk | No. Sick | No. Dead | Age | Breed | Sex | A^1 | LBC^2 | L_3 | ${f C}^4$ | ND^5 | $\mathrm{D}\mathrm{y}^6$ | Di ⁷ | \mathbf{C}^8 |
| | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | |
| | | | | | | | | | | | | | | | | | |

¹ A= Anorexia; ² LBC = Loss of Body Condition; ³ L= Lacrimation; ⁴ C = Corneal Opacity; ⁵ ND = Nasal Discharge; ⁶ Dy = Dyspnoea; ⁷ Di = Diarrhoea; ⁸ C = Classification (Suspect, Probable, or Confirmed)

| | Time | | | | | | | Place | Other | | |
|------------|------------|------------------|------------------------------|------------------------|---------------------------|---------|--------|---------------|--------------------|-------------------|----------------------|
| Premise ID | Herd ID | Date of Visit | Date Symptoms First Observed | Date of First Death | Date of Lab Submission | Country | Region | Local area | GPS Coordinates | Screening Test | Confirmatory Test |
| | | | | | | | | | | | |
| | | | | | | | | | | | |

Open the MS Excel Spreadsheet titled

"ISAVET_Exercise_5_Line_Listing_ECF_Participant". In the first worksheet tab, there are six rows for which herd-level cases should be created from the field case reports. Enter the data from the five field case reports into the line listing on your electronic spreadsheet.

The following laboratory results were obtained in the field from screening tests.

| | | Other | | |
|---------------|------------|-------------------|----------------------|--|
| Premise ID | Herd ID | Screening Test | Confirmatory Test | |
| 1 | 1 | Positive | | |
| 1 | 1 | | | |
| 2 | 1 | Negative | | |
| 3 | 1 | Negative | | |
| 3 | 1 | Positive | | |
| 3 | 1 | | | |

3. Add the screening test results to the line list and determine what other laboratory results should be included in the line list for confirmation.

What does this change the line list classification for each of these?

| Premise | Previous | Changed To |
|---------|----------------|------------|
| ID | Classification | |
| 1 | Suspect | |
| 1 | Not a Case | |
| 2 | Suspect | |
| 3 | Suspect | |
| 3 | Suspect | |
| 3 | Not a Case | |

Make the changes to the classification status in your spreadsheet

Using a PivotTable, identify the number of cases for the following questions.

- a) How many herds are suspect?
- b) How many herds are probable?
- c) How many herds are confirmed?

Samples were taken and submitted to the lab for confirmation. On the 5th day of the outbreak, the following results for the herd IDs came back with the following confirmatory laboratory results.

| | | Other | | |
|---------|------|-----------|--------------|--|
| Premise | Herd | Screening | Confirmatory | |
| ID | ID | Test | Test | |
| 1 | 1 | Positive | Positive | |
| 1 | 1 | | | |
| 2 | 1 | Negative | Positive | |
| 3 | 1 | Negative | Positive | |
| 3 | 1 | Positive | Positive | |
| 3 | 1 | | | |

- 4. Add the above test results to your electronic line listing.
- 5. After confirmation, make the correct changes to the classification status in your worksheet.
- 6. After confirmation testing, how many herds are confirmed positive for theileriosis (east coast fever)?

Lesson 4 – Data Quality Principles

| Estimated Lesson and Exercise Time | 1 hour |
|------------------------------------|--|
| Participant Materials | ISAVET Lesson 4 – Data Quality Principles Participant Guide. PDF Computer and Microsoft Word |
| | Pen or Pencil |

In Service Applied Veterinary Epidemiology Training (ISAVET)

Lesson 4: Data Quality Principles

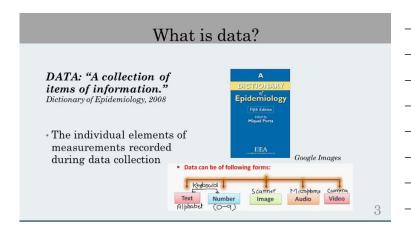


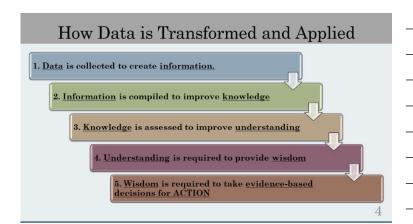


Learning Objectives

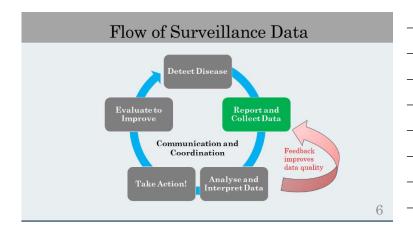
At the end of this lesson, you will be able to:

- 1. Apply the basic principles of ensuring data quality;
- 2. Review the quality of a data set systematically;
- 3. Provide feedback to improve data quality.





Data Quality Data quality: Refers to accuracy and completeness of data gathered and that they convey the intended meaning Begins by ensuring that data is gathered in a standard way Need to have a standard data collection form Garbage in – garbage out Good data is needed if we are to make good decisions



$Reasons \ for \ Poor \ Data \ Quality \\ {\tiny (Adapted: Frontline \ FETP, \ 2016)}$

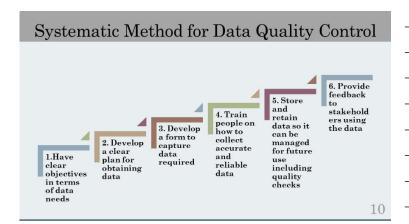
- · Poorly completed forms
- $\cdot \ Un\text{-entered forms}$
- · Underreporting
- Overreporting
- Duplicate reporting
- · Unsystematic data collection / reporting
- · Untruthful reporting
- Inconsistent reporting formats
- · Late submission/ reporting
- · Calculation errors
- · Lack of documentation
- · Data files are lost

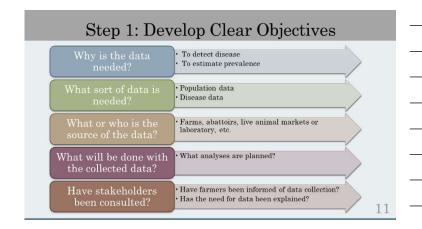
7

Example of Good Quality Data

| 1 | 1 | 1 | F | 2 | Friesian |
|---|---|---|---|---|----------|
| 1 | 1 | 2 | M | 4 | Jersey |
| 1 | 2 | 1 | M | 5 | zebu |
| 1 | 2 | 1 | F | 6 | zebu |

| Unique ID | Local Area | Farm ID | Animal ID | Sex | Age | Weight | Clinical signs | Date of visit |
|--------------|---------------|------------|--------------|--------|-------|---------|-------------------|---------------|
| 1 | 1 | 23 | 25 | f | | 64 | vomiting, pyrexia | 2017/1/12 |
| 2 | 1 | 23 | 25 | male | | 64 KG | pyrexia | 1/12/2017 |
| 3 | 1 | 1 | 2 | m | 1 | 100 lbs | diaroa | 11/1/2017 |
| 4 | 1 | 1 | 1 | female | 4 | | diarrhoea | 10/01/2017 |
| 4 | 1 | 12 | | 4 | 2 yrs | | death | 11/1/2017 |
| 6 | 1 | 25 | | femal | | | | 11/1/2017 |
| 7 | 1 | 2 | | mal | | | | 11/1/2017 |





| Step 2: Have a | Clear Data Collection Plan |
|----------------|--|
| What? | • What data needs to be gathered? |
| How? | Active versus passive collection Reporting methods |
| Who? | • Who will gather the data and are they trained in a standard way? |
| Where? | · Where will the data be stored? |
| When? | • Timelines of data collection |

| | Advantages | Disadvantages | |
|--------------------|---|---|--|
| 1. Hard copy form | Widely accessible for field use Retain a permanent copy | Handwriting quality varies Can be delayed, lost or damaged | |
| 2. Mobile phone | One step input process Timely submission No transcription required to electronic format | Variable phone signal access Input errors difficult to catch when entering data | |
| 3. Electronic mail | - Scanned copies can be sent directly to the destination | - Transcription required to electronic format | |

Step 4: Standardised Data Input Training

- Train data collectors to collect accurate and reliable data
 - Standard training to ensure personnel are given clear step-by-step instructions about each data element and how to enter the data $\,$
 - Standard training on interviewing methods to ensure data quality



Frontline ISAVET

Step 5: Store and Retain Data

· Hardcopy storage:

- Paper copies should be stored in a secure and safe location for a period of time specified by the veterinary authority
- Forms should be filed systematically so that they are accessible

$\cdot \ \underline{\textbf{Electronic storage:}}$

- Electronic data should be stored in a standard format (e.g. MS Excel, comma delimited, etc) $\,$
- Should be backed up and saved on an external drive at least weekly
- Ensure that anti-virus and security access is in working order

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Step 5: Data Management – Data Quality Checks

- ${}^{{}_{\diamond}}$ An important step is to check the quality of the data that has been gathered.
- . This includes:
- Ensuring the correct number of records have been entered
- Performing quality control checks on each data field related to:
- Errors: Incorrect values that have been recorded wrongly or entered/typed incorrectly into a database or spreadsheet
- ${\color{blue}\mathbf{Outliers}}.$ An extreme observation that does not appear to be in line with other observations
- · Missing data: Data not entered

| Data Quality Audit | | | | | |
|--|--|---|--|--|--|
| COMPONENT | DESCRIPTION | METRICS | EXAMPLES | | |
| Data Collection | Standarised line list for each priority, notifiable disease | Completeness, timeliness, accuracy, zero reporting, format | Sublocal area reports were 50% complete and 30% timely | | |
| Case Reporting | Classify cases as suspect (S), probable (P) or confirmed (C) | S,P, C case definitions exist for each priority, notifiable disease | Suspect case of HPAI: Hemorrhage, necrosis diarrheoa, discharges | | |
| Analysis and Interpretation | Graphic representations | Describe disease distribution, trendline with explanation | Increase in FMD cases spatially and temporally | | |
| Follow up Confirmatory Investigation | Laboratory and field investigations | Rapid confirmatory laboratory testing and field questionnaires | A suspect case of PPR confirmed by laboratory diagnosis | | |
| Reporting | Official reports for stakeholders | Internal and external reports and presentations | Report to national level Report to farmers | | |

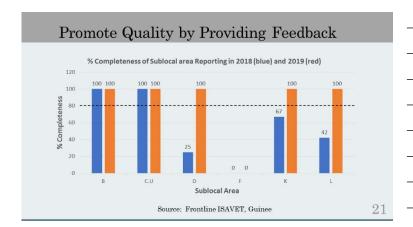
Step 6: Provide Feedback

- · All stakeholders:
 - Should be consulted when data collection is planned to ensure that the right data can be collected
 - Should be asked to provide feedback on the strengths and weaknesses of the data collection methods to ensure continuous quality improvement (COI)
- Results of data collected from stakeholders MUST be shared to provide an incentive to continue to support ongoing data collection

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Threats to Data Quality Lack of reporting No incentives to report leading to underestimation of disease cases reporting Design flaws Not collecting the right data Data collection Mot collecting data in the best way method Technical errors Incorrect entry, transfer and analysis of data errors Misinterpreting data Data sharing File formatting, data transfer and compiling data from different databases

Promote Quality by Providing Feedback! Table 1: Weekly Sublocal Area Reporting Summary of Completeness and Timeliness Subdistrict Name No. Reports Received This Week No.(%) Weekly Reports Received in [2019] Subdistrict/Facility A L 2 (20%) Subdistrict/Facility A Subdistrict/Facility B Subdistrict/Facility C T 7 (100%) Subdistrict/Facility B Subdistrict/Facility C T Subdistrict/Facility C Subdistrict/Facility C T Subdistrict/Facility C Subdistrict/Facility C Subdistrict/Facility C T Subdistrict/Facility C Subdistrict/Facility C Subdistrict/Facility C Subdistrict/Facility C T Subdistrict/Facility C Subdistrict



Exercise 6: Data Quality

- 1. This exercise will take 45 minutes.
- 2. Pair yourselves into groups of two.
- 3. Review the dataset provided.
- 4. Identify corrections from the dataset and identify all errors and omissions
- 5. Provide suggestions for improving data quality in the process just described in this lesson.
- 6. Plenary review with facilitated discussion with trainer

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In Summary...

- Understanding different data types allows us to correctly analyse and make use of the data
- \cdot High quality surveillance data is the basis of reliable information for good decision-making in animal health
- To ensure good quality data it is important to:
 - Have $\underline{\text{clear objectives}}$ about the data that is needed
 - Develop a <u>clear plan</u> about the best way of obtaining the data
- Use <u>standardised forms</u> or formats that can capture the data
- Train people about how to collect accurate and reliable data
- <u>Store, review and retain the data</u> so that you can retrieve it for future use
- Provide feedback to all stakeholders using the data

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Exercise 6 - Data Quality Principles

Review and Identify Corrections Required for Dataset with Errors and Omissions

Description of Exercise:

Using the data quality checklist, review and identify corrections required for a dataset with errors and omissions. Should you have any questions over the exercise, please ask a trainer for clarification before during, and after the exercise.

Allotted Time: 45 minutes

Exercise Components and Structure:

- 1. Work in pairs of two people.
- 2. You have 45 minutes to complete this exercise
- 3. For the dataset provided, review and identify corrections required for a dataset with errors and omissions

Materials, Data or Information:

- 1. Dataset of HPAI H5N1 Outbreaks in Nigeria
- 2. Record answers in MS Word or PowerPoint

Expected Outputs and Deliverables of Each Participant:

- 1. Follow a systematic method to review and correct data
- 2. List ways to improve the data set provided

Dataset Dictionary

HPAI H5N1 Outbreaks in Nigeria

Records from an H5N1 HPAI outbreak from Nigeria are presented in the dataset $Exercise_6.version\ 4.xls.$

Each row of the data in MS Excel spreadsheet represents an outbreak report (multiple rows per affected farm) and details of the fields can be found in Table 1.

Table 1. Details of variables in the *Exercise_6.xls*.

| Variable | Description |
|--------------------------|--|
| farmerid | Unique identifier for each farmer |
| LGA | Local government area |
| GPS location | Geographic coordinates of farm |
| Location | Farm name |
| farmtype | Type of farm |
| Type of birds | Bird types on farm |
| Age | Age of birds on farm |
| flocksize | Bird population |
| Susceptiblepop | Number of susceptible birds |
| Date of onset | Date clinical signs observed |
| Date reported | Date reported |
| Mortality | Number of dead birds |
| Number slaughtered | Number of birds slaughtered |
| Bases for diagnosis | How cases were diagnosed |
| Date samples sent to lab | Date samples were sent to the laboratory |
| Date of confirmation | Date outbreak confirmed |
| | Measures implemented to control the |
| Control measure | outbreak |
| Status | HPAI status |
| Remarks | Comments |

| farmerid | LGA | farmtype | Type of birds | Age | flocksize | Susceptiblepop | Date of onset | Date reported | | Mortality | Number slaughtered | Bases for diagnosis | Date samples sent to lab | Date of confirma tion | Control measure | Status | Remarks |
|----------|----------------------|----------|------------------------------|---|-----------|----------------|--------------------------|---------------|----------|--------------------------|--------------------|-------------------------------|--------------------------|-----------------------|--------------------|----------|---|
| 1 | Dala | layer | Layers | 22weeks, 10 wks, 4wks | 1568 | 1568 | 2/1/2015 | 2/1/2015 | 1/0/1900 | 1370 | 198 | Clincal and | 7/1/2015 | 8/1/2015 | Quarantine | Positive | |
| 1 | Dala | | Layers | WKS, 4WKS | | | 3/1/2015 | | | 350 | 66 | and | | | | | |
| 1 | Dala | | Layers | | | | 4/1/2015 | | | 50 | 63 | | | | | | |
| | | | Chickens | | | | | | | | | | | | | | |
| | | | (Local & exotic, | Different age | | | | | | | | Clincal and | | | | | |
| 2 | Sabon gari | mixed | geese, turkeys, | groups | 30,000 | 30,000 | 22/12/2014 | 24/12/2014 | | 919 | 450 | Laborator | 7/1/2015 | 8/1/2015 | Quarantine | Negative | |
| | | | peacock) | | | | | | | | | , | | | | | |
| 3 | Gwale | | Layers | 24wks | 570 | 570 | 11/1/2015 | 11/1/2015 | | 4 | 0 | Clincal and Laborator | 12/1/2015 | 13/1/201 5 | Quarantine | Positive | Farmer visited Rimi LBM on |
| | | | | | | | | | | | | У | | | | | 06/01/2015 |
| 3 | Gwale Gwale | | | | | | 12/1/2015 13/01/2015 | | | 48 120 | | | | | | | |
| | | <u> </u> | _ | | | | | - 1 - 1 | | | _ | Clincal | | 13/01/20 | | | |
| 3 | Manjibir | mixed | Growers | 12 weeks | 3,257 | | 8/1/2015 | 9/1/2015 | | 62 | 0 | and | 12/1/2015 | 15 | Quarantine | Positive | |
| 3 | Manjibir | | Pullets | 18 weeks | 2800 | | 13/01/2015 | | | 287 pullets | | | | | | | |
| 3 | Manjibir Manjibir | | Cocks Pullets | 10 weeks | 6000 | | 13/01/2015 15/01/2015 | | | 672 cocks 185 pullets | | | | | | Positive | |
| 3 | Manjibir | | cocks | | | | 15/01/2015 | | | 1105 cocks | | | | | | | |
| 4 | Kumbotso | | Pullets | 19 weeks | | | | | | | | Clincal and | 12/1/2015 | 13/1/201 5 | | Negative | |
| | | | | | | | | | | | | Clincal | 15/01/201 | | | | There are |
| 5 | Gwale | layer | Layers | 20 weeks | 3500 | 3500 | 11/1/2015 | | | 16 | 0 | and Laborator | 5 | Pending | Quarantine | Pending | some bags of feed |
| 5 | Gwale | | Layers in battery cage | Adult laying birds, Apparently healthy | 175 | | 12/1/2015 | | | 86 | | | | | Quarantine | | |
| 6 | Kumbotso | layer | Layers | Room 1- 19weeks (affected flock) | 1600 | 1600 | 9/1/2015 | 9/1/2015 | | 10 | 0 | Clinical and laboratory | 10/1/2015 | 15/01/20 15 | Quarantine | Positive | |
| 6 | Kumbotso | | Layers | Room 2 - 44 weeks | 1753 | | 10/1/2015 | | | 30 | | | 15/01/201 | Pending | | Pending | |
| 6 | Kumbotso | | Layers | Room 3 - 44 weeks | 680 | | 11/1/2015 | | | 102 | | | | | | | |
| 6 | Kumbotso | | Layers | Room 4 - 19 weeks | 1233 | | 12/1/2015 | | | 202 | | | | | | | |
| 6 | Kumbotso | | Layers | Room 5 - 7 weeks | 4884 | | 13/01/2015 | | | 578 | | | | | | | |
| 7 | Kumbotso | layer | Layers | 12 months | 1200 | 1200 | 7/1/2015 | 7/1/2015 | | 470 | 60 | Clinical and laboratory | 12/1/2015 | 13/01/20 15 | Quarantine | Positive | 670 live birds were taken to the livebird market, |
| 7 | Kumbotso | mixed | Pullets | 7 weeks | 300 | | | | | | | | | | | | 60 dressed birds were taken to Wudil LBM |
| 8 | Tofa | | Pullets (point of lay) | 20 weeks | 576 | | | | | | | | | | | | |
| 8 | Tofa | | Pullets (point of lay) | Geese | 7 | | | | | | | | | | | | |
| 8 | Tofa | | Pullets (point of lay) | Turkeys | 4 | | | | | | | | | | | | |
| 8 | Tofa | | Pullets (point of lay) | Guinea fowl | 40 | | | | | | | | | | | | |
| 8 | Tofa | | Pullets (point of lay) | Local chickens | 6 | | | | | | | | | | | | 77 |

1. Identify data quality issues that are found in the aforementioned dataset with a corresponding solution.

To be assigned as homework assignment and then follow the SOP provided:

Conduct data quality audit for any five selected variables and 10 randomly selected entries using the data quality audit tool provided and interpret your findings.



Lesson 5.1 – Describing and Acting upon Animal Disease and Health Data: Central Tendency

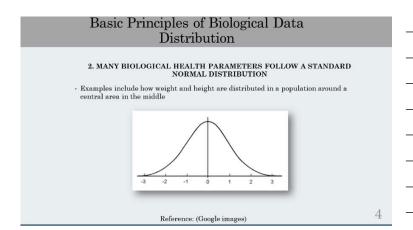
| Estimated Lesson and Exercise Time | 1 hour and 30 minutes |
|------------------------------------|--|
| Participant Materials | Frontline ISAVET Describing and Acting upon Animal Disease and Health Data: Central Tendency Participant Guide.doc Computer and Microsoft Word |
| | Pen or Pencil |
| Handout Materials for Exercises | MS Excel Lesson 5_Ex7 .xls (Lesson 7) |

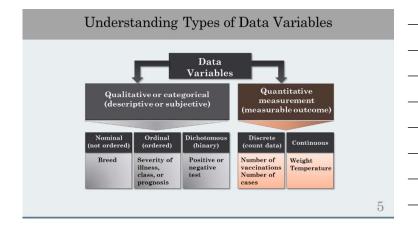
In Service Applied Veterinary Epidemiology Training (ISAVET) Lesson 5.1: Describing and Acting upon Animal Disease and Health Data: Central Tendency

Learning Objectives

At the end of this lesson, you will be able to: 1. Explain and calculate measures of central tendency: average, frequency, median, mode.

Basic Principles of Biological Data Distribution 1. DISEASE DOES NOT OCCUR RANDOMLY Epidemiological Triad: Risk factors create patterns of disease based on the relationship between agent, host and environment Host Agent Environment Reference: (Google images)





Describing Data: Frequency, Average, Median and Mode



Reference: https://www.africa-ugandahusiness.traval.mide.com/ A farm that you visit has 8 broiler chickens, and the farmer wants to find out how uniform the weights are since he suspects that the day old chicks are coming from different sources. You then weigh and record the weight of each of the 8 chickens.

| Animal | Weight |
|--------|--------|
| ID | (kg) |
| 1 | 2.5 |
| 2 | 1.0 |
| 3 | 2.5 |
| 4 | 2.0 |
| 5 | 1.5 |
| 6 | 1.5 |
| 7 | 6.0 |
| 8 | 2.0 |

6

7

Average

- · If you selected any one of the 8 chickens, about how much would it weigh?
- Weight is a continuous variable and we can calculate the arithmetic mean by doing the following:
- 1. Sum up all observation values
- 2. Divide the sum by the number of observations

Step 1: Add together the results of your observations

| Animal ID | Weight (kg) |
|--------------|----------------|
| 1 | 2.0 |
| 2 | 1.0 |
| 3 | 2.5 |
| 4 | 2.0 |
| 5 | 1.5 |
| 6 | 1.5 |
| 7 | 6.0 |
| 8 | 2.0 |
| Sum | 18.5 |

Average

Step 2: Calculate the average N = 8 chickens

Sum = 18.5 kg

Average = 18.5 kg /8 chickens X = 2.3 kg / chicken

What's wrong here?

Step 3: Plot the results

Chicken Weights

7

8

24

34

35

2 2

4 15

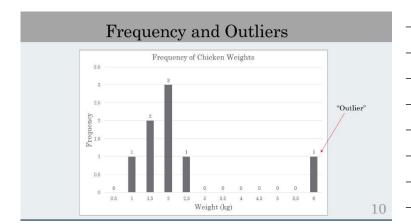
1 4 5 6 7 8

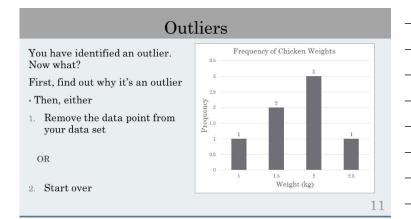
Animal ID

Frequency

Step 4: Calculate frequency How many times does each weight appear?

| Chicke | n Weights | | |
|----------------|-----------|---------------|---|
| Weight (kg) | Frequency | | |
| 0.5 | 0 | | |
| 1.0 | 1 | | |
| 1.5 | 2 | | |
| 2.0 | 3 | | |
| 2.5 | 1 | | |
| 3.0 | 0 | | |
| 3.5 | 0 | | |
| 4.0 | 0 | | |
| 4.5 | 0 | | |
| 5.0 | 0 | | |
| 5.5 | 0 | Reference: | C |
| 6.0 | 1 | Image: Google | č |





Class Exercise · Remove the outlier(s) Chicken Weights Animal Weight ID (kg) 1 2.0 Step 2: (Re)Calculate the average 1.0 3 2.5 N = 7 chickens 2.0 1.5 Sum = 12.5 kg6 1.5 Average = 12.5 kg /7 chickens 6.0 8 2.0 **X** = Sum 12.5 12

| Median | | | | | | | | | |
|--------------|----------------|-------------------|--------------|----------------|----------------------|--|--|--|--|
| Step 5: | Order th | e observatio | ons from l | owest to | o highest | | | | |
| Chicken | Weights | | Chicken | Weights | Step 6: Identify the | | | | |
| Animal ID | Weight (kg) | | Animal ID | Weight (kg) | middle value | | | | |
| 1 | 2.0 | | 2 | 1.0 | | | | | |
| 2 | 1.0 | | 5 | 1.5 | | | | | |
| 3 | 2.5 | Step 5 | 6 | 1.5 | Step 6 | | | | |
| 4 | 2.0 | \longrightarrow | 1 | 2.0 | | | | | |
| 5 | 1.5 | | 4 | 2.0 | | | | | |
| 6 | 1.5 | | 8 | 2.0 | | | | | |
| 8 | 2.0 | | 3 | 2.5 | | | | | |
| Sum | 12.5 | | Sum | 12.5 | | | | | |

| Class | s Exe | ercise | | |
|------------|-------------------------|----------------|--|--|
| What is th | at is the median value? | | | |
| Chi | hicken W | eights | | |
| An | | Weight (kg) | | |
| | 2 | 1.0 | | |
| | 5 | 1.5 | | |
| | 6 | 1.5 | | |
| | 1 (| 2.0 | | |
| | 4 | 2.0 | | |
| | 8 | 2.0 | | |
| s | Sum | 12.5 | | |
| | | | | |

Mode

Step 7: Calculate the mode

| Chicken | en Weights al Weight (kg) 1.0 1.5 1.5 2.0 2.0 2.0 2.5 |
|--------------|---|
| Animal ID | |
| 2 | 1.0 |
| 5 | 1.5 |
| 6 | 1.5 |
| 1 | 2.0 |
| 4 | 2.0 |
| 8 | 2.0 |
| 3 | 2.5 |
| Sum | 12.5 |

What weight is most commonly shared among my chickens?

Which data value occurs most frequently?

15

| H | varcica | 7. | T | Measures of | Central | Т | andancy |
|----|---------|----|----|-------------|---------|---|---------|
| IN | xercise | | 11 | reasures or | Central | | endency |

- 1. This exercise should take 60 minutes.
- 2. Work in pairs or groups of three.
- 3. Use the dataset titled, "Lesson 5_Ex7Ex8Ex9.xlsx".
- 4. Calculate the measures of central tendency: mean, median and mode for the following variables:
 - HHAge
 - $\hbox{-} \ Distance \ to \ main \ road$
 - Herd size
- $5.\ {\rm For\ each\ of\ these\ variables},$ create histograms of each of these variables.

Exercise 7 – Calculate and Apply Measures of Central Tendency: Mean, Median, Mode, and Range

Description of Exercise:

Calculate measures of central tendency and make frequency histograms.

Allotted Time: 60 minutes

Organisation of Group Work:

- This exercise will take 60 minutes
- Work in pairs or small groups of three.

Exercise Components and Structure:

- 1. Calculate the measures of central tendency: mean, median and mode for the following variables:
 - HHAge
 - Distance to main road
 - Herd size
- 2. For each of these variables, create histograms of each of these variables

Materials, Data or Information:

Use the dataset titled, "Lesson 5_Ex7Ex8Ex9.xlsx".

Expected Outputs and Deliverables of Each Participant:

1. Mean, median and mode value for the following variables:

| Measure of Central Tendency | HHAge | Distance to main road | Herd size |
|--------------------------------|-------|-----------------------|-----------|
| Mean | | | |
| Median | | | |
| Mode | | | |
| Min | | | |
| Max | | | |

 $2. \;\;$ Frequency histograms of the data from the same three variables.

Lesson 5.2 – Describing and Acting upon Animal Disease and Health Data: Disease Occurrence and Impact

| Estimated Lesson and Exercise Time | 2 hours |
|------------------------------------|---|
| Participant Materials | Frontline ISAVET Describing and Acting upon Animal Disease and Health Data: Disease Occurrence and Impact Participant Guide.doc |
| | Computer and Microsoft Word |
| | Pen or Pencil |
| Handout Materials for Exercises | MS Excel Lesson 5_Ex8.xlsx .xls (Lesson 8) |
| | MS Excel MS Excel Topic 6: Prevalence |
| | and Incidence.xls |

In Service Applied Veterinary Epidemiology Training (ISAVET) Lesson 5.2: Describing and Acting upon Animal Disease and Health Data: Disease Occurrence and Impact

At the end of this lesson, you will be able to: 1. Explain and calculate measures of disease occurrence

Learning Objectives

and impact.

Measuring Disease Occurrence and Impact

STEP 1: COUNT

affected units of interest such as animals, farms, villages, cases, etc.)

• Include: Animal, Place, Time



STEP 2: COMPARE

counts to measure disease occurrence and impact:

- Qualitative data: Proportion
- · Quantitative data: Proportion, rate, ratio

Reference: Gregg M. (ed). Field Epidemiology, Third Edition. Oxford University Press. New York. 2008.

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STEP 1: COUNT

A field epidemiologist counts 1020 sick ducks and 310 sick geese in one village.

STEP 2: COMPARE

To compare the number (count) of sick ducks to the number (count) of sick geese in the village we divide one count by another to give the ratio of ducks to geese: Ratio (ducks/geese) = 1020/310 = 3.3

Interpretation: There are 3.3 times more sick ducks than sick geese in the village

Measuring Disease Impact Using a Proportion

 $\label{lem:Approportion} A \ proportion \ is \ a \ fraction \ that \ includes \ the \ numerator \ in \ the \ denominator.$ Proportion = a/(a+b)

STEP 1: COUNT

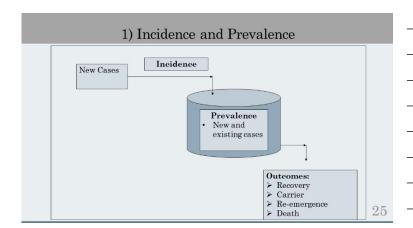
A field epidemiologist counts 1020 sick ducks and 310 sick geese in one village.

STEP 2: COMPARE

To calculate the proportion of sick water fowl in the village that are geese:

Proportion of water fowl that are sick geese = 310 / (310 + 1020) = 0.23 (23%)Proportion of water fowl that are sick ducks = 1020 / (310 + 1020) = 0.77 (77%)

| Measures of Disease Occurrence | | |
|---|-----|--|
| Measures of disease occurrence: | | |
| 1) Incidence and prevalence | | |
| 2) Defining the susceptible population at risk (PAR) | | |
| Incidence (new cases) Incidence risk (CI) – Approximate method Incidence rate (IR) – Exact method | | |
| 4) Prevalence (new and old cases) | | |
| 5) Other measures of disease risk: Attack risk | | |
| 6) Crude and Specific risk ("rate") | | |
| | 22 | |
| Caution about Epidemiology Terms: Risk versus Rate | | |
| | | |
| Some epidemiological terms used in public health apply the term "rate" when in fact it is measuring a "risk" expressed as a proportion: | | |
| • Example 1: Attack Rate is really an Attack Risk • Example 2: Case Fatality Rate is really a Case Fatality Risk | | |
| To account for this difference, the public health term "rate" will be included in parenthesis in this lesson so that you are aware of this difference in terminology Remember that a rate must refer to the number of events per unit time | 23 | |
| | | |
| Key Concepts: Risk versus Rate | | |
| | | |
| Risk Rate | | |
| | | |
| Is a probability expressed as a | | |
| proportion: Example 2 of 10 Measures risk per unit time animals are dead = 20% | | |
| Equation: Risk = a / (a+b) Equation: Count/Animal-time at risk | | |
| Units: Percentage affected (%) Units: Animal time at risk e.g. 40 cattle-weeks at risk | | |
| | 9.4 | |
| | 2.4 | |



2) Defining the Population at Risk (PAR)



- Risk is expressed as a proportion: R = P = a/(a+b)
- Coccidiosis is a parasitic disease that affects calves under the age of 90 days of age.
- A small herd of beef cattle contains 35 young calves under the age of 90 days and 40 mature cows.
- · What is the PAR for coccidiosis in this beef herd?
- Answer: The population at risk for coccidiosis is 35 and this will become the denominator

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3) Cumulative Incidence (CI) – Closed Population

 $\mathbf{CI} = \frac{\textit{Number count of new cases during a specified period}}{\textit{Number count of Invididuals initialy at risk during the period}}$

Process: Count and then compare the numerator and denominator

- · Numerator:
 - Counts the new cases that occur during a defined period
- · Denominator:
 - Closed populations (stable population):
 - Option 1: Count of healthy animals initially at risk at the start of the period
 - Open populations (dynamic population):
 - Option 2: Average count of population at beginning and end of the period

Example - Cumulative Incidence (CI)

Closed Population (stable):

There are 40 new cases of Rabies diagnosed in cattle in local area x over a one year period. The cattle population was estimated as 1000 in January at the start of the year. What is the incidence risk or the CI?

 $CI = rac{ ext{Number count of new Cases during a specified period}}{ ext{Initial count of PAR at begining of the period}}$

 $CI: \frac{40}{1000} = 0.04$ cases per animal-year at risk

- Multiply the incidence rate (IR) by either 100, 1000 or 10,000 or some other number (human health incidence rates are often compared per 100,000 population).
- The CI: [0.04 X 1,000] = 40 cases of rabies per 1,000 head of cattle in one year in local area X

 $\textbf{Reference:} \ \ \textbf{Thrusfield, 2008.} \ \ \textbf{Veterinary Epidemiology, Third Edition.}$

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Cumulative Incidence (CI) - Open Population

Open Population (dynamic):

To be used when the population at risk (PAR) is changing over time

 $CI = \frac{\textit{Count of new cases that occur in the population during a specified period}}{(\textit{Count at risk at the start of the period} + \textit{number at risk at the end of the study period})/2}$

- · Numerator: same as incidence risk
 - Counts the new cases that occur during a defined period
 - Excludes diseased individuals at the start of the period
- · Denominator:
 - Average population at risk during the period

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Example: Cumulative Incidence - Open Population

Open population:

There are 40 new cases of Rabies diagnosed in cattle in local area x over a one year period. The cattle population was estimated as 1000 in January at the start of the year. A number of Cattle were marketed in May, leaving 660 animals in December at the end of the year. What is the incidence rate?

 $CI = \frac{\textit{Count of new cases that occur during a specified period}}{(\textit{Count at risk at the start of the period} + \textit{number at risk at the end of period})/2}$

IR = $\frac{40}{(1000 + 660)/2} = \frac{40}{830} = 0.048$ cases per animal-year at risk

- Multiply the incidence rate (IR) by either 100, 1000 or 10,000 or some other number (human health incidence rates are often compared per 100,000 population).
- The CI : [0.048 X 1,000] = 48 cases of rabies per 1,000 head of cattle in one year in local area X

Incidence Rate (IR) - Exact Method

Measures how quickly new cases of disease occurs over time

 $IR = \frac{\textit{Number of new Cases during a specified period}}{\textit{Animal-time at risk duing the period}}$

- Numerator: same as incidence risk
 - Counts the new cases that occur during a defined period
 - Excludes diseased animals at the start of the period $\,$
- · Denominator:
 - Number of animals x the time that each animal is at risk ('animal-time at \mbox{risk} ')
 - e.g. 30 animals at risk for 2 years = 30 x 2 = 60 animal years at risk

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Summary: Prevalence, Incidence Risk, Incidence Rate

| Animalid | Jan | Feb | Mar | Apr | May | Jun | luc | Aug | des | Oct | Nov | Dec | diseased | months at risk |
|----------|-----|-----|---------|---------|-----|-----|---------|-----|-----|---------|-----|---------|----------|----------------|
| 1 | | | | Disease | | | | | | | | | Yes | 3 |
| 2 | | | | | | | | | | Disease | | | Yes | 9 |
| 3 | | | | | | | Removed | | | | | | No | 6 |
| 4 | | | | | | | | | | | | | No | 12 |
| 5 | | | Disease | | | | | | | | | | Yes | 2 |
| 6 | | | | | | | | | | | | Disease | Yes | 11 |
| 7 | | | | | | | | | | Disease | | | Yes | 9 |
| 8 | | | | | | | | | | | | | No | 12 |
| 9 | | | | | | | | | | | | | No | 12 |
| 10 | | | | | | | Removed | | | | | | No | 6 |
| Total | | | | | | | | | | | | | 5 | 82 |

No. of disease events: 5

Prevalence in June: 20%(2 cases in 10 animals)

Number present at start: 10 Number of removals: 2

Number present at end: 8

39

Example: Comparing Prevalence, Incidence Risk, Incidence Rate

| Animalid | Jan | æ | Mar | Apr | May | Jun | luc. | Aug | des | 55 Ost | Nov | Dec | diseased | months at risk |
|----------|-----|---|---------|---------|-----|-----|---------|-----|-----|-----------|-----|---------|----------|----------------|
| 1 | | | | Disease | | | | | | | | | Yes | 3 |
| 2 | | | | | | | | | | Disease | | | Yes | 9 |
| 3 | | | | | | | Removed | | | | | | No | 6 |
| 4 | | | | | | | | | | | | | No | 12 |
| 5 | | | Disease | | | | | | | | | | Yes | 2 |
| 6 | | | | | | | | | | | | Disease | Yes | 11 |
| 7 | | | | | | | | | | Disease | | | Yes | 9 |
| 8 | | | | | | | | | | | | | No | 12 |
| 9 | | | | | | | | | | | | | No | 12 |
| 10 | | | | | | | Removed | | | | | | No | 6 |
| Total | | | | | | | | | | | | | 5 | 82 |

No. of disease events: 5

Point Prevalence in June: 20% (2 cases in 10 animals)

Point Prevalence in December: 62.5% (5 cases in 8 animals)

Incidence risk (Approximate): 50% (5 cases in 10 animals)

Incidence rate (Exact) 5 cases per 84 animal-months at risk

Number present at start: 10

Number of removals: 2

Number present at end: 8

4) Prevalence Risk or Proportion (P)

 ${}^{\bullet}$ Refers to the number of <code>existing cases</code> including those previously existing and new cases that have developed at some point during a given time period.

P = (Count of existing cases) (Total count of population at risk)

- Process: Count and Compare the numerator and denominator
- · Point prevalence:
 - Counting the existing cases at one brief point in time divided by the population at risk (PAR) at that time
- · Period prevalence:
- Counting cases over a longer period of time

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Example: Prevalence risk (P)

If 100 cows in a herd of 800 cows are positive for mastitis during one month, what is the <u>period prevalence</u> of mastitis for that month?

 $P = \frac{\textit{Count of existing cases during time period}}{\textit{Total count of population at risk during time period}}$

 $P = \frac{100}{800} = 0.125$ or 0.125 x 100 = 12.5%

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5) Other Measures of Disease Risk

- A rate is a risk (probability) that is calculated over a given time period and includes the following:
 - A numerator: \mathbf{count} of animals diseased or dead in a specified time period
 - Denominator: total ${f count}$ of all animals at risk or animal-time
 - Specified time period
 - Example
 - In a chicken flock, the rate of new cases of infectious coryza in the flock over a 12 month period:

Incident rate is 25 cases/100 birds/12 months

Attack (Rate) Risk

- * Refers to incidence risk with a short duration
- Generally used during an outbreak situation
- Commonly referred to as "attack rate" in Public Health it is really is a measure of <u>risk</u> since there is no time component

 $AR = \frac{\textit{Number of new Cases of disease during a specified period}}{\textit{Number of animals initialy at risk during the period}}$

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6) Crude and Specific (Rates) Risks

- $\mathbf{Crude}\ \mathbf{risks};$ are risks that are expressed for the entire population at risk
 - · Morbidity or illness
 - · Mortality or deaths
 - * Example : Crude morbidity and mortality risks
- Stratum specific risks: are those risks expressed for specific subpopulations based on factors such as age, breed, sex, production type, etc
- · Example: Age-specific mortality risk

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Crude (Rates) Risk – Morbidity Risk

 Morbidity risk: refers to the number of cases that are clinically affected of the population at risk over an identified time period

 $Crude \ Morbidity = \frac{Number \ clinically \ ill}{Population \ at \ risk \ (PAR)}$

- · Example:
- · In a flock of Pekin ducks, 120 clinically ill and 50 deaths were observed out of 1100 ducks.
- The crude morbidity risk was:

Crude Morbidity Risk = $\frac{120}{1100}$ = 1.0%

Crude (Rates) Risk - Mortality Risk

· Mortality risk - the number of deaths in a population over a specific time

Crude Mortality Risk= Number of animals that die during the period

- $\cdot Example$
 - · In a flock of Pekin ducks, 120 clinically ill and 50 deaths were observed out of 1100 ducks.
- The crude mortality rate was:

Crude Mortality Risk = 50/1100 = 4.5%

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Crude (Rates) Risk – Case Fatality Risk (CFR)

- Case fatality risk (CFR) – the number of deaths amongst all infected cases during a specified time period

> Number of deaths $CFR = \frac{Number \ of \ access}{Number \ clinically \ ill \ (cases)}$

- · Example:
- · In a flock of Pekin ducks, 120 clinically ill and 50 deaths were observed out of 1100 ducks. The case fatality rate was:

CFR = 50/120 = 41.6%

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Stratum-Specific (Rates) Risks

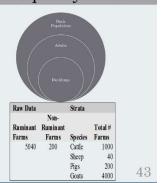
- \bullet These refers to description of clinical disease and death rates according to specific parts of the population such as :
 - Age, sex, breed, production type, etc.
- The crude mortality risk in a flock of Peking ducks was 50/1100 = 4.5%
- We know that more ducklings died than adult ducks. Prior to the occurrence of deaths in the flock, 20% of the flock were ducklings. We also know that 30 out of 50 duck deaths were ducklings. We can calculate age-specific risks as follows:

Duckling Age - specific risk = $\frac{30}{1100 \times 0.2}$ = 13.6% of ducklings died

Adult duck Age-specific risk = $\frac{20}{1100 \times 0.8}$ = 2.3% of adult ducks died 42

Stratification of Disease Frequency Data

- Many risk associations are hidden when the population data is considered as a whole
- It is necessary to separate out or stratify the data into layers or levels as we did for different age groups
- Assumption: The PAR is well defined and we can identify and count members of each strata
- In this example ruminant farms are stratified by cattle, sheep and goat farms



Exercise 8: Measures of Disease Occurrence and Impact

- 1. This exercise should take 60 minutes.
- 2. Work in pairs or in groups of three.
- 8. Part 1: Calculate proportion and ratio to assess disease risk.
 - a) Use dataset titled, "Lesson 5_Ex7Ex8Ex9.xlsx".
 - b) Calculate the proportion of HH with outbreaks
 - c) Calculate the ratios of dog bites among regions of your choice.
- 4. Part 2: Calculate incidence and prevalence to assess disease impact.
- Copy of Day 2 MS Excel Topic 6_Prevalence and Incidence_instructor.xls
- Refer to MS Excel Topic 6_Prevalence and Incidence_instructor.xls and MS Excel Video Topic 6: Prevalence and Incidence.xls

Exercise 8 – Calculate Measures of Disease Occurrence (Incidence and Prevalence) and Disease Impact (Important Rates and Ratios)

Description of Exercise:

• Calculate measures of disease occurrence and disease impact

Allotted Time: 60 minutes

Organisation of Group Work:

- This exercise will take 60 minutes
- Work in pairs or in groups of three.

Exercise Components and Structure:

Part 1: Proportion and ratio

- 1) Calculate the proportion of HH with outbreaks
- 2) Calculate the ratios of dog bites among regions of your choice.

Part 2: Incidence vs prevalence

Follow the instructions for MS Excel Topic 6: Calculating incidence and prevalence.

Materials, Data or Information:

Part 1. Use dataset titled, "Lesson 5_Ex7Ex8Ex9.xlsx".

Part 2. Copy of Day 2 MS Excel Topic 6_Prevalence and Incidence_instructor.xls

Expected Outputs and Deliverables of Each Participant:

Part 1. Use dataset titled, "Lesson 5_Ex7Ex8Ex9.xlsxIsigiro.xlsx".

| 1) | Calculate the | proportion | of HH | with | outbreaks |
|----|----------------|------------|---------|---------|---------------|
| , | Carcarate tric | proportion | OI IIII | ** 1011 | O a corr cars |

2) Calculate the ratios of dog bites among regions of your choice.

Part 2. Copy of Day 2 MS Excel Topic 6_Prevalence and Incidence_instructor.xls

Lesson 5.3 – Describing and Acting upon Animal Disease and Health Data: Descriptive Analysis by Animal-Place-Time

| Estimated Lesson and Exercise Time | 1 hour and 30 minutes |
|------------------------------------|---|
| Participant Materials | Frontline ISAVET Describing and Acting upon Animal Disease and Health Data: Descriptive Analysis by Animal-Place- Time. Participant Guide.doc |
| | Computer and Microsoft Word |
| | Pen or Pencil |
| Handout Materials for Exercises | MS Excel Lesson 5_Ex7Ex8Ex9.xlsx .xls (Lessons 7, 8 and 9) |

In Service Applied Veterinary Epidemiology Training (ISAVET)

Lesson 5.3: Describing and Acting upon Animal Disease and Health Data: Descriptive Analysis by Animal-Place-Time

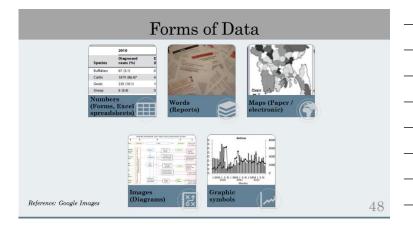
| A DIFFETTION ANIMAL DISPASES | Food and Agriculture Organization of the United Nations |
|------------------------------|---|

Learning Objectives

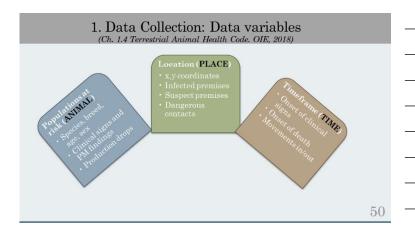
At the end of this lesson, you will be able to:

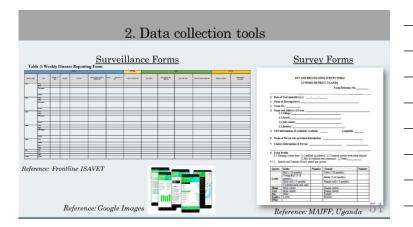
- Describe how to prepare and follow a plan to capture and describe data by animal-place-time.
- 2. Prepare a plan to describe data by animal-place-time.
- 3. Perform descriptive data analysis by animal-place-time.

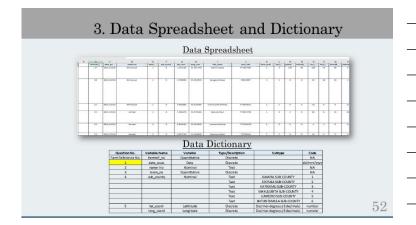
Usefulness of Data to Explain Disease Events | What happened? (DISEASE) | When did it occur? (TIME) | Where did happen? (PLACE) | Who is affected (pigs, poultry, etc.)? (ANIMAL) | | Analytical Epidemiology | Why did it happen (research hypothesis)? (RISK FACTORS) | How will the data be processed and used? (INTERVENTIONS)



Part 1: Elements of a Plan to Capture and Describe Data 1. What data variables to collect: 1. Enter data into the spreadsheet - Animal - Place - Time 2. Assess data quality (Lesson 4) 3. Measure central tendency: Quantitative data: mean, median, 2. Data collection tool mode - Design and test ques-Design and test questionnaire Qualitative data: count, proportion, 3. Design spreadsheet/database to record data - Create a data dictionary - Create a line list with variable 4. Measure disease occurrence/impact: - Mortality headings 5. Display data - tables, graphs, maps 4. Collect data and ensure all data is complete and of high quality 6. Interpret and report results 49







4. Data Collection Plan

Case Example

 $A\ retrospective\ study\ was\ conducted\ to\ understand\ the\ geographic\ and\ seasonal\ distribution\ of\ anthrax\ in\ Bangladesh\ using\ passive\ surveillance\ data\ from\ 1\ January\ 2010\ to\ 31\ December\ 2012.$

Scope of the Data Collection Plan What disease = anthrax (laboratory data)

Who (population) = livestock

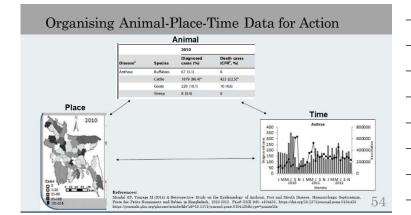
Where = Bangladesh (map data)

When = 3 years (1 January 2010 to 31 December 2012)

 $\mathbf{Why/how} = \text{to understand the geographic and seasonal distribution of endemic diseases of }$ livestock/retrospective study using data collected through passive surveillance

rences:
al SP, Yanage M (2014) A Retrospective Study on the Epidemiology of Anthrax, Foot and Mouth Disease, Haem
dadesh, 2010-2012. PLoS ONE 9(6): e104435. https://doi.org/10.1871/journal.pone.0104435
d/doi.org/10.1871/purnal.pone.0104436
//journal.pone.org/1046716/10471/purnal.pone.01044358type=printable

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Descriptive Data Analysis and Display of Data Variables

Count data can include quantitative or qualitative data.

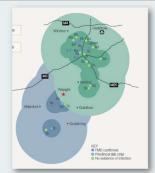
Table 1. Estimated number of diagnosed cases, death cases and vaccination coverage of arthrax, foot and mouth disease, haemorrhagic septicaemia, peste des petits ruminants and dog bite/rables in livestock (cattle, goats, sheep and buffaloes) in Bangladesh, 2010-2012.

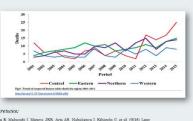
| Disease | 2010 | 2011 | 2012 | Total |
|--------------------------------|-------------------------------|--------------------|-------------------|---------------------|
| Anthrax | | | | |
| Diagnosed cases | 2,174 | 1,668 | 2,095 | 5,937 |
| Prevalence rate, % (95% CP) | 0.14 (0.13-0.15) ^a | 0.09 (0.08-0.09) | 0.17 (0.16-0.18) | 0.13 (0.12-0.13) |
| Death cases | 433 | 173 | 195 | 801 |
| Case fatality rate, % (95% CI) | 19.92 (18.23-21.61) | 10.37 (8.90-11.84) | 9.31 (8.06-10.56) | 13,49 (12,62-14,37) |
| Vaccination | 2,602,967 | 3,417,136 | 3,325,525 | 9,345,628 |
| Vaccination rate, % (95% CI) | 6.11 (6.10-6.12) | 8.02 (8.01-8.03) | 7.81 (7.80-7.82) | 7.31 (7.31-7.32) |

The table shows continuous variables include quantitative number counts and values.

lage M (2014) A Retrospective Study on the Epidemiology of Anthrax, Foot and Mouth Disease, Haemorrhagic Septicaemia, Ruminants and Rabies in Bangladesh, 2010-2012. PLoS ONE 9(8): e104435. https://doi.org/10.1371/journal.pone.0104435.

Examples: Displaying Data by Place and Time





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Exercise 9: Prepare a Plan to Analyse Data

- 1. This exercise should take 45 minutes.
- 2. Work in pairs or in small groups of three.
- 3. Prepare a plan to analyse a dataset called "Lesson 5_Ex7Ex8Ex9.xlsx"

In Summary...You now know how to:

- 1. Calculate measures of central tendency in data including:
 - · Mean (average), frequency, median and mode; and
- 2. Calculate measures of disease occurrence and impact from data including:
 - · Ratios and proportions
 - Incidence
 - $\cdot \, {\rm Prevalence}$
 - Attack risk
 - $\boldsymbol{\cdot}$ Crude and specific morbidity and mortality risks.
- 3. Prepare and follow a plan to capture and describe data by animal-place-time.

| ISAVE | T Contributing Universities | | |
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| | | 59 | |

Exercise 9 – Prepare a Plan to Analyze Data by Animal-Place-Time

Description of Exercise:

Prepare and follow a plan to capture and describe data by animal-place-time using the MS Excel file L5 Ex7Ex8Ex9.xls.

Allotted Time: 45 minutes

Organisation of Group Work:

- Total time 45 minutes
- Work in groups of two or three people.
- For the first 20 minutes groups will answer the questions below.
- During the last 25 minutes, the instructor will lead a plenary discussion.

Exercise Components and Structure:

1. Under the tab labelled "data", list the variable names in the following table that describe characteristics of animal, place and time.

| ANIMAL | PLACE | TIME |
|--------|-------|------|
| | | |

- 2. Define a data dictionary and explain how it is used.
- 3. Review the data in the spreadsheet. Do you see any errors or omissions?
- 4. How many total animals are there in the 20 herds? What is the size of the smallest and largest herds?

Materials, Data or Information:

MS Excel file L5 Ex7Ex8Ex9.xls. contains all data under the tab labelled "data".

Expected Outputs and Deliverables of Each Participant:

1. Under the tab labelled "data" list the variable names in the following table that describe characteristics of animal, place and time. Note that record ID and occupation pertain to the line list number and the occupation of the animal owner, respectively.

| ANIMAL | PLACE | TIME |
|--------|-------|------|
| | | |
| | | |
| | | |
| | | |
| | | |

- 2. Define a data dictionary and explain how it is used.
- 3. Review the data in the spreadsheet. Do you see any errors or omissions?
- 4. How many total animals are there in the 20 herds? What is the size of the smallest and largest herds?

Lesson 6 - Display Data for Decision-Making

| 1 0 | |
|------------------------------------|--|
| Estimated Lesson and Exercise Time | 3 hours |
| Participant Materials | Frontline ISAVET Lesson 6 Display Data |
| | for Decision-Making Participant |
| | Guide.PDF |
| | Computer Microsoft Word and Excel |
| | Pen or pencil |
| Handout Materials for Exercises | MS Excel PivotTable.doc |
| | |

In Service Veterinary Epidemiology Training (ISAVET)

Lesson 6: Display Data for Decision-Making





Learning Objectives

At the end of this lesson, you will be able to:

- 1. Create and interpret one and two variable tables;
- 2. Create and interpret a line graph;
- 3. Create and interpret a histogram;
- 4. Create and interpret one and two variable bar charts;
- 5. Create and interpret pie charts; and
- 6. Describe when to use each type of table, graph, and chart.

Tables

- \bullet Frequencies (counts) of qualitative data are often best presented in a table \bullet Single variable
- An advantage of tables over graphs is that actual values can be presented (i.e., specific amounts rather than approximations)
- Sometimes tables communicate information better than graphs
- · Do not provide a method to showcase trends in data

3

Tables: Data Representation

Cases of FMD in Country X by Species, October, 2018 Species Number of Cases Caprine Ovine 30 Bovine (Beef) 25 Bovine (Dairy) 12 30 Porcine Total 107 Note: Notional data What type of variable is being presented here?

и

Table Components

Cases of Disease FMD in Country X by Species, October, 2018 Number of Cases Species Caprine Ovine 10 30 Bovine (Beef) 25 Bovine (Dairy) 12 Porcine 30 Total 107 Note: Notional data Component 1: There is a labelled descriptive title.

Cases of Disease FMD in Country X by Species, October, 2018 | Species | Number of Cases | | Caprine | 10 | Ovine | 30 | Bovine (Beef) | 25 | Bovine (Dairy) | 12 | Porcine | 30 | Total | 107 | Note: Notional data | Component 2: Each row and column are clearly labelled and data is arranged in rows and columns

| | ID in Country X by Spe tober, 2018 |
|----------------|---------------------------------------|
| Species | Number of Cases |
| Caprine | 10 |
| Ovine | 30 |
| Bovine (Beef) | 25 |
| Bovine (Dairy) | 12 |
| Porcine | 30 |
| Total | 107 |

Cases of Disease FMD in Country X by Species, October, 2018 Species Number of Cases Caprine 10 Ovine 30 Bovine (Beef) 25 Bovine (Dairy) 12 Porcine 30 Total 107 Note: Notional data What is another item that is not included on this table which may help to describe the data?

Two Variable Table

Cases of Disease FMD in Country X by Species and District, October, 2018

| Species | Number of Cases | | | |
|---------------------|-----------------|------------|--|--|
| | District A | District B | | |
| Caprine | 20 | 10 | | |
| Ovine | 3 | 30 | | |
| Bovine (Beef) | 4 | 25 | | |
| Bovine (Dairy) | 20 | 12 | | |
| Porcine | 25 | 30 | | |
| Total | 72 | 107 | | |
| Note: Notional data | | | | |

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Summary of Table Components

- 1. Components 1: There is a labelled descriptive title.
- 2. Component 2: Each row and column are clearly labelled and data is arranged in rows and columns.
- 3. Component 3: Totals for rows and columns are clearly defined
- Component 4: List any codes used in the table, (e.g., yrs, or mg/dl), need to be explained in a footnote: yrs = Years; mg/dl = milligrams per deciliter.

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Exercise 10: Display Data in a Table

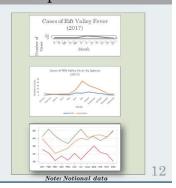
- Refer to MS Excel YouTube Video #7 Creating a Pivot Table
- 2. This exercise should take 30 minutes.
- 3. Open the Exercise 10 MS Word participant document located in your Exercise 10 participant folder on your flash drive.
- Work in pairs.
- 5. Construct tables in MS Excel using the dataset provided in Exercise 10.

Examples of Line Graphs

- Represent a trend over time

 - Years Months
- Days
- · Showcases a change in direction

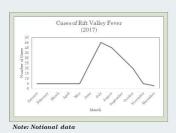
- Simple:
 One trend based on cumulative values per unit time
- Compare trend lines of different strata from the same variable over time
 Comparative:
 Shows stratification of multiple variables



Simple Line Graph

- · Shows the relationship between two variables
 - X-axis: time
- Y-axis: variable you are looking at over time to monitor changes

Examples: number of cases, prevalence and, incidence etc.

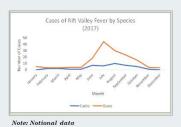


Stacked Line Graph

- · Includes multiple variables of data
 - X-axis: time
 - Y-axis: variable you are looking at over time to monitor changes

Examples: number of cases and prevalence

- Shows stratification of $\underline{\text{one}}$ categorical variable
 - Examples: species, country and gender

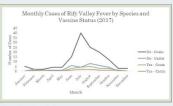


Comparative Line Graph

- · Includes multiple variables of data
 - X-axis: time
- Y-axis: variable you are looking at over time to monitor changes

 $Examples: number of \ cases \ and \ prevalence$

- · Shows stratification of multiple categorical variables
- Examples: species and vaccination status



Note: Notional data

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Line Graphs: Advantages/Disadvantages

- · Advantages
- Easy to read and create
- Can visually see the trend of the data (how the data increases or decreases)
- Useful for making comparisons of two sets of data
- Useful for showing periods of change over time
- · Disadvantages
 - $\dot{}$ If axis are not properly labeled, it may lead to an inaccurate representation of the data
- · Usually can only apply to continuous data points

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Exercise11: Display Data Using a Line Graph

- 1. Refer to MS Excel YouTube Video #8 Pivot Table Line Graphs
- 2. This exercise should take 45 minutes.
- 3. Open the Exercise 11 MS Word participant document located in your Exercise 11 participant folder on your flash drive.
- 4. Work in pairs of two.
- 5. Review handouts for Pivot Tables.
- Construct line graphs in MS Excel using the dataset provided in Exercise 11.

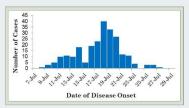
Histograms

- · Graph of the frequency distribution of a quantitative variable
- · Useful for summarising statistical properties of the data set, including the shape of the frequency distribution, modality, and
- · Columns are adjoining
- · Height of each column is proportional to number of observations in that interval

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Frequency Histogram

- Epidemiologic curves are histograms that show the frequency of cases over
- · They are fundamental tools of an outbreak investigation

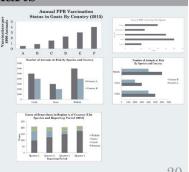


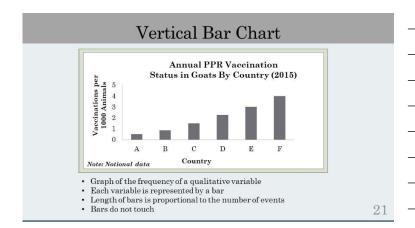
Note: Notional data

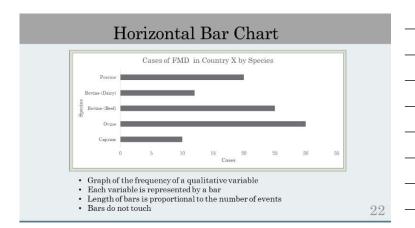
19

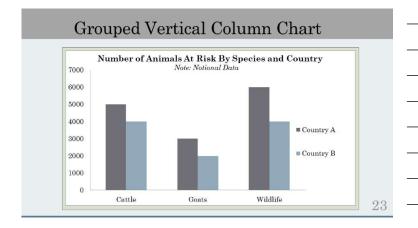
Bar Charts

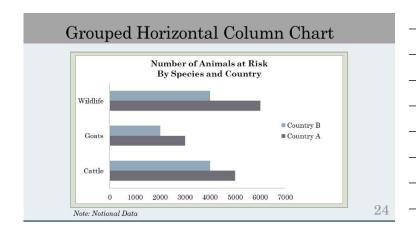
- $\hbox{-}\ Use\ with\ qualitative/categoric}$
- · Can vertically or horizontally displayed:
 - · Simple
 - · Grouped
 - · Stacked

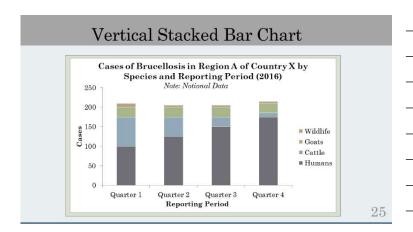


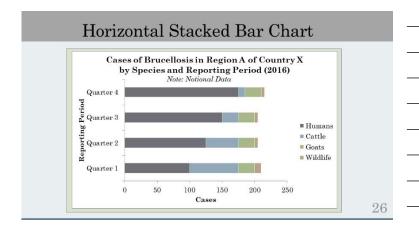












Bar Charts: Advantages/Disadvantages

· Advantages

- Shows each category in a frequency distribution
- Displays relative numbers/proportions of each category
- Easy to understand

Disadvantages

- · Fail to expose patterns
- ${}^{\bullet}$ Can be easily manipulated to give false impressions
- · Less effective than line graphs in showing trends over time
- *They do not provide a good showcase for "acceleration" of the data, a quick rise in the number of cases

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Presents information that shows categories as parts of a whole Usually represented in percentages Population Size Cattle, 36% Cattle, 36% Goats Wildlife, 43%

Note: Notional Data

■ Goats, 21%

-

Annual PPR Vaccination Status in Goats By Country (2015) Annual PPR Vacci

Pie Charts: Advantages/Disadvantages · Advantages - Shows percent of total for each category · Disadvantages - Hard to compare to datasets - No exact numerical data - Use only with discrete data - Best for 4 to 7 categories 30 Exercise12: Display Data in a Bar Chart and Pie Chart 1. Refer to MS Excel YouTube Videos: #9 Pivot Table Bar Charts and #10 Pie Charts 2. This exercise should take 30 minutes. Open the Exercise 12 MS Word participant document located in your Exercise 12 participant folder on your flash drive. 4. Work in teams of two persons. Review handouts for Pivot Tables. Construct bar charts and pie charts in MS Excel using the dataset provided in Exercise 12. In Summary... • Tables are used for both qualitative and quantitative variables. · Frequencies (counts) of qualitative data, especially those with a single variable, are best presented in a table. $\dot{}$. There are several components which should be incorporated into a table to best describe the data for the user. \cdot Quantitative data is best represented by tables, line graphs, or histograms $\boldsymbol{\cdot}$ Qualitative data is best represented by tables, bar graphs, or pie charts. \cdot Line graphs provide a temporal function and display trends in the data over · Bar charts are used to present and compare qualitative data · Pie charts represent categorical variables or values of variables

| ISAVE | T Contributing Universities | | |
|--------------|-----------------------------------|----|--|
| Partners | TEXAS ASM. ATEXAS ASM. AGRILIFE | | |
| Contributors | MAKERHE UNIVERSITY | 33 | |

Exercise 10 - Display Data in a Table

Description of Exercise:

Use the tables provided to construct an appropriate two-variable and multi-variable table of brucellosis and theileriosis (east coast fever) cases by species, local area, region, and test results. Should you have any questions over the exercise, please ask a trainer for clarification.

Allotted Time: 30 minutes

Exercise Components and Structure:

- 1. You have 30 minutes to complete this exercise
- 2. Construct the following tables based on the data provided
 - a. Two-variable table
 - **b.** Multi-variable table
- Work in pairs for this exercise;
- Each pair should review the provided tables for brucellosis and theileriosis (east coast fever) field testing;
- Create the appropriate two-variable and multi-variable tables from the data table provided.
- Copy and paste the data tables from this exercise into an MS Excel worksheet.

Materials, Data or Information:

- 1. MS Word
- 2. MS Excel
- 3. Paper and pen

Expected Outputs and Deliverables of Each Participant:

1. Depict animal-place-time by creating tables using MS Excel.

Brucellosis Data

| Village | Local | Species | Total | Total Blood | Total Milk | Card | Brucellosis | Brucella |
|---------|----------|--------------|---------|-------------|------------|------|--------------|---------------|
| ID | area | | Animals | Samples | Samples | Test | confirmation | Strain Typed |
| 1 | Luwero | Goat | 5 | 4 | 2 | + | + | B. Melitensis |
| 2 | Luwero | Sheep | 7 | 7 | 4 | + | + | B. Melitensis |
| 3 | Luwero | Dairy Cattle | 10 | 10 | 5 | + | + | B. abortus |
| 4 | Luwero | Goat | 15 | 15 | 10 | + | + | B. Melitensis |
| 5 | Luwero | Goat | 12 | 12 | 8 | - | - | |
| 6 | Luwero | Sheep | 10 | 10 | 6 | + | + | B. Melitensis |
| 7 | Luwero | Sheep | 5 | 4 | 3 | + | + | B. Melitensis |
| 8 | Luwero | Beef Cattle | 3 | 3 | | + | + | B. Abortus |
| 9 | Luwero | Dairy Cattle | 9 | 6 | 4 | + | + | B. Melitensis |
| 10 | Luwero | Dairy Cattle | 6 | 5 | 4 | + | + | B. Melitensis |
| 11 | Luwero | Goat | 9 | 9 | 6 | + | + | B. Melitensis |
| 12 | Luwero | Sheep | 3 | 3 | 1 | - | + | B. Melitensis |
| 13 | Luwero | Sheep | 7 | 7 | 5 | + | - | |
| 14 | Luwero | Goat | 12 | 12 | 4 | + | - | |
| 15 | Luwero | Goat | 15 | 15 | 9 | - | + | B. Melitensis |
| 16 | Kayunga | Goat | 17 | 17 | 15 | + | + | B. Melitensis |
| 17 | Kayunga | Dairy Cattle | 9 | 9 | 6 | + | + | B. Abortus |
| 18 | Kayunga | Dairy Cattle | 4 | 4 | 2 | + | + | B. Melitensis |
| 19 | Kayunga | Goat | 3 | 3 | | + | + | B. Melitensis |
| 20 | Kayunga | Sheep | 5 | 5 | 2 | + | + | B. Melitensis |
| 21 | Kayunga | Dairy Cattle | 7 | 7 | 4 | + | + | B. Melitensis |
| 22 | Kayunga | Goat | 15 | 15 | 10 | + | + | B. Melitensis |
| 23 | Kayunga | Sheep | 10 | 7 | 5 | - | + | B. Melitensis |
| 24 | Kayunga | Sheep | 10 | 7 | 5 | + | + | B. Melitensis |
| 25 | Nakasake | Beef Cattle | 5 | 4 | | + | + | B. Abortus |
| 26 | Nakasake | Beef Cattle | 7 | 6 | | + | + | B. Abortus |
| 27 | Nakasake | Beef Cattle | 3 | 3 | | + | + | B. Abortus |
| 28 | Nakasake | Beef Cattle | 4 | 4 | | - | + | B. Abortus |
| 29 | Nakasake | Beef Cattle | 5 | 4 | | - | - | |
| 30 | Nakasake | Beef Cattle | 2 | 2 | | - | - | |

Data Dictionary

| Variable | Description | Variable Type | Coding |
|-----------------------------|---|--------------------------|--|
| Village ID | Village number | Quantitative, Discrete | None |
| Local area | Name of local area where sample was collected | Qualitative, Nominal | 1 = Luwero; 2= Kayunga; and 3 = Nakasake |
| Species | Species | Qualitative, Nomina1 | 1 = Goat; 2 = Sheep; 3 = Dairy Cattle; 4 = Beef Cattle |
| Total Animals | The total number of animals in the herd | Quantitative, Discrete | None |
| Total Blood Samples | The total number of blood samples collected | Quantitative, Discrete | None |
| Total Milk Samples | The total number of milk samples collected | Quantitative, Discrete | None |
| Card Test | A positive or negative reading of the brucellosis card test from field collection | Qualitative, Categorical | 1 = +; 0 = - |
| Brucellosis Confirmation | Laboratory confirmation of a brucellosis positive case | Qualitative, Categorical | 1 = +; 0 = - |
| Brucella Strain Typed | Laboratory confirmation of the Brucella strain typed | Qualitative, Nominal | 1 = B. Melitensis; $2 = B.$ Abortus |

- 1. Open a new MS Excel workbook and enter the above data into the spreadsheet.
- 2. Using the sort and filter function in MS Excel, group total animals into the following categories. Insert a new variable in the spreadsheet titled operation size.

| Category | Total Animals |
|----------------------|---------------|
| 1 – No animals | 0 |
| 2 – Small operation | 1-10 |
| 3 – Medium operation | 11-20 |
| 4 – Large operation | >21 |

- 3. Construct an appropriate table showing all confirmed cases of brucellosis by *Brucella* strain type. Copy and paste your table from MS Excel into this document.
- 4. Construct an appropriate table showing all confirmed cases of brucellosis stratified by operation type and local area. Copy and paste your table from MS Excel into this document.
- 5. Construct and appropriate table showing the total number of animals sampled by blood stratified by species and local area. Copy and paste your table from MS Excel into this document.

East Coast Fever (Theileriosis) Laboratory Submissions Dataset

| | East Coast Fever (Theileriosis) Laboratory Submissions Dataset | | | | | | | | |
|--------|--|---------|--------|---------------------------|--|--|--|--|--|
| Sample | Local area | Species | Gender | ECF ¹ Positive | | | | | |
| ID | | | | Lab | | | | | |
| | | | | Confirmation | | | | | |
| 1 | Dokolo | Cattle | Male | - | | | | | |
| 2 | Dokolo | Cattle | Male | + | | | | | |
| 3 | Dokolo | Cattle | Female | + | | | | | |
| 4 | Dokolo | Cattle | Male | + | | | | | |
| 5 | Dokolo | Cattle | Female | - | | | | | |
| 6 | Dokolo | Cattle | Female | - | | | | | |
| 7 | Dokolo | Cattle | Female | - | | | | | |
| 8 | Kibuku | Cattle | Female | - | | | | | |
| 9 | Kibuku | Cattle | Male | - | | | | | |
| 10 | Kibuku | Cattle | Female | - | | | | | |
| 11 | Kibuku | Cattle | Male | - | | | | | |
| 12 | Kiruhura | Cattle | Female | - | | | | | |
| 13 | Kiruhura | Cattle | Male | + | | | | | |
| 14 | Kiruhura | Cattle | Male | - | | | | | |
| 15 | Kiruhura | Cattle | Female | + | | | | | |
| 16 | Kiruhura | Cattle | Male | + | | | | | |
| 17 | Kiruhura | Cattle | Female | + | | | | | |
| 18 | Kiruhura | Cattle | Female | + | | | | | |
| 19 | Kiruhura | Cattle | Female | - | | | | | |
| 20 | Budaka | Cattle | Male | + | | | | | |
| 21 | Budaka | Cattle | Female | + | | | | | |
| 22 | Budaka | Cattle | Female | + | | | | | |
| 23 | Luwero | Cattle | Male | + | | | | | |
| 24 | Luwero | Cattle | Female | + | | | | | |
| 25 | Luwero | Cattle | Male | - | | | | | |
| 26 | Luwero | Cattle | Female | + | | | | | |
| 27 | Budaka | Cattle | Female | + | | | | | |
| 28 | Isingiro | Cattle | Male | + | | | | | |
| 29 | Isingiro | Cattle | Male | + | | | | | |
| 30 | Isingiro | Cattle | Male | + | | | | | |
| 31 | Luwero | Cattle | Female | + | | | | | |
| 32 | Luwero | Cattle | Male | - | | | | | |
| 33 | Luwero | Cattle | Female | - | | | | | |
| 34 | Luwero | Cattle | Female | - | | | | | |
| 35 | Luwero | Cattle | Female | + | | | | | |
| 36 | Luwero | Cattle | Male | + | | | | | |
| 37 | Luwero | Cattle | Female | + | | | | | |
| 38 | Amolatar | Cattle | Female | + | | | | | |
| 39 | Amolatar | Cattle | Male | + | | | | | |
| 40 | Amolatar | Cattle | Male | + | | | | | |
| 41 | Amolatar | Cattle | Female | + | | | | | |
| 42 | Amolatar | Cattle | Male | + | | | | | |

¹ = ECF (East Coast Fever)

Data Dictionary

| Variable | Description | Variable Type | Coding |
|--------------|----------------------|---------------|-------------------------|
| Sample ID | Sample number | Quantitative, | None |
| | | Discrete | |
| Local area | Name of local area | Qualitative, | 1 = Dokolo; 2= Kibuku; |
| | where sample was | Nominal | and 3 = Kiruhura; 4 = |
| | collected | | Budaka; 5 = Luwero; 6 = |
| | | | Isingiro; 7 = Amolatar |
| Species | Species | Qualitative, | 1 = Cattle |
| | | Nominal | |
| Gender | The gender of the | Quantitative, | 1 = Male; 2 = Female |
| | species | Discrete | |
| ECF Positive | Laboratory | Qualitative, | None |
| Lab | confirmation of east | Nominal | |
| Confirmation | coast fever | | |

- 6. Open a new worksheet in your MS Excel Workbook and enter the above East Coast Fever Data.
- 7. Using the sort and filter function in MS Excel, group local areas into the following categories.

| Category | Local areas |
|---------------------|------------------------|
| 1 – Northern region | Amolatar and Dokolo |
| 2 – Central region | Luwero and Nakasongola |
| 3 – Western region | Kiruhura and Isingiro |
| 4 – Eastern region | Budaka and Kibuku |

- **8.** Construct an appropriate table showing cases of East Coast Fever (Theileriosis) by gender. Copy and paste your table from MS Excel into this document.
- 9. Construct an appropriate table showing cases of East Coast Fever (Theileriosis) by region. Copy and paste your table from MS Excel into this document.

Exercise 11 - Display Data Using a Line Graph

Description of Exercise:

Use the tables provided to construct line graphs from cases of brucellosis and theileriosis (east coast fever) in a quarterly fashion and over a ten-year period. Should you have any questions over the exercise, please ask a trainer for clarification.

Allotted Time: 45 minutes

Exercise Components and Structure:

- 1. You have 45 minutes to complete this exercise.
- 2. Work in pairs of two for this exercise.
- 3. Each pair should review the provided tables for brucellosis and east coast fever field testing and create the appropriate line graphs from the data table provided.
- 4. Manipulate data to calculate prevalence and prepare data for graphical display.
- 5. Construct the line graph based on the data provided using MS Excel graphing function.
 - a. Line graph (prevalence of brucellosis stratified by species over 7 quarters)
 - b. Line graph (theileriosis (east coast fever) cases over a 10-year period) in several local areas
- 6. Construct the line graph based on the data provided using MS Excel PivotTable function.
 - a. Line graph prevalence of brucellosis stratified by species over 7 quarters
 - b. Line graph theileriosis (east coast fever) cases over a 10-year period

Materials, Data or Information:

- 1. MS Word
- 2. MS Excel
- 3. Paper and pen

Expected Outputs and Deliverables of Each Participant:

1. Depict animal-place-time by creating line graphs using MS Excel.

Brucellosis Cases in Luwero Local area (January 2017 - September 2018) for Dairy Cattle, Sheep, and Goats

| 2514 | <u> </u> | | o Local area (| Juliudi y = | 2017 | 2010) 101 | Buily cuttie, | Sicop, and G | - Catis |
|----------------------|----------------|--------------------------------|----------------|----------------|--------------------------------|----------------|---------------|--------------------------------|----------------|
| | | Dairy Cattl | e | | Sheep | | | Goats | |
| Month | Total Cases | Total Population (Herds) | Prevalence (%) | Total Cases | Total Population (Herds) | Prevalence (%) | Total Cases | Total Population (Herds) | Prevalence (%) |
| January | 25 | 1000 | | 50 | 1000 | | 60 | 1000 | |
| February | 35 | 1000 | | 65 | 1000 | | 85 | 1000 | |
| March | 22 | 1000 | | 55 | 1000 | | 65 | 1000 | |
| April | 10 | 1000 | | 45 | 1000 | | 75 | 1000 | |
| May | 45 | 1000 | | 25 | 1000 | | 65 | 1000 | |
| June | 30 | 1000 | | 25 | 1000 | | 65 | 1000 | |
| July | 25 | 1000 | | 45 | 1000 | | 80 | 1000 | |
| August | 45 | 1000 | | 35 | 1000 | | 80 | 1000 | |
| September | 40 | 1000 | | 33 | 1000 | | 80 | 1000 | |
| October | 37 | 1000 | | 30 | 1000 | | 80 | 1000 | |
| November | 35 | 1000 | | 25 | 1000 | | 75 | 1000 | |
| December | 29 | 1000 | | 24 | 1000 | | 50 | 1000 | |
| | | | | | 2018 | | | | |
| | | Dairy Cattl | | | Sheep | | | Goats | |
| Month | Total Cases | Total Population (Herds) | Prevalence (%) | Total Cases | Total Population (Herds) | Prevalence (%) | Total Cases | Total Population (Herds) | Prevalence (%) |
| January | 25 | 1000 | | 18 | 1000 | | 45 | 1000 | |
| February | 23 | 1000 | | 15 | 1000 | | 35 | 1000 | |
| March | 20 | 1000 | | 10 | 1000 | | 33 | 1000 | |
| April | 15 | 1000 | | 5 | 1000 | | 30 | 1000 | |
| May | 18 | 1000 | | 2 | 1000 | | 22 | 1000 | |
| June | 11 | 1000 | | 1 | 1000 | | 20 | 1000 | |
| July | 5 | 1000 | | 0 | 1000 | | 10 | 1000 | |
| | 2 | 1000 | | 0 | 1000 | | 7 | 1000 | |
| August | | | | | | | | | |
| September | 0 | 1000 | | 0 | 1000 | | 7 | 1000 | |
| September October | | | | 0 | 1000 | | 7 | 1000 | |
| September | 0 | 1000 | | | | | | | |

Data Dictionary

| Variable | Description | Variable Type | Coding |
|-------------|-----------------------|---------------|---------------------------------|
| Year | Year sample collected | Quantitative, | None |
| | | Discrete | |
| Month | Month sample | Qualitative, | 1 = January; 2 = |
| | collected | Nominal | February; 3 = March; 4 = |
| | | | April; $5 = May$; $6 = June$; |
| | | | 7 = July; $8 = $ August; $9 =$ |
| | | | September; 10 = October; |
| | | | 11 = November; 12 = |
| | | | December |
| Species | Species | Qualitative, | 1 = Dairy Cattle; 2 = |
| | | Nominal | Sheep; 3 = Goats |
| Total Cases | Total cases positive | Quantitative, | None |
| | | Discrete | |
| Total | Total population in | Quantitative, | None |
| Population | herd | Discrete | |
| (Herds) | | | |
| Prevalence | Prevalence | Quantitative, | None |
| | | Continuous | |

- 1. Open a MS Excel Workbook and enter in the aformentioned data into the spreadsheet.
- 2. Conduct data manipulation and insert columns to where you six variables across the top row with the necessary data from the above tables in each column by variable. Identify those six variables.
- 3. In your spreadsheet, calculate the prevalence of cases for each species by month. Prevalence should be showcased as a percentage in the spreadsheet.

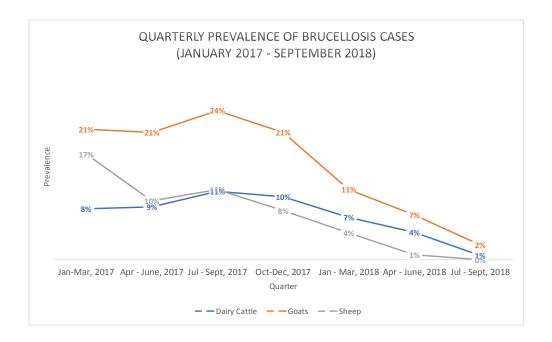
| 2017 and 2018 Prevalence by Species | | | | | | | |
|-------------------------------------|--------------|------|------|------|-------|------|--|
| | Dairy Cattle | | She | ep | Goats | | |
| | 2017 | 2018 | 2017 | 2018 | 2017 | 2018 | |
| January | | | | | | | |
| February | | | | | | | |
| March | | | | | | | |
| April | | | | | | | |
| May | | | | | | | |
| June | | | | | | | |
| July | | | | | | | |
| August | | | | | | | |
| September | | | | | | | |
| October | | | | | | | |
| November | | | | | | | |
| December | | | | | | | |

4. In Excel, add a new variable to your spreadsheet. Use the sort and filter function to code the new "Quarter Variables" by the following numbers.

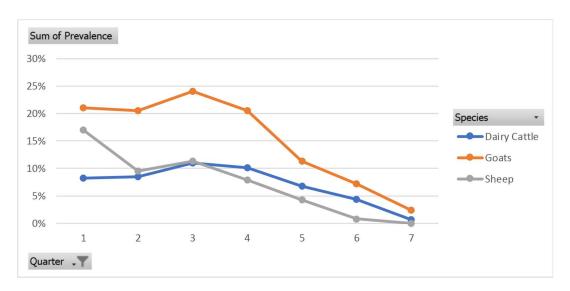
| Variable Code Category | |
|------------------------|--------------------------|
| 1 - Quarter 1 | January – March, 2017 |
| 2 – Quarter 2 | April – June, 2017 |
| 3– Quarter 3 | July – September, 2017 |
| 4– Quarter 4 | October – December, 2017 |
| 5– Quarter 5 | January – March, 2018 |
| 6– Quarter 6 | April – June, 2018 |
| 7– Quarter 7 | July – September 2018 |

5. Construct a line graph which demonstrates the prevalence of brucellosis stratified by species over a seven-quarter period. Title the graph "Quarterly Prevalence of Brucellosis Cases (January 2017 – September 2018)". The x axis should be labeled "Prevalence" and the y axis should be labeled "Cases". Copy and paste your answer into this document.

a) Use the MS Excel line graph function to develop this.



b) Use the MS Excel PivotTable function to develop this. Refer to Pivot Table handout for how to use this function in Excel.

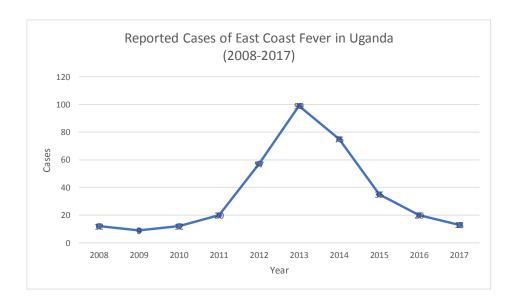


Cases of Theileriosis (East Coast Fever) from 2008-2017

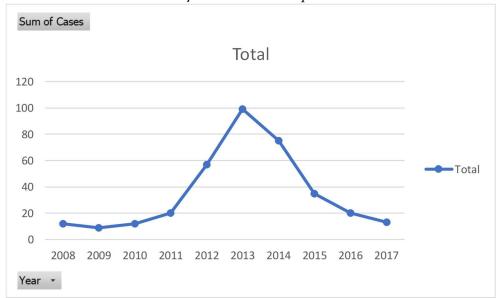
| Region | Local area | 2008 | 2009 | 2010 | 2011 | 2012 | 2013 | 2014 | 2015 | 2016 | 2017 |
|----------|-------------|------|------|------|------|------|------|------|------|------|------|
| Northern | Amolatar | 0 | 0 | 0 | 0 | 0 | 20 | 15 | 5 | 2 | 0 |
| Northern | Dokolo | 0 | 0 | 0 | 0 | 10 | 15 | 12 | 7 | 3 | 1 |
| Central | Luwero | 0 | 0 | 0 | 5 | 10 | 7 | 0 | 0 | 0 | 3 |
| Central | Nakasongola | 0 | 0 | 3 | 5 | 15 | 20 | 10 | 5 | 2 | 1 |
| Eastern | Budaka | 5 | 4 | 4 | 5 | 12 | 15 | 13 | 4 | 4 | 4 |
| Eastern | Kibuku | 7 | 5 | 5 | 5 | 10 | 22 | 7 | 6 | 6 | 3 |
| Western | Kiruhuna | 0 | 0 | 0 | 0 | 0 | 0 | 10 | 5 | 2 | 0 |
| Western | Isingiro | 0 | 0 | 0 | 0 | 0 | 0 | 8 | 3 | 1 | 1 |

- 6. Click on a new Excel worksheet in your MS Excel workbook and enter in the above data into the spreadsheet.
- 7. Construct a line graph which demonstrates the cases of Theileriosis (East Coast Fever) across all regions over a 10-year period. Title the graph "Reported Cases of East Coast Fever in Uganda (2008-2017)". The x axis should be labeled "Year" and the y axis should be labeled "Cases" Copy and paste your answer into this document.

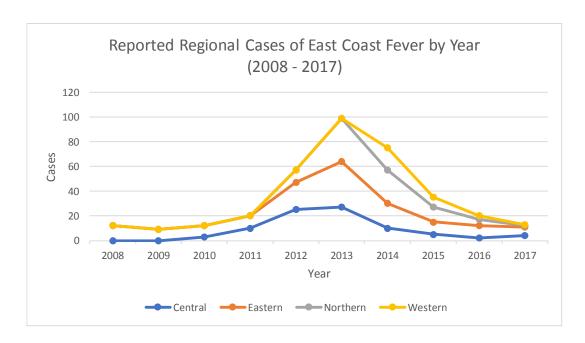
a.) Use the MS Excel line graph function to develop this.



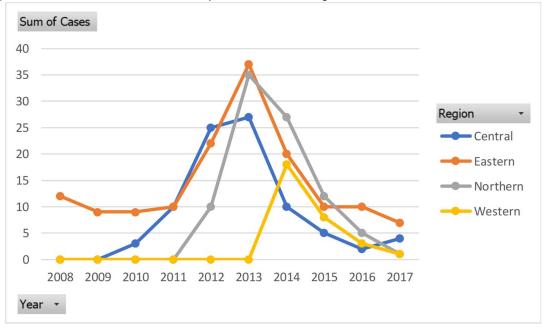
b.) Use the MS Excel PivotTable function to develop this.



- 8. Construct a line graph which demonstrates the cases of Theileriosis (East Coast Fever) stratified by each region over a 10-year period. Title the graph "Reported Regional Cases of East Coast Fever by Year (2008-2017)". The x axis should be labeled "Year" and the y axis should be labeled "Cases" Copy and paste your answer into this document. Copy and paste your answer into this document.
 - a) Use the MS Excel line graph function to develop this.



b) Use the MS Excel PivotTable function to develop this.



Exercise 12 - Display Data Using a Bar Chart and Pie Chart

Description of Exercise:

Based on the information provided in each of the following tables, use the following exercise to develop and display a bar chart and pie chart. Should you have any questions over the exercise, please ask a trainer for clarification.

Allotted Time: 30 minutes

Exercise Components and Structure:

- You have 30 minutes to complete this exercise
- Work in pairs for this exercise;
- Each pair should review the provided tables for brucellosis and east coast fever field testing;
- Create the appropriate bar graphs and pie charts from the data table provided.
- Copy and paste the line graphs from this exercise into an MS Excel worksheet.

Materials, Data or Information:

- 1. MS Word
- 2. MS Excel
- 3. Paper and pen

Expected Outputs and Deliverables of Each Participant:

1. Depict animal-place-time by creating bar and pie charts using MS Excel.

Brucellosis Data

| | | | | | airy Cattle | 4 | | | |
|--------------|---------------|---------|----------|------|----------------|-------|--------------------------|--------------------------------|-----------------------------|
| Sample ID | Local area | Region | Country | Year | Rose Bengal | ELISA | Brucellosis Confirmed | Farmer Consumes Raw Milk | Farmer Boils Raw Milk |
| 1 | Luwero | Central | Uganda | 2016 | + | + | + | + | - |
| 2 | Luwero | Central | Uganda | 2016 | - | + | + | + | - |
| 3 | Luwero | Central | Uganda | 2016 | + | + | + | + | - |
| 4 | Isingiro | Western | Uganda | 2016 | + | + | + | + | - |
| 5 | Marsabit | Eastern | Kenya | 2016 | - | - | - | - | + |
| 6 | Marsabit | Eastern | Kenya | 2016 | + | + | + | + | - |
| 7 | Nyeri | Central | Kenya | 2016 | + | + | + | + | - |
| 8 | Nyeri | Central | Kenya | 2016 | + | + | + | + | + |
| 9 | Chifra | Afar | Ethiopia | 2016 | - | + | + | + | - |
| 10 | Chifra | Afar | Ethiopia | 2016 | - | + | + | - | - |
| 11 | Dubti | Afar | Ethiopia | 2016 | - | + | + | - | - |
| 12 | Banja | Amahara | Ethiopia | 2016 | - | - | - | - | + |
| 13 | Longido | Arusha | Tanzania | 2016 | + | + | + | + | - |
| 14 | Longido | Arusha | Tanzania | 2016 | + | + | + | + | - |
| 15 | Longido | Arusha | Tanzania | 2016 | + | + | + | + | + |
| 16 | Luwero | Central | Uganda | 2017 | + | + | + | + | + |
| 17 | Luwero | Central | Uganda | 2017 | + | + | + | + | - |
| 18 | Isingiro | Western | Uganda | 2017 | + | + | + | + | - |
| 19 | Meru | Eastern | Kenya | 2017 | + | + | + | - | - |
| 20 | Meru | Eastern | Kenya | 2017 | - | + | + | - | - |
| 21 | Meru | Eastern | Kenya | 2017 | + | + | + | + | - |
| 22 | Nyeri | Central | Kenya | 2017 | + | + | + | + | - |
| 23 | Nyeri | Central | Kenya | 2017 | + | + | + | + | - |
| 24 | Nyeri | Central | Kenya | 2017 | + | + | + | + | + |
| 25 | Banja | Amahara | Ethiopia | 2017 | + | + | + | + | + |
| 26 | Banja | Amahara | Ethiopia | 2017 | + | + | + | + | + |
| 27 | Longido | Arusha | Tanzania | 2017 | + | + | + | - | - |
| 28 | Longido | Arusha | Tanzania | 2017 | + | + | + | + | + |
| 29 | Longido | Arusha | Tanzania | 2017 | + | + | + | + | + |

Brucellosis Data Continued

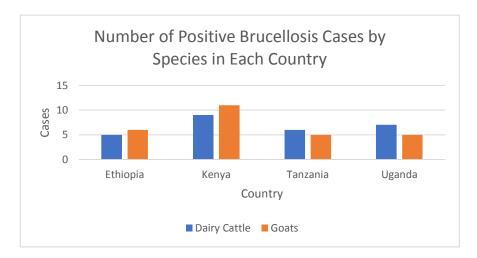
| | | | | | Goats | | | | |
|--------------|---------------|---------|----------|------|----------------|-------|--------------------------|--------------------------------|-----------------------------|
| Sample ID | Local area | Region | Region | Year | Rose Bengal | ELISA | Brucellosis Confirmed | Farmer Consumes Raw Milk | Farmer Boils Raw Milk |
| 30 | Isingiro | Western | Uganda | 2016 | + | + | + | + | - |
| 31 | Isingiro | Western | Uganda | 2016 | - | + | + | + | - |
| 32 | Marsabit | Eastern | Kenya | 2016 | + | + | + | + | - |
| 33 | Nyeri | Central | Kenya | 2016 | + | + | + | + | - |
| 34 | Nyeri | Central | Kenya | 2016 | - | - | - | - | + |
| 35 | Nyeri | Central | Kenya | 2016 | + | + | + | + | - |
| 36 | Marsabit | Eastern | Kenya | 2016 | + | + | + | + | - |
| 37 | Marsabit | Eastern | Kenya | 2016 | + | + | + | + | + |
| 38 | Dubti | Afar | Ethiopia | 2016 | - | + | + | + | - |
| 39 | Dubti | Afar | Ethiopia | 2016 | - | + | + | - | - |
| 40 | Dubti | Afar | Ethiopia | 2016 | - | + | + | - | - |
| 41 | Karatu | Arusha | Tanzania | 2016 | - | - | - | - | + |
| 42 | Karatu | Arusha | Tanzania | 2016 | + | + | + | + | - |
| 43 | Karatu | Arusha | Tanzania | 2016 | + | + | + | + | - |
| 44 | Karatu | Arusha | Tanzania | 2016 | + | + | + | + | + |
| 45 | Luwero | Central | Uganda | 2017 | + | + | + | + | + |
| 46 | Luwero | Central | Uganda | 2017 | + | + | + | + | - |
| 47 | Luwero | Central | Uganda | 2017 | + | + | + | + | - |
| 48 | Marsabit | Eastern | Kenya | 2017 | + | + | + | - | - |
| 49 | Marsabit | Eastern | Kenya | 2017 | - | + | + | - | - |
| 50 | Marsabit | Eastern | Kenya | 2017 | + | + | + | + | - |
| 51 | Nyeri | Central | Kenya | 2017 | + | + | + | + | - |
| 52 | Nyeri | Central | Kenya | 2017 | + | + | + | + | - |
| 5 3 | Nyeri | Central | Kenya | 2017 | + | + | + | + | + |
| 54 | Karatu | Arusha | Tanzania | 2017 | + | + | + | + | + |
| 55 | Dubti | Afar | Ethiopia | 2017 | + | + | + | + | + |
| 56 | Dubti | Afar | Ethiopia | 2017 | + | + | + | - | - |
| 57 | Dubti | Afar | Ethiopia | 2017 | + | + | + | + | + |
| 58 | Karatu | Arusha | Tanzania | 2017 | + | + | + | + | + |

Data Dictionary

| Variable | Description | Variable Type | Coding |
|--------------|-----------------------|---------------|-----------------------------|
| Sample ID | Sample ID number | Quantitative, | None |
| | | Discrete | |
| Local area | Local area sample | Qualitative, | 1 = Luwero; 2 = Isingiro; 3 |
| | collected in | Nominal | = Marsabit; 4 = Nyeri; 5 = |
| | | | Chifra; 6 = Banja; 7 = |
| | | | Longido; 8 = Marsabit; 9 = |
| | | | Dubti; 10 = Karatu |
| Region | Region local area is | Qualitative, | 1 = Central; 2 = Eastern; |
| | located in | Nominal | 3 = Afar; 4: Amahara; 5: |
| | | | Arusha |
| Year | Year | Quantitative, | 1 = +; 0 = - |
| | | Discrete | |
| Rose Bengal | Positive Pose Bengal | Qualitative, | 1 = +; 0 = - |
| | test | Binary | |
| ELISA | Positive ELISA test | Qualitative, | 1 = +; 0 = - |
| | | Binary | |
| Confirmed | Confirmed Brucellosis | Qualitative, | 1 = +; 0 = - |
| Brucellosis | positive | Binary | |
| Farmer | Farmer consumes raw | Qualitative, | 1 = +; 0 = - |
| Consumes Raw | milk from herd | Binary | |
| Milk | | | |
| Farmer Boils | Farmer boils raw milk | Qualitative, | 1 = +; 0 = - |
| Raw Milk | collected from herd | Binary | |

- 1. Open a MS Excel Workbook and enter in the above data into the spreadsheet.
- 2. Develop a bar graph to display the relationship of Species to the number of cases of diagnosed brucellosis through confirmatory testing stratified by country. Show countries separately from each other. Title the graph "Number of Positive Brucellosis Cases by Species in each Country" The X axis should be titled "Country" and the Y axis should be titled "Cases" The legend should include each species.

a. Use the MS Excel bar graph function to develop this.

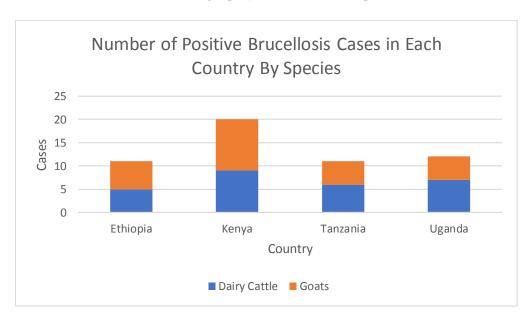


b. Use the MS Excel PivotTable function to develop this.

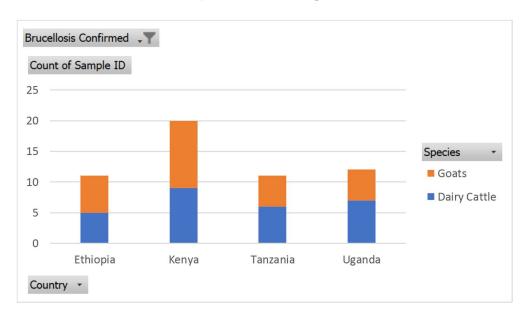


3. Develop a bar graph to display the relationship of Species to the number of cases of diagnosed brucellosis through confirmatory testing stratified by country. Show countries separately from each other. Title the graph "Number of Positive Brucellosis Cases by Species in each Country" The X axis should be titled "Country" and the Y axis should be titled "Cases" The legend should include each species.

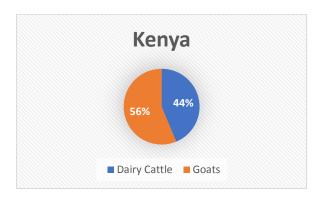
a. Use the MS Excel stacked bar graph function to develop this.



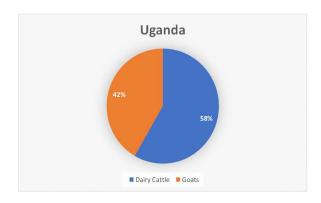
b. Use the MS Excel PivotTable function to develop this.



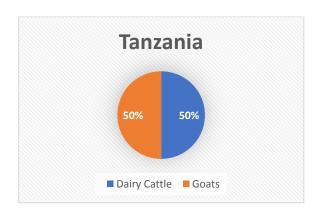
- 4. Develop a pie graph that shows the percentage of farmers who consumed raw milk from all herds that were brucellosis positive by species for each country.
 - a. Kenya:



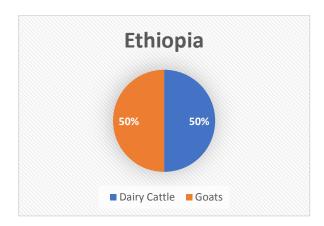
b. Uganda:



c. Tanzania:



d. Ethiopia:



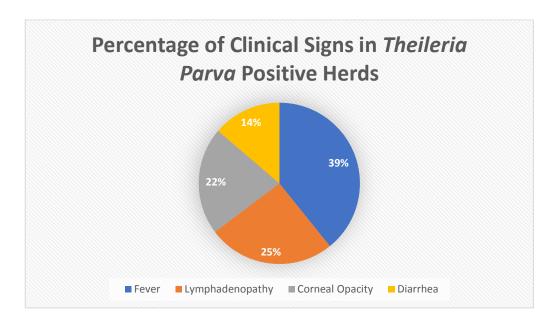
Theileriosis (East Coast Fever) Data

| Herd | Fever | Lymphadenopathy | Corneal | Diarrhoea | Theileria parva |
|------|-------|-----------------|---------|-----------|-----------------|
| ID | | | Opacity | | sp. Positive |
| 1 | + | + | + | - | + |
| 2 | - | - | + | - | - |
| 3 | + | - | + | - | + |
| 4 | + | - | + | - | + |
| 5 | + | - | + | - | + |
| 6 | + | + | + | - | + |
| 7 | + | + | + | - | + |
| 8 | + | + | + | - | + |
| 9 | - | + | + | - | - |
| 10 | + | + | + | - | + |
| 11 | + | + | - | - | + |
| 12 | + | + | - | - | + |
| 13 | + | + | - | - | + |
| 14 | + | + | - | - | + |
| 15 | + | - | - | - | + |
| 16 | + | - | - | + | + |
| 17 | + | - | - | + | + |
| 18 | + | + | - | + | + |
| 19 | + | + | - | - | - |
| 20 | - | + | _ | - | - |
| 21 | + | + | - | - | - |
| 22 | + | - | - | - | - |
| 23 | + | - | - | - | - |
| 24 | - | + | - | - | - |
| 25 | + | + | - | + | - |
| 26 | + | - | - | + | + |
| 27 | - | - | - | + | - |
| 28 | + | + | + | + | + |
| 29 | + | + | + | + | + |
| 30 | + | + | + | + | + |

Data Dictionary

| Variable | Description | Variable Type | Coding |
|------------------|----------------------|---------------|--------------|
| Herd ID | Herd ID number | Quantitative, | None |
| | | Discrete | |
| Fever | Local area sample | Qualitative, | 1 = +; 0 = - |
| | collected in | Binary | |
| Lymphadenopathy | Region local area is | Qualitative, | 1 = +; 0 = - |
| | located in | Binary | |
| Corneal Opacity | Year | Quantitative, | 1 = +; 0 = - |
| | | Discrete | |
| Diarrhoea | Positive Pose Bengal | Qualitative, | 1 = +; 0 = - |
| | test | Binary | |
| Theirleria parva | Positive ELISA test | Qualitative, | 1 = +; 0 = - |
| sp. positive | | Binary | |

5. Develop a pie graph that shows the percentage of clinical signs (i.e., fever, lymphadenopathy, corneal opacity and diarrhea) which were in *theileria parva* positive herds.



Case Study 1: PPR

Lesson 7 – Data Interpretation and Reporting to Improve Situational Awareness and Decision-Making

| Estimated Lesson and Exercise Time | 2 hours |
|------------------------------------|---|
| Participant Materials | Frontline ISAVET Lesson 7 – Data |
| | Interpretation and Reporting to Improve |
| | Situational Awareness and Decision- |
| | Making Participant Guide.PDF |
| | Computer and Microsoft Word |
| | Pen or Pencil |

In Service Applied Veterinary Epidemiology Training (ISAVET)

Lesson 7: Data Interpretation and Reporting to Improve Situational Awareness and Decision-Making



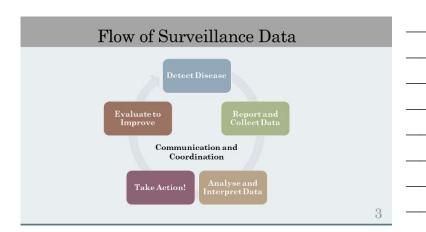
Learning Objectives

At the end of this lesson, you be able to:

- Apply methods used to interpret data to improve situational awareness;
- 2. Describe reasons that could lead to true or artefactual increase in surveillance cases.
- 3. Describe the use of thresholds.

-

4



Data Interpretation

- The main aim is to differentiate between $\underline{\text{real}}$ vs $\underline{\text{artifacts}}$ in surveillance data
 - To detect first signals of outbreaks
- How are outbreaks detected?
- · Passive surveillance:
- Farmers reporting of cases
- Frontline veterinarians/clinicians who see multiple cases and make an association
- $\dot{\,}$ For certain diseases, one case may be enough to raise an alarm
- · Active surveillance systems:
- ${\boldsymbol{\cdot}}$ Syndromic surveillance
- Abattoir
- Laboratory
- Production

Principles of Data Interpretation

1. Assess the quality of your data before proceeding further.

2. Identify the limitations in your data.

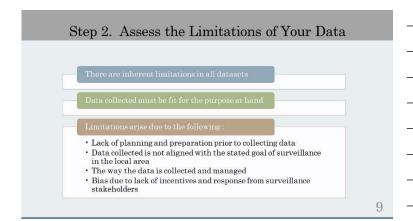
3. Create simple statements that summarise each point of analysis.

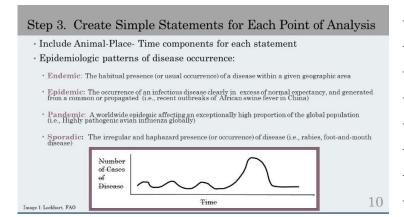
4. Compare the results of your data analysis to what you would expect based on the results of previous analysis.

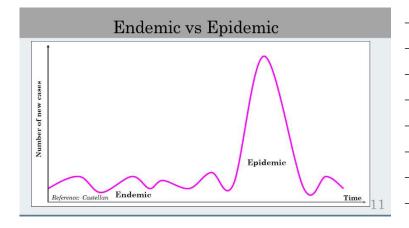
5. Explain possible reasons why there is a difference between observed and expected results.

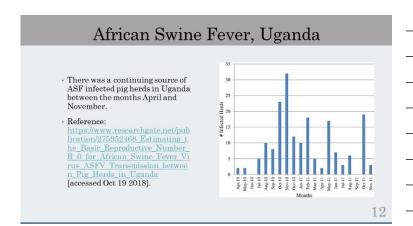
6. Explain what the results infer for those making decisions.

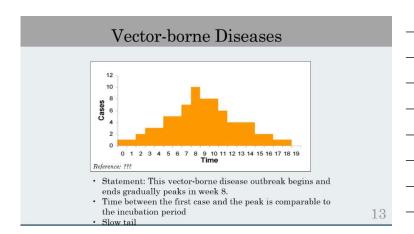
Basic Assumptions · Assumption 1: Have you collected high quality data? • Assumption 2: Do you have high quality data to compare it to? · If you cannot answer yes to both questions, it's not too late to begin the process of improving data quality in your local area office. 6 Step 1. Assess the Quality of Your Data · What are the considerations for data quality? - Refers to accuracy and completeness of data gathered and that they convey the intended meaning - Begins by ensuring that data is gathered in a standard way - Standardised application based on a standard data collection form · Data quality measures and indicators: - Must be agreed upon and written down - Provide insights to assess data quality Assess the Quality of Your Data · Data quality measures and indicators - some guidelines in assessing data quality: Relevance: to stated objectives Accuracy: to what we intend to measure Integrity: data is stable and reliable Timeliness / time lag: we require data for immediate response and action Accessibility and clarity: clear and accessible data Comparability: can compare with existing data Coherence: the data makes sense Coverage / representativeness: data represents the population Redundant / duplicates: remove duplicate values

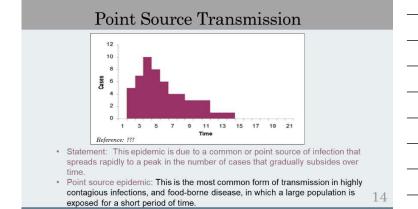




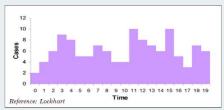








Continuing Common Source or Intermittent Exposure

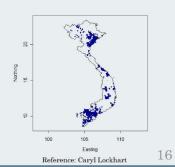


- Statement: This epidemic is due to a common or point source of infection that spreads rapidly followed by a peak in the number of cases that gradually subsides great lim.
- $\bullet \quad \text{In this case, there are several peaks, and the incubation period cannot be identified.} \\$

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Spatial Distribution of Highly Pathogenic Avian Influenza H5N1 in Viet Nam

- Maps provide complementary information to animal and time data to describe disease occurrence.
- Statement: Highly Pathogenic Avian Influenza H5N1 detection in Viet Nam is concentrated near the major cities in North and South Viet Nam, as well as along transportation routes.

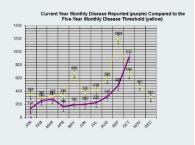


Step 4. Compare Observed Data with Expected Data

- Example
 - If the expected (i.e. normal) prevalence of disease X in country Y is 0.02
- Local area 1 has population of 20,000 animals
 - * The expected number of cases of disease would be = 0.02 X 20,0000 = 400
 - * If the number of cases observed in local area X $\,$ is 500, then we have 100 more cases than we expect based on previous estimates
 - ACTION STEP: Consult with the national epidemiologist to discuss the interpretation of the results and the action to be taken. Remember that disease crosses boundaries and needs to be considered at both the local and the higher levels.

Making Decisions Based on Thresholds

- A threshold is an expected value that is calculated for a given time period. We then compare our current data with the threshold value to determine whether there is an unexpected increase in the disease.
- · EXAMPLE: No. Reported Suspect Animal Cases of FMD
- Yellow line 5 year threshold line:
- Blue line current year: Note that the level in October exceeds the threshold value and action must be taken



Reference: Frontline ISAVET

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Step 5. Explain the Reasons for the Difference Between Observed and Expected Results

Real increase based on statistical evidence

Apparent increase

- · Reporting error
- Massurament arro
- Reporting bias (population changes over time)
- Change in reporting procedures / change in surveillance system
- New personnel
- · Change in case definition
- Improvements in diagnostic procedures
- Increased awareness/interest
- In or out migration of populations
- Change in denominator or population size

19

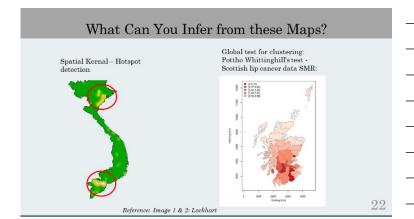
Exercise 13: Calculation of a 5-Year Average

- 1. Refer to MS Excel YouTube Video #12: Calculating Thresholds in Excel
- $_{\rm 2.}$ This exercise should take 45 minutes to complete.
- 3. Work in pairs.
- 4. Calculate the 5-year average incidence of lung lesions from abattoir data provided.
- 5. Describe the use of thresholds.

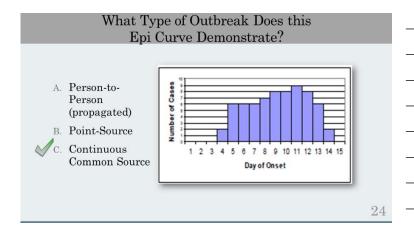
Step 6. Explain What the Results Infer for Decision Makers

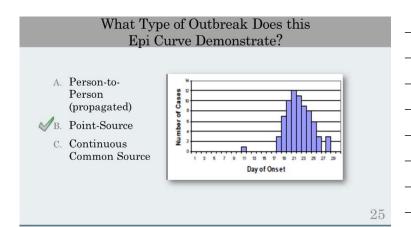
- · What do the results infer about the disease?
 - a. Is it an epidemic or endemic?
 - b. What are the spatial and temporal patterns of disease?
 - c. Is this a single incident or should you look further for more cases?
 - d. Are the results realistic and biologically plausible?
 - e. Is one location or part of the population affected more than others?
- · How do the results reflect the quality of your data?
 - a. Have you accounted for the limitations of the data?
 - b. Is your data consistent and comparable with existing data?
 - c. Do your recommendations reflect the limitations of your data?

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What Type of Outbreak Does this Epi Curve Demonstrate? A. Animal-to-Animal (propagated) B. Point-Source C. Continuous Common Source Day of Onset





Exercise 14: Identify the Challenges in Threshold Deviations

- 1. This exercise will take 45 minutes to complete.
- 2. Divide into groups of four.
- 3. Answer the guiding questions and identify specific challenges in interpreting thresholds and making recommendations.

In Summary...

- 1. Data quality is fundamental to good decision making.
- 2. Make it a habit of providing clear and simple statements for each one of your descriptive analysis results.
- 3. Understand the limitations of your data.
- 4. Thresholds are used to make recommendations for action by comparing observed and expected data results; and
- 5. Bew aware of challenges in interpreting thresholds and making recommendations.

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Partners Contributors Contributors Contributors

Exercise 13 – Calculation of a 5-Year Average

Description of Exercise:

Calculate a 5-year average of the lung lesion disease count from the abattoir data provided. Should you have any questions over the exercises, please ask a trainer for clarification before, during, and after the exercises.

Allotted Time: 45 minutes

Exercise Components and Structure:

- 1. Refer to MS Excel YouTube Video #12: Calculating Thresholds in Excel
- 2. This exercise should take 45 minutes to complete.
- 3. Work in pairs.
- 4. Calculate a 5-year average incidence of lung lesions from abattoir data provided.
- 5. Describe the use of thresholds.
- 6. Copy data into the MS Excel spreadsheet provided as follows:
 - a. Copy the following number of cases of cattle lung lesions for each month between 2000 and 2004 below.

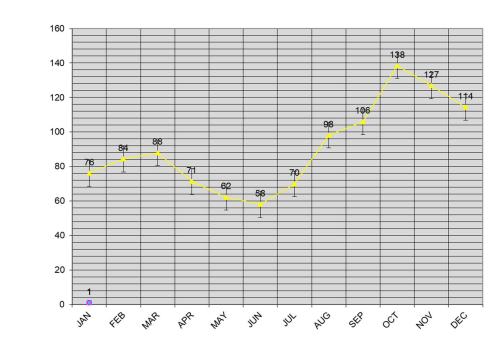
Materials, Data or Information:

- 1. Data set
- 2. MS Excel File: L7_Ex13

1. Using the MS Excel spreadsheet, titled "ISAVET Exercise 13 Participant", calculate the following.

| Number of | Cattle with L | ung Lesions | | | | | | | | | | |
|-----------|---------------|-------------|-------|-------|-----|------|------|--------|-----------|---------|----------|----------|
| Year | January | February | March | April | May | June | July | August | September | October | November | December |
| 2000 | 45 | 34 | 42 | 56 | 48 | 38 | 29 | 26 | 27 | 23 | 23 | 24 |
| 2001 | 31 | 39 | 47 | 54 | 57 | 50 | 37 | 30 | 27 | 30 | 36 | 38 |
| 2002 | 50 | 56 | 65 | 63 | 57 | 39 | 27 | 19 | 19 | 25 | 28 | 34 |
| 2003 | 57 | 73 | 77 | 68 | 55 | 35 | 34 | 55 | 66 | 99 | 88 | 91 |
| 2004 | 66 | 61 | 65 | 57 | 57 | 52 | 66 | 86 | 88 | 103 | 99 | 82 |

Five-Year Monthly Disease Threshold (yellow)

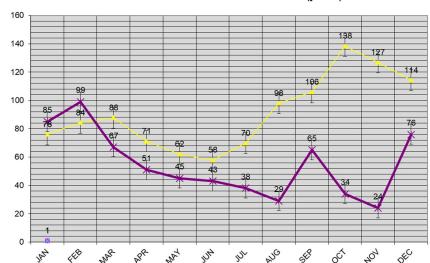


No. Disease Reports

b. Copy the number of cases of cattle lung lesions for the current year below:

| Current year | JAN | FEB | MAR | APR | MAY | JUN | JUL | AUG | SEP | ОСТ | NOV | DEC |
|--------------|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| | 85 | 99 | 67 | 51 | 45 | 43 | 38 | 29 | 65 | 34 | 24 | 76 |

Current Year Monthly Disease Reported (purple) Compared to the Five-Year Monthly Disease Threshold (yellow)



No. Disease Reports

2. Using the MS Excel spreadsheet, titled "ISAVET Exercise 13 Participant", calculate the 5-year average disease counts from abattoir data.

Exercises 14 – List the Challenges in Recognizing Deviations Above Thresholds

Description of Exercise:

Identify the challenges in recognising deviations above the threshold. Should you have any questions over the exercises, please ask a trainer for clarification before, during, and after the exercises.

Allotted Time: 45 minutes

Organisation of Group Work:

• Divide into groups of four.

Exercise Objective(s):

- 1. Apply methods used to interpret data to improve situational awareness.
- 2. Describe signals and thresholds, and how they are used to make recommendations for action.
- 3. Answer the guiding questions and identify specific challenges in interpreting thresholds and making recommendations.

Exercise Components and Structure:

1. Discuss and list the challenges in recognising deviations above thresholds.

Guiding Questions:

- 1. Which months of the current year recorded the highest number of lung lesions?
- 2. List possible reasons for the decline in the number of lung lesions from September to November in the current year as compared with the five year average?
- 3. At this stage, how would you interpret the results of this year's lung lesions to your supervisor?
- 4. What additional information do you need to assess in order the determine if the results reflect a real change in the number of lung lesions or if they reflect a change in how we detect lung lesions?

Response Checklist:

Lesson 8 - Elements of a Surveillance Report

| Estimated Lesson and Exercise Time | 2 hours |
|------------------------------------|---|
| Participant Materials | Frontline ISAVET Lesson 8 Elements of a Surveillance Report Participant Guide Version 5.PDF |
| | Computer Microsoft Word and Excel |
| | Pen or pencil |

| In Service Applied Veterinary Epidemiology Training (ISAVET) | |
|---|---|
| Lesson 8: Elements of a Surveillance Report | |
| | |
| INSTITUTE FOR SHOULD DISFORE | Food and Agriculture Organization of the United Nations |

Learning Objectives

| A | t the end of this lesson, you will be able to: |
|----|--|
| 1. | Describe the structure and list the components of a basic form for surveillance reporting in terms of animal, place and time; |

2. Produce a brief summary report with recommendations for action.

Objectives and Use of Animal Health Surveillance (Epidemiological Surveillance in Animal Health, OIE) Objectives Detect new introductions and emerging diseases: e.g. Nipah virus, Ebola virus Assess disease trends over time: e.g. TB, rabies, brucellosis Estimate the burden of animal diseases – estimate disease prevalence: e.g. FMD, Newcastle disease Define important priority diseases: e.g. RVF, anthrax Evaluation of animal disease control programmes – establish freedom from disease: e.g. rinderpest, PPR

Surveillance Objectives with Examples

- · Determine prevalence of an existing disease
 - Endemic brucellosis, rabies, highly pathogenic avian influenza (HPAI)
- Prove freedom from disease
 - Foreign animal diseases such as bovine spongiform encephalopathy (BSE)
- · Detect a new disease
 - Bat viruses, H1N1 in swine

| | ents of Survei Terrestrial Animal He | | | |
|---------------------------------|---|--------------------------------|------------|---|
| 1. A-P-T | 2. Definitions | 3. D | ata | |
| Populations at risk (Animal) | Epidemiological Unit | Data Col and Manage | d | |
| Clustering (Place) | Case Definition | Applica the Appr Data An | opriate | |
| Time Period (Time) | Diagnostic Testing | Quality Assurance | Validation | 5 |

Sources of Surveillance Data (Ch. 1.4 Terrestrial Animal Health Code. OIE, 2018) 1. Sub-local area and local area level office data 2. Wildlife data 3. Farm production records ${\it 4. Targeted testing programmes (e.g. swill feeding)}\\$ $5.\ Ante-mortem\ and\ post-mortem\ inspections\ (abattoirs,\ markets)$ 6. Laboratory investigation records 7. Sentinel surveillance sites 8. Field observation and investigation reports 9. National disease control programmes 6

Scenario: Laboratory Surveillance of Anthrax, Nakuru, Kenya

- This retrospective study includes a data review of anthrax cases in livestock and wildlife at RVIL, Nakuru Kenya from January 2013 to December 2018. This facility serves all livestock and wildlife populations in Nakuru County and its border counties like Baringo, Nyandarua, Laikipia and Narok.
- · The National Census of 2009 showed that livestock populations in Nakuru were:
 - Cattle 513,286
- Sheep 567,009 Goats 355,306
- Poultry 1,844,180
- Pigs 20,137
- Rabbits 89,813
- Donkeys 51,935

| Species | Production Class | Production System | Demographic Details | Number Counts |
|--|---|---|---|---------------------------------|
| Cattle | Dairy Beef Dual purpose Breeder | Intensive Semi-intensive Extensive | Breed Age Sex | |
| • Sheep | Milk Meat | | Health and vaccination status | |
| • Goats | Dual purpose Breeder | | Feed and water source | |
| Chickens Ducks Geese Other | Meat Eggs Dual purpose Breeder | Closed house Semi-closed Extensively raised smallholder | Breed Strain Age Sex Health and vaccination status, Feed and water source | Total at risk Sick Treated Dead |
| Horse Donkey Mule Other | Draft Pleasure Meat Dual purpose | Intensive, Semi-intensive Extensive | Breed Age Sex Health and vaccination | |
| • Pets | Dogs, cats, other | Domestic Feral Mixed | status Food and water source | 8 |

Scenario: Anthrax Animal Data

(Frontline ISAVET

 Animal data is disaggregated (stratified) by species to permit analysis and comparison of the number of samples received

| Species Screened | No. Samples tested |
|------------------|--------------------|
| | 128 |
| | 1 |
| | 2 |
| | 1 |
| | 16 |
| Rhino | 8 |
| | 4 |
| | 9 |
| | 169 |

9

1b. Place Data Elements

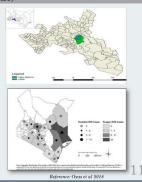
- · GPS latitude and longitude coordinates
- · Administrative boundaries
- · Nearby roads, railways, rivers and water bodies
- · Housing and mobility
- · Value chain maps for each production class
- Do you have a map with important animal data in your office to show disease distribution and clusters?

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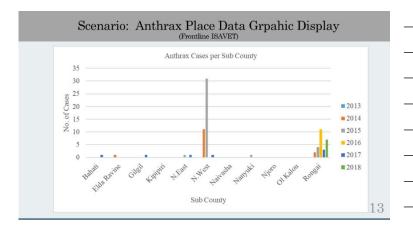
Scenario: Anthrax Place Data (Frontline ISAVET)

- The top map displays the anthrax surveillance area but does not include GPS spot map coordinates of the locations of all positive and negative samples
- The bottom map is an example of what a spot map looks like based on GPS coordinates of the spatial distribution of Rift Valley Fever (RVF) cases in Kenya
- Kenya

 The size of each circle is proportional to a range in the number of positive cases of RVF



| | anu | by Y | ear (| rontline | ISAVET |) | |
|---------------------------------|------|------|-------|----------|--------|------|-------|
| | | | | | | | |
| Table 4: Summ anthrax (B.ant | | | | | | | |
| Sub County | 2013 | 2014 | 2015 | 2016 | 2017 | 2018 | Total |
| Bahati | 0 | 0 | 0 | 0 | 1 | 0 | 1 |
| Elda Ravine | 0 | 1 | 0 | 0 | 0 | 0 | 1 |
| Gilgil | 0 | 0 | 0 | 0 | 1 | 0 | 1 |
| Kipipiri | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| N.East | 0 | 0 | 1 | 0 | 1 | 0 | 2 |
| N.West | 0 | 11 | 31 | 0 | 1 | 0 | 43 |
| Naivasha | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Nanyuki | 0 | 0 | 1 | 0 | 0 | 0 | 1 |
| Njoro | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Ol Kalou | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | 0 | 5 | 4 | 11 | 3 | 4 | 27 |
| Total | 0 | 17 | 37 | 11 | 7 | 4 | 76 |



| Reporting | Observation | Diagnostie Testing | Treatment | Movement (since last report) |
|--------------------------------|--|--|-------------------------------------|---|
| ☐ Date report submitted | ☐ Date of visit or inspection: either active or passive | ☐ Date test collected | Date of last antibiotic treatment | Date of new introductions into the flock or herd |
| ☐ Date reviewed | ☐ Date clinical signs observed | ☐ Date test submitted | Date of recent vaccinations | Date of movement from the flock or herd |
| ☐ Date feedback provided | ☐ Evidence of temporal clustering | Date test result received | ☐ Tick and insect control | ☐ Movement control measures |
| ☐ Seasonal events | □ Date of on farm post- mortem | ☐ Date result shared with farmer | ☐ Date last given supplements | |

| | * | 2220 | |
|------------|----------------------------|--------------------------|---------------|
| Table 2: S | Summary of temporal dist | ribution of reported liv | estock and |
| | nthrax (B. anthracis) posi | | |
| | | tive cases at KVIL, Nai | turu, 2015 to |
| 2018 n=7 | 6 | | |
| Year | Number of sample tested | B.anthracis (Positive) | Number dead |
| 2013 | 7 | 0 | 0 |
| 2014 | 48 | 17 | 16 |
| 2015 | 54 | 37 | 19 |
| 2016 | 26 | 11 | 7 |
| 2017 | 22 | 7 | 7 |
| 2018 | 12 | 4 | 3 |
| Total | 169 | 76 | 52 |

| Epidemiological unit | "Animals with a defined epidemiological relationship that shar approximately the same likelihood of exposure to a pathogenic agent." (Terrestrial Animal Health Code. OIE, 2018) | | |
|-------------------------|---|--|--|
| Case definition | Suspect: clinical signs | | |
| | Probable: screening test and epidemiological links | | |
| | Confirmed: gold standard test e.g. PCR | | |
| Diagnostic testing | Preliminary: screening test e.g. Rose bengal | | |
| | Interim: e.g. ELISA | | |
| | Confirmed: e.g. culture, PCR | | |

| Table A: Disaggregated Data (detailed data provided) | | | | | | | | |
|--|--------------|--------------|--------------------------|------------------|--|--|--|--|
| | Dairy Cattle | Beef Cattle | Dual Purpose Cattle | Local Area Total | | | | |
| Sublocal area A | 1, 510 | 10,101 | 8,000 | 18,101 | | | | |
| Sublocal area B | 150 | 8,555 | 6,544 | 15,249 | | | | |
| Sublocal area C | 3,226 | 5,498 | 2,563 | 11,287 | | | | |
| Sublocal area D | 2,500 | 3,400 | 876 | 6,776 | | | | |
| Sublocal area Total | 5,876 | 27,554 | 17,983 | 51,413 | | | | |
| | Table B: | Aggregated D | ata (detailed data lost) | | | | | |
| | Dairy Cattle | Beef Cattle | Dual Purpose Cattle | Local Area Total | | | | |
| Local Area Total | 5,876 | 27,554 | 17,983 | 51,413 | | | | |

| | Fable 1: Summary of Livestock and wildlife species distribution of | | | | | | | | | | |
|------|---|----|-----|---------|--|--|--|--|--|--|--|
| | anthrax (B. anthracis) at RVIL, Nakuru, from 2013 to 2018 (n=76) | | | | | | | | | | |
| sted | Species No. Samples B. anthracis % samples tested Screened tested (Positive) positive | | | | | | | | | | |
| | 57.9 | 44 | 128 | Bovine | | | | | | | |
| | 1.3 | 1 | 1 | | | | | | | | |
| | 2.6 | 2 | 2 | Ovine | | | | | | | |
| | 0 | 0 | 1 | | | | | | | | |
| | 18.4 | 14 | 16 | Buffalo | | | | | | | |
| | 10.5 | 8 | 8 | Rhino | | | | | | | |
| | 5.3 | 4 | 4 | Zebra | | | | | | | |
| | 4 | 3 | 9 | | | | | | | | |
| | 100 | 76 | 169 | Total | | | | | | | |

Steps in Completing the local area Weekly Surveillance Report

STEP 1:

 \checkmark Complete Table 1: Weekly Metrics

STEP 2

√Colour Code "No. of Reports" and "Year to Date" columns in Table 1

STEP 3

 \checkmark Complete Table 2: Weekly Disease Reporting Form

STEP 4:

√ Complete Table 3: Summary of Key Notifiable Diseases this Week and Cumulatively

STEP 5:

Create weekly and monthly graphs of two major animal diseases of concern in your local area to show the disease trend over time

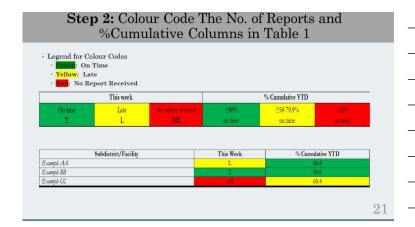
19

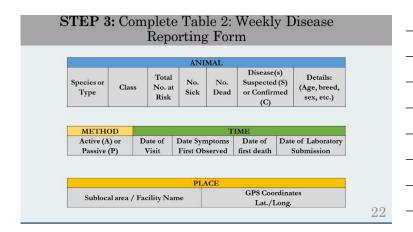
Step 1: Complete Table 1 Weekly Surveillance Report Metrics

 $A\ metric\ is\ something\ we\ measure\ to\ assess\ the\ performance\ of\ the\ surveillance\ system$

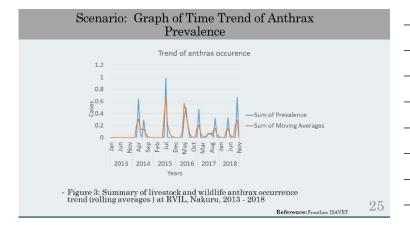
Table 1: Weekly Sublocal area Reporting Summary of Completeness and Timeliness

| Subdistrict Name | No. Reports Received This Week | Cumulative YTD* No. (%) Weekly Reports Received in [Year] | Mode No. Days from Laboratory Submission to District Reporting | Mode No. Days from District Notification to Farmer Reporting (Days) | No. of Surveillance Events Reported this Year |
|----------------------------|--------------------------------------|---|---|---|---|
| Subdistrict/Facility A | | | | | |
| Subdistrict/Facility B | | | | | |
| Subdistrict/Facility C | | | | | |
| Subdistrict/Facility D | | | | | |
| Subdistrict/Facility E | | | | | |
| Subdistrict/Facility F | | | | | |
| Subdistrict/Facility G | | | | | |
| Subdistrict/Facility H | | | | | |
| % Reports Received to Date | Total: | % Cumulative YTD: | District Mode: | District Mode: | District Total = |





| | | nis W | COL | | | |
|---------------------|---------------------|--------------|------------------------|---------|--------------|--------------------------|
| List the key noti | fiable diseases in | your lo | cal area | | | |
| 2. Count and record | d the number of s | uspecte | d and cor | firme | d cases | |
| . Count and record | d the number of d | eaths d | lue to the | diseas | e | |
| . Calculate the cas | se-fatality rate (N | lo. deat | hs / No. o | f Cases | s) | |
| | | | | | | |
| | | | | | | |
| | | Current Week | k No. | Cı | unulative We | eekly No. |
| | | Current Wee | k No. Case Fatality | Cı | mulative Wo | eekly No. Case Fatality |



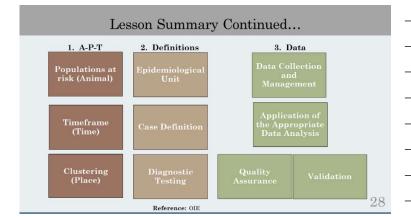
Exercise 15a: Weekly Surveillance Report

- 1. This exercise will take 60 minutes.
- 2. Form yourselves into groups of roughly equal size.
- 3. Create a surveillance report with the information provided.
- 4. Each group will make comments on what the data reveals.

Lesson Summary...

- There are three elements required in an animal disease surveillance report:
 - Animal-Place-Time
 - Definitions
- Data Management
- \cdot Raw, disaggregated data is required in a surveillance report in order to analyse data and take action!
- There are three steps for completing a Weekly local area Surveillance Report
 - STEP 1: Complete Table 1: Weekly Metrics
 - $^{\circ}$ STEP 2: Colour Code "No. of Reports" and "Year to Date" columns in Table 1
 - * STEP 3: Complete Table 2: Weekly Disease Reporting Form

Reference: OIF





Exercise 15a - Local Area Disease Surveillance Report

Description of Exercise:

Create a surveillance report from information provided in tables below. Determine what the data reveals and provide comments in the surveillance report. Should you have any questions over the exercise, please ask a trainer for clarification before during, and after the exercise.

Allotted Time: 1 hour

Exercise Components and Structure:

- 1. This exercise will take 60 minutes to complete.
- 2. Form yourselves into 5 groups of roughly equal size.
- 3. Review the below local area disease surveillance report.
- 4. Conduct calculations from report data.
- 5. Conduct calculations from animal-place-time data.
- 6. Develop definitions from data provided in the local area disease surveillance report.

Materials, Data or Information:

- 1. Computer
- 2. Microsoft Word
- 3. Microsoft Excel
- 4. Pen or Pencil

Expected Outputs and Deliverables of Each Participant:

1. Surveillance report.

Local Area Disease Surveillance Report

1. Monthly local area reporting summary (Table 1):

Monthly Data and Metrics

| Sublocal area Name | No. Monthly Reports Received in 2020 | No. Days from Laboratory Submission to Local area Reporting | No. Days from Local area Notification to Farmer Reporting (Days) | No. of Surveillance Events this Month |
|-------------------------------|---|---|--|--|
| Sublocal area A | 5 | 2 | 1 | 1 |
| Sublocal area B | 1 | 1 | | |
| Sublocal area C | 4 | 3 | 5 | 5 |
| Sublocal area D | 5 | | 1 | |
| Sublocal area E | 3 | 2 | | |
| Sublocal area F | 5 | 2 | 1 | |
| Sublocal area G | 4 | 2 | 1 | |
| Sublocal area H | 0 | 0 | | 5 |
| % Reports Received to Date | | Mode: | Mode: | |

A. With the data provided from the Monthly Reporting Summary (Table 1):

- 1. Calculate the percentage of Reports received to date in 2020.
- 2. Calculate the mode for the No. of days from laboratory submission to local area reporting.
- 3. Calculate the mode for the No. of days from local area notification to farmer reporting (days).
- 4. Calculate the total No. of surveillance events this month in the local area.

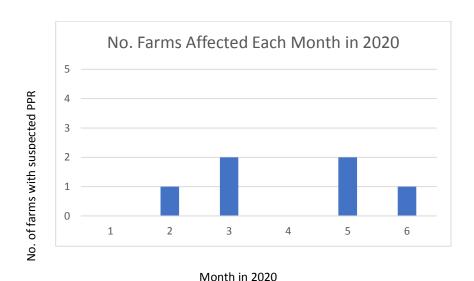
2. Active (A) and Passive (P) surveillance data by animal-place-time (Table 2):

| | 2. AC | <i>1110</i> (11 | | IMAL | e (F) surveilland | oc aata sy | Active | lace till | | ME | | PLACE | |
|-----------------------|-----------------|----------------------------|-------------|-------------|---|---|--------------------------|---------------------|---------------------------------------|------------------------------|-------------------------------------|-----------------------|--------------------|
| Species or Type | Class | Total No. at Risk | No. Sick | No. Dead | Disease(s) Suspected (S) or Confirmed (C) | Notes: (Age groups, breed and sex, etc.) | (A) or Passive (P) | Date of Visit | Date Symptoms First Observed | Date of first death | Date of Laboratory Submission | Sublocal area Name | GPS Coordinates |
| Cattle | Dairy | | | | | · | | | | | | | |
| | Beef | 45 | 2 | 0 | Trypanosomiasis (S) | Mature cows | P | June 10 | June 9 | NA | June 10 | С | |
| | Dual purpose | 82 | 4 | 2 | Trypanosomiasis (S) | Mature cows | A | June 6 | June 5 | June 5 | None | С | |
| | Breeder | | | | , , | | | | | | | | |
| Sheep | Meat | | | | | | | | | | | | |
| | Milk | | | | | | | | | | | | |
| | Dual | | | | | | | | | | | | |
| | purpose | | | | | | | | | | | | |
| | Breeder | | | | | | | | | | | | |
| Goats | Meat | | | | | | | | | | | | |
| | Milk | | | | | | | | | | | | |
| | Dual purpose | | | | | | | | | | | | |
| | Breeder | 150 | 10 | 5 | PPR (S) | 5 dead kids | P | June 11 | June 5 | June 5 | None | Н | |
| Poultry | Meat | | | | | | | | | | | | |
| | Eggs | | | | | | | | | | | | |
| | Dual Purpose | 5,000 | 0 | 2 | Marek's Disease (S) | 50-week old layers (daily mortality) | P | June 5 | Not applicable (NA) | June 5 | None | В | |
| | Breeder | | | | | | | | | | | | |
| Equine | Horse | | | | | | | | | | | | |
| | Donkey | | | | | | | | | | | | |
| | Mule | | | | | | | | | | | | |
| | Other | | | | | | | | | | | | |
| Pets | Dog | | | | | | | | | | | | |

| | ANIMAL | | | | | | Active | | TI | ME | | PL | ACE |
|-----------------------|---------|----------------------------|-------------|-------------|---|--|--------------------------|---------------------|---------------------------------------|------------------------------|-------------------------------------|-----------------------|--------------------|
| Species or Type | Class | Total No. at Risk | No. Sick | No. Dead | Disease(s) Suspected (S) or Confirmed (C) | Notes: (Age groups, breed and sex, etc.) | (A) or Passive (P) | Date of Visit | Date Symptoms First Observed | Date of first death | Date of Laboratory Submission | Sublocal area Name | GPS Coordinates |
| | Cat | 1 | 1 | 0 | Rabies (S) | 1 feral dog | P | June19 | June 18 | NA | June 19 | Е | |
| | Other | | | | | | | | | | | | |
| Wildlife | Specify | | | | | | | | | | | | |

B. Review Table 2. With the surveillance data provided data by animal-place-time (Table 2):

- 1. Calculate the percent (%) mortality for:
 - a. Suspected Trypanosomiasis reports in Beef in June 2020.
 - b. Suspected PPR in Goats in June 2020.
- 2. Interpret the following graph of PPR in the local area for Goats so far in 2020. What pattern do you observe and what does it mean for the occurrence of PPR in the local area?



3. Definitions (Table 3):

| Disease | Species | Epidemiological Unit | Case Definition | Diagnostic Testing |
|---------|---------|-------------------------|--------------------|-----------------------|
| | | | | |
| | | | | |
| | | | | |
| | | | | |
| | | | | |

| Disease | Species | Epidemiological Unit | Case Definition | Diagnostic Testing |
|---------|---------|-------------------------|--------------------|-----------------------|
| | | | | |
| | | | | |
| | | | | |
| | | | | |

C. Review the definitions table (Table 3).

Try to gradually complete the definitions table with the assistance of a veterinary epidemiologist from your national animal health agency (Ministry of Agriculture).

Lesson 9 – Making Recommendations for Animal Disease Prevention and Control

| Estimated Lesson and Exercise Time | 2 hours |
|------------------------------------|--|
| Participant Materials | Frontline ISAVET Lesson 9 Making Recommendations for Animal Disease Prevention and Control Participant Guide.PDF MS Excel Exercise 15b |
| | Computer Microsoft Word and Excel |
| | Pen or pencil |

In Service Applied Veterinary Epidemiology Training (ISAVET)

Lesson 9: Making Recommendations for Animal Disease Prevention and Control





Learning Objectives

At the end of this lesson, you will be able to:

- Describe and apply the four main aspects to conducting a data quality audit.
- 2. Make recommendations from a monthly surveillance reporting form used for animal disease surveillance related to:
 - The quality of the data;
 - The meaning of the data; and
 - The initial assessment of the surveillance system.

Assessment of Surveillance Data Using the Four Components of a Data Quality Audit 1) Data collection 2) Data analysis 3) Data use 4) Laboratory data Reference: Chen et al., Int. J. Environ. Res. Public Health 2014, 11, 5170-5207; doi:10.3390/ijerph1105051703

1. Data Collection

| UDIT | | ATTRIBUTE | MEASURE | OUTPUT |
|--------|---|---------------------------|--|--------|
| | 1 | Collector | Name of Surveillance Focal Points: Indicate whether training | |
| | | | has been provided on data collection | |
| D | 2 | Collection Method | Specify the method of data collection from the farm, village or | |
| _ | | | facility (e.g. abattoir) to the district level including: 1) Field | |
| A T | | | observation; 2) Interview; 3) Survey (strcutured, unstructured); | |
| A | | | and 4) Audit of existing field data Attach forms if possible | |
| A | 3 | Priority Diseases | List the priority diseases under surveillance | |
| С | 4 | Completeness | Completeness: the percentage of blank or unknown data, not | |
| ō | | | zero/missing; or proportion of filling in all data elements in the | |
| ĭ | | | facility report form. ALL DATA SHOULD BE IN A | |
| î. | | | DISAGGREGATED FORM TO PERMIT FURTHER ANALYSIS | |
| Ē | 5 | Timeliness | Timeliness: the percentage of reports from the subdistricts, | |
| c | | | abattoirs and facilities that were received on time | |
| Ť | 6 | Accuracy | Accuracy: the percentage of data variables on the collection | |
| 1 | | | form without an error: EXAMPLES - missing data, incorrect | |
| ò | | | coding, transposed error, incorrect units, incorrect/inconsistent | |
| N | | | format | |
| | 7 | Action | What actions are taken to correct late, absent, or incomplete | |
| | | | reporting from the reporting sites? | |
| | 8 | Data Storage and Security | How is the data stored and maintained and backed up? | |
| | | | | |

4

Data Collection Tools

- Plan and develop the collection tool with stakeholders – sublocal area and national
- Pre-test the data collection tool
 - Leads to good quality data
- Leads to reliable epidemiological results for decision-making and action
- The data can be a primary and secondary source



What is Primary and Secondary Data?

- \bullet It is data you collect yourself.
- ${\mbox{\footnotemath{\bullet}}}$ You understand how the data was collected.
- · You input the data yourself.
- · You understand the limitations of the collection tool and of the data.

- It is data that already exists from a field office or from a laboratory.
- ${\boldsymbol \cdot}$ You do not fully understand how the data was collected (you were not part of the collection).
- · You did not input the data yourself.
- · You do not fully understand the limitations of the collection tool and of

6

Integrity and Quality of the Surveillance Data

Data Quality

Missing data

Transcription errors

Misplaced data

Formatting errors

Coding errors

Copy errors

Omission of animal-place-time

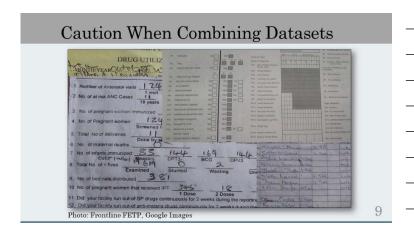
Not providing disaggregated data

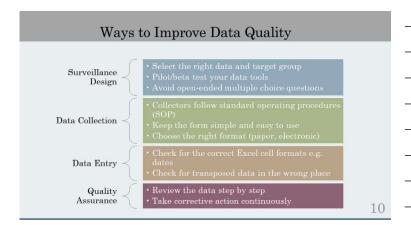
What Do You Think of this Dataset?

| | | Brucello | sis Testir | g for Janua | ry 2018 | |
|---------|------------------|----------|------------|-------------|-------------------------------|-----------------------------|
| Herd ID | Date of Onset | Age | Sex | Abortion | Brucellosis Screening Test | Brucellosis Confirmation |
| 796 | 5-Jan-18 | 1 | Female | No | Neg | Pos |
| 797 | 5-Jan-18 | 4 | Male | | Neg | Pos |
| 799 | 5-Jan-18 | 2 | Male | | Neg | Pos |
| 800 | 5-Jan-18 | 5 | Male | | Neg | Pos |
| 801 | 5-Jan-18 | 4 | Female | Yes | X | Pos |
| 802 | 5-Jan-18 | (33) | Female | No. | Neg | Neg |
| 803 | 5-Jan-18 | 2 | Male | Yes | | |
| 804 | 5-Jan-18 | 6 | Female | Yes | Neg | Pos |
| 805 | 5-Jan-18 | 3 | Female | Yes | Neg | Pos |
| 806 | 5-Jan-18 | - 1 | Male | 100 | Neg | Neg |
| 807 | 5-Jan-18 | 2 | Male | 111 | Neg | Nog |
| (8008) | 5-Jan-18 | 3 | Female | No | Z | Pos |
| 809 | 5-Jan-18 | 1 | Female | No | Neg | Neg |
| 810 | 5-Jan-18 | 4 | Female | Yes | Neg | Pos |
| 811 | 5-Jan-18 | 2 | Male | *** | Neg | Neg |
| 812 | 5-Jan-18 | . 5 | Male | | Neg | Neg |
| 813 | 5-Jan-18 | 3 | | Yes | Pos | Pos |
| 814 | 5-Jan-18 | 3 | Female | No | Neg | Neg |
| 815 | 5-Jan-18 | 1 | Male | 4441 | Neg | Neg |
| 816 | ő-Feb-18 | 1 | Male | 140 | Neg | Neg |
| 817 | 5-Jan-18 | (0.5) | Female | Yes | Pos | Pos |
| 818 | 5-Jan-18 | 4 | Male | 200 | Neg | Neg |
| 818 | 5-Jan-18 | 2 | Female | Yes | Pos | Pos |

Identify the data quality issues with this dataset

Reference: Heather Simmons





2. Data Analysis MEASURE AUDIT ATTRIBUTE OUTPUT A 9 Data Tools Describe the <u>tools used for analyzing data</u> at: 1) the farm or village to the district level D A 10 Software Describe the computer software used A L T Y A S Calculate percentage (%) Calculate percentage (%) and create groupings/categories Analyse disaggregated data according to animal, place and time Quantitative Data 13 Animal-Place-Time Tables, graphs, maps, flow diagrams, SWOT table, Fishbone

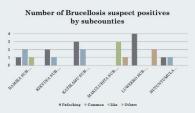
2. Data Analysis and Interpretation

- 1. Analyse and display your data
 - · Calculate measures of central tendency and disease occurrence that illustrate the impact of disease
 - \cdot Create graphs or maps to illustrate the meaning of the data
- 2. Describe each analysis and compile a list of the main findings
- Make inferences only from data that is of sufficient quality to analyse
- 4. Make recommendations based on the surveillance data analysis

12

Example: Brucellosis sub-counties at risk

- · All sub counties sampled had at least 1 farm Brucellosis positive
- · MEANING: % brucellosis positivity is higher among paddocked than communal farms



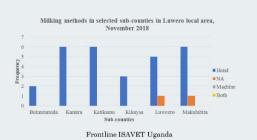
Frontline ISAVET Uganda

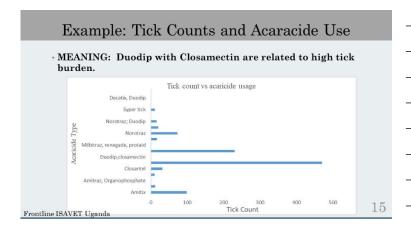
13

Example: Milking method

Milking practice is one of the predisposing factors for brucellosis

 MEANING: In most of the subcounties, hand milking is the most popular practice that farmers use





| AUDIT | Τ | ATTRIBUTE | MEASURE | OUTPUT |
|---------------------------------|----|--------------------|---|--------|
| D A T A U S E | 15 | Data Sharing | Describe the frequency and kind of reports used to share data from one level to the next: 1) the farm or village to the district level; 2) the district to the subnational level; from the province to the number of the state of the subnational level. Describe, or attach if possible. | |
| | 16 | Data for Action | Calculate how many surveuillance data reports led to conducting field investigations, initiated further training at the subdistrict level, were shared with other agencies such as public health and wildlife health | |
| | 17 | Data for Planning | Describe the <u>frequency</u> and how analysed data contributed to <u>any planning reports</u> developed by the district, subnational or national offices related to animal health, public health or wildlife health. Describe, or <u>attach if possible</u> . | |
| | 18 | Data for Research | Describe the <u>frequency and how analysed data contributed to</u> <u>any research reports</u> developed by the district, subnational or national offices related to animal health, public health or wildlife health. Describe, or attach if possible . | |
| | 19 | Feedback Mechanism | Describe the data feedback mechnism among subdistrict/local, | |

| How will the Data be Used? | | | | |
|---------------------------------------|---|--|--|--|
| USE | EXAMPLES | | | |
| DATA SHARING | - District reports - National laboratory reports | | | |
| DATA FOR ACTION | - A threshold is exceeded that prompts and outbreak investigation | | | |
| DATA FOR PLANNING | - Burden of disease estimates serve as a basis for disease prioritisation | | | |
| DATA FOR RESEARCH | $\hbox{-}\ Validation of laboratory tests}$ | | | |
| FEEDBACK MECHANISM | - Provide a PowerPoint presentation for farmers on a priority disease in the district | | | |
| AWARENESS OF DATA USE BY STAKEHOLDERS | - Maintain confidentiality by aggregating data results into groups rather than individuals | | | |

| T + T + T + T + T + T + T + T + T + T + | | + |
|--|-------------|----|
| Table 3: Summary of livestock and wildlife anthrax occurrence trend at RVIL. Nakuru, 2013 to 20 YM Jan Feb Mar Apr May Jun Jul Aug Sep Oct Nov T + T + T + T + T + T + T + T + T + T | W27 HA U 10 | + |
| Table 3: Summary of livestock and wildlife anthrax occurrence trend at RVIL. Nakuru, 2013 to 20 YM Jan Feb Mar Apr May Jun Jul Aug Sep Oct Nov T + T + T + T + T + T + T + T + T + T | W27 HA U 10 | + |
| T + T + T + T + T + T + T + T + T + T + | Dec T | + |
| | | |
| | T + | |
| | 0 0 7 | 0 |
| 0 0 5 5 11 4 7 2 9 4 0 0 6 2 0 0 7 0 0 0 3 0 | 0 0 48 | 1 |
| 2015 0 0 5 0 1 0 1 0 0 0 2 2 39 34 1 0 2 0 0 0 1 1 | 2 0 54 | 31 |
| 2016 0 0 0 0 1 0 2 0 7 3 7 5 5 3 2 0 0 0 2 0 0 0 | 0 0 26 | 1 |
| 2017 6 2 3 0 2 0 0 0 0 0 1 1 1 1 3 1 2 0 4 2 0 0 | 0 0 22 | 7 |
| 2018 0 0 0 0 3 0 0 0 4 2 0 0 1 0 0 0 0 4 2 0 0 | 0 0 12 | 4 |

| Making Recommendations from Data Quality | Audits |
|--|----------|
| • There are three types of recommendations arising from and analysis of surveillance data. | a review |
| 1 Integrity and quality of the data | |
| 2 Meaning of the surveillance data | |
| Assessment of the surveillance system | |
| ~ | 20 |

Methods to Improve the Surveillance Data Quality We need to measure certain "metrics" or measures in order to assess the performance of a surveillance system We can assess the data flow in terms of timeliness and completeness of the surveillance reports received from the sublocal area level. Recommended ways to improve the surveillance system include the following: - Holding regular meetings with stakeholders and staff Staff Training Reviewing the surveillance format and protocols - Asking for technical assistance from the national level 21 In Summary... • There are four main aspects to conducting a data quality audit: 1) Data collection; 2) Data analysis; 3) Data use; and 4) Laboratory Data ${\raisebox{0.5ex}{$\scriptscriptstyle\bullet$}}$ The quality of data affects the entire surveillance system · Many types of errors can affect data quality · You can put measures in place to ensure quality data collection, merging and storage · Feedback works in order to improve: · The quality of the data: · The meaning of the data; and · The assessment of the surveillance system. 22 Exercise 15b: Local Area Disease Surveillance Report This exercise will take 75 minutes. Part I: Work in groups of 3 to 4 individuals. Review the data provided, answer the guiding questions Draw conclusions and recommendations for further action related to: - The quality of the data - The meaning of the data - The assessment of the surveillance system PART II: Work alone or in pairs. Based on your knowledge of the surveillance system in your district, do an initial assessment the quality of data in your district using the following Data Quality Audit Tool:

| ISAVE | T Contributing Universities | | |
|--------------|-----------------------------|----|--|
| Partners | TEXAS ASM. GRILIFE | | |
| Contributors | MAKERIKE UNIVERSITY | 24 | |

Exercise 15b - Local Area Disease Surveillance Report

Description of Exercise:

Review a local area disease surveillance report and draw conclusions and recommendations for further action related to the quality of the data, the meaning of the data, and assessment of the surveillance system. Should you have any questions over the exercise, please ask a trainer for clarification before during, and after the exercise.

Allotted Time: 1.25 hours

Exercise Components and Structure:

- 1. You have 1 hour and 15 minutes to complete this exercise
- 2. Work in groups of 3-4 individuals and answer the questions in the exercise.
- 3. Review the exercise and work in groups to assess the quality of the local area surveillance report.

Materials, Data or Information:

- 1. Microsoft word
- 2. Pen and paper

Expected Outputs and Deliverables of Each Participant:

1. Applying critical thinking for making conclusions and recommendations.

Local Area Disease Surveillance Report

4. Monthly local area reporting summary (Table 1):

Annual Data and Metrics

December 31, 2020

| Sublocal area Name | No. Monthly Reports Received in 2020 | No. Days from Laboratory Submission to Local area Reporting | No. Days from Local area Notification to Farmer Reporting (Days) | No. of Surveillance Events this Year |
|----------------------------|---|---|--|---|
| Sublocal area A | 11 | 2 | 1 | 12 |
| Sublocal area B | 1 | | | 1 |
| Sublocal area C | 8 | 3 | 5 | 2 |
| Sublocal area D | 12 | 2 | 3 | 5 |
| Sublocal area E | 7 | 2 | 3 | 3 |
| Sublocal area F | 12 | 2 | 0 | 15 |
| Sublocal area G | | | | 2 |
| Sublocal area H | 0 | Not applicable | | 12 |
| % Reports Received to Date | | Mode: Mean: | Mode: | Total = |

- 1. Make the calculations on the bottom row of the table. Assess the surveillance system in this local area with regards to:
 - a) Timeliness
 - b) % Reports Received to Date
 - c) Give your assessment of surveillance in each sublocal area listed below.

Sublocal area A -

Sublocal area B -

| Sublocal area C – | | |
|-------------------|--|--|
| Sublocal area D – | | |
| Sublocal area E – | | |
| Sublocal area F – | | |
| Sublocal area G – | | |
| Sublocal area H – | | |

| | ANIMAL | | | | | Active | | T | ME | | PLACE | | |
|-----------------------|-----------------|----------------------------|-------------|-------------|---|--|--------------------------|------------------|---------------------------------------|---------------------------|-------------------------------------|-----------------------|----------------------------------|
| Species or Type | Class | Total No. at Risk | No. Sick | No. Dead | Disease(s) Suspected (S) or Confirmed (C) | Notes: (Age groups, breed and sex, etc.) | (A) or Passive (P) | Date of Visit | Date Symptoms First Observed | Date of first death | Date of Laboratory Submission | Sublocal area Name | GPS Coordinates Lat./Long. |
| Cattle | Dairy | 1251 822 | 3 5 | 2 | Trypanosomias (S) RVF (C) | Mature cow Mature | P P | 12/2/20 | 11/29/20 12/20/20 | 11/3/20 12/20/20 | Not applicable (NA) | A | |
| | Beef | 744 | | 10 | Anthrax (C) | Mature Females | Р | 12/12/20 | 12/7/20 | 12/7/20 | 12/12/20 | A | |
| | Dual purpose | | | | | | | | | | | | |
| | Breeder | 5 | 0 | 1 | Brucellosis (S) | Bull | S | 1212/20 | 11/24/20 | 12/12/20 | NA | H | |
| Sheep | Meat Milk | | | | | | | | | | | | |
| | Dual purpose | 230 | 1 | 1 | PPR (C) | Lamb on pasture | Р | 12/3/20 | 12/3/20 | 12/3/20 | NA | D | |
| | Breeder | | | | | 1 | | | | | | | |
| Goats | Meat | | | | | | | | | | | | |
| 1 | Milk | | | | | | | | | | | | |
| | Dual purpose | 451 | 3 | 2 | PPR (S) | Kids on tether | Р | 12/17/20 | 12/15/20 | 12/17/20 | NA | Е | |
| | Breeder | | | | | | | | | | | | |
| Poultry | Meat | 4525 | 62 | 45 | Newcastle disease (S) | 28 day broilers | A | 12/5/20 | 12/5/20 | 12/5/20 | 12/5/20 | F | |
| | Eggs | | | | | | | | | | | | |
| | Dual | | | | | | | | | | | | |
| | Purpose | | | | | | | | | | | | |
| | Breeder | | | | | _ | | | | _ | | | |
| Equine | Horse | | | | | | | | | | | | |
| | Donkey | | | | | | | | | | | | |
| | Mule | | | | | | | | | | | | |

| | ANIMAL | | | | Active | | TI | ME | | PL | ACE | | |
|-----------------------|---------|----------------------------|-------------|-------------|---|--|--------------------------|------------------|---------------------------------------|---------------------------|-------------------------------------|-----------------------|----------------------------------|
| Species or Type | Class | Total No. at Risk | No. Sick | No. Dead | Disease(s) Suspected (S) or Confirmed (C) | Notes: (Age groups, breed and sex, etc.) | (A) or Passive (P) | Date of Visit | Date Symptoms First Observed | Date of first death | Date of Laboratory Submission | Sublocal area Name | GPS Coordinates Lat./Long. |
| | Other | | | | | | | | | | | | |
| Pets | Dog | 23 | 0 | 1 | Rabies (S) | 4-month puppy | P | 12/31/20 | 12/25/20 | 12/31/20 | 12/31/20 | F | |
| | Cat | | | | | | | | | | | | |
| | Other | | | | | | | | | | | | |
| Wildlife | Specify | | | | | | | | | | | | |

| 2. Assess the data quality for each disease line item in the December local area report. |
|--|
| Sublocal area A – |
| |
| Sublocal area B – |
| |
| Sublocal area C – |
| |
| Sublocal area D – |
| |
| Sublocal area E – |
| |
| Sublocal area F – |
| |
| Sublocal area G – |
| |
| Sublocal area H – |
| |

| 3. | What is the meaning of the data following your analysis? |
|----|--|
| 4. | What is the disease with the highest impact in this local area? Please explain your answer. |
| 5. | What type of surveillance is most commonly used in the local area? Why is this important to record and what does it say about the ability to have early disease detection? |
| 6. | What do you recommend for improving the surveillance in this local area? |

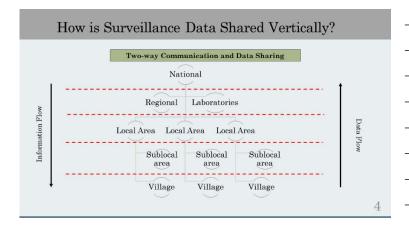
Lesson 10 – Sharing Surveillance Information in a Network for Animal Disease Prevention and Control

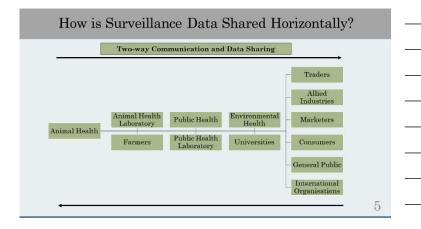
| Estimated Lesson and Exercise Time | 2 hours and 30 minutes |
|------------------------------------|---|
| Participant Materials | Frontline ISAVET Lesson 10 Sharing Surveillance Information in a Network for Animal Disease Prevention and Control Participant Guide.PDF |
| | Flip chart Markers |

In Service Applied Veterinary Epidemiology Training (ISAVET) Lesson 10: Sharing Surveillance Information in a Network for Animal Disease Prevention and Control

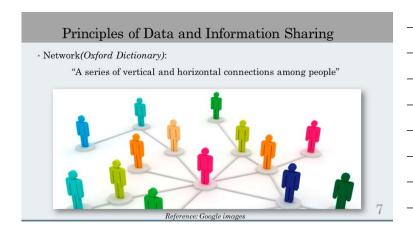
Learning Objectives At the end of this lesson, you will be able to: 1. Describe the stakeholders of surveillance information; and 2. Describe the principles of sharing animal health surveillance data and information. 3. Describe One Health approaches needed for sharing surveillance information.

Why is it Important to Share Data and Information from Surveillance Systems? To use a network of people involved in animal health, food inspection and extension education to continuously improve surveillance of animal health at the local area level To interpret animal health data and contribute to appropriate follow-up actions to improve livestock animal health To link, summarize, interpret and act upon data that connects animal health, food safety and public health using a broad network of partners To raise awareness and reduce the risk of threats to animal health from beyond the local area's borders To support animal health surveillance of the national government, to maintain and demonstrate the health of national livestock 3





Who are the Key National Animal Health Stakeholders? 1. Ministry of Agriculture and Ministry in charge of Wildlife animals 2. Animal owners 3. Independent farmers 4. Farmer cooperatives 5. Livestock private enterprises 6. Allied industries – feed, drugs, waste removal 7. Trade groups 8. Universities and research organizations 9. Public health and wildlife partners e.g. Tour companies, Fisheries 10. General Public



| Why is it | Important to Share Data and Information from Surveillance Systems? |
|--|---|
| Who should the information be shared with? | Those from the public and private sector who have a stake in improving animal health $$ |
| Why is it important to share information? | Cannot act if unaware of the problems or if data not available to support efforts |
| information: | With knowledge comes responsibility to take action |
| How can the | Real-time alerts during an emergency |
| information be | Surveillance summaries / reports |
| disseminated? | Bulletins and newsletters |
| | Press releases |
| | Veterinary / epidemiologic journal articles |
| | Meetings |

Principles of Data and Information Sharing

- · The animal health local area office plays an critical and necessary role, namely:
- Gathering disaggregated raw data from the sublocal area level; Compiling data; and
- Sharing data from the local area to the regional and national levels.
- · What is the incentive to collect and compile data?

Key Principle

If data is not improved in quality so that it can be analysed and interpreted, the incentive to collect the data is reduced.

9

Principles of Data and Information Sharing

- The animal health local area office plays an important role:
 - By making meaning and recommendations to sublocal area and national
- · What is the incentive to make recommendations?
- Improve capacity for detection and response
- Enables national and international collaboration, capacity strengthening, insight into public health system performance, and ultimately better control of infectious diseases

Key Principle

Evidence-based decision-making depends on making sound recommendations based on sound data.

10

Basic Building Blocks of Network Data Sharing Build Trust Explain the Value and Benefits Plan for Data Sharing Improve Data Quality High-quality data enables the generation of high-quality evidence leading to better animal health outcomes Understand the Legal Context Establish Data Sharing Agreements

Principles of Data and Information Sharing

- There are two additional things you can do right now to improve the usefulness of surveillance data in your local area:
 - 1. Evaluate the timeliness of reporting; and
- 2. Evaluate the completeness of data you receive.

Provide Feedback to and from Other Levels.

12

Principles of Data and Information Sharing

- Results must be shared with people and officials who are in a position to take action to improve animal health
 - Optimal sharing requires an understanding of the roles and responsibilities of these stakeholders
- TWO WAY FLOW: Feedback <u>must always</u> be shared with those who provide the data!
- · Determine the best method to communicate results
 - Official communication channels
- · Determine when and how often results will be shared
 - During peace time
- In an emergency



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Take Action

- The purpose of a surveillance system is to create evidence for action
- Optimal sharing requires an understanding of the roles and responsibilities of these stakeholders
 - Do you understand your local area stakeholders?

Key Question

How could you change the way you use surveillance data so that it leads to action?

Scenario · You are waiting for a laboratory report to arrive in order to share results with a farmer. The farmer calls you with news that the situation is getting worse. What action will you take? 15 Reference: Google images

Exercise 16: Sharing Surveillance Information

- Form four groups
- $\dot{}$ This exercise will take 60 minutes followed by 30 minutes of plenary discussion
- · Country-based group work using MS Word, MS PowerPoint or flip charts
- - secribe:

 The key stakeholders in your country/local area for animal health surveillance data including public health and wildlife health partners. The existing information / data sharing mechanisms a) focus on data quality, interpretation of surveillance data and attributes of an efficient surveillance system

 How you will improve information sharing to support fact-based decisions by conducting a gap analysis to identify challenges that hinder efficiency of the existing surveillance system suggest solutions to the identified gaps that are within local area's reach.

16

In Summary...

- · Data sharing is guided by a set of a well defined principles.
- · Sharing surveillance data to appropriate stakeholders improves the effectiveness of animal health interventions and response, trust and transparency
- · If data is not improved in quality so that it can be analysed, the incentive to collect the data is reduced.
- · Provide feedback to other levels and seek feedback from other levels.

| ISAVE | T Contributing Universities | | |
|--------------|-------------------------------|----|--|
| Partners | TEXAS ASM TEXAS ASM TEXAS ASM | | |
| | TEXAS ARM CRILIFE | | |
| | XXX | | |
| | M. DE Sets | | |
| Contributors | | | |
| | MAKERER UNIVERSITY | 18 | |
| | | 10 | |

Exercise 16 – Sharing Surveillance Information in a Network for Animal Disease Prevention and Control

Description of Exercise:

Participants will describe the key stakeholder in your country/local area for animal health surveillance data while focusing on data quality, meaning of the surveillance data and discussion of attributes on an efficient surveillance system. Should you have any questions over the exercise, please ask a trainer for clarification.

Allotted Time: 60 minutes + 30 minutes for plenary

Exercise Components and Structure:

• Form into four groups will

Materials, Data or Information:

- 1. MS Word, MS PowerPoint or Flip charts
- 2. Markers

Expected Outputs and Deliverables of Each Participant:

- 1. List the key stakeholders who need animal surveillance information in your country and show how surveillance data is shared.
- 2. Provide suggestions on how to improve sharing surveillance information in your country under a One Health approach.

| 1. | Describe the key stakeholders in your country for animal health surveillance data |
|----|---|
| | including public health and wildlife health partners. |

2. Describe the existing information / data sharing mechanisms in your country.

3. Describe how you will improve information sharing to support fact-based decisions by conducting a gap analysis to identify challenges that hinder efficiency of the existing surveillance system - suggest solutions to the identified gaps that are within local area's reach.

Lesson 11 – Assessing Surveillance in Your Local Area to Improve Response to Animal Disease and Health Events

| Estimated Lesson and Exercise Time | 1 hour and 30 minutes |
|------------------------------------|---|
| Participant Materials | ISAVET Lesson 11 Assessing Surveillance in Your Local Area to Improve Response to Animal Disease Events Participant Guide.doc |
| | Computer Microsoft Word |

In Service Applied Veterinary Epidemiology Training (ISAVET)

Lesson 11: Assessing Surveillance in Your Local Area to Improve Response to Animal Disease and Health Events

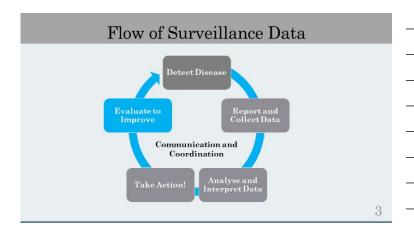
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|----|------------|--|--|
| | B. BIZZERN | | |



Learning Objectives

At the end of this lesson, you will be able to:

- 1. Use surveillance data to improve timeliness of outbreak response based on monitoring and evaluation; and
- $_{\rm 2.}$ Explain the attributes of surveillance system evaluation.



| 1 | Monitoring |
|---|---|
| | and continuous tracking of planned |
| | and continuous tracking of planned nce activities Evaluation |

| | and Targets ontline FETP) |
|---|--|
| Indicator | Target |
| | |
| Statement to measure achievement of an activity objective | Desired level of achievement |
| Example: Is reporting done on time? | Example: 80% of monthly reports have been sent on time to national level |
| | 5 |

What Are Some Surveillance Indicators For Your Local Area in These Areas? 6 What Are Some Surveillance Indicators in Your Local Area? Median number of Percentage of Time to respond to Time from sample monthly reports active surveillance farmer request for collection to received from samples collected per field investigation laboratory reporting sublocal area offices month Number of secondary Time from laboratory Percentage of local Percentage of field investigations generated from a area field report received by complete disease data investigations with a from each sublocal local area until primary field complete report farmer notification investigation What Are Some Surveillance Indicators in Your Local Area? Local Area Response Target Percentage of monthly reports received from sublocal area offices Median number Time from Time to respond to farmer request for field investigation 4 hours of active sample collection to laboratory surveillance samples collected per 48 hours 80% 200 reporting month Time from Percentage of secondary field investigations generated from Percentage of Local Area field investigations with a complete laboratory report received by Percentage of complete disease data from each 4 hours

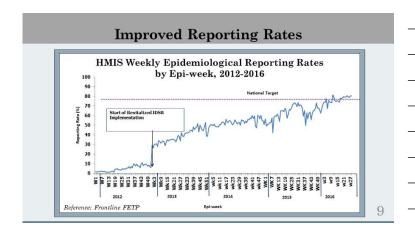
a primary field investigation

Local Area until farmer

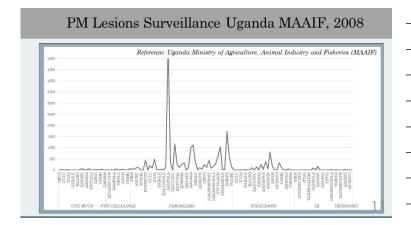
notification

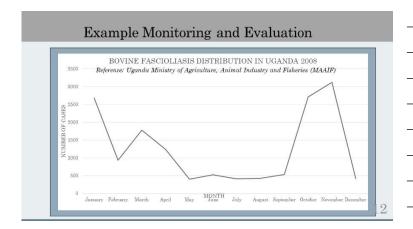
report

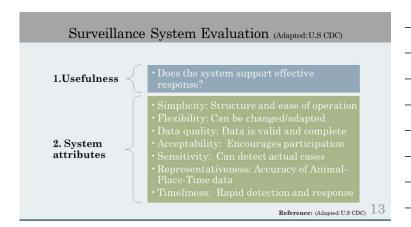
sublocal area











| Examples: Timeliness, Completeness, Data Quality | | | |
|---|--|---|---|
| Attribute | Performance Indicator | Target | Finding |
| Timeliness | 4 out of 8 (50%) sublocal areas have reported on time this week | 80% on time | Deficient in timely reporting for half of the sublocal areas |
| Completeness | Last week, 7 out of 8 sublocal area reports provided incomplete information | 90% complete reports | 88% of Local Areas provided a report generally meeting the target |
| Data Quality | The % of reports with blank data for at least one variable was 80% this week | Maximum 10% of reports with blank data for at least one variable | The data quality is short of the target by 70% |

Exercise 17: Assessing Surveillance Information in a Network for Animal Disease Prevention and Control

- 1. This exercise will take 60 minutes.
- 2. This is a small group discussion and sharing in plenary discussion.
- Divide into three groups by surveillance purpose (Group A. Reduce disease burden; Group B. Freedom from disease; and Group C. detection of disease).
- 4. Use flip charts to address the items below.
 - List the ways you can improve surveillance in your Local Area and develop 5 key indicators, including targets for each.
 - Describe how you can use surveillance data to improve outbreak detection and response in your Local Area.
- 6. Have one individual from each group present in plenary session.

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In Summary...

- Monitoring is essential to maintain the function and quality of the surveillance system;
- Develop indicators to monitoring performance of reporting sites for timeliness and completeness;
- 3. Surveillance system evaluation can be used to develop strategies for improvement; and
- 4. Improved performance of the surveillance system will lead to improvements in outbreak detection and response.

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Partners Contributors Contributors ISAVET Contributing Universities TEXAS A&M TEXAS A&M GRILIFE

Exercise 17 – Assessing Surveillance Information in a Network for Animal Disease Prevention and Control

Description of Exercise:

Participants will develop five indicators and targets for specific surveillance objectives. Should you have any questions over the exercise, please ask a trainer for clarification before during, and after the exercise.

Allotted Time: 60 minutes

Exercise Components and Structure:

- 1. You have 60 minutes to complete this exercise.
- 2. Divide into 3 groups by surveillance purpose.
 - Group A: Reduce disease burden
 - Group B: Freedom from disease
 - Group C: Detection of disease in an area previously not known to have a disease burden
- 3. Each group should develop 2 objectives for their surveillance system. They can choose the disease of their choice.
- 4. Use flip charts to:
 - List the ways you can improve surveillance in your local area and develop 5 key indicators, including targets for each.
 - Describe how you can use surveillance data to improve outbreak detection and response in your local area.

Materials, Data or Information:

- 1. Flip chart and markers
- 2. Pen and paper

Expected Outputs and Deliverables of Each Participant:

- 1. A list of the ways to improve surveillance in your local area.
- 2. Describe how you can use surveillance data to improve outbreak response in your local area.

1. List up to five ways you can improve surveillance in your local area.

| List up to five ways you can improve surveillance in your local area. | | | |
|---|-------------------------|--|--|
| Surveillance Purpose | Surveillance Objectives | | |
| Reducing Disease Burden | | | |
| Freedom from Disease | | | |

Detection of Disease (Area previously does not have a disease burden)

2. Identify 5 indicators with specific targets for your groups surveillance system.

| Group A: Surveillance Syst Indicator(s) | tem Purpose: Reducing Disease Burden Target(s) |
|---|---|
| Indicator 1: | Target 1: |
| Indicator 2: | Target 2: |
| Indicator 3: | Target 3: |
| Indicator 4: | Target 4: |
| Indicator 5: | Target 5: |

| Group | B: Surveillance System Purpose: F | |
|--|--|-----------------------------|
| | Indicator(s) | Target(s) |
| Indicator 1: | Targe | : 1: |
| Indicator 2: | Targe | 2: |
| Indicator 3: | Targe | 3: |
| Indicator 4: | Targe | z 4: |
| Indicator 5: | Targe | 5: |
| | | |
| Gro | un C. Surveillance System Purnose | Disease Detection |
| Gro | up C: Surveillance System Purpose: Indicator(s) | Disease Detection Target(s) |
| Ground Indicator 1: | up C: Surveillance System Purpose: Indicator(s) Targe | Target(s) |
| | Indicator(s) | Target(s) |
| Indicator 1: | Indicator(s) Targe | Target(s) 2: |
| Indicator 1: Indicator 2: | Indicator(s) Targe | Target(s) 2 1: 2 2: |
| Indicator 1: Indicator 2: Indicator 3: | Targe Targe Targe | Target(s) 21: 22: 33: 44: |

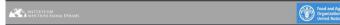
3. Describe how you can use surveillance data to improve outbreak detection and response in your local area

Section II: Week 2 – Field Investigation and Response

Lesson 12 - Animal Field Investigations

| Estimated Lesson and Exercise Time | 2 hours |
|------------------------------------|---|
| Participant Materials | ISAVET Lesson 12 Animal Field Investigations Participant Guide.doc |
| | Computer |
| | Microsoft Word |

In Service Applied Veterinary Epidemiology Training (ISAVET) Lesson 12: Animal Field Investigations



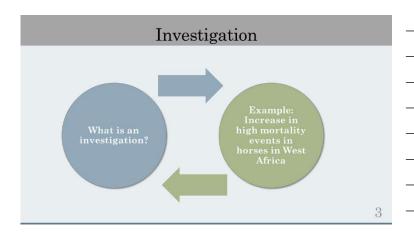
Learning Objectives

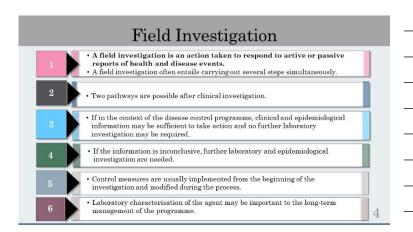
At the end of this lesson, you will be able to:

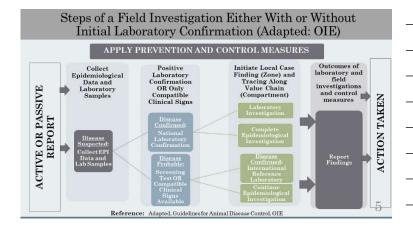
- 1. Define field investigation.
- 2. Understand the transition from a field investigation to an outbreak investigation.
- 3. Describe how veterinary field investigations can be utilised under the following settings:
 a) Animal production or disease events;
 b) Value chain events;

 - c) Zoonoses, food safety and public health events;
 d) Import and export events; and

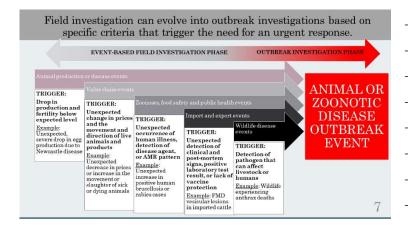
 - e) Wildlife disease events.

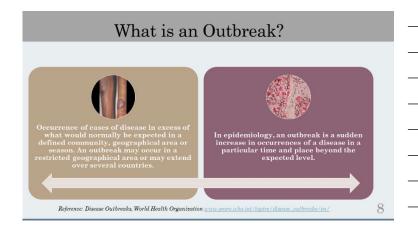


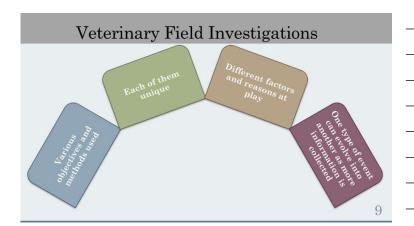




| Objectives and Methods Used for Various Types of Investigations | | | |
|--|---|--|---|
| | | | |
| Animal production | Determine reasons for drops in reproductive performance | Nutrition and disease and production history | Review historical surveillance records to assess trends Conduct a survey to estimate the burden of disease |
| Priority national endemic diseases | Determine the cause for increased morbidity and mortality | Conduct an investigation: measure the impact of disease; review records | Address methods of disease entry and transmission; disease |
| Priority national zoonotic diseases | Confirm and contain the disease from spreading | Apply biosafety and biosecurity practices; collect and submit samples | Sample collection using PPE; Test samples and respond by taking preventive action |
| Internationally reportable high-impact animal diseases | Investigate a potentially high-impact disease | Treat the investigation as a potential high- impact animal disease; Follow the standard steps for an animal disease outbreak investigation | Farm quarantine; Area movement controls; Aggressive case finding Detailed epidemiological investigation of index farm; Notification of farmers; Preparation for possible ring vaccination. |







Animal Production of Disease Event



Scenario 1

A dairy farmer calls you to investigate some recent late-term abortions and reduced fertility in his herd of 10 cows.

What is the objective of this investigation?

Reference: Google Images

10

Animal Production or Disease Event



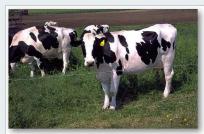
Scenario 1

A dairy farmer calls you to investigate some recent late-term abortions and reduced fertility in his herd of 10 cows.

What methods would you use to investigate the drop in production?

Reference: Google Images

Zoonoses, Food Safety and Public Health Event



Scenario 1

The laboratory has confirmed that the late-term abortions and reduced fertility in the herd is due to *Brucella abortus*.

Brucellosis in cattle is thought to be endemic in this local

area.
What actions would you take next to deal with this at the population level?

Reference: Google Image:

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Zoonoses, Food Safety and Public Health Event



Scenario 1

Brucellosis represents a zoonotic disease risk to the farmer's family and the local community.

In the case of zoonosis, the outbreak investigation should be done in coordination with the public health authorities.

What is the objective of a zoonotic disease field investigation?

Reference: Google Image

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Animal Production Event



Scenario 2

A small poultry broiler producer has experienced an increase in the amount of morbidity and mortality in his flock of 5,000 broilers. In the past, he has noticed some respiratory signs when chickens are 4-weeks old.

What is the objective of this investigation?

Reference: Google Images

Animal Production or Disease Event



Scenario 2

A small poultry broiler producer has experienced an increase in the amount of morbidity and mortality in his flock of 5,000 broilers. In the past, he has noticed some respiratory signs when chickens are 4-weeks old.

What methods would you use for this investigation?

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Animal Production or Disease Event



Scenario 2

Laboratory confirmation reveals that the flock is infected with mesogenic Newcastle disease virus. Avian influenza testing is in progress, but not yet complete. Newcastle disease is likely endemic in this local area.

What actions would you take next to deal with this at the population level?

Reference: Google Images

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Possible Zoonotic Disease Event Due to AI



Scenario 2 Newcastle disease is not a zoonotic disease risk to the farmer's family and the local community aside from a mild conjunctivitis.

However, laboratory confirmation has just been received for low pathogenic avian influenza (LPAI) subtype H7N3 was confirmed through egg inoculation and RT-PCR tests

What action should be taken?

Reference: Google Images

Scenario 3: Animal Production or Disease Event How would you go about investigating the sudden death of a cow reported by a farmer who had foot skin lesions? What is the objective of the field investigation? Reference: Situational Report to the National Task Force, Uganda on the status of Anthrax outbreaks in Arua, Kirihura and Kween districts, July 2018. 18 Scenario 3: Animal Production or Disease Event OR Zoonoses, Food Safety and Public Health Event - Zoonotic - High-Impact TAD What method of field investigation should be conducted? re: Situational Report to the National Task Force, Uganda on the status of Anthrax outbreaks in Arna, Kirihura o Scenario 3: Zoonoses, Food Safety and Public Health Event - Zoonotic List the methods for the investigation. port to the National Task Force, Uganda on the status of Anthrax outbreaks in Arua, Kirihura and

Scenario 3: Zoonoses, Food Safety and Public Health Event





Identify 3 field components to conduct during a zoonotic disease investigation?

- 1. Sample collection using PPE
- 2. Testing of samples
- 3. Respond by taking preventive action (community awareness, risk communication, treatment and vaccination)

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Situational Report to the National Task Force, Uganda on the status of attreaks in Arua, Kirihura and Kween districts, July 2018.

Scenario 3: Zoonotic Investigations Community Awareness is Done Based on a One Health Approach



Reference: Preliminary Report on Suspected Anthras Outbreak, Kween District, Ministry of Health, 26/07/2018 to the National Task Force: Prepared by: Esther Kisaakye and Kenneth Bainomugisha, Supervised by: Lilian Bulage. Report to the One health Platform, Ugant

Animal Disease Events: Internationally Reportable Animal Diseases



Internationally Reportable High-Impact Animal Diseases



Scenario 4

A beef farmer in your local area has 200 cross-bred cows currently nursing 3-month old calves. The farmer reports a very sudden increase of 20% mortality among the calves in the herd, as well as, some lameness and recumbence among cows.

He also reports seeing blisters and raw areas on the mucosal membranes of the mouth, as well as, interdigital lesions.

What is the objective of this investigation?

Reference: Google Images

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Internationally Reportable High-Impact Animal Diseases



Scenario 4

A beef farmer in your local area has 200 cross-bred cows currently nursing 3-month old calves. The farmer reports a very sudden increase of 20% mortality among the calves in the herd, as well as, some lameness and recumbence among cows.

He also reports seeing blisters and raw areas on the mucosal membranes of the mouth, as well as, interdigital lesions.

What are the methods you would use for this field investigation?

Reference: Google Images

25

Internationally Reportable High-Impact Animal Diseases



Scenario 4

The original field investigation objective and methods may remain.

Alternatively, the field investigation may turn into an outbreak investigation with different objectives and methods.

What action needs to be taken?

Reference: Google Images

Exercise 18: Animal Disease Investigations

- 1. This exercise will take 60 minutes.
- 2. Form four groups.
- Using the below examples, complete a table that describes the
 - objectives, methods and actions for each type of investigation.

 a) Animal production field investigation milk production loss due to
 - b) Endemic disease field investigation Marek's disease
 - c) Zoonotic disease field investigation Rabies
 - d) Internationally reportable high-impact field investigation $\ensuremath{\mathsf{PPR}}$
- 4. Each group will report their output.
- 5. Additional feedback will be provided.

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Summary...

- 1. A field investigation is an action taken to respond to active or passive reports of health and disease events.
- A field investigation can evolve into an outbreak investigation based on specific criteria that trigger the need for an urgent response.
- 3. There are five (5) types of veterinary field events:
 - a) Animal production or disease events;
 - b) Value chain events;
 - c) Zoonoses, food safety and public health events;
 - d) Import and export events; and
 - e) Wildlife disease events.

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ISAVET Contributing Universities TEXAS A&M **Partners** GRILIFE Contributors

Exercise 18 – Animal Field Investigations

Description of Exercise:

Determine the objectives methods and actions for four different field investigation types. Should you have any questions over the exercise, please ask a trainer for clarification before during, and after the exercise.

Allotted Time: 1 hour

Organisation of Group Work:

Divide into four groups of roughly equal size.

Exercise Objective(s):

1. Describe four (4) types of veterinary field investigations including, objectives and the methods and actions to be taken for each.

Exercise Components and Structure:

- 1. Complete a table that describes the objectives, methods and actions for each type of investigation.
 - a) Animal production field investigation Milk production loss due to bovine mastitis
 - b) Endemic disease field investigation Marek's disease
 - c) Zoonotic disease field investigation Rabies
 - d) Internationally reportable high-impact field investigation PPR
- 2. Each group will report their output and additional feedback will be provided.

Materials, Data or Information:

1. Word and PPT

Expected Outputs and Deliverables of Each Participant:

- 1. Description of the types of investigation related to:
 - a) Animal health and production Milk production loss due to bovine mastitis
 - b) Endemic diseases affecting trade Marek's disease
 - c) Zoonotic diseases Rabies
 - d) Internationally reportable diseases Peste des Petits Ruminants (PPR)

| Situation/issue | Objective | Methods | Action |
|-----------------|-----------|---------|--------|
| Milk production | | | |
| loss due to | | | |
| mastitis | | | |
| | | | |
| | | | |
| | | | |
| | | | |
| | | | |
| | | | |
| | | | |
| Endemic | | | |
| Marek's disease | | | |
| field | | | |
| investigation | | | |
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| D 1: 1: | | | |
| Rabies disease | | | |
| field | | | |
| investigation | | | |
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| | | | |
| High impact | | | |
| disease | | | |
| investigation | | | |
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Table 1. Stages of Outbreak Investigation and Response

| Outbreak Investigation | Outbreak Response |
|---------------------------|---------------------------|
| 1. Preparation | 1. Outbreak control |
| 2. Surveillance | 2. Outbreak communication |
| 3. Outbreak investigation | 3. Outbreak documentation |
| a. Confirmation and | |
| assessment | |
| b. Observation and | |
| description | |
| 4. Full investigation | |
| a. Analytical component | |
| b. Environmental | |
| component | |
| c. Laboratory component | |

Lesson 13 – Investigation Strategies for Early Prevention and Control of Animal Disease Transmission

| Estimated Lesson and Exercise Time | 1 hour and 30 minutes |
|------------------------------------|--|
| Participant Materials | ISAVET Lesson 13 Investigation Strategies for Early Prevention and Control of Animal Disease Transmission Participant Guide.PDF |
| | Computer and Microsoft Word Pen or Pencil |

In Service Applied Veterinary Epidemiology Training (ISAVET) Lesson 13: Investigation Strategies for Early Prevention and Control of Animal Disease Transmission

Learning Objectives

At the end of this lesson you will be able to:

- Describe zonal and compartmental approaches;
- 2. Explain the strengths and weaknesses of each approach for animal disease investigations; and
- 3. Apply zonal and compartmental approaches for case finding.

Investigation Scenario

Report:

A medium size pig producer in your local area notifies you that he is experiencing sudden spiking mortality in his herd.

- Is this an active or passive disease report?
- 2. What are important differential diagnoses to consider?
- What immediate action could you take prevention and control spread of this disease assuming it could be due to ASF?
- 4. What is your strategy to investigate the extent or scope of pig mortality in your local area?

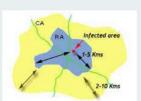


Reference: Google images

3

Strategy 1: Investigate the <u>zone</u> immediately surrounding the affected farm

- Looking within the <u>zone</u> immediately surrounding the affected farm we can understand how disease spreads to adjacent pig farms that are in the same geographic area
- Disease movement may begin from the first affected farm to neighbouring farms within the $\underline{\text{zone}}$
 - Mechanisms include tick vectors
 - · Scavenging animals (wildlife, domestic)
- * Pigs, people, equipment movement



Reference: Google OIE

4

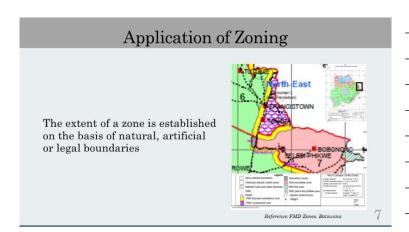
Strategy 2: Investigate the Pig Production System Compartment

- If we look within the pig production system <u>compartment</u> we can understand how disease spreads within the system
- A compartment is defined as an animal subpopulation that shares a common biosecurity management system
- Disease movement begins from the first affected farm to all farms and markets along the value chain by way of farmers and marketers of live animals, meat, feed and other products and services



Reference: FAO, Designing and Implementing Value Chain Studies 5

Disease Investigations Require Both Zonal and Compartmental Approaches Think and act in terms of zonal and compartment disease spread OIE definitions in Ch.1.1.1 Oie • Zone/Region • a clearly defined part of a country containing an animal subpopulation with a distinct health status with respect to a specific disease... • Compartment • one or more establishments (premises in which animals are kept) under a common biosecurity management system containing an animal subpopulation with a distinct health status ... Reference: Google images, OIE



A zone is defined based on the establishment of three areas: Infected area Buffer area Disease free area

Example of Zoning FMD zoning in Southern Africa · Ref: OIE, 2017. OIE standards on zoning and compartmentalization and $their\ implementation$ · FMD free zones are based on freedom with or without vaccination (green and blue) • FMD zones without a defined disease status are shown as a striped diagonal

Assessment of the Zonal Approach to Case Finding

Strengths

- May provide a date for the
- Defines the geographic extent of an outbreak
 Identifies spatial disease
- Identifies spatial disease clusters
 Assesses local high risk areas
 Guides prevention and control activities
 Permits contact tracing

Weaknesses

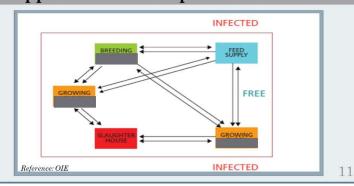
- Does not focus on value chain contacts like traders who travel extensively
 Limits case finding to

- Limits case inding to local spread
 Does not consider value chain actors who transmit viruses widely
 Does not consider transboundary transmission

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Application of Compartmentalisation



Example of Compartmentalisation: · Ref: ABPA, 2017. http://www.abpa-· Poultry value chain map within the broiler compartment in one area of Brazil Note that the subpopulation is: commercial broiler chickens with associated links in the value chain ABPA = 12

Assessment of the Compartmental Approach to Case Finding

Strengths

- Extends beyond the geographic boundaries of an outbreak
- Focuses on value chain decision makers like traders Considers the economic motives for animal movement
- Considers animal market movement and transboundary transmission
- Provides sites in which to conduct risk-based surveillance including high risk gathering points (abattoirs)

Weaknesses

- Does not provide a date for the index case
 Provides a limited
- amount of information about local spread Traders and marketers distrust governments
- and may not cooperate Does not assess local high-risk disease clusters

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How can we be sure the disease does not exist in a zone or a compartment?

Apply the Iceberg Principle:

- · Only part of the disease picture is apparent to the epidemiologist.
- · It is always safest to assume that we are likely arriving sometime after disease transmission began.
- Therefore, aggressive case finding is absolutely essential to assess the extent of disease transmission zone or a compartment!



Reference: Google Images

Case Finding and Data Collection

· Active Case Finding

- · Aggressive
- · Door-to-door and farm-to farm
- · Direct observation of animals
- Tracing movement in and out of a farm or premises
- * Tracing movement along the value chain

· Systematic Data Collection

- · Questionnaire
- · Taking samples
- · Animals, people, equipment, etc.
- \cdot Movement



Reference: Google Images

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How do we conduct case finding using zonal and compartmental approaches?

- Both approaches are important and should be considered for every outbreak investigation
- For zonal case finding:
- * Begin at the index farm and conduct surveillance on all neighbouring farms in an outward direction.
- · For compartmental case finding:
 - Trace and record all incoming and outgoing movements during the high risk period as defined by OIE and investigate each contact along the value chain.
 - High risk gathering points (e.g. markets, abattoirs)
 - · Contact tracing

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Exercise 19: Investigating an Outbreak

- 1. This exercise should take 1 hour to complete.
- 2. Divide into groups of four.
- 3. Explain which strategies your would use to investigate the following:
 - a) FMD in a communally pastured beef herd
 - b) Avian influenza in a commercial poultry flock
 - c) African swine fever in a smallholder pig herd

In Summary....

- ${}^{\raisebox{3.5pt}{\text{\circle*{1.5}}}}$ Disease investigation in a zone is based upon finding disease in a subpopulation in a geographic area.
- Disease investigation in a compartment is based upon finding disease links in a subpopulation that are part of the same biosecurity system along the value chain.
- Know the advantages and disadvantages of each approach and use both methods to increase the chance of finding the disease when it exists.
- Case finding should be conducted using both zonal and compartmental approaches to get a more complete picture of the extent of the disease transmission.

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Partners Contributors Contributors Contributors

Exercise 19 – Investigating an Outbreak

Description of Exercise:

Explain the strategies you would use to investigate different disease situations by production type. Should you have any questions over the exercise, please ask a trainer for clarification.

Allotted Time: 1 hour

Organisation of Group Work:

Divide into three groups.

Exercise Objective(s):

Working in groups, explain which strategies you would use to investigate the following:

- 1. FMD in a communally pastured beef herd
- 2. Avian influenza (AI) in a commercial poultry flock
- 3. African swine fever (ASF) in a smallholder pig herd

Exercise Components and Structure:

1. Flip Chart

Materials, Data or Information:

1. No data

Expected Outputs and Deliverables of Each Participant:

- 1. Description of traditional zonal case finding and tracing approach.
- 2. Description of compartmental approach including value chain mapping.

| 1. | Explain which strategies you would use to investigate FMD in a communally pastured beef herd. |
|----|--|
| 2. | Explain which strategies you would use to investigate avian influenza in a commercial poultry flock. |
| 3. | Explain which strategies you would use to investigate African swine fever in a smallholder pig herd. |
| | |

Lesson 14 – Establish Two-Way Linking Between Laboratory Disease Diagnosis and Field Investigation

| Estimated Lesson and Exercise Time | 2 hours and 30 minutes |
|------------------------------------|--|
| Participant Materials | Frontline ISAVET Lesson 14 – Establish Two-Way Linking Between Laboratory Disease Diagnosis and Field Investigation.PDF |
| | Computer and Microsoft Word Pen and Pencil |

In Service Applied Veterinary Epidemiology Training (ISAVET) Lesson 14: Establish Two-Way Linking Between Laboratory Disease Diagnosis and Field Investigation

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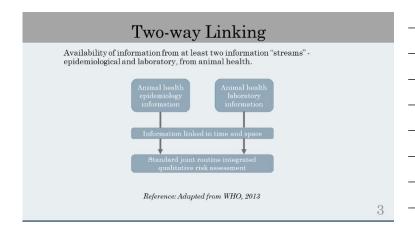
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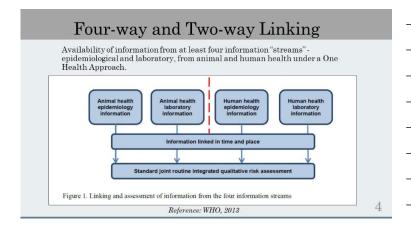
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Learning Objectives

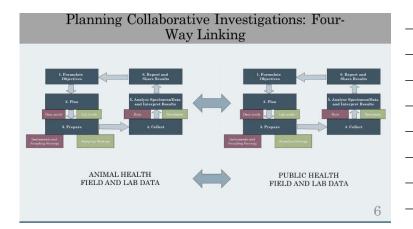
At the end of this lesson, you will be able to:

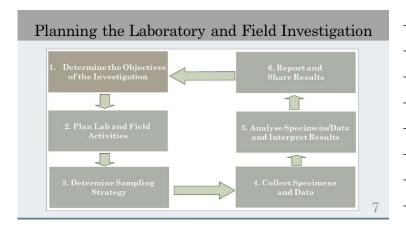
- Coordinate with the laboratory to collect, manage and interpret test results;
- $2. \ \,$ Use appropriate diagnostic methods for case detection and verify the diagnosis; and
- 3. Collect, label, package and transport samples for laboratory diagnosis using accepted methods.

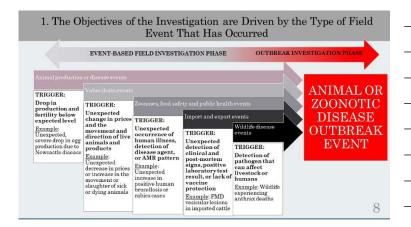












1. Determine the Objectives of the Investigation · Objectives of the Investigation: - Determine the presence or absence of disease - Determine the source of a disease - Determine the distribution of disease (Animal-Place-Time) - Determine the impact of disease 9 2. Plan Laboratory and Field Activities Field Laboratory Support the objectives of the investigation. • Confirm the diagnosis · Prepare for field work: Biosafety Biosecurity Transportation Detect common source Clinical case management (AMR) Laboratory supplies Use only validated (pre-tested) standardised data collection forms and questionnaires Coordinate with farmers, laboratory, national epidemiologists and local health officials Standard Operating Procedures Select the appropriate tests and laboratory forms Select the appropriate number and type of samples to collect in collaboration with the national epidemiologists officials Standard Operating Procedures are in place for biosafety and biosecurity Not to exceed daily laboratory surge capacity Provide and ensure that Standard Operating Procedures are in place for sample collection and submission 10 3. Determine Sampling Strategy Field Laboratory · Consult with an Prepare to collect either single or pooled samples based on proper laboratory protocol epidemiologist and the laboratory to determine the number and type of samples · Laboratory confirmation * Use appropriate laboratory tests Select the appropriate animals to sample:

• Healthy

 Collect descriptive data for the laboratory form and the

epidemiology questionnaire

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· Sick

· Dead

Use routine procedures
 Standard collection and

transport procedures
Know the biosafety level
laboratory required for the

• Establish laboratory role in

4. Collect Specimens and Data

Laboratory

- · Provide materials to collect samples, reagents and ice packs to maintain cold chain
- Follow written Standard Operating Procedures (SOP) for sample collection, handling and submission
- Follow written SOP for laboratory data collection
- Identify each sample with a unique sequential set of numbers that includes the farm No. and the Animal No.
 Example: Sample 3-1

Field

- Select the appropriate number of samples from healthy, sick and dead animals based on:

 Advice of an epidemiologist

 Advice of a laboratory scientist
- Follow written Standard Operating Procedures (SOP) for sample collection, handling and submission
- Follow written SOP for laboratory and epidemiology data collection
- Single (mortality) versus repeated (serology) sampling
- Confirm that the number of samples do not exceed the daily surge capacity of the laboratory

4.1 Laboratory Involvement in the Field

- · Presence in the field ideal
- Can provide timely input based on direct involvement and observation
 - Time consuming and expensive
 - · Most useful in complex investigations, unusual clinical presentations or unknown pathogens
- Remote participation as part of outbreak team (more common)
 - Optimal value if involved early
 - Need to exchange appropriate and sufficient information
 - Efficient for routine investigations (known, common pathogens)

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4.2 Labelling Samples

Includes:

- Flock/herd and animal unique ID number
- Test(s) ordered
- Time and date of collection
- Sample collector's initials



4.3 Sample Collection • Types of sample specimens to be collected in the field Collected specimens (one or more for the following): Whole blood Fresh tissue (lymph node, liver, spleen, Swabs kidney and lung) Whole blood (1-2 mL) Tissue specimens - Whole blood (1-2 mL) - Erroneous results can occur through: - Specimen mismanagement in the field - Improper labelling of specimen in the field - Incorrect date and time the specimen was collected - Incorrect transport medium is used for sample collection - Specimen is stored at the incorrect - temperature and degrades the sample - Specimen is not packaged and shipped in the correct manner - Retroner Tem A&A Wetgring Medical Biagnostic Laborators - Retroner Tem A&A Wetgring Medical Biagnostic Laborators - Temperature A&A Wetgring Biagnostic Biagnostic Laborators - Temperature A&A Wetgring Biagnostic B - Whole carcass Follow Frontline ISAVET Field Training SOP: Laboratory Specimen Submission, Packaging and Shipment

e: Texas A&M Veterinary Medical Dia

4.4 Information to accompany samples...1/2

- The following information should accompany samples to the laboratory;
- · Name and address of owner of the farm/animal (s) sampled
- · Contact information (telephone and fax numbers, e-mail address) of owner
- Geo-location (latitude and longitude, if available)
- · Animal details (species, breed, sex, age and identity of the animals sampled, and traceability number when available)
- · Herd details (numbers in herd, affected and dead)
- · Case history:
 - The clinical signs and symptoms observed
 - · Findings of the ante- and post-mortem examinations

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4.4 Information to accompany samples...2/2 TVMDL · Disease agent suspected · Differential diagnosis Fei BH-109-HIN Below REQUIRED · Tests requested List, type and quantity of samples submitted with transport media used · Date samples were collected and submitted · Any medication given to the animals, and when given Vaccination history describing the type of vaccines used and dates of application The type and standard of husbandry, including feeding and biosecurity measures · Contacts of person to receive results Contact Deposits The research is a deposit or all authorisists for a complete and of temp, unline, printing out against approving place and according to the contact and according to the con nce: Texas A&M Veterinary Medical Diagnostic Laborato







5. Analyse Specimens/Data and Interpret Results Laboratory Field · Coordinates submissions from · Transfers field collection forms the field to create a line listing in the local area or national level as Lab analysis is based on epidemiological information, objectives and laboratory appropriate · Initiates descriptive analysis consultations based on Animal-Place-Time characteristics including graphic Selects appropriate tests: · Screening · Confirmatory Interpret findings and · Applies the appropriate testing limitations Shares data with the laboratory · Shares laboratory test results in and national epidemiologists a timely manner Two-Way Communication with the Laboratory and the Field · Share preliminary information early - Time, place and animal characteristics - Suspect pathogens and differential diagnoses · Ensure on-going communication between the laboratory and the field - Identify focal person(s), obtain contact information - Generate an ID number for each source location - Provide regular daily updates - Send regular epidemiological report for input and revisions 22

6. Report and Share Results

Field

Clarify:

Provides results to the local veterinarian and national epidemiologist and as soon as possible

Laboratory

- Interprets the findings of the laboratory analysis
- Treats all laboratory results as sensitive information
- · Reports results to only designated recipients
- Exercises caution with the media -sensitive issues (e.g. RVF)
- · Epidemiology debriefing should include the laboratory methods and results
- · Interpret and update epidemiological results in light of the laboratory evidence
- · Coordinate the final report with the laboratory and national epidemiologists based on which group will take the lead on writing and sharing the final report

| | · | | | | | |
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Exercise 20: Specimen Collection, Diagnostic Testing and Interpretation of Diagnostic Tests

Wet Lab Instructions:

- Form 4 groups and complete this exercise in 1 hour and 30 minutes.
- Describe the steps you would take with the laboratory in order to establish a disease diagnosis of Pullorum in poultry
- 3. Practice safe sample collection, packaging, handling, submission and testing of samples for the disease: Pullorum in poultry.
- 4. Interpret results and propose measure to prevent and control each of the four diseases listed above.

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In Summary...Two-Way Linking is a MUST!

- 1. Engage in positive interactions with laboratory specialists
- 2. Identify appropriate samples to collect
- 3. Appropriately collect, label, package and transport specimens maintaining the cold chain and reducing biosafety risks
- ${\it 4.} \quad I dentify when and which laboratory methods to use e.g. to confirm existence of outbreak, identify causative agent, conduct phylogeny, etc.$
- 5. Identify the need for and use of follow-up testing. e.g. antimicrobial susceptibility
- 6. Identify the role of the laboratory in animal health surveillance
- 7. Understand laboratory quality assurance principles
- Establish two-way information sharing throughout the investigation and publish results jointly.

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Partners Contributors Contributors ISAVET Contributing Universities ATEXAS A&M AGRILIFE Contributors

Exercise 20 – Specimen Collection, Diagnostic Testing and Interpretation of Diagnostic Tests

Description of Exercise:

This exercise, is a wet lab that will be supervised by a laboratory diagnostician and two assistants and where you will discuss specimen collection, packaging shipment and diagnostic testing and interpretation for *Salmonella pullorum*. Should you have any questions over the exercise, please ask a trainer for clarification before during, and after the exercise.

Trainer Note: Pullorum was chosen since it is endemic in many countries, is not zoonotic and can be collected from poultry in many locations. Other diseases and SOPs would need to be developed if trainers want to conduct a wet lab for testing other diseases in other species.

Time: 1 hour and 30 minutes

Organisation of Group Work:

Assemble into four groups.

Wet Lab Instructions:

- 1. Describe the steps you would take with the laboratory in order to establish a disease diagnosis of Pullorum in poultry.
- 2. Discuss the information that would need to be included on the laboratory submission form from the field.
- 3. Practice safe sample collection, packaging, handling, submission, shipment and testing of samples for pullorum.
- 4. Interpret results and describe how you will report and share information that links the laboratory and the field activities.

Exercise Components and Structure:

Wet Lab Demonstration and Discussion:

Group A: Pullorum disease in poultry

- 1. The laboratory supervisor will review the laboratory diagnosis of pullorum disease and then a Frontline ISAVET field trainer demonstrate how to handle the chickens, select and prepare the site and then collect blood.
- 2. The laboratory supervisor will review a laboratory submission form and demonstrate the specific information that should be included and how to properly label and secure this information, so it does not get damaged during transport.
- 3. The laboratory supervisor will demonstrate how to perform, interpret and record the pullorum test and trainees will then perform, interpret and record the results of the slide agglutination test themselves.
- 4. The laboratory supervisor will demonstrate how to safely prepare and protect the quality of a blood sample tube for labelling, packing, handling and submission.

5. Based on the learning objectives of Lesson 14, the laboratory supervisor will ask trainees to explain how to report and share the findings of laboratory tests between laboratory and field including farmers, national epidemiologists and other stakeholders. Two trainees will be asked to volunteer to take notes to record all group discussion.

Materials, Data or Information:

- 1. Microsoft Word
- 2. Penn State video on recommended blood sample collection from chickens.
- 3. Standard Operating Procedure (SOP) for collection of blood from poultry
- 4. SOP for pullorum diagnostic testing in poultry
- 5. SOP for proper sampling handling, packaging and shipping.

Expected Outputs and Deliverables of Each Participant:

- 1. Explain the laboratory diagnosis of a specific disease (e.g. Pullorum);
- 2. Conduct steps of safe sample collection, packaging, handling, submission and testing of samples; and
- 3. Interpret results and how to report and share the findings of laboratory tests between laboratory and field including farmers, national epidemiologists and other stakeholders.

Pullorum Disease

Part 1: Diagnostic Testing for Pullorum Disease

OIE Terrestrial Manual Chapter 2.3.1

Description and importance of the disease: Pullorum disease of chickens is a bacterial infection caused by Salmonella enterica subspecies enterica serovar Gallinarum biovar Pullorum (Salmonella Pullorum). At this time the serovar is referred to as Gallinarum in some parts of the world and Pullorum in others; in this chapter the serovar will be referred to as Gallinarum or Pullorum according to the biovar under discussion as this is more meaningful from a clinical and epidemiological perspective.

In its acute form, Pullorum disease is almost exclusively a septicaemic disease of young chickens. However, the organism may also be associated with disease in turkey poults and may be carried subclinically or lead to reduced egg production and hatchability, plus a range of atypical signs in older birds. Ovarian transmission is a major route by which the organism can spread. Game birds and 'backyard' poultry flocks may act as reservoirs of infection, and wild birds may act as vectors for the organism and as such are important in the epidemiology of the disease.

Fowl typhoid in chickens and turkeys is caused by S. Gallinarum biovar Gallinarum and is more often observed in the later growing period and in mature stock. Disease is often characterised by rapid spread with high morbidity and acute or subacute mortality. Red mites may be involved in the transmission of disease and persistence in poultry houses. Clinical signs in chicks and poults include anorexia, diarrhoea, dehydration, weakness and death. In mature birds, Pullorum disease is less severe but decreased egg production, poor hatchability and some increased mortality may occur. Fowl typhoid is a more acute septicaemic condition which mainly affects mature birds and may be particularly severe in commercial laying flocks.

Identification of the agent: Samples should not be taken from birds or eggs that have recently been treated with antimicrobial drugs. Swabs or aseptically collected samples from infected tissues, or intestinal and cloacal contents should be used for diagnostic testing. Other materials that may be sampled include eggs, embryos, faecal droppings and hatcher debris, especially fluff, dust and broken eggshells and chick box linings. Samples of tissues such as caecal tonsils, liver, gall bladder and spleen from infected birds are preferable to faecal and environmental samples. Tissue samples should be inoculated into non-selective and selective enrichment broths and on selective agar medium, such as brilliant green agar, as soon as possible after collection.

In case of delay, samples should be stored at 4°C. Typical colonies can be identified by serological and biochemical

tests. Molecular approaches can also be used to identify and differentiate S. Gallinarum and S. Pullorum. Final serological confirmation of suspect isolates can normally only be completed in a Salmonella Reference Typing Laboratory.

Serological tests: These are satisfactory for identifying the presence and estimating the prevalence of infection within a flock. The test used in the field is the rapid whole blood plate agglutination test. This test is unreliable in turkeys and ducks as many uninfected birds may give positive reactions. In the laboratory a serum agglutination test is used, either as a rapid plate test or

as a tube test. These can be applied as macro- or microagglutination tests, though the latter may be more likely to give false-positive results with turkey sera. Any positive reactors should be confirmed as being infected by culture at post-mortem examination. Enzyme-linked immunosorbent assays have been reported but no commercial test is available.

The use of vaccines to control S. Enteritidis or S. Gallinarum infections in chickens may cause problems in the interpretation of serological results.

Requirements for vaccines: Live and inactivated vaccines are available for fowl typhoid in some countries. The most commonly used vaccine is a commercial live vaccine derived from the stable rough strain of S. Gallinarum known as '9R'.

Part 2. Specimen Collection-Implications for Testing

Instructions

Please review the information on specimen collection provided below. After carefully reading the information, provide written answers for questions that follow (All Groups).

Specimen Collection

Planning for Specimen Collection

The process of collecting, storing, and transporting a clinical specimen directly affects the quality of the specimen and the ability to produce quality results from a laboratory test. Many outbreak investigations will involve the collection of animal clinical specimens from outbreak-associated cases. Laboratory confirmation of an aetiologic agent is a critical component to a successful outbreak investigation. For this reason, it is important to remember that the ability of a laboratory to successfully identify a pathogen depends on appropriate specimen collection and transportation. Communication with the laboratory before specimen collection is critical to ensure appropriate collection technique and maintenance of sample so that accurate results can be obtained.

Performing Specimen Collection

It is important to obtain an adequate amount of the specimen and handle it with care, as this may be the only opportunity to obtain a specimen during the outbreak. A sample must be collected properly in order to ensure that the pathogen or infectious agent can be recovered in a viable form for laboratory analysis. Again, communication with the laboratory is vital.

Labelling and Identification:

Ensuring that specimens are accurately labelled at collection time is absolutely essential. Misidentification of a specimen leads to misidentification of a patient, which can result in improper diagnosis and treatment.

Storage and Transportation:

Appropriate storage of specimens before and during transportation must be maintained in order to preserve the integrity of the specimen. Most specimens need to be transported in sterile containers, although containers for faeces do not need to be sterile. Specimens transported in incorrect containers (e.g. non-sterile when sterility is required) may be rejected by the laboratory. Remember, all specimen containers should be closed tightly. Laboratories may reject the specimen if it shows signs of leakage/seepage, since this could potentially expose laboratory personnel to the contents.

Packaging of clinical specimens must comply with regulations (e.g., mail and commercial) for transport of infectious material. These regulations depend upon the

| con befo | e of transport (like ground or air delivery) and should be determined in sultation with the laboratory and carrier prior to specimen collection. Finally, ore transport, the receiving laboratory should be notified of the pending shipment hat they can prepare the facilities and workforce to handle the samples. |
|-------------|---|
| Gu | iding Questions |
| 1. | Before collecting a specimen: |
| | a. How do you determine what clinical and/or other samples you will need to collect? |
| | |
| | b. How do you determine which laboratory you will send your specimen(s) to? |
| | c. What logistics do you need to consider in terms of collection, packing, and transportation of the specimen? |
| | d. List the safe handling procedures of using the pullorum antigen in the field. |
| | When is the best time to collect a specimen from sick animal (s) during a course of an outbreak? |

| 3. | Why is proper sample collection technique so important to the laboratory? |
|----|---|
| 4. | What pieces of information should be included when labelling/identifying a laboratory specimen? |
| 5. | If a specimen contains a particularly dangerous or infectious organism, should this be noted in the labeling and identification process? Should the laboratory be notified in advance about what type of specimens to expect? |
| 6. | True or False: Most specimens stored at room temperature are suitable (or remain suitable) for laboratory diagnostic testing. Explain. |
| | |

Part 3: Interpreting and Reporting Laboratory Data

Group A: Poultry

Instructions

1. Interpret and record the test results in the following table:

| Chicken | Test Result |
|---------|-------------|
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2. How would you interpret, report and share the results from this flock?

3. You have a flock that has 9 of 10 poultry that have tested positive in the field. None of the birds are showing clinical signs. You are suspicious that the test may be producing false positive results. What are some reasons for why the test result might be unreliable under field circumstances.

Lesson 15 – Biosafety and Biosecurity for Animal Disease Investigations

| Estimated Lesson and Exercise Time | 2 hours and 30 minutes |
|------------------------------------|--|
| Participant Materials | ISAVET Lesson 15 Biosafety and Biosecurity for Animal Disease Investigations Participant Guide.doc |
| | Microsoft Word and Computer Pen and Pencil |

In Service Applied Veterinary Epidemiology Training (ISAVET)

Lesson 15: Biosafety and Biosecurity for Animal Disease Investigations

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Learning Objectives

At the end of this lesson, you will be able to:

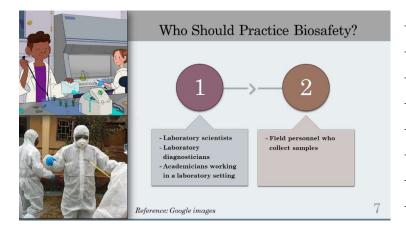
- 1. Explain and apply the principles of biosafety;
- 2. Explain and apply the four principles of biosecurity; and
- 3. Describe the three components of situational awareness for field epidemiologists.

What is Biosafety? "The containment principles, technologies and practices that are implemented to prevent unintentional exposure to pathogens and toxins, or their accidental release." Reference: WHO, Laboratory Biosafety Manual - Third Edition, http://www.who.int/csr/resources/publications/biosafety/WHO_CDS_CSR_LYO_2004_11/en/ 3 What is Biosecurity? Biosecurity is defined as "Procedures or measures designed to protect the population against harmful biological or biochemical substances." (The Oxford Dictionary, 2018) Laboratory biosecurity is defined as protection, control and accountability for valuable biological materials within laboratories in order to prevent their unauthorized access, loss, theft, misuse, diversion or intentional release." (Laboratory Biosafety Manual, 3rd Edition WHO 2004 and Biorisk Management – Laboratory Biosecurity guidance, WHO 2006) 4 $Reference: Google\ images$ Biosafety and Biosecurity are Required to Protect Ourselves and the Animals and Farmers That We Work With **Biosafety and Biosecurity**

5

Reference: Google images

Four Principles of Biosafety for Field Epidemiologists 1. Understand the agent 2. Understand the transmission risk 3. Manage the risk 4. Communicate the risk



Biosafety Principle 1: Understand the Agent (Example: Chapter 3.5 of the Terrestrial Manual. OIE, 2014).

- · Hazard:
- Bacillus anthracis



- Pathogen and disease:
- Anthrax is an acute bacterial disease primarily of herbivores and is transmissible to humans
- It is caused by $B.\ anthracis$, a gram-positive spore-forming rod-shaped bacterium

ference: Google images

Biosafety Principle 2: Understand the Transmission Risk

- Animals become infected by ingesting spores or possibly by being bitten by flies that have fed on an infected animal or carcass
- · Infected animals are usually found dead
 - Death can occur within 24 hours
- To avoid environmental contamination, post-mortem examinations conducted in the field (outside of laboratory containment) of carcasses of animals suspected to have died of anthrax is discouraged
- More than 95% of human anthrax cases take the cutaneous form and result from handling infected carcasses or hides, hair, meat or bones from such carcasses

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Biosafety Principle 3: Manage Risk

- Follow the following administrative procedures:
 - An emergency response plan
 - A communication plan with public and animal health authorities
 - A health and safety plan for personnel who may be exposed
 - A waste management plan
 - Training of field and laboratory personnel



Reference: Google images

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Biosafety Principle 3: Manage Risk

Follow standard operating procedures (SOP) $\,$











Prohibit postmortems in the field Safe carcass handling storage and disposal

g prescribed nd packaging l and shipping instructions Disinfect postmortem and laboratory areas and equipment

Wear
personal
protective
equipment
(PPE)
including
respiratory
protection

Photos: Google Images

Biosafety Principle 3: Manage Risk

- ${\mbox{-}}$ Follow the following standard operating procedures (SOP) (continued):
 - Follow safe handling practices of anthrax suspect carcasses including burning and burial





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Biosafety Principle 3: Manage Risk

- · Consider economic and environmental security risks:
 - Restrict movement of wool and hides
 - Avoid high risk contaminated soil where spores are stabile for years
 - Safely handle and inactivate microbiology laboratory liquid waste and solid waste before release
- Be aware of the possibility of intentional misuse of contaminated waste $\,$

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Biosafety Principle 3: Manage Risk

| ECONOMIC RISKS | ENVIRONMENTAL RISKS |
|--|--|
| Direct losses for farmers from reduced meat and milk | High risk contaminated soil where spores are stable for years |
| Restricted movement of wool and hides | Contaminated microbiology laboratory liquid waste and solid waste before release from laboratory |
| Healthcare costs for affected humans | Uintentional misuse of contaminated waste |

Activity Spectrum of Select Detergents and Disinfectants (Department of Epidemic and Pandemic Alert and Response of the World Health Organization, 2007)

| | BG+ | BG- | Myco B | Spores | Yeast | Virus | Prions |
|-----------------|-----|-----|-------------|--------------|-------------|-------------|--------|
| Alcohol 70° | ++ | ++ | ++ | 0 | + | + | 0 |
| Aldehydes | +++ | +++ | ++ | + | +++ | ++ | 0 |
| Ammonia IV | +++ | + | 0 | 0 | + | + | 0 |
| Anilides | + | 0 | NP | NP | 0 | NP | 0 |
| Chlorhexidine | +++ | ++ | 0 | 0 | + | + | 0 |
| Cl compounds | +++ | +++ | ++ | ++ | ++ | ++ | +(a) |
| Iodine (+ der.) | +++ | +++ | ++ | ++ | ++ | ++ | 0 |
| Hg compounds | +++ | ++ | 0 | 0 | + | 0 ou + | 0 |
| Phenols | | 7 | ariable act | ivity depend | ding on con | ponents (b) | |
| Hexachlorophe | +++ | + | 0 | 0 | + | 0 | 0 |

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Biosafety Principle 4: Communicate Risk

- In consultation with the national animal health authority:
 - $1. \ \ Contact the \ national \ office \ and \ request \ communication \ support$
- 2. The national office will develop protocols including identifying responsible person(s) and develop a message to be activated in the event of a positive anthrax diagnosis, including a press release, contact lists and a questions and answers
- ${\it 3.} \quad Identify\ responsible\ person(s)\ and\ develop\ a\ communication\\ message\ to\ be\ activated$

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| 1. Understand | A zoonotic orthomyxovirus found in wild birds that are low and high pathogenicity | | | | | |
|----------------------------|---|--|--|--|--|--|
| the agent | to poultry | | | | | |
| | The virus mutates and re-assorts rapidly and constantly with 16 H and 9 N subtypes | | | | | |
| 2. Understand transmission | Direct: Contact with wild birds and among poultry at farms, households and markets | | | | | |
| risk | Indirect: Movement of people, equipment, vehicles, cages, egg cartons among farms and markets along the value chain | | | | | |
| 3. Manage risk | Get annual influenza vaccine if you work with poultry | | | | | |
| | Reduce exposure: Wear clean PPE at every location and shower | | | | | |
| | Post-exposure anti-viral treatment | | | | | |
| 4.Communicate | e Although handling infected birds can result in human infection, cooking destroys the virus in meat and eggs | | | | | |
| one risk | Develop and share messages on how to prevent and control avian influenza in | | | | | |

Health Interventions Model Biosecurity is part of the Health Intervention Model 18

Five Principles of Biosecurity

- 1. Isolation e.g. quarantine
- 2. Traffic and movement control
- 3. Cleaning and disinfection
- 4. Disposal of dead animals and animal waste
- 5. Herd and flock health



Source: USDA

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Biosecurity Principle 1: Isolation

Ideal Isolation Scenario



Reference: Google images

Isolate Different Animal Species

Real Isolation Scenario

• Separation of species and production types from each other:

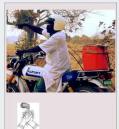


Reference: Google images

- Pigs
- Wild birds
- Ducks
- Fighting Cocks
- Other animal species

21

Isolate Yourself



Establish

Establish clean and dirty areas of your vehicle, clean boots and launder clothing daily. Practice good personal hygiene!

Leave dirty personal protective equipment (PPE) behind at the site you just visited.

Visit a maximum of one farm per day and wash yourself and your belongings after each visit.

Clean vs Dirty Once you are exposed to the disease agent, you are "dirty". You need to clean yourself and wait 24 hours to be "clean".

Reference: Google images

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Isolate Yourself: Wear Personal Protective Equipment (PPE)



- $\boldsymbol{\cdot} \, \mathbf{Supplies:} \,$
 - Chemical-resistant tape
 - Blunt-nosed scissors
 - Inner and outer gloves
 - Tyvek coveralls
 - Rubber boots
- N95 or reusable APR
- Goggles
- Biohazard bag
- Supplies for tasks

2. Traffic and Movement Control Prevent and control disease transmission both locally and through the value chain CA RA Infected area 2-10 Kms Reference: Google images, OIE



3. Cleaning and Disinfection

- · Cleaning
 - Most important step!
 - Must remove ALL faecal and organic matter
- · Disinfection
- Final Step

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References: Disinfectants

· FAO

MANUAL ON PROCEDURES FOR DISEASE ERADICATION BY STAMPING OUT

http://www.fao.org/docrep/004/Y0660E/Y0660E03.htm

· OIE

2018 © OIE - Terrestrial Animal Health Code - 10/08/2018 CHAPTER 4.13. GENERAL RECOMMENDATIONS ON DISINFECTION AND DISINFECTION http://www.oie.int/fileadmin/Home/eng/Health_standards/tahc/current/chapitre_disinfect_disinsect.pdf



4. Disposal of Dead Animals and Waste

- · Types include:
 - Carcasses
 - Faecal matter
 - Offal
 - Feathers
 - Hair
 - Horns
 - Contaminated feed
 - Contaminated fomites
- Dead animals and waste products are major sources of animal disease pathogens.

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Disposal of Dead Animals and Waste

| Disposal Method | Advantages | Disadvantages | | |
|------------------|--|---|--|--|
| Burial | Rapid on-site removal of carcasses and topsoil | High water table in rainy season; depends on soil and bedrock | | |
| Burning | Rapid on-site removal | Environmental air pollution | | |
| Composting | Permits re-cycling | Specific guidelines required wild and feral animals can poach | | |
| Alkali Digestion | Complete digestion of carcass | Expensive to construct | | |

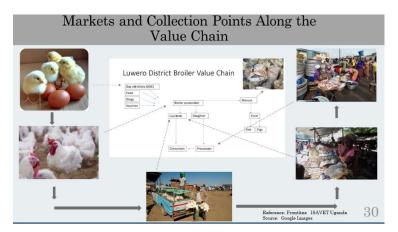
Each situation is different and options must be evaluated carefully to meet local conditions

28

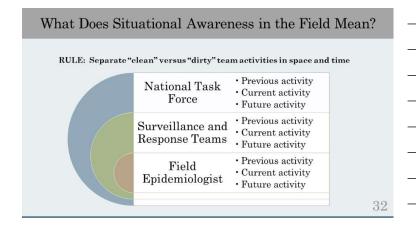
5. Flock and Herd Health



- Biosecurity includes preventive practices
- $1.\ Prevent\ exposure$
- 2. Protect susceptible animals from infection
- 3. Prevent progression from infection to disease
- · Prevent death and disability
- $\cdot \ \text{Prevent further transmission}$
- 5. Interrupt transmission
- Some activities targeted
 Environmental; Social/behavioural; Legal; Genetics; Nutrition; Management; Veterinary care and Vaccination







| Fi | ontline curriculum – Participant ma | nual |
|--|---|------|
| Examples of | Situational Awareness | |
| Level | Sequence of Events and Interventions | |
| 1. Individual | You came from a "clean" location You just entered an "infected" location You must take action to clean yourself and clothing before visiting other farms | |
| 2. Field Team | Your field team is doing case finding for FMD You enter a farm with infected animals Your team is now considered "dirty" and may be assigned to culling duties or other action | |
| 3. National Task Force | A positive case is discovered in the "surveillance" zone without a previous case The area is now designated as "infected" zone All teams and individuals in this "infected" zone must take interventions as noted above | |
| | osafety and Biosecurity for | |
| Animai | Field Investigations | |
| Instructions: | | |
| 1. This exercise will take 60 | | |
| This exercise is divided in minutes each). | to three parts and take 60 minutes (20 | |
| | he principles of biosafety, biosecurity and | |
| 4. Form yourselves into grou | ups of roughly equal size. | |
| 5. Read the scenario and an | swer the questions for each section. | |
| 6. Groups will report their a | nswers in 60 minutes. | |
| | 34 | |
| | | |
| | | |
| | | |
| In S | Summary | |
| | principles, technologies and practices that are tentional exposure to pathogens and toxins, or | |
| | res or measures designed to protect the iological or biochemical substances | |
| | g an investigation prevents disease ealth surveillance and response teams by ' activities. | |

| ISAVE | T Contributing Universities | - | |
|--------------|-----------------------------|---------------------|--|
| Partners | TEXAS A&M GRILIFE | - | |
| Contributors | MAKERINE UNIVERSITY | - - - 36 - | |

Exercise 21 – Biosafety and Biosecurity for Animal Disease Investigations

Description of Exercise:

Review the following three scenarios and answer biosafety and biosecurity questions with respect to situational awareness and decision-making. Should you have any questions over the exercise, please ask a trainer for clarification before during, and after the exercise.

Allotted Time: 60 minutes and an additional 30 minutes for plenary discussion

Organisation of Group Work:

• Form yourselves into 5 groups of roughly equal size.

Exercise Objective(s):

- 1. Explain the principles of biosafety.
- 2. Explain the four principles of biosecurity.
- 3. Describe three components of situational awareness for field epidemiologists.

Exercise Components and Structure:

- 1. Review each of the 3 scenarios (20 minutes/scenario).
- 2. Apply the principles of biosafety, biosecurity and situational awareness.
- 3. Answer the questions from each scenario within the allotted time of 20 minutes for each scenario.

Materials, Data or Information:

- 1. Computer
- 2. Microsoft Word
- 3. Pen or Pencil

Expected Outputs and Deliverables of Each Participant:

1.Understand decision-making processes for biosafety and biosecurity while in the field.

Scenario 1

You are aware of a large die-off of wild birds that has been confirmed as highly pathogenic avian influenza (HPAI) subtype H5N1 at a lake near a Community X poultry cooperative with 20,000 egg layers. You are asked to investigate the death of the some of the egg layers that has just been reported by the sublocal area officer.

A. Biosafety in the field

- 1. Using the 4 principles of biosafety, explain how you would how would you protect field epidemiologists investigating the poultry mortality?
 - a) The agent:
 - b) Disease transmission:
 - c) Risk management:
 - d) Risk Communication:
- 2. Before you leave to go to the field, give three ways you would you prepare yourself to ensure biosafety, assuming the poultry could be infected with avian influenza virus:

B. Biosecurity in the field

1.

2.

Scenario 2

You are in Community X that is experiencing pig mortality due to African Swine Fever and you learn that the pig farms in Community X sell pigs regularly to markets in nearby villages and in Town A located nearby.

| gularly to markets in nearby villages and in Town A located nearby. |
|--|
| Using the five principles of biosecurity, give one example of control measures you would take for each of the following: |
| a) Isolation: |
| b) Traffic and movement control: |
| c) Cleaning and sanitation: |
| d) Disposal of dead animals and animal waste: |
| e) Herd health: |
| State one prevention measure you would take to prevent transmission of African Swine Fever virus to: |
| a) Other pigs: |

- b) Humans working at the local markets:
- a) Consumers:

C. Situational awareness in the field

Scenario 3

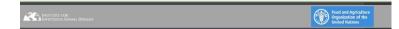
Foot and Mouth Disease outbreaks have been occurring recently in your local area. Dead and sick cattle have not yet been reported in Town A due to Foot and Mouth Disease (FMD), near to the cattle market. You are asked to lead a surveillance team to investigate beef and dairy cattle of unknown health status at the market near to Town A.

1. Before going to Town A, you recall that you have just returned from investigating Foot and Mouth Disease (FMD) positive infected cattle in Town B where cattle were sick and dying. Using your knowledge of situational awareness, what will you do next? Is it safe to conduct surveillance in the market near to Town A?

Lesson 16.1 – Apply the Steps of an Animal Health Outbreak Investigation for Animal-Specific and Zoonotic Diseases

| Estimated Lesson and Exercise Time | 3 hours and 30 minutes |
|------------------------------------|---|
| Participant Materials | ISAVET Lesson 16.1 Apply the Steps of an Animal Health Outbreak Investigation for Animal-Specific Diseases Participant Guide.doc |
| | Computer and Microsoft Word |
| | Pen or Pencil |

In Service Applied Veterinary Epidemiology Programme (ISAVET) Lesson 16.1. Apply the Steps of an Animal Health Outbreak Investigation for Animal-Specific and Zoonotic Diseases



Learning Objectives

At the end of this lesson, you will be able to...

- 1. Define the objectives for an animal health outbreak investigation.
- 2. Define the steps for conducting an animal health outbreak investigation.

Field Epidemiology

"...The early investigative activities surrounding the identification of a possibly emergent disease must be carried out in the field and not the laboratory.

This is the world of shoe-leather epidemiology..."

(Murphy, 1998)

3

Outbreak Investigation

 $Outbreak\ Investigation-Field\ and\ laboratory\ investigations\ that\ are\ linked\ to\ the\ following\ events\ must\ be\ considered\ as\ potential\ outbreak\ events:$

- $a.\ Unusual\ drops\ in\ milk,\ meat\ and\ egg\ production$
- $b. \ Suspicion \ or \ detection \ of \ unusual \ high-risk \ antimic robial \ resistance$ profiles in humans and animals
- c. Unusual high morbidity and high mortality animal disease events
- d. Zoonotic disease events linked to human and animal disease
- e. Isolation of internationally reportable high impact animal diseases as defined by the World Organisation for Animal Health (OIE)

4

Objectives of an Outbreak Investigation

- $\cdot \ Determine \ the \ presence \ or \ absence \ of \\ disease$
- · Determine the source of a disease
- · Determine the distribution of disease (Animal-Place-Time)
- · Determine the impact of disease
- $\begin{array}{c} \cdot \ Determine \ the \ magnitude \ of \ the \\ outbreak \end{array}$
- Determine risk factors associated with the outbreak
- · Propose future prevention and control measures and next steps

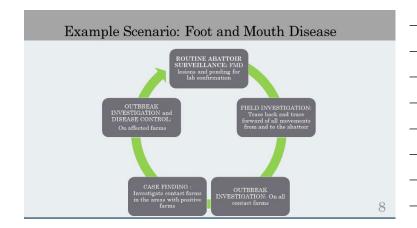


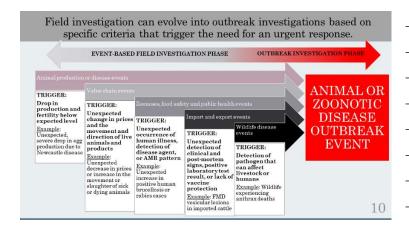
ce: Google Image

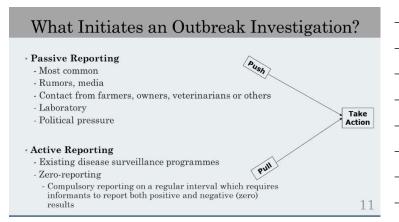
 $\begin{tabular}{ll} Reference: \\ U.S.~CDC. & Morbidity~Mortality~Weekly~Reports,~2004. \end{tabular}$

Describe the objectives of the outbreak investigation Develop a case definition Assist in developing and apply a useful questionnaire Plan the data collection Conduct outbreak investigations Data entry Data analyses

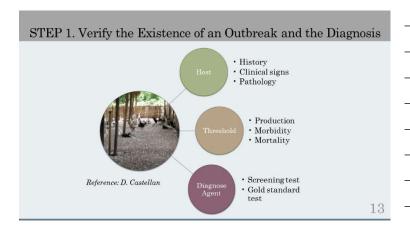








Steps of an Outbreak Investigation (Adapted From: Mazet, UC Davis and Reingold, UCLA) 1. Verify the existence of an outbreak and the diagnosis 2. Establish working case definition(s) 3. Prepare for field work 4. Verify the diagnosis 5. Case finding and data collection 6. Describe the outbreak by Animal, Place, Time 7. Develop a hypotheses 8. Intensive follow-up including analytical studies to test the hypotheses 9. Conduct special studies (e.g. environmental, value chain) 10. Implement control measures 11. Communicate findings



1. Verify: Population Disease Thresholds Greater or less than "normal": - 2 standard deviations (SD) above or below 5-year mean value Production - Decrease in milk, meat, egg production - Decrease in feed and water intake Morbidity: - Daily, weekly, etc. - Village, farm – field data

STEP 2. Establish Working Case Definition(s) Classification (3-Tiers) Suspect case Probable case Confirmed case Define Based on the Following Criteria: Animals at risk Place of occurrence Time of occurrence Unit of interest Clinical signs Lesions Screening and Gold Standard Tests Morbidity & Mortality 15

STEP 3. Prepare for Field Work

- · Local area profile information
- · Resources and equipment
- · Logistics
- · Planning
 - Objective(s)
 - Essential data and specimens to collect
- Roles and responsibilities:
 - Epidemiologist
 - · Laboratory diagnostician
 - ${\color{blue} \bullet} \ \, \textbf{Environmental specialist}$
 - · Government ministrie
 - ${\color{gray} \cdot} \ {\bf Communications} \ {\bf officer}$
 - · Often, one person will play several roles
- · Others.



Reference: Castellan

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STEP 4. Verify the Diagnosis

- Field situation is compatible with the laboratory diagnosis
 Proper interpretation of the case definition
- · Submit additional samples to confirm
- Use an international reference laboratory for final confirmation
- Include molecular characterisation of field isolates e.g. FMD, AI

When to Initiate Control Measures?

- An outbreak is an emergency...therefore apply control measures continuously from the first day onwards!
- Timely response can control and prevent further cases
- ${}^{\bullet}$ Must often take action with incomplete knowledge but must take action!
- · Is it a known agent?
- · Review transmission routes daily
- · Review spatial and temporal disease patterns daily
- · Provide immediate recommendations to the farmer

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STEP 5. Case Finding and Data Collection

- · Active Case Finding find it, don't miss it!
 - Direct observation
 - Take samples
 - Tracing and following movement in and out of a population
 - Animals, people, equipment, etc.

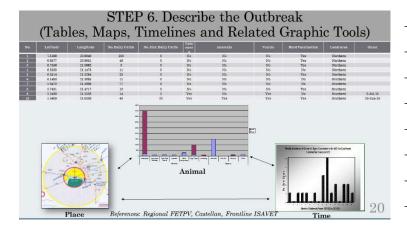
- Systematic Data Collection

- Use a pretested Questionnaire
- Develop a line listing of data collected





Reference: Regional FETP



Descriptive epidemiology

- $\begin{array}{l} \bullet \; \underline{\mathbf{What}} \; \mathrm{events} \; \mathrm{occurred:} \\ \bullet \; \mathrm{Production, movement} \; \mathrm{and} \; \mathrm{molecular} \; \mathrm{changes} \end{array}$
- Who is involved (animals and humans): Animal
 Individual animal/human
 Herd
 Flock

- When events occurred in time (critical time periods): Time
 Onset of clinical signs
 Movements animals, people, equipment
 Management changes
 Contact with other farms, villages and markets



Essential Disease Investigation Data Reference: Google Images

 $\bullet \ \underline{\mathbf{Where}} \ \text{events occurred including man-made and natural environments: } \mathbf{\underline{Place}} \\$

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STEP 7. Develop a Hypotheses from Data:

Hypothesis can be set to determine

- · Source of pathogen
- Mode of transmission
- · Factors related to the outbreak
 - * Intrinsic factor i.e. age, sex, breed, health condition
 - ${}^{\bullet}$ Extrinsic factor i.e. humidity, temperature

May need to do further study such case-control study

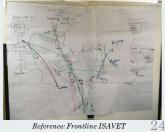
7. Develop a Hypotheses from Data: $\substack{ \text{(Example: Classical swine fever)} }$

| Herd Name | Location | % Morbidity | Disease Period | share boar | swill feed | trader | no vaccine |
|-----------------|----------|-------------|-------------------|---------------|------------|--------|------------|
| N | Dist. 4 | 88 (14/16) | April | 1 | 0 | 1 | 1 |
| A | Dist. 8 | 7 (1/14) | April | 1 | 0 | 1 | 1 |
| 5 | Dist. 4 | 100 (2/2) | May | 1 | 0 | 0 | 1 |
| w | Dist. 4 | 88 (14/16) | May | 1 | 0 | 1 | 1 |
| c | Dist. 4 | 8 (2/25) | May | 1 | 0 | 1 | 0 |
| Y-F (confirmed) | Dist.12 | 8 (5/66) | May | 0 | 0 | 1 | 1 |
| н * | Dist. 12 | 3 (3/84) | May | 1 | 1 | 1 | 0 |
| Ac | Dist. 8 | 26 (7/26) | May | 1 | 0 | 1 | 1 |
| So | Dist. 11 | 18 (11/60) | June | 0 | 0 | 1 | 0 |
| 5u* | Dist. 4 | 71 (5/7) | July | 1 | 0 | 1 | 1 |
| 50p* | Dist. 8 | 5 (3/52) | July | 0 | 0 | 1 | 1 |
| | | | | 72.73% | 9.09% | 90.91% | 72.737 |

Reference: Regional FETPV

STEP 8. Conduct Intensive Follow-up Studies STEP 9. Conduct Special Studies

- · Follow-up Studies
 - Observational: cross-sectional studies or case-control studies are commonly done
- · Special Studies
- Environmental
 - · Wild birds, bats, water and feed
- Value Chain
- Follow the flow into and out of each place
- Price fluctuations



Reference: Frontline ISAVET

STEP 10. Implement Control Measures

- · Start immediately and apply continuously
- · Risk communication is important
- · Movement control
 - Zone <u>and</u> compartment-based
- · Humane culling and disposal
- Surveillance
 - Aggressive case finding
 - Investigate value chain relationships
- · Vaccination when appropriate

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STEP 11. Report Findings

- · Methods

 - Informal
 Verbal report or briefings
 Written briefings including recommendations

 - <u>Formal</u> Publications Presentations
 - After action review
- · Reasons to Report
- Share experience with others to improve surveillance and response systems ${\bf Advance}$ our understanding of the disease

Exercise 22: Outbreak Investigation

- 1. This exercise is a plenary panel discussion will take 90 minutes.
- Following both presentations, a facilitator will guide a panel discussion directed at the following key points:
- a) Describe similarities and differences between animal health and public health outbreak investigations.
- b) Describe the strengths and weaknesses, opportunities and strengths (SWOT analysis) showing how animal health and public health agencies currently collaborate at the local level in your area.
- Propose follow up action you will take to support joint outbreak investigations when you return to your local area following Frontline ISAVET.

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In Summary...

- · Every outbreak is unique
- · Know the steps of an outbreak investigation
- · Apply them flexibly
- · Keep a wide perspective using a One Health approach through joint investigation with public health and wildlife health experts
- · Share and report your findings with stakeholders

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ISAVET Contributing Universities

Partners



Contributors

Lesson 16.2 – Apply the Steps of a Public Health Outbreak Investigation for Animal-Specific and Zoonotic Diseases

| Instructor Materials | ISAVET Lesson 16.2 Apply the Steps of a Public Health Outbreak Investigation for Animal-Specific Diseases.pptx |
|-----------------------|--|
| | ISAVET Lesson 16b Apply the Steps of a Public Health Outbreak Investigation for Animal-Specific Diseases Instructor Guide.doc |
| | Computer |
| | Microsoft Word |
| Participant Materials | ISAVET Lesson 16.2 Apply the Steps of a Public Health Outbreak Investigation for Animal-Specific Diseases Participant Guide.doc |

Exercise 22 - Field Investigation and Response

Description of Exercise:

Identify key stakeholders who will need to be involved in an animal disease outbreak in your local area and at the national level. Should you have any questions, please ask a trainer for clarification.

Allotted Time: 90 minutes

Exercise Objective(s):

 Describe the stakeholders in your local area and national level and the methods you will use to collaborate on an outbreak investigation for an animal disease event.

Exercise Components and Structure:

- 1. This exercise will take 90 minutes.
- 2. Form groups of equal size.
- 3. Complete the table provided to:
 - a) list the local area level stakeholders;
 - b) their roles and responsibilities; and
 - c) how they coordinate their activities together to deal with an animal disease event in your local area and with the national level.
- 4. Groups will report their answers in plenary discussion.

Materials, Data or Information:

- 1. Computer
- 2. MS Word or MS PowerPoint
- 3. Pen or Pencil

Expected Outputs and Deliverables of Each Participant:

- 1. List of stakeholders.
- 2. Coordination mechanisms.

Coordination of Animal Disease Events at Local area and National Levels

| Local area Stakeholders | Roles and Responsibilities of Local area Stakeholders | Coordination Mechanisms at Local area and National Levels |
|----------------------------|--|--|
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Case Study 2: Highly Pathogenic Avian Influenza

Case Study of an Outbreak Investigation: Highly Pathogenic Avian Influenza

Acknowledgements:

This scenario was originally developed by the Faculty of Veterinary Medicine, Kasetsart University and the Thailand Department of Livestock Development. The scenario for the outbreak investigation for this exercise is adapted from an outbreak situation in which only 3 households were involved. The names and places in the exercise are fictional and have been modified from their original version for teaching purposes. The views expressed in this exercise are solely those of the authors.

General Instructions:

This is a table-top outbreak investigation exercise that will take three hours to complete.

Audience: This case study is designed for Frontline In-service Applied Veterinary Epidemiology Training (ISAVET) trainees.

Level of Training: Basic level (frontline) training in epidemiology

Language: English and French

Prerequisites:

- 1. Enrollment in Frontline ISAVET, Week 2, Field and Outbreak Investigation.
- 2. Knowledge of avian influenza virus.
- 3. Skills in descriptive epidemiology to analyse animal-place-time components related to an outbreak investigation.

Time Required: Approximately 3 hours

Learning Objectives:

At the end of this case study, participants will follow the steps of an outbreak investigation and be able to:

- 1. Verify the existence of an outbreak and the diagnosis
- 2. Establish working case definition(s)
- 3. Prepare for field work
- 4. Verify the diagnosis
- 5. Case finding and data collection
- 6. Describe the outbreak by Animal, Place, Time
 - a. Develop an epidemic curve
 - b. Calculate the impact of disease morbidity and mortality
- 7. Develop a hypotheses for risk factors
- 8. Suggest follow-up studies including analytical studies to test the hypotheses
- 9. Implement control measures

Part I - Background

Highly pathogenic avian influenza (HPAI) subtype H5N1 is an infectious disease which can cause high mortality rates in poultry and restricts international trade in poultry products. Outbreaks of HPAI were first reported in Africa in February 2006. The disease has been classified as a notifiable disease under national health law. Passive and active surveillance systems are in place along with control and preventive measures that include cleaning and disinfection, education, farm bio-security, compartmentalization, movement control, fighting cock identification, stamping out and compensation.

Scenario – Outbreak Investigation of Avian Influenza

Place: Village 1, Crater Subdistrict, Volcano District, Island Region

On 28 October 2010, the Volcano District Livestock Office was informed by a local livestock health inspector of high mortality from an unknown cause in poultry in Village 1, Crater subdistrict, Volcano district, Island region. The inspector asked about the symptoms the birds exhibited before they died and the owner (Mr. A) reported that the affected chickens were depressed with torticollis and had swollen heads, cyanotic wattles and combs, and respiratory dyspnea. There was also some morbidity and mortality in chickens near to Mr. A's house. The District Livestock Office therefore informed the Regional Livestock Office of the event.

<u>Question 4.1a:</u> In the aforementioned scenario, are Regional or National Veterinary Officers required to investigate this report of disease?

- A. Yes
- B. No
- C. Not sure

Answer:

Question 4.1b: Is this an outbreak? Please provide any reasons to support your selection.

Answer:

Question 4.2: The district livestock officer collected and sent samples to the laboratory. It will take 24 hours before confirmatory results are available using RT-PCR. Poultry mortality was not limited in only one household and seemed to have spread. This disease event needs to be investigated, What would not be a priority objective of the disease investigation?

- A. Search for the cause of the disease outbreak.
- B. Identify the magnitude of outbreak.
- C. Perform disease control measures to stop any spread of the disease.
- D. Set up protocols for prevention of future outbreaks.

Answer:

Next, the investigator needs to specify what the 'case definition' is. In order to do that, the officer must go to the outbreak area to examine affected animals and collect the epidemiological information that helps to characterise this particular disease event.

Preliminary findings in the affected area:

The affected area is in Village 1, Crater subdistrict, Volcano district, Island region. The villagers were predominantly crop farmers and their houses were scattered throughout the village. The poultry were mainly backyard chickens. Within a radius of 3 km from the notified case. There were 50 households with poultry, having a total of 800 chickens. The notified case was in Mr. A's property which is 16,000 m² in area. Mr. A. had forty (40) backyard chickens and six (6) beef cattle. He had no activities related to cock fighting and he had not gone to the cock fighting arena. On 27 October 2017, he observed that four (4) chickens died from an unknown cause and that egg production was reduced. On 28 October 2017, there were 11 more deaths and all of the 11 dead chickens had swollen heads and cyanotic wattles and combs. Mr. A, therefore promptly notified the district veterinary officer on the same day. After the investigation team entered the affected area, they also found dead chickens at Mr. D's house. Of 10 mature chickens, five (5) showed dyspnoea, lacrimation and swollen faces prior to their death. Some also has petechia on their shanks. Two chicks are still healthy with no signs of disease.

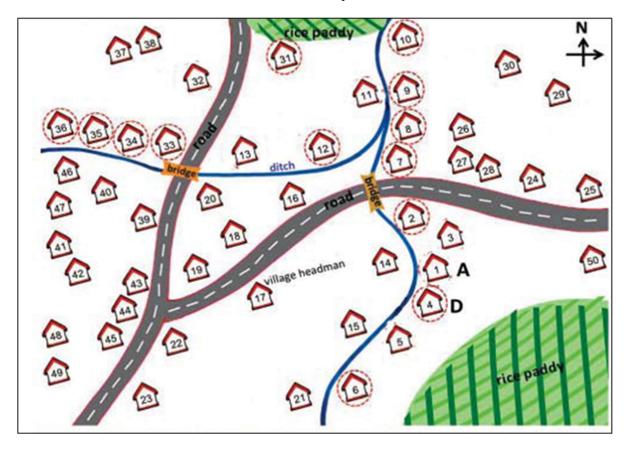
Question 4.3a: From the information provided, what is your preliminary "case definition"?

Answer:

Question 4.3b: What poultry diseases are among your primary differential diagnoses?

Answer:

Before entering the affected area to investigate, you are called from the neighbouring district to assist. You want to get the history of recent poultry disease events and the district livestock officer from the Volcano District provided you with a map. The local livestock health care volunteer has also circled the locations of places on the map, where the cases were preliminarily reported.



<u>Question 4.4:</u> You want to collate the information about the outbreak that is set out on the disease investigation forms for individual households. You need to do this to obtain a better understanding of the epidemiology of the outbreak. What information do you need to collect from poultry owner?

Answer:

•

You have decided to visit all households in the affected area and investigation team was divided into 3 small sub-teams (A, B and C) after taking the clusters and locations of cases in the area into consideration.

Team A investigated the area that included house numbers 1 to 13. Team B was allocated house numbers 14 to 36, and Team C house numbers 37 to 49. The teams were instructed to enter only uninfected farms first to avoid contact with the virus and further spread.

The assumption is made from the first data collected that the incubation period is 3 days.

| House No. | House | No. Poultry | No. Adults | No. Chicks | Respiratory | Neurological | Gastroenteric | No. sick | No. dead | Date of |
|-----------|-------|-------------|------------|------------|-------------|--------------|---------------|----------|----------|------------|
| | Name | at Risk | | | symptoms | symptoms | symptoms | observed | observed | Onset of |
| | | | | | | | | | | Clinical |
| | | | • | | • | ~ | ~ | | | Signs 🔻 |
| 1 | Α | 40 | 30 | 10 | 11 | 0 | 0 | 11 | 15 | 27/10/2010 |
| 2 | В | 20 | 10 | 10 | 0 | 0 | 5 | 5 | 5 | 26/10/2010 |
| 3 | С | 38 | 20 | 18 | 0 | 20 | 0 | 15 | 20 | 25/10/2010 |
| 4 | D | 12 | 10 | 2 | 5 | 0 | 0 | 0 | 10 | 28/10/2010 |
| 5 | Е | 18 | 10 | 8 | 8 | 0 | 0 | 8 | 8 | 27/10/2010 |
| 6 | F | 12 | 12 | 0 | 6 | 0 | 6 | 0 | 12 | 28/10/2010 |
| 7 | G | 16 | 8 | 8 | 8 | 4 | 0 | 0 | 16 | 26/10/2010 |
| 8 | Н | 22 | 12 | 10 | 0 | 10 | 0 | 10 | 10 | 19/10/2010 |
| 9 | I | 17 | 10 | 7 | 0 | 4 | 0 | 4 | 10 | 18/10/2010 |
| 10 | J | 10 | 2 | 8 | 2 | 0 | 0 | 0 | 10 | 17/10/2010 |
| 11 | K | 10 | 2 | 8 | 2 | 4 | 4 | 0 | 10 | 18/10/2010 |
| 12 | L | 16 | 8 | 8 | 8 | 4 | 0 | 0 | 16 | 18/10/2010 |
| 13 | М | 18 | 10 | 8 | 6 | 0 | 0 | 6 | 10 | 19/10/2010 |
| 31 | EE | 40 | 35 | 5 | 15 | 5 | 5 | 15 | 25 | 24/10/2010 |
| 32 | FF | 14 | 10 | 4 | 6 | 0 | 0 | 6 | 8 | 27/10/2010 |
| 33 | GG | 15 | 10 | 5 | 8 | 7 | 0 | 0 | 15 | 24/10/2010 |
| 34 | НН | 16 | 10 | 6 | 5 | 0 | 5 | 0 | 16 | 25/10/2010 |
| 35 | = | 15 | 5 | 10 | 5 | 0 | 0 | 5 | 5 | 28/10/2010 |
| 36 | IJ | 10 | 8 | 2 | 2 | 2 | 0 | 2 | 8 | 27/10/2010 |

Question 4.5: An epidemic curve can be constructed at an early stage of the investigation and before laboratory confirmation. Please draw an epidemic curve using the data presented in the previous Table (Provide graph-drawing paper) and please interpret this epidemic curve. Correct X-axis interval: 1/3 of average incubation period (assume the incubation period is 3 days).

| | J | , | • | • | • / |
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<u>Question 4.6a:</u> Calculate the total mortality rate for all affected houses for all chickens and chicks. Is the total mortality rate above the expected threshold of 5%?

Answer:

Investigations carried out within a radius of 3 km from the notified case found that there had been deaths of chickens at Mr. B's house. The house was located 300 meters away from Mr. A's house. Mr. B had 20 backyard chickens and had a retail restaurant business which involved cooking and food sales. There was a small ditch alongside his house and a pond behind. On 26 October 2010, 5 chickens with symptoms of white colored diarrhoea had died. Mr. B buried these dead chickens and did not notify the veterinary authorities.

The investigation team found that Mr. A (house number 1) usually visited the village shop belonging to Mr. B (house number 2), who had experienced chicken deaths in this backyard poultry. Mr. B told the investigation team that Mrs. EE (house number 31) had visited his shop to buy live chickens for slaughter one month before the death of Mr. B's chickens. He also reported that there had been some wild birds behind his house and that a flock of free-grazing ducks passed by the front of his house one week ago.

We decided that if we want to find the origin and cause of the outbreak we should start by investigating the first point of the outbreak, which was in Ms. J's house (House number 10). Ms. J was visited at her house and it was noticed that her house was located close to a 2 meter-wide ditch. The water passed through other neighbouring villages before going past her house. The rate at which the water flowed was rather strong through the outbreak village despite it being reduced after passing through a bifurcation. All villagers used the water from this ditch for their agricultural activities.

Ms. J did not know where her father worked. In the weekend, her father sometimes went to the cock fighting arena but he did not own any fighting cocks.

Before this outbreak, Ms. J's house had 10 backyard chickens and 2 of them showed dyspnea on 17 October 2010, and finally died. Other chickens died suddenly. The weather was rather cool at the time the chickens were affected and Ms. J disposed of all the dead chickens by throwing them into the ditch immediately after they died.

Mrs. H's house (House No. 8) used to have a flock of 1,000 free-grazing ducks which were kept for 6 months until they were sold on 21 October 2010. The ducks were apparently healthy and did not show any signs of illness or abnormalities. Because the ducks had been sold, it was not possible to test them for avian influenza related to the outbreak at Ms. J's house.

<u>Question 4.6b:</u> From the data in the Table, which would be the first household you should visit to collect some more information?

- A. A's house
- B D's house
- C. EE's house
- D. J's house

Please provide an explanation to support your selection.

Answer:



<u>Question 4.7:</u> Please describe the outbreak in terms of animal, place and time, in words with supportive evidence. Are there any value chain components to this outbreak?

What is your hypothesis or theory about what the risk factors for this outbreak should be assessed further?

What control measure should be put into place?

| | Description | Supportive Evidence and Data | | | |
|-------------|-------------|------------------------------|--|--|--|
| Animal | | | | | |
| Place | | | | | |
| Time | | | | | |
| Value Chain | | | | | |

| Answer: | | |
|---------|-------------|------------------------------|
| | Description | Supportive Evidence and Data |
| | | |
| | | |
| | | |
| | | |
| | | |
| | | |

<u>Question 4.8</u>: From the descriptive information provided below, please <u>calculate the proportion of each risk factors among the HPAI confirmed positive case homes and summarize some possible risk factors for this outbreak.</u>

| House No. | Nam e | No. Poultr y | Adul t | Chic k | Proximit y to Road | Proximit y to Ditch | Cock Fightin g | New Introducti on | Free Ducks Passin g By |
|----------------|----------|--------------------|-----------|-----------|--------------------------|---------------------------|----------------------|-------------------------|---------------------------------|
| 1 | Α | 40 | 30 | 10 | 0 | 1 | 0 | 1 | 1 |
| 2 | В | 20 | 10 | 10 | 1 | 1 | 0 | 0 | 1 |
| 3 | С | 38 | 20 | 18 | 1 | 1 | 0 | 0 | 0 |
| 4 | D | 12 | 10 | 2 | 0 | 1 | 0 | 0 | 1 |
| 5 | E | 18 | 10 | 8 | 0 | 1 | 1 | 0 | 1 |
| 6 | F | 12 | 12 | 0 | 0 | 1 | 1 | 0 | 1 |
| 7 | G | 16 | 8 | 8 | 1 | 1 | 0 | 0 | 1 |
| 8 | Н | 22 | 12 | 10 | 0 | 1 | 0 | 0 | 1 |
| 9 | 1 | 17 | 10 | 7 | 0 | 1 | 0 | 0 | 0 |
| 10 | J | 10 | 2 | 8 | 0 | 1 | 1 | 1 | 0 |
| 11 | K | 10 | 2 | 8 | 0 | 1 | 1 | 0 | 0 |
| 12 | L | 16 | 8 | 8 | 0 | 1 | 1 | 0 | 0 |
| 13 | М | 18 | 10 | 8 | 1 | 1 | 0 | 0 | 0 |
| 31 | EE | 40 | 35 | 5 | 0 | 0 | 1 | 0 | 0 |
| 32 | FF | 14 | 10 | 4 | 1 | 0 | 0 | 1 | 0 |
| 33 | GG | 15 | 10 | 5 | 1 | 1 | 1 | 0 | 0 |
| 34 | НН | 16 | 10 | 6 | 0 | 1 | 0 | 0 | 0 |
| 35 | II | 15 | 5 | 10 | 0 | 1 | 1 | 0 | 0 |
| 36 | JJ | 10 | 8 | 2 | 0 | 1 | 0 | 0 | 0 |
| Proportio n | | | | | | | | | |

| Answers: | | | |
|----------|--|--|--|
| | | | |
| | | | |

Results from the laboratory were sent back on 5 November 2010. Intestinal parasites were reported and E. coli was cultured from liver samples.

On the same day, avian influenza (H5) was confirmed by real-time PCR and by egg inoculation. The virus sequences were similar to those for the virus reported in two neighbouring provinces two years before and which had high virulence for chickens.

We have decided that a cross-sectional study would be the most appropriate study design to identify associations between possible (hypothetical) risk factors and occurrence of cases of disease caused by HPAI H5 virus in household poultry.

Crude Odds Ratios with 95% confident intervals were calculated to evaluate possible risk factors for this outbreak. Note that positive cases are more likely to be located near to the drainage ditch than negative, non-affected homes in Village 1.

| Factors | Cases | Not Affected | Odds Ratios | 95% CI | |
|---|-------------|--------------|----------------|--------|------------|
| Proximity to road | Present | 6 | 16 | 0.43 | 0.13 - 1.4 |
| | Not present | 13 | 15 | | |
| Proximity to ditch | Present | 17 | 6 | 35.41 | 6.4-196.8 |
| | Not present | 2 | 25 | | |
| Having fighting cock | Present | 8 | 9 | 1.77 | 0.54 - 5.9 |
| activities (include entering fighting cock arena) | Not present | 11 | 22 | | |
| Introduction of new | Present | 3 | 4 | 1.27 | 0.25 - 6.4 |
| poultry | Not present | 16 | 27 | | |
| Having free grazing | Present | 7 | 9 | 1.36 | 0.4 - 4.6 |
| duck walking nearby the house | Not present | 12 | 21 | | |

<u>Question 4.10</u>: Are there any special studies that you would like to do to confirm the descriptive and analytic result?

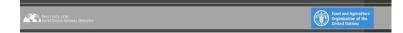
| Α | n | c | ١, | ۵ | r | |
|---|---|---|----|----|---|---|
| м | П | 5 | w | /₩ | | ı |

| Question 4.11: What would be your recommendation to prevent and control the outbreak egarding to you descriptive and analytic results? | |
|--|--|
| Answer: | |

Lesson 17.1 – Managing Outbreak Investigation Data: Collect Data and Create a Line Listing

| Estimated Lesson and Exercise Time | 1 hour and 15 minutes |
|------------------------------------|---|
| Participant Materials | Frontline ISAVET Lesson 17.1 Managing Outbreak and Investigation Data: Collect Data and Create a Line Listing Participant Guide.PDF MS Excel Training Videos, on creating a line list (3.1, 3.2), and outbreak histogram (13.1, and 13.2). |
| | Computer and Microsoft Word |
| | Pen or Pencil |
| _ | · |

In Service Applied Veterinary Epidemiology Training (ISAVET) Lesson 17.1: Managing Outbreak Investigation Data: Collect Data and Create a Line Listing



Learning Objectives

At the end of this lesson, you will be able to:

- 1. Find cases and complete a questionnaire systematically; and
- 2. Create a line list of all potential cases.

Data Collection and Recording During an Outbreak Collect high quality data at all stages of the outbreak investigation • Manual · Farm observations and records \cdot Electronic capture of data · Outbreak investigation forms $\circ \ Fit-for-purpose$ Laboratory forms o Spreadsheet format Community information Existing data o Passive and active surveillance data $\circ\, Value\, chain\, data$ 3 Outbreak Scenario: Classical Swine Fever (CSF) Suspected deaths due to CSF reported by farmer at index farm in Luwero local area, Uganda Sudden deaths, abortions, · CSF Field confirmed by RT-PCR at investigation initiated abortions, stillbirths, mummification in pigs on index farm in Luwero local area begins • Symptoms: fever, anorexia, conjunctivitis, skin cyanosis, diarrhea, norvous system tremors Objectives: 1.Confirm this is an outbreak national laboratory • Initiate planning for outbreak investigation 2.Determine the extent and magnitude of the outbreak • Samples submitted Plan: Design a questionnaire Conduct case finding and data laboratory collection Case Finding · Develop and follow the suspect case definition: • Unit of Interest: Pig Herd • Animal: Pig herd from commercial and backyard experiencing: • Sudden death OR * Abortion, still birth, mummification OR • At least 4 of the following clinical signs: fever, anorexia, conjunctivitis, skin cyanosis, diarrhea, nervous system tremors

Reference: Frontline ISAVET

Place: Luwero local area, Uganda
 Time: April 15 – July 28, 2018
 A confirmed case of CSF

A suspect case that is confirmed by RT-PCR
 Initiate Active Case Finding – find it using the suspect case definition, don't miss it!

Need to have access to standard questionnaire Need to have access to standard questionnaire form appropriate for the objectives of the investigation Generally forms should be available at the local area level Questionnaires can be modified with the input of an epidemiologist to meet the objectives of the investigation Develop data collection tools Create sections of an outbreak investigation form: 1. Background information on the farm/village/abattoir 2. Management and demographics 3. Potential risk factors 4. Beta (pre-) test the questionnaire before using it (if it is new)

Data Collection and Recording During an Outbreak Investigation

- · Use your direct observations
- · Take samples
- Tracing and following movement in and out of a population
- Consider movement of animals, people, equipment, etc.
- · Use a pretested Questionnaire
- Collect data systematically and in a standardised way for every location
- Develop a line listing of data collected



Reference: Google Images

| Location | No. Sick | No. | Morbidity | Disease Period | Share | Swill |
|----------|-------------|-----|-----------|-------------------|-------|-------|
| Dist. 4 | 14 | 16 | 88% | April | 1 | 0 |
| Dist. 8 | 1 | 14 | 7% | April | 1 | 0 |
| Dist. 4 | 2 | 2 | 100% | May | 1 | 0 |
| Dist. 4 | 14 | 16 | 88% | May | 1 | 0 |
| Dist. 4 | 2 | 25 | .8% | May | 1 | 0 |
| Dist.12 | 5 | 66 | 8% | May | 0 | 0 |
| Dist. 12 | 3 | 84 | 4% | May | -1 | .1 |
| Dist. 8 | 7 | 26 | 27% | May | 1 | 0 |
| Dist. 11 | 11 | 60 | 18% | June | 0 | 0 |
| Dist. 4 | 5: | 7 | 71% | July | 1 | .0 |
| Dist. 8 | 3 | 52 | 6% | July | .0 | 0 |

How to Administer the Questionnaire (Refer to SOP for Conducting Surveys and KAP Studies)

Introduce yourself to the owner and explain the purpose of your visit

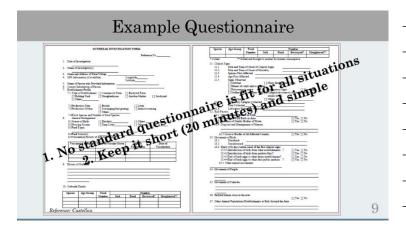
Make sure you are interviewing the person who manages the animals

Ask some general questions about the owner's disease concerns

Stand or sit at the same level as the person you are interviewing and lister

Deliver each question in the same way for each interview

End by asking the owner if he/she has any questions, concerns or requests



Create a Line List By Hand

- Refer to YouTube Training Video $3.1\,$
- · Enter the variable names in order across the top row of a paper form.

- Variables collected on each location include: Herd name, location, %morbidity, disease period, sharing a boar, swill feeding, visits from traders and use of CSF vaccine $\,$
- · You will enter selected data from the questionnaires used in your interviews under the titles and then input the data into Excel or share this data with the national epidemiologist

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Create a Line List Using Excel

- 1. Refer to YouTube Training Video 3.2
- Open and a blank spreadsheet in MS Excel and save with the file name: "Outbreak" $\,$
- On the first tab (at the bottom) create a data dictionary using including each variable in the questionnaire: - Farm reference number

 - Location Latitude and longitude
- 4. Name the tab (right click > rename) "Dictionary"
- Start a new spreadsheet and rename "Data"
- Enter the variable names in order across the first row of the "Data spreadsheet" $\,$
- You will enter selected data from the questionnaires used in your interviews into the "Data" spreadsheet and then merge the data with your colleagues \mathbb{I}_1

The Uses of a Line List

- $\dot{\,\,}$ To tabulate preliminary information on cases, such as demographic information, test results and exposure risks
- To rapidly summarise the outbreak in terms of animal, place and time $\dot{}$ Example: The spatial and temporal distribution of affected pig farms in Luwero local area
- $\boldsymbol{\cdot}$ To determine further refine symptoms and other parameters of the case
 - Example: More precise symptoms based on frequency of occurrence
- ${}^{\textstyle \cdot}$ As the investigation progresses, the line listing is the basis for analysis of the outbreak data
 - · Example: Create an outbreak histogram in time

Example: Classical Swine Fever Line Listing

| Herd Name | Location | % ! | Morbidity | Disease Period | share boar | swill feed | trader | no vaccine |
|-----------------|-----------|-----|-----------|-------------------|---------------|------------|--------|------------|
| N | SDist. 4 | 88 | (14/16) | April | 1 | 0 | 1 | 1 |
| A | SDist. 8 | 7 | (1/14) | April | 1 | 0 | 1 | 1 |
| s | SDist. 4 | 100 | (2/2) | May | 1 | 0 | 0 | 1 |
| w | SDIst. 4 | 88 | (14/16) | May | 1 | 0 | 1 | 1 |
| С | SDIst. 4 | 8 | (2/25) | May | 1 | 0 | 1 | 0 |
| Y-F (confirmed) | SDist.12 | 8 | (5/66) | May | 0 | 0 | 1 | 1 |
| H * | SDist. 12 | 3 | (3/84) | May | 1 | 1 | 1 | 0 |
| Ac | SDist. 8 | 26 | (7/26) | May | 1 | 0 | 1 | 1 |
| So | SDist. 11 | 18 | (11/60) | June | 0 | 0 | 1 | 0 |
| Su* | SDist. 4 | 71 | (5/7) | July | 1 | 0 | 1 | 1 |
| Sop* | SDIst. 8 | 5 | (3/52) | July | 0 | 0 | 1 | 1 |
| Percentage | | | | | 72.73% | 9.09% | 90.91% | 72.73% |

Exercise 23a: Collect and Enter Data in a Line List

- Exercise 23a: 30 minutes
 - · Each of you should have a questionnaire, work in pairs take turns asking questions to each other.
- $\boldsymbol{\cdot}$ Using the data you will create a line listing generate, organise the data, assess for quality, perform calculations, display and interpret findings of the data.

Exercise 23a - Collect and Enter Data in a Line List

Description of Exercise:

A questionnaire will be provided for individuals to role-play the field investigator and the farmer. Information gleaned will be used to develop a frequency histogram in MS. Excel. Should you have any questions over the exercise, please ask a trainer for clarification before during, and after the exercise.

Allotted Time: 30 minutes

Organisation of Group Work:

This exercise will be done independently.

Exercise Objective(s):

- 1. Each person will receive a questionnaire to fill in asking questions of a classmate from the villagers.
- 2. Develop a line listing in MS Excel. Exercise Components and Structure:
- 3. Questionnaire

MS Excel Training Videos, on creating a line list (3.1, 3.2),

Materials, Data or Information:

MS Excel

Expected Outputs and Deliverables of Each Participant:

- Experience collecting data.
- Line listing.

OUTBREAK INVESTIGATION FORM

| Fa | rm Reference No |
|----------------|--|
| 1. | Date of Investigation |
| 3. | Name of Investigator(s) Name and Address of Farm/Village GPS Information (if available) a. Latitude |
| 5. 6. 7. | b. Longitude Name of Person who Provided Information Contact Information of Person Establishment Profile 1.1 Type of Establishment |
| | [] Commercial Farm [] Backyard Farm [] Holding Yard [] Slaughterhouse [] Auction Market [] Stockyard [] Others |
| | 1.2 Production Type [] Broiler [] Layer |
| | 1.3 Production System [] Scavenging/free grazing [] Intensive rearing [] Others |
| 8. | 7.4 Flock Species and Number of Each Species General Management 8.1 Source of Birds [] Hatchery [] Others 8.2 Housing System [] Deep Litter [] Slatted 8.3 Feed Types |
| | 8.4 Feed Source(s) 8.4 Vaccination History of Affected Flocks |

| Vaccinated | Against: N | ame of Vac | cine Given | Age at Vaccination | | Dat | e of Vaccination |
|---------------|--------------------------------|---------------|---------------|--------------------|----------|-------|------------------|
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| 9. History of | f Outbreak | | ı | | | | |
| | | | | | | | |
| | | | | | | | |
| 10. Outbrea | k Details | | | | | | |
| Producti | Age | Total | | | Number | | |
| on Type | (weeks) | Numbe r | Morbidit y | Mortalit y | Destro | yed* | Slaughtered** |
| Meat | | | | | | | |
| Eggs | | | | | | | |
| Dual | | | | | | | |
| | | | | | | | |
| * Culled | ** I | Killed and b | rought to m | arket for hur | nan cons | umpti | on |
| 11. Clinical | Signs | | | | | | |
| 11.1 | Date of Ons | set of Clinic | al Signs | | | | |
| 11.2 | Date of Ons | set of Morta | ality | | | | |
| 11.3 | Species Firs | st Affected | | | | | = |
| 11.4 | Age First A | ffected | | | | | _ |
| 11.5 | Signs Obser | rved | | | | | |
| | [] Diarrhea | ı | | | | | |
| | [] Many de | aths over 3 | days | | | | |
| | [] Edema o | f comb and | or wattles | | | | |
| | [] Reluctan | ice to move | | | | | |
| | [] Respirat | ory signs | | | | | |
| | [] Sneezing | g | | | | | |
| | [] Sudden (| deaths of m | any birds | | | | |
| | [] Congesti | on/cyanosis | s of comb, wa | ittles or shar | ks/hocks | 3 | |
| | [] Eye Opa | city | | | | | |
| | $[\]$ Others $_$ | | | | | | |
| 11.6 | Laboratory | Samples C | ollected | | | | |
| 11.7 | Date Collec | ted | | | | | |
| 11.8 | = | Reference | Number | | | | |
| 12. Risk Fact | | , | | | | | |
| | er Animals Pr sence of Wild | | | | | | [] Yes [] No |

| | | Presence of Nearby Bodies of Water Disposal and Management of Manure | [] Yes | [] No | |
|------|-------|--|--------|--------|-------------|
| | 19 5 | Close to Border of AI-Affected Country | | [] Yes | |
| 13 | | ement of Birds | | [] res | [] NO |
| 10. | 13.1 | Traceback | | | |
| | 13.2 | Traceforward | | | |
| | | Thirty (30) days before onset of the first clinical signs | | | |
| | | 13.4.1 Introduction of birds from other establishments? | | []Yes | [] No |
| | | 13.4.2 Introduction of birds from markets/fairs? | | [] Yes | |
| | | 13.4.3 Exit of birds/eggs to other farms/establishment? | | [] Yes | |
| | | 13.4.4 Exit of birds/eggs to other fairs/public markets? | | [] Yes | |
| | | 13.5 Other animal movements | | | |
| 14. | Move | ement of People | | | |
| 16. | Relat | ement of Vehicles red human cases in the area [] Yes [] No r Animal Populations/Establishments at Risk Around the Ar | ea | | |
| 18. | Addi | tional Observations/Comments Other than Indicated | | | |
| 17. | Мар | and Photos of the Area | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| Sign | natur | e of Investigator: | | | |

Lesson 17.2 - Managing Outbreak Investigation Data

| Estimated Lesson and Exercise Time | 1 hour |
|------------------------------------|---|
| Participant Materials | Frontline ISAVET Lesson 17.2 Managing Outbreak and Investigation Data Participant Guide.PDF MS Excel Training Videos, on creating a line list (3.1, 3.2), and outbreak histogram (13.1, and 13.2). |
| | Computer and Microsoft Word |
| | Pen or Pencil |

| In Service Applied Veterinary Epidemiology Training (ISAVET) | |
|---|---|
| Lesson 17b: Managing Outbreak Investigation Data | |
| | |
| PROTECTION STREET DAYS | Food and Agriculture Organization of the United Nations |

Learning Objectives

At the end of this lesson, you will be able to:

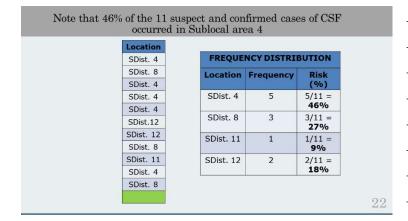
- Calculate measures of central tendency (mean, median and mode);
- Calculate and interpret measures of disease occurrence (rates, ratios, proportion, incidence and prevalence);
- 3. Create a standard outbreak histogram and interpret the results.

| Herd Name | Location | % Morbidity | Disease Period | share boar | swill feed | trader | no vaccino |
|-----------------|-----------|-------------|-------------------|---------------|------------|--------|------------|
| N | SDist. 4 | 88 (14/16) | April | 1 | 0 | 1 | 1 |
| A | SDist. 8 | 7 (1/14) | April | 1 | 0 | 1 | 1 |
| S | SDist. 4 | 100 (2/2) | May | 1 | 0 | 0 | 1 |
| w | SDist. 4 | 88 (14/16) | May | 1 | 0 | 1 | 1 |
| С | SDist. 4 | 8 (2/25) | May | 1 | 0 | 1 | 0 |
| Y-F (confirmed) | SDist.12 | 8 (5/66) | May | 0 | 0 | 1 | 1 |
| H * | SDist. 12 | 3 (3/84) | May | 1 | 1 | 1 | 0 |
| Ac | SDist. 8 | 26 (7/26) | May | 1 | 0 | 1 | 1 |
| So | SDist. 11 | 18 (11/60) | June | 0 | 0 | 1 | 0 |
| Su* | SDist. 4 | 71 (5/7) | July | 1 | 0 | 1 | 1 |
| Sop* | SDIst. 8 | 5 (3/52) | July | 0 | 0 | 1 | 1 |

| easures of Central Tendency: By Hand | | | |
|---|-----------------------|--|------|
| easures of Central Tendency: By Hand | Mean | 67 / 11 | 6.09 |
| Mean, Median, Mode, Range, Minimum, Maximum, Sum, Count | Standard Error | $\frac{1}{\left[(z-\bar{z})\right]} \underbrace{\sum_{(1-\bar{z})^2} \underbrace{\frac{\left[\sum_{(1-\bar{z})^2} (z-\bar{z})\right]}{\sum_{(1-\bar{z})^2}}}_{}$ | 1.45 |
| | Median | Middle value of the 11 values is 5 | 5 |
| | Mode | Most common value is 14 | 14 |
| | Standard Deviation | $S = \sqrt{\frac{\sum (x_i - \overline{x})^2}{n-1}}$ | 4.81 |
| | Range | Lowest – Highest Values | 13 |
| | Minimum | Lowest Value | 1 |
| | Maximum | Highest Value | 14 |
| | Sum | Add all Values | 67 |
| | Count | No. of values | 11 |

| | Statistic | Morbidity | |
|---|-----------------------|-----------|-----|
| Refer to YouTube Training Video 5 for instructions Mean, Median, Mode, Range, Minimum, Maximum, Sum, Count | Mean | 6.09 | |
| | Standard Error | 1.45 | |
| | Median | 5.00 | |
| | Mode | 14.00 | |
| | Standard Deviation | 4.81 | |
| | Range | 13 | |
| | Minimum | 1 | |
| | Maximum | 14 | |
| | Sum | 67 | |
| | Count | 11 | 1 (|

Measures of Disease Frequency: Ratio Comparison Among Ratio Suspected Farms Share boar: Swill Feed 8:1 Share boar: Trader 0.8



Cumulative Incidence Risk: (Approximate Method)

- · In real terms the risk per 1,000 pigs is 0.18 x 1000 = 180 cases per 1,000 pigs during this time period
- Conclusion: 67 / 368 pigs at risk = 0.18 cumulative incident risk occurred between April 15 and July 31, 2018.

| Herd Name | No. Sick | Total | |
|------------------------------|-------------|------------|--|
| N | 14 | 16 | |
| A | 1 | 14 | |
| S | 2 | 2 | |
| W | 14 | 16 | |
| C | 2 | 25 | |
| Y-F (confirmed) | 5 | 66 | |
| H * | 3 | 84 | |
| Ac | 7 | 26 | |
| So | 11 | 60 | |
| Su* | 5 | 7 | |
| Sop* | 3 | 52 | |
| Total Affected | 67 | 368 | |
| Cumulative Incidence Risk | 0.18 | 0.18 cases | |

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Measuring Disease Period Prevalence

 NOTE: The denominator is reset at the beginning of each month based on the number of healthy pigs that remain.

| Time Period | No. Sick | Population at Risk | Period Prevalence |
|----------------|-------------|-----------------------|----------------------|
| April | 15 | 368 | 15/368 = 4% |
| May | 33 | 353 (368-15) | 33/353 = 9 % |
| June | 11 | 320 (353-33) | 11/320 = 3 % |
| July | 8 | 309 (320-11) | 8/309 = 3 % |
| Total | 67 | 339 | 20 % (67/339) |

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Crude Morbidity, Crude Mortality and Case Fatality Risk

| % Morbidity | | |
|-------------|---------|--|
| 88 | (14/16) | |
| 7 | (1/14) | |
| 100 | (2/2) | |
| 88 | (14/16) | |
| 8 | (2/25) | |
| 8 | (5/66) | |
| 3 | (3/84) | |
| 26 | (7/26) | |
| 18 | (11/60) | |
| 71 | (5/7) | |
| 5 | (3/52) | |
| (6 | 7/368) | |

- · NOTE: No mortality was reported by farmers that were interviewed
- Crude morbidity risk = # sick / total population at risk

= 67 / 368 = 18%

• Crude mortality risk = # dead / total population at risk

= 0 / 368 = 0%

- Case fatality risk = # dead / # sick

= 0 / 67 = 0%

nce: Regional FETPV

Describe Disease Occurrence Over Time Number of New Pig Herds Suspected or · Create an outbreak histogram Confirmed with Classical Swine Fever, ${\boldsymbol{\cdot}}$ Plot as frequency histogram April to July 2018 · y-axis = number of cases in population (animal/farm) in population • x-axis time of clinical sign observed/death • Ideal interval of x-axis 1/4-1/3 of incubation period - Hours (food poisoning) $\,$ - Days (influenza) - Months (rabies) May April MONTH

Exercise 23b: Outbreak histogram, Measures of central tendency and disease occurrence.

- · Exercise 23b: 60 minutes
- With the data provided, create an outbreak histogram, calculate measures of central tendency and measures of disease occurrence.

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In Summary...you have learned

- Questionnaires are flexible tools to undertake investigations on a wide range of topics
- · How to create
 - · a line list of all potential cases using Excel or by hand
- · Calculate
- * measures of central tendency (mean, median and mode);
- and interpret measures of disease occurrence (rates, ratios, proportion, incidence and prevalence);
- ${\boldsymbol{\cdot}}$ To create a standard outbreak histogram and interpret the results.

| ISAVE | T Contributing Universities | |
|--------------|---|----|
| Partners | TEXAS ARM. TEXAS ARM. ATEXAS ARM. AGRILIFE | |
| Contributors | MAKEBUR UNIVERSITY | 29 |

Exercise 23 - Outbreak Histograms

Description of Exercise:

A questionnaire will be provided for individuals to role-play the field investigator and the farmer. Information gleaned will be used to develop a frequency histogram in MS. Excel. Should you have any questions over the exercise, please ask a trainer for clarification before during, and after the exercise.

Allotted Time: 1 hour

Organisation of Group Work:

This exercise will be done independently.

Exercise Objective(s):

- 1. Each person will receive a questionnaire to fill in asking questions of a classmate from the villagers.
- 2. Using the data provided, organise the data, assess for quality, perform calculations, display and interpret findings of the data.
- 3. Include: line listing, an outbreak histogram, measures of central tendency and measures of central tendency and measures of disease occurrence.

Exercise Components and Structure:

- 1. Questionnaire
- 2. MS Excel Training Videos, on creating a line list (3.1, 3.2), and outbreak histogram (13.1, and 13.2).
- 3. Development of a frequency histogram

Materials, Data or Information:

1. MS Excel

Expected Outputs and Deliverables of Each Participant:

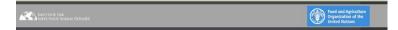
- 1. Experience collecting data.
- 2. Line listing, an outbreak histogram, measures of disease occurrence.

Lesson 18 - Follow-up Investigations and Special Studies

| Estimated Lesson and Exercise Time | 1 hour and 30 minutes |
|------------------------------------|---|
| Participant Materials | Frontline ISAVET Lesson 18 Follow-up Investigations and Special Studies Participant Guide Version 4.doc |
| | Computer and Microsoft Word |
| | Pen or Pencil |

In Service Applied Veterinary Epidemiology Training (ISAVET)

Lesson 18: Follow-up Investigations and Special Studies

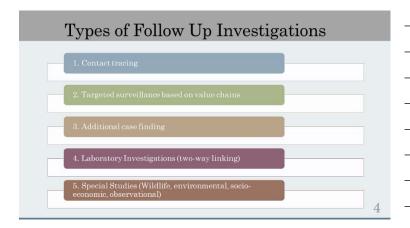


Learning Objectives

At the end of this lesson, you will be able to:

- 1. Describe the purpose and types of follow-up animal disease investigations related to an initial outbreak investigation; and
- 2. Describe the types of special studies that contribute to an outbreak investigation.

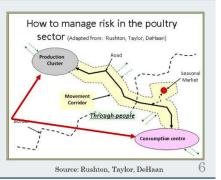
Purpose of Follow-Up Investigations What is the purpose of a follow-up investigation? 1. To trace disease transmission into and from an affected location 2. To target surveillance based on high-risk points along the value chain 3. To create an updated timeline of new cases discovered 4. To perform laboratory investigations of affected and unaffected locations



Review questionnaire and line listing data to look for connections to the affected Farm B. Draw diagrams linking animal movement including epidemiological information about animal-place-time. Part of the face of the lines are lined as a line line listing between the lines are lined as a line lines are lines as a lines are lines are lines are lines as a lines are lines a

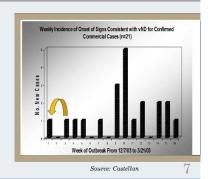
2. Targeted Surveillance Along the Value Chain

- · Target surveillance based on the frequency, direction and volume of movement along the value chain.
- Targeted surveillance along key risk points along the value chain includes:
- Production centers Trading channels
- Sales yards
- Live animal markets and slaughter facilities
- · Take immediate action
- Risk communication with stakeholders



3. Additional Case Finding

- · When a case is discovered that occurred earlier than previously thought, it is important to open new lines of investigation.
- · Do additional case finding related to the earlier cases discovered
- · What would you do next?



4. Laboratory Investigations

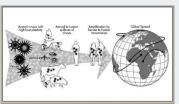
Collect the same samples from all locations while conducting case finding

- Confirm the presence or absence of the disease agent
- Identify the molecular subtype at each location and in each area e.g. ASF, HPAI, FMD

Some examples of special studies: Wildlife studies: Sampling wild birds, bats, deer, wildebeest, jackals, primates and hyaenas, etc. Retrospective analysis of existing surveillance data Environmental studies: Socio-economic studies: Antimicrobial resistance studies Socio-economic studies: Cross-sectional and case-control studies Observational Studies: Socio-economic studies assessing price fluctuations along the value chain, impact and benefit/cost analysis

Wildlife Studies

- · Recall that 75% of EID originate in wildlife e.g. rabies
- Include ecological approaches based on a One Health approach that include domestic animals, humans and wildlife
- Consult with a wildlife expert for all diseases involving domestic and wild animals



Source Johnson et al. Nature, Special Reports, 2015.

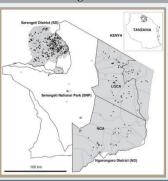


Source: Google Images

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Example: Rabies virus variant is maintained in spotted hyaenas in the Serengeti

 Rabies is transmitted to several 'spillover' hosts that might be considered target populations of concern, including humans (Knobel et al., 2005), endangered wildlife (Randall et al., 2006; Vial et al., 2006) and livestock



Environmental Studies

Environment and climate change impacts the emergence of infectious diseases $\,$

These studies measure temperature, altitude, soil type, deforestation, human density, housing, water, and feed or food sources etc.

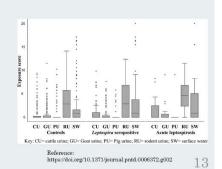
Analysis of water, feed, soil and food products can provide useful information linking exposure sources and affected animals and people

The analysis could be bacteriological (anthrax), virological (avian influenza) or toxicological (aflatoxin)

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Example: Leptospirosis in Urine and Surface Water

- Leptospirosis is an important zoonotic disease that is associated with livestock and rodent urine as well as surface water contamination of rivers and streams
- All possible sources need to be sampled to determine possible sources of infection for humans
- The graph shows the participant scores of humans exposed to animal urine and surface water, northern Tanzania, 2012–14 (N = 844).



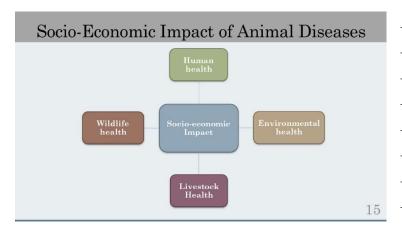
Socio-economic Studies

Socio-economic studies can be used to measure the following:

- ${\small 1.}\ Cost-benefit\ analysis$
- 2. Economic impact on animal production and livelihoods
- 3. Price and market changes
- 4. Impact on import/export trade
- 5. Costs to humans and the environment

| | | | (Contraction) |
|--------------------|---------------|---------------|---------------|
| | Solution A | Solution B | Solution |
| Total Costs | €10,000 | €15,000 | €20,000 |
| Total Benefits | €12,000 | €19,000 | €23,000 |
| Cost-Benefit ratio | 1.20 | 1.27 | 1.15 |

Source: Google Images



5. Follow-up Observational Studies

Collect high
quality
data from
all locations
visited
while
conducting
case finding

Cross-sectional studies

- · Most commonly done
- Collect risk factor data at the same time we collect laboratory samples from all locations
- Case-control studies
- Once we find a confirmed positive case, we compare with confirmed negative locations based on their responses to risk factor questions

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Exercise 24: Follow-Up Investigations

Instructions:

- 1. The exercise will take 45 minutes to complete.
- 2. Form into 3 groups.
- 3. Recommend which type of follow up investigation(s) or special studies would be useful for the following disease outbreaks.
 - a) Leptospirosis in cattle, wildlife and humans
 - b) Aflatoxicosis in poultry feed
 - c) Trichinellosis in pigs and humans
- 4. Explain the objective of each investigation and study.
- 5. Explain the expected contribution/outcome of these investigations and special studies.

In Summary...

- The types of follow-up animal disease investigations include:
 - 1. Contact tracing
- 2. Targeted surveillance based on value chains
- 3. Additional case finding
- Laboratory Investigations (two-way linking)
 Special Studies (Wildlife, environmental, socio-economic, observational)
- The types of special studies include:

 1. Wildlife studies
- 2. Environmental studies
- 3. Socio-economic studies
- 4. Observational studies

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Exercise 24 – Follow-up Field Investigations

Description of Exercise:

This exercise will focus on different types of follow-up investigations and making recommendations for which type of follow-up investigation(s) or special studies should be conducted for various types of disease outbreaks. Should you have any questions over the exercise, please ask a trainer for clarification before during, and after the exercise.

Allotted Time: 45 minutes

Organisation of Group Work: Form into three groups, one for each disease.

Exercise Objective(s):

- 1. Recommend which type of follow-up investigation(s) or special studies would be useful for the following disease outbreaks
 - Leptospirosis in cattle and humans
 - Aflatoxicosis in poultry in feed
 - Trichinellosis in pigs and humans
- 2. Explain the objective of each investigation and study
- 3. Explain the expected contribution/outcome of these investigations and special studies
- 4. Choose a follow up investigation including:
 - Contact tracing investigations forward and tracing backwards
 - Targeted risk-based surveillance at high risk points along the value chain
 - When the timeline for the index case is revised through the discovery of an earlier outbreak
 - Observational studies cross-sectional and case control
 - Other
 - 5. Choose a special study including:
 - Wildlife studies
 - Environmental studies
 - Socio-economic studies
 - Other

Exercise Components and Structure:

- 1. Form into three (3) groups, one for each disease.
- 2. Create and complete an electronic table for each disease noted above.

Materials, Data or Information:

1. Microsoft Word and PowerPoint

Expected Outputs and Deliverables of Each Participant:

- 2. Recommendations with justification
- 1. Using Microsoft Word or PowerPoint, create and complete and electronic table for your specific groups' disease.

| Group A: Leptospirosis in Cattle and Humans | | | |
|---|------------|-------------------|--|
| Type of Follow Up Investigations | Objectives | Expected Outcomes | |
| Type of Special Studies | Objectives | Expected Outcomes | |

| Group B: Aflatoxin in Poultry | | | |
|-------------------------------------|------------|-------------------|--|
| Type of Follow Up Investigations | Objectives | Expected Outcomes | |
| | | | |
| Type of Special Studies | Objectives | Expected Outcomes | |
| | | | |
| | | | |

| Group C: Trichinellosis in Pigs and Humans | | | |
|--|------------|-------------------|--|
| Type of Follow Up | Objectives | Expected Outcomes | |
| Investigations | | | |
| | | | |
| | | | |
| | | | |
| | | | |
| | | | |
| Type of Special Studies | Objectives | Expected Outcomes | |
| | | | |
| | | | |
| | | | |
| | | | |
| | | | |
| | | | |

Lesson 19 - Surveillance Situation Assessment for Prevention and Control

| Estimated Lesson and Exercise Time | 1 hour and 30 minutes |
|------------------------------------|---|
| Participant Materials | Frontline ISAVET Lesson 19 Surveillance Situation Assessment for Prevention and Control Participant.PDF |
| | Computer and Microsoft Word |
| | Pen or Pencil |

In Service Applied Veterinary Epidemiology Training (ISAVET)

Lesson 19: Surveillance Situation Assessment for Prevention and Control

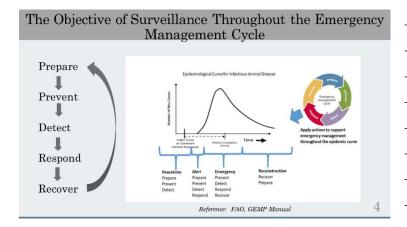
| INSTITUTE FOR ANIMAL DISSASS | Food and Agriculture Organization of the United Nations |
|------------------------------|---|
| | |

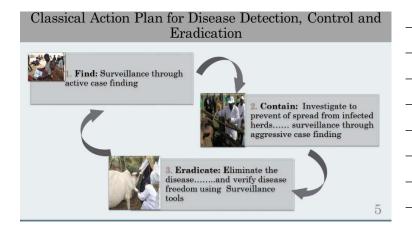
Learning Objectives

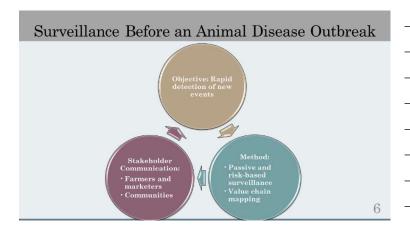
At the end of this lesson, you will be able to:

- 1. Review of basic surveillance concepts;
- 2. Explain the classical action plan for disease detection, control and eradication; and
- 3. Explain the importance of conducting surveillance **before**, **during** and **following** an animal disease outbreak investigation to determine prevention and control efforts.

What is Surveillance? - Surveillance is the systematic ongoing collection, collation and analysis of data and the timely dissemination of information to those who need to know so that action can be taken. (OIE Terrestrial Animal Health Code, Salman) - Desired outcomes of animal disease surveillance: Prevent Control Eradicate References: Google images; USDA; OIE







Critical Actions to Take Before An Outbreak

Conduct active and passive surveillance

- ullet Ensure high sensitivity for disease detection e.g. broad case definition
- \bullet Good performance of the surveillance e.g. daily evaluation of data
- Type of surveillance suitable for the situation i.e. sentinel surveillance-vector borne disease, risk-based surveillance-diseases with clear risk factors
- Prepare to conduct zero-reporting should always be used as soon as an outbreak begins

Create Value Chain Maps

- Based on stakeholder input
- Objective: Identify high risk points in the value chain on how to prepare before an outbreak

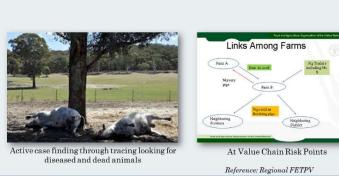
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Surveillance During an Animal Disease Outbreak Objective: Active aggressive case finding Measure disease incidence Stakeholder Communication: Government officials Field teams Farmers and marketers Communities Objective: Active aggressive case Finding Delivers Plaily zero reporting Daily update of outbreak curve 8

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How to Conduct Surveillance During an Animal Disease Outbreak



Aggressive case finding • Enhanced surveillance to find all possible cases as early as possible Movement controls • Adjust the size of infected, restricted and control areas • Communicate and update stakeholders daily Investigate locations in contact with suspect and confirmed cases • Direct tracing and at high risk points along the value chain Humane culling and disposal

 ally summary of disease situation and trends
 Daily briefing and debriefing including update of zero reporting and the outbreak histogram

Surveillance Following an Animal Disease Outbreak Objective: Estimate freedom from disease Maintain prevention and control efforts Stakeholder Communication: Government officials Field teams Farmers and marketers Communities Method: Prepare an after action report Case definition that is both sensitive and specific Grid based surveillance Manage risks identified during the outbreak

Critical Actions to Take Following An Outbreak Prepare an after action report Apply lessons learned during the outbreak to improve prevention and control measures – Refer to local area Outbreak After Action Report SOP Estimate freedom from disease Conduct grid based surveillance Maintain prevention and control efforts Ensure that farmers, traders and all primary stakeholders maintain prevention and control efforts using biosecurity best practices Case definition that is both sensitive and specific Detect disease early while avoiding false positives that may affect trade and movement Manage risks identified during the outbreak Utilise risk factors identified to prevent future re-entry and spread of disease Stakeholder Communication Communicate findings and lessons from the after action report Have field teams improve procedures and field practices

Examples: Surveillance Required Following an Animal Disease Outbreak

· Minimum requirements for a country/region to be declared free from disease (OIE Animal Health Code)

| | No Vaccination | | Vaccination | |
|---------------------------------|--------------------|--------------------------------|--------------------|---|
| DISEASE | 1st Recognition | After Outbreak | 1st Recognition | After Outbreak |
| Foot and mouth disease (FMD) | 12 months | 3 months (stamping out) (a) | 24 months | 12 months (stamping out) 24 months (no stamping out) |
| Classical swine fever (CSF) | 24 months | 6 months (stamping out) | 24 months | 12 months (stamping out) |
| African swine fever (ASF) | 36 months | 12 months (stamping out) | | |
| Avian influenza (AI) | 36 months | 6 months (stamping out) | 36 months | 6 months (stamping out) |

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Exercise 25: Surveillance Situation Assessment

- 1. This exercise will take 45 minutes.
- Explain the use of surveillance and how it affects prevention and control efforts before, during and following an animal disease outbreak event.
 - $\cdot \ Group \ 1: \ Your \ local \ area \ is \ located \ adjacent to \ a \ neighboring \ country \ that \ is \ experiencing \ large \ outbreaks \ of \ Rift \ Valley \ Fever \ (RVF). \ What \ are the \ surveillance \ objectives \ and \ methods \ you \ will \ use \ to \ detect \ the \ disease \ early? \ What \ actions \ will \ be \ taken?$
 - ${\boldsymbol \cdot}$ Group 2; RVF has been found in a beef herd from your local area. What are the surveillance objectives and methods you can use to find the disease and estimate the incidence and prevalence of the disease in your local area? What actions will be taken?
 - \cdot Group 3: No new cases of RVF have been reported during the past 2 weeks. What are the surveillance objectives and methods you can use to demonstrate either control or freedom from RVF in your local area? What actions will be taken?

In Summary...

 $Surveillance \ can \ be \ used \ differently \ before, \ during \ and \ animal \ disease \ outbreak...$

- · Before an outbreak:
 - Detect disease early and report it rapidly
- · During an outbreak:
 - $\operatorname{Aggressive}$ case finding
 - Risk based surveillance along the value chain
 - Estimate incidence and prevalence of disease $\,$
- $\cdot \ \underline{\text{After an outbreak}} :$
 - Prepare an after action report
 - Establish freedom from disease
 - Manage risk identified during outbreak investigation
 - $\hbox{-}\ Maintain\ effective\ surveillance}$

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Partners Contributors Contributors

Exercise 25 - Surveillance Situation Assessment

Description of Exercise:

Explain the use of surveillance and how it affects prevention and control efforts before, during and following an animal disease outbreak event. Should you have any questions over the exercise, please ask a trainer for clarification before during, and after the exercise.

Allotted Time: 45 minutes

Organisation of Group Work:

• Work in three groups for this exercise, one per each scenario.

Exercise Objective(s):

- 1. Explain the surveillance objectives in the scenario
- 2. Explain the methods that can be used to find disease in the scenario
- 3. Explain what actions that will be taken in each scenario

Exercise Components and Structure:

- 1. Form into three groups, one for each scenario
- 2. Answer the questions for each scenario

Explain the use of surveillance and how it affects prevention and control efforts before, during and following an animal disease outbreak event.

Materials, Data or Information:

- 1. MS Word
- 2. Paper and pen

Expected Outputs and Deliverables of Each Participant:

- 1. Surveillance objectives, methods of case findings and recommended actions for control and prevention.
- 2. Use Microsoft word or power point to record your responses.

Scenario 1: Your local area is located adjacent to a neighboring country that is experiencing large outbreaks of Rift Valley Fever (RVF).

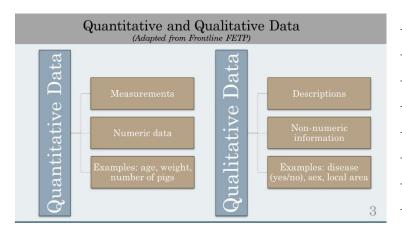
| a.) | What are the surveillance objectives and methods you will use to detect the disease early? |
|--------|---|
| b.) | What actions will be taken? |
| Scenar | rio 2: RVF has been found in a beef herd from your local area. |
| a.) | What are the surveillance objectives and methods you can use to find the disease and estimate the incidence and prevalence of the disease in your local area? |
| | |
| b.) | What actions will be taken? |
| Scenar | rio 3: No new cases of RVF have been reported during the past 2 weeks. |
| a.) | What are the surveillance objectives and methods you can use to demonstrate either control or freedom from RVF in your local area? |
| b.) | What actions will be taken? |
| | |

Lesson 20 – Display Outbreak Investigation Findings and Make Relevant Recommendations for Prevention and Control

| Estimated Lesson and Exercise Time | 1 hour and 30 minutes |
|------------------------------------|---|
| Participant Materials | ISAVET Lesson 20 Display Outbreak Investigation Findings and Make Relevant Recommendations for Prevention and Control Participant Guide Version 4.PDF Computer and Microsoft Word |
| | Pen or Pencil |

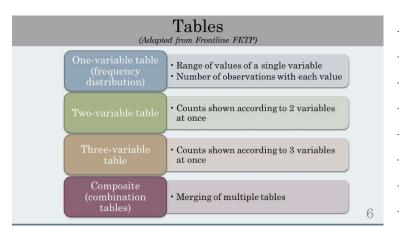
In Service Applied Veterinary Epidemiology Training (ISAVET) Lesson 20: Display Outbreak Investigation Findings and Make Relevant Recommendations for Prevention and Control

At the end of this lesson, you will be able to: 1. Display and interpret outbreak investigation findings using tables, graphs and maps including creating a spot map by hand. 2. Make relevant recommendations for prevention and control.



| Outbreak investigation data is most often displayed using: | 1. Tables |
|---|---------------------------------|
| | 2. Graphs and bar charts |
| | 3. Maps |
| | 4. Timelines and other displays |

| Quantitative Data Example: isplay and Interpretation of Count Data | Week | No. New RT- PCR Positive H5N1 Samples |
|--|-------|---|
| | 1 | 66 |
| G 6 1 4 4 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 | 2 | 60 |
| Confirmed cases of HPAI H5N1 have occurred during the past | 3 | 68 |
| three months | 4 | 83 |
| | 5 | 297 |
| The number of RT-PCR samples | 6 | 136 |
| detected each weeks 1-13 are | 7 | 115 |
| recorded in the right hand column | 8 | 48 |
| column | 9 | 36 |
| We will next make a histogram | 10 | 21 |
| graph of the data | 11 | 10 |
| | 12 | 6 |
| | 13 | 4 |
| | Total | 950 |



Example: One Variable Table of Brucellosis Prevalence at Sublocal area Level (Quantitative)

| Sublocal area | Prevalence |
|---------------|------------|
| В | 33% |
| Kam | 33% |
| Kat | 25% |
| Kik | 31% |
| L | 7% |
| М | 32% |
| Total | 26% |

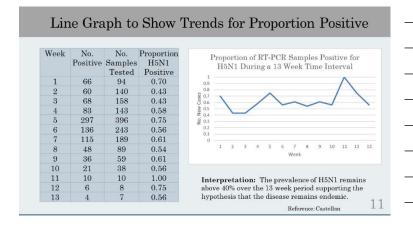
Example: Two Variable Table of Frequency of Animal Purchases at Sublocal area Level (Quantitative)

Purchase of new animals in selected sub-counties

| Sublocal area | Rarely | Sometimes |
|---------------|--------|-----------|
| В | 3% | 3% |
| Kam | 13% | 7% |
| Kat | 17% | 3% |
| Kik | 7% | 3% |
| L | 17% | 3% |
| М | 20% | 0% |
| Total | 77% | 20% |

| farmref_no | purch_freq | milk_how | wash_when |
|------------|------------|----------|-----------|
| 37 | 1 | 2 | 1 |
| 25 | 1 | 2 | 1 |
| 22 | 1 | 2 | 1 |
| 31 | 2 | 2 | 1 |
| 24 | 2 | 2 | 1 |
| 23 | 1 | 2 | 0 |

| Outbreak | Line graph | |
|-----------------------|------------|--|
| investigation data | Integraph | |
| | Histograms | |
| | Bar charts | |



Steps to Create a Line Graph (Adapted from Frontline FETP)

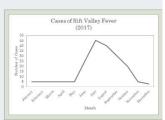
- · Make x-axis longer than y-axis (best ratio 5:3)
- X-axis:
- Match x-axis scale to intervals used during data collection
- · Y-axis:
- Always start y-axis with 0
- Identify largest value, round up for max. Y value
- Select reasonable interval size for y-axis $\,$
- · Plot data
- · Add axis labels and title (What, where, when)
- · Add comments and footnotes

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Arithmetic Line Graph

(Adapted from Frontline FETP)

- · Use for plotting rates over time
- · X-axis almost always time (rarely, age)
- Can be counts, proportions, or rates $% \left(-1\right) =-1$
- Begin at 0
- End with next round number larger than largest value needed to plot
- Divide into equal intervals
- $\boldsymbol{\cdot}$ On either axis, the intervals should be equal
- \cdot Good for comparing $% \left(t\right) =\left(t\right)$ two or more sets of data.



Note: National data showing the seasonal incidence of animal RVF cases between May and November in 2017.

Frequency Histogram: No. positive laboratory cases of HPAI by RT-PCR $\,$ What is your interpretation? Is the H5N1 outbreak under control? No. New Cases 14

Histogram

- Frequency distribution of quantitative data
- X-axis
- Continuous, usually time (onset or diagnosis date)
 Y-axis
- Represents frequency (number of cases)
- No spaces between adjacent columns
 i.e., adjacent columns "touch"
- Easiest to interpret with equal class (x) intervals
 In MS Excel, create "bins" to interpret
- new intervals Column height proportional to number of observations in that interval
 "Epidemic curve" in outbreak investigations



Note: National data

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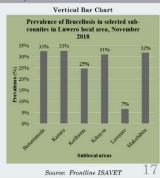
Making a Histogram (Adapted from Frontline FETP)

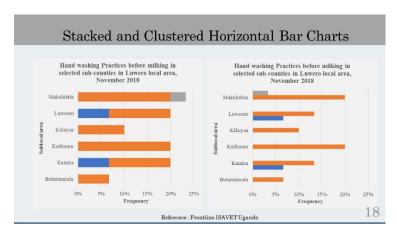
- For continuous numeric data, assign equal width, nonoverlapping categories
- 2. Count the number of times each category appears
- 3. Assign one bar to each category
- 4. Make the bar height equal to the frequency for each category
- 5. Include axis labels with units and a descriptive title

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Bar Charts

- · Can be vertical or horizontal
- Use for variables with discrete, non-linear categories (qualitative), such as local areas
- · Bars have same width
- · Bars have space ("gaps") between them, since categories are not continuous
- · 4 types simple, grouped, stacked,
- $\boldsymbol{\cdot}$ Best type depends on desired emphasis





Maps

- Describe geographic distribution of disease, services, etc.
- · Types:
 - Spot or dot maps
 - Area maps
 - Others
- Symbols represent events, disease
- · Size of circle can be proportionate to disease burden

KEY STEPS IN MAPPING LOCAL OUTBREAKS

- Laminate a local map in plastic so you can add affected locations/cases
- affected locations/cases
 2. Collect GPS Latitude and
 Longitude Coordinates and
 record them on paper or in
 MS Excel
- 3. Add the affected locations/cases on the wall map OR
- 4. Enter GPS coordinates into QGIS or other freeware such as Google Maps

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Spot or Dot Maps

- Spot map (also known as dot map)
 - Often used to show geographical location of individual cases rather than rates
 - You can draw maps by hand or using specialized spatial software showing the location of positive premises



Area Maps: Chloropeth

- Choropleth map
 A method of mapping to display quantitative information, such as rates, in defined jurisdictions such as provinces, regions or countries.

They give a good visual impression of change over space

- Disadvantages
 There is abrupt change at the boundaries Interest admits of shaded units.
 Usually not good for showing total values.
 Different shades not easily distinguishable.

 - Variations within map units are hidden.



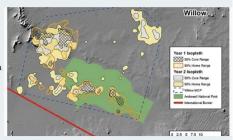
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Area Maps: Isopleth

· Isopleth map

- A method of mapping to show gradual change over space and avoids abrupt changes which boundary lines produce on choropleth
 - maps.

 Example: MAP OF THE
 ELEPHANT INDIVIDUAL
 "WILLOWS" ANNUAL
 RANGES IN THE AMBOSELI
 NATIONAL PARK KENYA,
 DURING THE TWO-YEAR
 TRACKING PERIOD.



Reference: Sowers, M., Fishlock, V. & Manor, T. (2015).

Timeline: Qualitative Data Names and sublocal area location are qualitative nominal variables SDist. 4 SDist. 8 SDist. 4 A <u>timeline</u> of CSF outbreak events is also qualitative display of an outbreak period with key events that can be added SDist. 4 SDIst. 4 Y-F (confirmed) SDist.12 28 july 09 H* SDist. 12 SDist. 8 SDist. 11 Peak Outbreak Phase Early signs Su* SDist. 4 SDist. 8 Sop* 23

Making Recommendations

Guidelines for Making Recommendations

- Recommendations are directly based on interpretation of results obtained from data analysis
- · Recommendations should follow SMART principles and be:
 - 1. Specific to the result of the investigation and local context
 - 2. Measurable based on the data that was collected
- 3. Achievable based on resources and capacity available
- 4. Realistic based on opportunities and limitations
- 5. Time-specific based on specific timelines and milestones

2.4

Exercise 26: Generate Graphs and Tables

- 1. This exercise will take 45 minutes.
- 2. Use the data set Spatial data rabies Table 1. Cases of animal bite injuries by region; 2001–2015 to generate graphs and tables and interpret the data.

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In Summary...

- Rationale behind the display strategy of the findings;
- How to display outbreak investigation data in a table, graph, chart or map;
- How to interpret the findings displayed in a table, graph, chart or map; and
- How to make "SMART" recommendations based directly on study results.

| ISAVE | T Contributing Universities | | |
|--------------|---|----|--|
| Partners | TEXAS ASM. TEXAS ASM. ATEXAS ASM. AGRILIFE | | |
| Contributors | MAKERIKE UNIVERSITY | 27 | |

Exercise 26 - Generate Tables and Graphs

Description of Exercise:

From the dataset, display findings and make relevant recommendation for prevention and control. Should you have any questions over the exercise, please ask a trainer for clarification before during, and after the exercise.

Allotted Time: 45 minutes

Organisation of Group Work:

• Work in pairs of three.

Exercise Objective(s):

1. Display data to be provided.

Exercise Components and Structure:

1. Use the data set Spatial data rabies Table 1. Cases of animal bite injuries by region; 2001–2015 to generate graphs and tables and interpret the data.

Materials, Data or Information:

- 1. Microsoft PowerPoint
- 2. Microsoft Excel

Expected Outputs and Deliverables of Each Participant:

1. Generate tables, graphs and maps.

Instructions

- 1. Open the spreadsheet titled, "X". Review the variables in the data dictionary. Rotate the time lines to the left side of the table.
- 2. Describe the measures of central tendency (Mean, Median, Mode, Range, Minimum and Maximum) for each region in the dataset.

Using the Fx function:

Mean Fx =AVERAGE(Cell:Cell)
 Median Fx =MEDIAN(Cell:Cell)
 Mode Fx =MODE(Cell:Cell)

• Range Ex =MAX(Cell:Cell)-MIN(Cell:Cell)

Minimum Fx = MIN(Cell:Cell)
 Maximum Fx = Max (Cell:Cell)

| | Central | Eastern | Western | Northern |
|--------------------|----------|----------|----------|----------|
| Mean | 3816.8 | 3116.133 | 3222.933 | 3758.8 |
| Standard Error | 281.5967 | 201.2011 | 263.5051 | 301.4981 |
| Median | 4230 | 3078 | 3434 | 4031 |
| Mode | #N/A | #N/A | #N/A | #N/A |
| Standard Deviation | 1090.619 | 779.2485 | 1020.551 | 1167.697 |
| Sample Variance | 1189451 | 607228.3 | 1041524 | 1363517 |
| Kurtosis | 0.312982 | -0.15366 | 0.651834 | 0.747517 |
| Skewness | -1.10762 | -0.1142 | -1.2548 | -0.91746 |
| Range | 3700 | 2918 | 3316 | 4225 |
| Minimum | 1330 | 1601 | 953 | 1049 |
| Maximum | 5030 | 4519 | 4269 | 5274 |
| Sum | 57252 | 46742 | 48344 | 56382 |
| Count | 15 | 15 | 15 | 15 |

3. Display the following:

Line graphs

- a) Line graph for the central region
- b) Add the rest of the regions to the graph. Use mean values of each region to plot the graph

Bar graph

- a) Plot a bar graph for the central region
- b) Add the rest of the regions to the graph (simple, grouped, stacked_

Pie charts

a. Compare the total number of dog bites per region using pie charts

Lesson 21 – One Health Panel Discussion: Multi-disciplinary Outbreak Investigation

| Estimated Lesson and Exercise Time | 1 hour and 30 minutes |
|------------------------------------|---|
| Participant Materials | ISAVET Lesson 21 One Health Panel Discussion: Multi-disciplinary Outbreak Investigation Version 4.doc |
| | Computer and Microsoft Word |
| | Pen or Pencil |
| Handout Materials for Exercises | |

Description of Lesson:

Animal health, human health and wildlife/environmental experts will discuss One Health implementation gaps and opportunities and interact with ISAVET trainees to demonstrate why coordination of surveillance, outbreak investigation and response is essential at the human-animal-wildlife interface in Africa, at the regional, national and local levels.

Facilitator Instructions:

The facilitator is expected to cover all questions in this exercise within the time allotted. He/she will also summarize key points made during the discussion at the end of the session.

Allotted Time: 90 minutes

Organisation:

There will be brief overviews of important One Health activities in Africa followed by questions posed by the facilitator and ISAVET trainees.

Exercise Objective(s):

A. Panelist Input (45 minutes total or 15 minutes per panelist):

1. Describe One Health activities that are underway in East or West and Central Africa.

- 2. Identify gaps in the implementation of One Health:
 - 1. in Africa;
 - 2. in regional level;
 - 3. at the national level:
 - 4. at the local level.
- 3. Describe opportunities are present to use the One Health approach:
 - 5. in Africa;
 - 6. in regional level;
 - 7. at the national level;
 - 8. at the local level.

B. Frontline ISAVET Trainees (30 minutes)

- 1. Describe how Frontline ISAVET can contribute to One Health implementation in your local area.
- 2. Describe your role as a Frontline ISAVET trainee when it comes to implementing One Health at the local level.
- C. General discussion (15 minutes)

Exercise Components and Structure:

- A. Facilitator guiding questions related to the participating countries as well as Africa as a whole:
 - 1. Describe One Health activities that are underway in East or West and Central Africa.
 - 2. Identify gaps in the implementation of One Health in Africa, in regional level, at the national level and at the local level.
 - 3. Describe opportunities are present to use the One Health approach in Africa, in regional level, at the national level and at the local level.
 - 4. Describe your role as a Frontline ISAVET trainee when it comes to implementing One Health at the local level.
- B. Open the floor to questions, answers and further discussion.

Expected Outputs and Deliverables of Each Participant:

1. Active listening and participation in asking questions, providing examples related to the improved implementation of One Health in Africa.

Supplemental Resources for Facilitators:

Survey results of One Health activities in Africa, at the regional, national and local levels provided by FAO regional and country team members.

Lesson 22 - Preparing MS PowerPoint Presentations

| Estimated Lesson and Exercise Time | 2 hours |
|------------------------------------|--|
| Participant Materials | ISAVET Lesson 22 Preparing MS PowerPoint Presentations Participant Guide.PDF |
| | Computer and Microsoft Word |
| | Pen or Pencil |

| In Service Applied Veterinary | |
|--------------------------------------|---|
| Epidemiology Training (ISAVET |) |

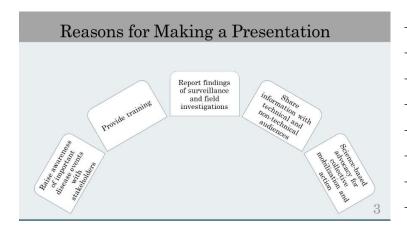
Lesson 22: Preparing MS PowerPoint Presentations

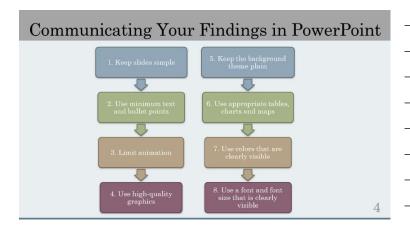
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| | |

Learning Objectives

At the end of this lesson, you will be able to:

- Apply PowerPoint formatting principles for clear presentations; and
- 2. Describe basic graphic design principles.





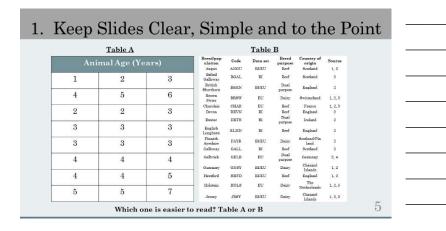


Table Components

Clear title, two variable table organised by species of animal, number of cases as well as the total number of cases with source of data.

| | ID in Country X by Species, tober, 2018 |
|---------------------|--|
| Species | Number of Cases |
| Caprine | 10 |
| Ovine | 30 |
| Bovine (Beef) | 25 |
| Bovine (Dairy) | 12 |
| Porcine | 30 |
| Total | 107 |
| Note: National data | |

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2. Use Minimum Text and Bullet Points

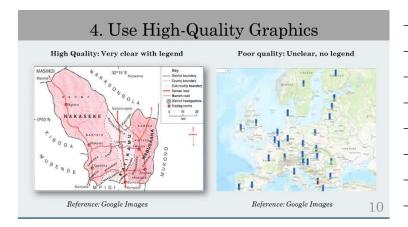
- May 23, 2019: Laboratory report received
- May 24, 2019: local area office reports results to farmer

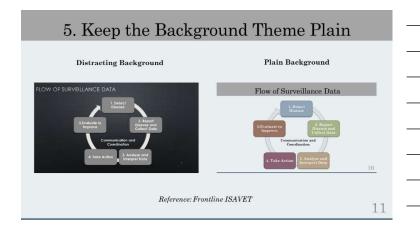
-7

3. Limit Animation

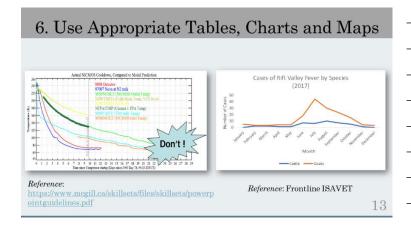
- ${\boldsymbol{\cdot}}$ Don't use slides transitions from one to another too often, this can be distracting
- \cdot If animation is used, use it to represent a flow diagram or cycle while providing your discussion

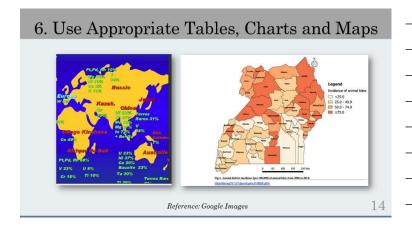
Example: Surveillance Data Analysis Limited animal to demonstrate calculation of risk proportion Production Class Risk Proportion (No. Dead/Total Dead) Population at Risk (PAR) Risk Proportion (No. Dead/PAR) Local Area A No. Dead 8,000 0.80 a 0.995 Egg Layers Broilers 250 0.08 0.003 Village Chickens 3,226 2 0.001 0.0003Village Ducks 1,175 11 0.01 0.0017Local area Total 12,651 6,418 0.51 $Reference: Frontline\ ISAVET$ 9



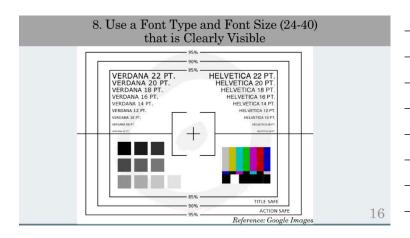


6. Use Appropriate Tables, Charts and Maps | Section | Compared |





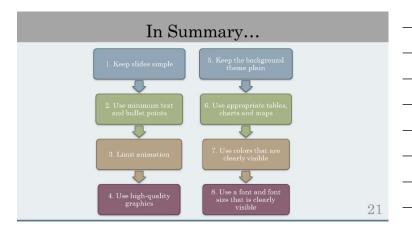
7. Use colors that are clearly visible This is a good mix of colors. Readable! This is a good mix of colors. Avoid bright colors on white. Unreadable! Reference: https://www.mcgill.ca/skillsets/files/skillsets/powerpointguidelines.pdf 15



Format for PowerPoint Slides

- Plan your presentation:
 - Allow 2 to 3 minutes per slide to a maximum of 30 slides
- Title slide is in large font (maximum 40 font):
 - Include title, name, organisation and meeting name, location, date
- Include an overview or objectives slide after the title slide
- · Create sections with section headings
- ${\boldsymbol{\cdot}}$ End with a conclusion or summary slide

Public Speaking Guidelines Some rules of public speaking: Know your subject Know your audience Tell the audience what the talk is about. In other words, tell the audience what you want to tell them (outline of presentation) Keep slides simple and use visuals to illustrate what you are saying Tell the audience what you just told them (conclusion) Acknowledge all contributors and thank the audience 18 Exercise 27: Developing a PowerPoint Presentation from a Report 1. This exercise will take 1 hour. 2. Work individually. Complete a brief 8-slide presentation using the template and report provided. 4. Review your slides with the person sitting next to you. 19 References: · http://www.garrreynolds.com/preso-tips/design/ $\bullet \underline{https://www.mcgill.ca/skillsets/files/skillsets/powerpointg}$ uidelines.pdf





Exercise 27 – Developing a PowerPoint Presentation from a Report

Description of Exercise:

Complete a brief 8-slide presentation using the template and data provided. Should you have any questions over the exercise, please ask a trainer for clarification before during, and after the exercise.

Allotted Time: 1 hour

Organisation of Group Work:

Work individually for this exercise.

Exercise Objective(s):

- 1. Apply PowerPoint formatting principles for clear presentation.
- 2. Describe basic graphic design principles.
- 3. Display data clearly using tables, graphs and maps.

Exercise Components and Structure:

- 1. Work individually.
- 2. Complete a brief PowerPoint presentation from the template provided.
- 3. Share your presentation with the individual sitting next to you.

Materials, Data or Information:

- 1. MS PowerPoint Template: "Frontline ISAVET Exercise 27.ppt"
- 2. Report on Anthrax Outbreak investigation: "RVF Kabale outbreak_report.pdf"
- 3. Paper and pen

Expected Outputs and Deliverables of Each Participant:

1. Complete a brief 8 slide presentation using the template and report provided.

1. Use Microsoft power point to create an 8 slide presentations based on the Rift Valley Fever Outbreak Report "RVF Kabale outbreak_report.pdf.

Lesson 23 - Guidelines for Outbreak Investigation Reports

| Estimated Lesson and Exercise Time | 1 hour |
|------------------------------------|--|
| Participant Materials | ISAVET Lesson 23 Guidelines for Outbreak Investigation Reports Participant Guide.doc |
| | Computer and Microsoft Word Pen or Pencil |

In Service Applied Veterinary Epidemiology Training (ISAVET) Lesson 23: Guidelines for Outbreak Investigation

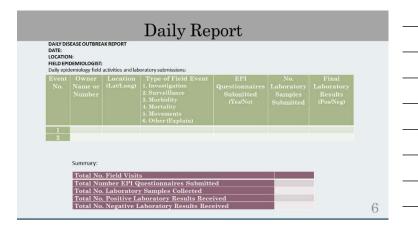
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|--|---|

Learning Objectives

At the end of this lesson, you will be able to:

- Describe the standard sections of an outbreak investigation report for a technical audience, including:
 - Abstract, Introduction, Objective(s), Methods, Results, Discussion, Limitations, Conclusions and Recommendations, Acknowledgements, and
- Describe the kind of report to give to a non-technical audience approach for disease prevention and control.
- 3. Describe how to develop an abstract.

Why is it Important to Prepare an Outbreak Investigation Report? $Reference: Adapted from \ https://wiki.ecdc.europa.eu/fem/w/wiki/definition-of-an-outbreak-investigation-report$ 3 How is an Outbreak Investigation Report Usefull? 4 Reference: Adapted from https://wiki.ecdc.europa.eu/fem/w/wiki/definition-of-an-outbreak-There are two Kinds of Outbreak Investigation Reports · In the early stage of an outbreak investigation, as you are collecting data, you need to do daily updates · Daily debriefing of outbreak events are reviewed each day · Informs follow up action required for the following day · Based on the daily reports The final report follows completion of the outbreak investigation $% \left(1\right) =\left(1\right) \left(1\right) \left$ including measures taken to demonstrate freedom from disease



Outbreak Scenario

Twenty cattle have died in the past 5 days in Kambe local area which is a major dairy and beef raising area. The first cattle where deaths were reported is a village communal grazing herd. It is the second occurrence of increased mortality during the past 12 months. The disease has resulted in the death of young cattle under the age of 6 months of age. The lesions are compatible with Foot and Mouth Disease (FMD).

We will build an outline of an outbreak investigation report based on this scenario.

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The Structure of a Final Outbreak Investigation Report (Reference: Adapted Regional FETPV) 1. Abstract or Executive Summary 2. Introduction 3. Objective(s) 4. Methods 5. Results 6. Discussion 7. Limitations 8. Conclusions 9. Recommendations

1a. Abstract

- · Briefly describe the context of the outbreak:
- 1. Animal-Place-Time components
- 2. Laboratory diagnosis
- 3. Previous disease history in the area
- 4. Disease impact
- 5. Prevention and control measures taken
- 6. Outcomes and lessons learned
- 7. Recommendations for future prevention and control

OUTBREAK SCENARIO: Twenty cattle have died in the past 5 days in Kambe local area which is a major dairy and beef raising area. The first cattle where deaths were reported is a village communal grazing herd. It is the second occurrence of increased mortality during the past 12 months. The disease has resulted in the death of young cattle under the age of 6 months of age. The lesions are compatible with Foot and Mouth Disease (FMD).

1b. Introduction

- Briefly describe the context of the outbreak:
 - 1. Past history of the disease (time)
 - 2. The area(s) currently affected (place)
 - 3. The population at risk (PAR) (animal)
 - 4. Describe the early events at the index location
 - 5. Explain the initial impact of the disease outbreak

OUTBREAK SCENARIO: Twenty cattle have died in the past 5 days in Kambe local area which is a major dairy and beef raising area. The first cattle where deaths were reported is a village communal grazing herd. It is the second occurrence of increased mortality during the past 12 months. The disease has resulted in the death of young cattle under the age of 6 months of age. The lesions are compatible with Foot and Mouth Disease (FMD).

2. Objective(s)

Options for objectives could include:

- 1. Find the source of the outbreak
- 2. Investigate risk factors affecting the frequency and distribution of the disease
- 3. Assess disease impact

In practical terms, the immediate need is to:

- 1. Confirm the diagnosis of FMD
- 2. Identify risk factors for disease introduction and to contain, control and prevent further spread of the virus

3. Methods

Mothoda

Describe:

- 1. The population at risk
- 2. The investigation methods utilised
- 3. Specific data sources and collaborations
- 4. Data collection, quality control, data analysis (software) and data display
- 5. Explain specifically how the methods address the objective(s) of the outbreak investigation

Kambe local area contains 10,000 diary and beef cattle raised on pasture. Clinical exams will be done and laboratory samples will be submitted immediately. Active case finding using a questionnaire and community participatory methods will trace animal movement and other risk factors for introduction and possible spread of FMD.

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4. Results

Results

- · Each result addresses an objective of the outbreak investigation
- Results are displayed in formats (i.e. tables, graphs and maps) that best show the data
- · Include descriptive statistics: Animal-Place-Time
- Use words sparingly: let the data you have displayed speak to the audience
- · Avoid explaining the meaning, only present results
- The results provide the evidence for the rest of the report
- Type O FMD virus confirmed is similar to the previous year
 Hypothesis for introduction is recent cattle movement into
- Hypothesis for introduction is recent cattle movement into Kambe local area from a neighbouring country.
- 3. A higher proportion of confined herds have been protected from FMD outbreaks so far.
- 4. Two neighbouring local areas just reported FMD.

1.9

5. Discussion

Discussion

- · Interpret and explain the meaning of your data
- Describe the significance of the results of the outbreak investigation data
- Compare your results with previous outbreaks and other published reports

The seroptye and molecular pattern is used to qualify the lesions, morbidity and mortality experienced and whether there is a worsening trend compared with previous outbreaks. The impact and reason for higher risk among pastured herds compared with confined herds. The challenges of the field investigation are also presented including lack of field personnel and lack of reporting. Participatory data was very useful and confirmed the questionnaire responses for possible risk factors.

6. Limitations

Limitations

- Clearly and briefly list the limitations of the methods, including:
 - 1. The outbreak investigation
 - 2. Effectiveness of the response
 - 3. The data collected
 - 4. The data that could not be collected
 - 5. Sources of bias in your data

The outbreak likely began at least one month prior to the outbreak investigation. Case finding was difficult because neighbouring local areas could not assist with tracing out. Cattle dealers were not interested in providing information that might endanger their livelihood. Responses from questionnaires was somewhat limited but data collected using participatory epidemiology was complete.

1.

7. Conclusions

Conclusions

- Clearly and briefly state the main conclusions of your report on the data you provided
- Explain how the limitations encountered affect the results

Persistence of the same serotype means that either cattle or wildlife are acting as reservoirs for the virus. Pastured animals are also at higher risk of contact with wildlife and so this must be considered in future prevention and control programmes. There is improved data access from participatory community engagement as compared with questionnaire based interviews with individual farmers.

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9. Acknowledgements

Acknowledgements

Provide acknowledgement of key internal stakeholders in the outbreak event:

- 1. Public sector
- 2. Private sector
- 3. Farmers
- 4. Communities
- 5. Collaborating agencies and partners e.g. Emergency Management, Police, Military

8. Making Recommendations

Making Recommendations

- The MOST IMPORTANT SECTION OF YOUR REPORT!
- · Recommendations should follow SMART principles and be:
- 1. Specific to the local context
- 2. Measurable based on the data that was collected
- 3. Achievable based on resources and capacity available
- 4. Realistic based on opportunities and limitations
- 5. Time-specific based on specific timelines and milestones
- Preventive action is required to trace cattle sold out to other farms, sales yards and abattoirs from Kambe local area during the past 30 days.
- Present findings of the outbreak investigation report so that farmers and marketers are aware of the impact and reasons for introduction and spread

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Report Writing Revise.... revise.... Be persistent and hone your writing skills until the report is clear and simple Acknowledge support received from all collaborators and institutions Prepare an abstract or "Executive Summary" of the report for those who will not likely read the whole report

Sharing the Report with Non-technical Audiences

Sharing the Report Share an abstract or "Executive Summary" of the report for those who will not likely read the whole report

Once official permission is granted, publish... publish...

Convert the report into a brief PowerPoint of brief written report to share with colleagues and farmers

Frontline curriculum - Participant manual Share Lessons Learned Use your report to begin an after action review of the outbreak Always meet and provide feedback to the farmers who were affected by the animal disease outbreak Hold discussions centered on how to improve surveillance, control and prevention 21 Elements of an Outbreak Investigation Report for Technical and Non-Technical Audiences TECHNICAL AUDIENCE NON-TECHNICAL AUDIENCE 1. Title Use the same headings and structure 2. Background 1. Use simple, clear language free of jargon Limit the report to 2 pages in 3. Objectives of the Outbreak Investigation length as an executive summary of 4. Methods for disease investigation

the technical report Include any maps, graphs or tables that are important that

describe the outbreak

Exercise 28: Report Writing Assessment

- This exercise will take 60 minutes to include 30 minutes to write an abstract of
- the report provided and 30 minutes for general discussion.

 Write a brief abstract of 300 words maximum in 30 minutes of the outbreak
 - investigation report provided.

 Briefly describe the context of the outbreak:
 - $^{\circ}$ Animal-Place-Time components

5. Results – Extent and impact 6. Limitations and conclusions

7. Recommendations 8. Acknowledgements

- · Laboratory diagnosis · Previous disease history in the area
- ${}^{\centerdot}$ Disease impact
- Prevention and control measures taken
- ${\boldsymbol{\cdot}}$ Outcomes and lessons learned ${\boldsymbol{\cdot}}$ Recommendations for future prevention and control
- The following outbreak investigation report is provided:
- Reports on Investigation and Response To Rift Valley Fever Outbreak in Kabale local area, Uganda, March 10 April 27, 2016 (RVF Kabale outbreak_report.pdf)
- 4. Participate in the general discussion of the abstract.

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In Summary...

- $\boldsymbol{\cdot}$ There are two kinds of outbreak investigation reports: Daily and Final Reports
- $\boldsymbol{\cdot}$ Outbreak investigation reports are important for a number of reasons:

 - Inform decision makersInform prevention and control actions
 - Document evidence of steps taken during an outbreak investigations
- $\boldsymbol{\cdot}$ For technical audience an outbreak investigation report should consist
- of these standard sections:
 Introduction, Objective(s), Methods, Results, Discussion, Limitations, Conclusions and Recommendations.
- $\boldsymbol{\cdot}$ For non-technical audience, the report should consist of:
 - An abstract of the outbreak investigation report $% \left(x\right) =\left(x\right) +\left(x\right) +\left$
 - ${\bf Brief\,PowerPoint\,presentation\,to\,initiate\,discussion}$

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ISAVET Contributing Universities TEXAS A&M Partners Contributors

Exercise 28 - Report Writing Assessment

Description of Exercise:

Group discussion including questions to provide reports to technical and nontechnical audiences. Should you have any questions over the exercise, please ask a trainer for clarification before during, and after the exercise.

Allotted Time: 1 hour

Organisation of Group Work:

Work in groups for this exercise.

Exercise Objective(s):

- 1. Describe the standard sections of an outbreak investigation report for a technical audience: Introduction, Objective, Methods, Results, Discussion, Limitations, Conclusion, and Recommendations
- 2. Describe the kind of report to give to a non-technical audience for disease control and prevention

Exercise Components and Structure:

- 1. Form into 3 groups
- 2. Review an example Outbreak Investigation Report provided

Materials, Data or Information:

- 1. The following Reports are provided:
 - a. Reports on Investigation and Response To Rift Valley Fever Outbreak in Kabale Local area, Uganda, March 10 April 27, 2016 (*RVF Kabale outbreak_report.pdf*)
 - b. One Health Anthrax Surveillance In Arusha Local area, April 25 May 4, 2018 (*Anthrax One Health Surveillance.pdf*)
- 2. Paper and pen

Expected Outputs and Deliverables of Each Participant:

1. Provide a review of an outbreak investigation report in terms of its content and

Two-outbreak investigation Reports are provided as follows:

- c. Reports on Investigation and Response To Rift Valley Fever Outbreak in Kabale Local area, Uganda, March 10 April 27, 2016 (*RVF Kabale outbreak_report.pdf*)
- d. One Health Anthrax Surveillance In Arusha Local area, April 25 May 4, 2018 (Anthrax One Health Surveillance.pdf)

Groups 1 + 2:

Reports on Investigation and Response To Rift Valley Fever Outbreak in Kabale Local area, Uganda, March 10 – April 27, 2016

Group 3:

One Health Anthrax Surveillance In Arusha Local area, April 25 – May 4, 2018

- 1. Assess the reports provided to see if the structure and components are presented and fulfill what is expected of a standard outbreak investigation report.
 - a. Introduction
 - b. Objective (s)
 - c. Methods
 - d. Results
 - e. Discussion
 - f. Limitations
 - g. Conclusions
 - h. Recommendations

Section III: Week 3 – Preparedness, Disease Prevention and Response, Communication, Ethics, and Professionalism

Lesson 24 – Communicating Disease Transmission Risk to Diverse Audiences

| Estimated Lesson and Exercise Time | 2 hours |
|------------------------------------|--------------------------------------|
| Participant Materials | ISAVET Lesson 24 Communicating |
| | Disease Transmission Risk to Diverse |
| | Audiences Participant Guide.PDF |
| | Computer and Microsoft Word |
| | |

| In Service Applied Vete | rinary |
|------------------------------|--------|
| Epidemiology Training | |

Lesson 24: Communicating Disease Transmission Risk to Diverse Audiences





Learning Objectives

At the end of this lesson, you will be able to:

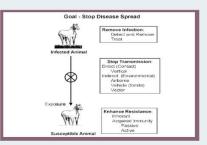
- 1. Define the terms hazard and risk.
- 2. Describe the kinds of animal disease transmission risk.
- $^{\rm 3.}$ Communicate disease transmission risk using a simple risk pathway diagram.

What is a Hazard? HAZARD: means a biological, chemical or physical agent in, or a condition of, an...animal or ...animal product with the potential to cause an adverse effect on aquatic, animal health or public health. Reference: Adapted, OIE. https://www.oie.int/fileadmin/Home/eng/Health_standards/aahc/2010/en_glossaire.htm. Accessed 11/09/2019 • Examples Biological Hazards Chemical Hazards Physical Hazards Highly pathogenic avian influenza virus Botulinum toxin Earthquakes, hurricanes, typhoons Bacillus anthracis Aflatoxin Foreign bodies

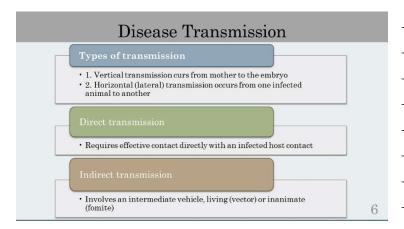
| V | What is Risk? | | |
|--|------------------------------------|--|--|
| RISK | | | |
| means the <u>likelihood of the occurrence</u> and the <u>likely magnitude of the biological and</u> <u>economic consequences</u> of an adverse event or effect to animal or human health. | | | |
| We now understand that there are two ways to define Risk: | | | |
| 1. Probability (P) of an event occurring | | | |
| 2. Probability (P) of an event occurring times (x) the consequences (C) of an event | | | |
| Reference: OIE. https://www.oie.int/fileadmi tm. Accessed 11/09/2019 | n/Home/eng/Health_standard | ds/aahc/2010/en_glossaire.h | |
| | Epidemiological Risk Definition | Risk Analysis Risk Definition | |
| Equation | P = a / a+b (A proportion) | $R = P \times C$ (The product of a proportion and a consequence) | |
| Example | P = Prevalence = 15/30 = 0.5 | $R = 0.5 \times 100 = 100$ | |

Mechanisms of Disease Transmission

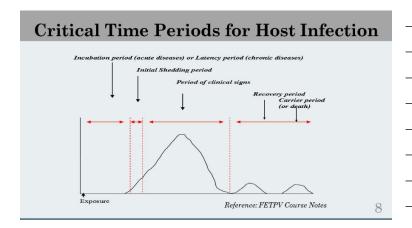
 Transmission of infectious disease agents require a susceptible host, and their ability to replicate in order to maintain the cycle of infection



Reference: Thrusfield



| Mode of Transmission | HPAI ¹ | PPR ² | RVF ³ |
|-----------------------------------|-------------------|------------------|------------------|
| Faecal-oral | + | + | |
| Respiratory | + | + | (Blood aerosols) |
| Tears, saliva, nasal discharge | + | + | |
| Insect vector | | | .+ |
| Reproductive tract | + | | + (Foetus) |
| Milk | | | +/- |
| Meat | + | | + |
| Eggs | + | | |



Disease Transmission Pathways MOVEMENT OF DISEASE AGENTS People and Equipment Domestic Animals Wildlife Insect Vectors Viranshumance - conflict zones Along animal value chains Livestock-wildlife interface Seasonality Trade and Commerce— animal feed, drugs Example: Cattle transporters Across man-made boundaries Example: Cattle grazing with gazelles Example: Ticks related to Rift Valley Fever Seasonality Example: Cattle grazing with gazelles

Disease Transmission Follows Trade and Human Conflict

- Drivers for movement of animals and animal products:
- Established trading routes (domestic and export trades)
- Price differences (profit)
- Transhumance (pastoralism) production system
- Conflict areas

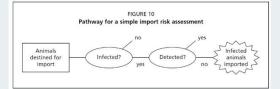


Reference: Google images

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What is a Risk Pathway?

"A risk pathway is a series of conditions that must be met, or events that have to occur, in order for the unwanted outcome to occur. " $(FAO,\,2011)$



Communicating Disease Transmission Risk

- $\cdot \ Veterinary \ field \ epidemiologists \ are \ not \ communication \ specialists;$
- · Use risk pathways to visualise how disease would likely enter, spread and impact the local populations using risk pathways

1. Describe the disease hazard

· Disease X



2. Describe the risk transmission pathway

· Introduction of Disease X via export of animals from country x to country y

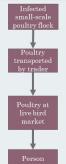
12

Let's Create a Risk Pathway for Avian Influenza

- 1. Hazard
- Family Orthomyxoviridae
- Influenza virus A
- Enveloped virus
- · 8 Strands of RNA
- · Code for 10 Proteins
- * 2 Surface Glycoproteins Hemagglutinin (H1-16) and Neuraminidase (N1-9)
- Example: H5N1
- Affects 17 orders of birds
- Low pathogenic or highly pathogenic types based on chicken challenge model
- Zoonotic with varying impact on human health depending on subtype

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2. Risk pathway diagram for a person to become infected with virus X as a result of visiting a live bird market in geographic area Y during time period Z



Risk question: What is the risk of viable X virus being present at least one live bird market enters into at least one geographic area Y during time

Risk question:

What is the risk of at least one person becoming exposed to viable X virus as a result of visiting at least one live bird market in at least one geographic area Y during time period Z?

Consequence assessment:

Risk question:
What is the risk of at least one person becoming infected with viable X virus and developing clinical disease as a result of visiting at least one live bird market in at least one geographic area Y during $time\ period\ Z?$

 $Source:\ Dirk\ Pfeiffer$

Let's Create a Risk Pathway for PPR

1. Hazard:

- * Peste des Petits Ruminants (PPR) virus six structural proteins
- *One serotype/four lineages related to Africa (I-III) and Middle East and
- ${}^{\centerdot}$ Morbilivirus (related to rinderpest and distemper viruses)
- · Detection: Antigen (F protein) ELISA and RT-PCR (F and N genes) can differentiate PPR from rinderpest
- \cdot Hosts: Affects sheep and goats as well as some zoological animal species affected; buffaloes and pigs may be sub-clinically infected
- · Transmission: Mainly direct through secretions, aerosols and feces; limited indirect through water and objects

2. Risk pathway diagram for importation of PPR virus as a result of sheep and goat trade from geographic area Y during time period Z



Entry assessment: Risk question:

Mas is the risk of PPR from at least one sheep or goat entering from xxxx to at least one neighbouring location from official and non-official trade in mm yyyy?

Exposure assessment:

Risk question:

What is the risk of PPR exposure from at least one imported sheep or goat for susceptible animals in at least one location if it enters in

Consequence assessment:

Risk question:

What is the risk of a major outbreak of PPR in at least one ${\it destination \, location \, if \, at \, least \, one \, susceptible \, animal \, is \, exposed \, in}$

 $Source:\ Dirk\ Pfeiffer$

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Exercise 29: Risk Flow Diagrams

- 1. This exercise will take 60 minutes.
- 2. Form two groups.
- 3. Using the risk analysis principles, draw risk flow diagrams that describe:
 - · The risk of Peste des Petits Ruminants (PPR) in sheep and goats; or
 - * The risk of highly pathogenic avian influenza (HPAI) for poultry and humans.

In Summary...

- Knowledge about the mode of transmission and spread of infectious diseases is the basis and essential input for designing and implementing effective risk mitigation measures;
- Understanding the socio-economic dynamics that drive movement of livestock and livestock products help to develop disease transmission pathways;
- Application of risk pathways enable us to anticipate, prevent, mitigate and effectively manage animal disease risks in a systematic fashion; and
- Risk pathways are is an integral part of risk communication that field epidemiologists are expected to undertake involving different groups of target audiences.

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Partners Contributors Contributors Contributors

Exercise 29 - Risk Flow Diagrams

Description of Exercise:

Develop risk flow diagrams for two infectious diseases. Should you have any questions over the exercise, please ask a trainer for clarification before during, and after the exercise.

Allotted Time: 1 hour

Organization of Group Work:

Form into two groups.

Exercise Objective(s):

- 1. Describe the kinds of animal disease transmission risk.
- 2. Describe disease transmission risk using risk analysis principles: hazard identification, risk assessment, risk communication and risk management.

Exercise Components and Structure:

Using the risk analysis principles, draw risk flow diagrams that describe:

- 1. The risk of peste des petits ruminants (PPR); or
- 2. The risk of highly pathogenic avian influenza (HPAI) for poultry and humans.

Materials, Data or Information:

- 1. Microsoft Word
- 2. Microsoft PowerPoint

Expected Outputs and Deliverables of Each Participant:

1. Two disease risk flow diagrams.

Group 1: PPR

Scenario 1: The Risk of PPR in Sheep and Goats

A pastoral flock of sheep and goat from Region X regularly travels during dry season to the nearest Region Y, which is a mid-altitude area with relatively better pasture and water availability and where sheep and goat flocks graze in communal grazing and water in common watering bodies. Develop a risk pathway for possible spread of PPR from Region X to Region Y of Country A during the dry season, which usually lasts for 2-months in Region X.

- 1. Create a risk pathway diagram for a sheep and goat population to become infected with PPR as a result of movement of live shoats from Region X to Region Y in Country A during the dry 2-months.
- 2. Develop one risk question that should be addressed for PPR when conducting the entry assessment, exposure assessment and consequence assessment.

| Risk Analysis | Risk Question |
|------------------------|---------------|
| Entry Assessment | |
| Exposure Assessment | |
| Consequence Assessment | |

Group 2: HPAI

Scenario 2: The Risk of HPAI for Poultry and Humans

Country X was hit by H5N8 HPAI involving 3 local areas of its Region Y. As a result, the veterinary authorities imposed movement restrictions on poultry and poultry products from the affected local areas and region. Despite the restriction, poultry producers smuggle, through informal markets, broilers into a neighbouring Country Z affected by political crisis. Please develop a risk transmission pathway for the likely introduction and risk of H5N8 HPAI spread to the poultry and human population of the neighbouring country through the informal trading of broiler chickens from the infected country for a period of 1 month until the situation was discovered and stopped by cooperative efforts of authorities in both countries.

- Create a risk pathway diagram for a poultry and human population to become
 infected with H5N8 as a result of informal training of broiler chickens from a
 HPAI infected region of Country X neighbouring Country Y during time period
 Z.
- 2. Develop one risk question that should be addressed for PPR when conducting the entry assessment, exposure assessment and consequence assessment.

| Risk Analysis | Risk Question |
|------------------------|---------------|
| Entry Assessment | |
| Exposure Assessment | |
| Consequence Assessment | |

Lesson 25 – Stakeholder Risk Communication Before, During and Following an Animal Disease Event

| Estimated Lesson and Exercise Time | 2 hours |
|------------------------------------|--|
| Participant Materials | Frontline ISAVET Lesson 25 Stakeholder Communications Before, During and Following an Animal Disease Event |
| | Participant Guide.PDF Computer and Microsoft Word |
| | Pen or Pencil |

In Service Applied Veterinary Epidemiology Training (ISAVET) Lesson 25: Stakeholder Risk Communication Before, During and Following an Animal Disease Event

Post and Agriculture
Organization of the
United National Diseases

Learning Objectives

At the end of this lesson, you will be able to:

- 1. Identify and map important stakeholders in your local area related to animal disease disease events; and
- Explain ways to establish two-way risk communication before, during and following animal disease events affecting animal health and public health.

National Structure for Animal Health Communications (Example: Uganda) Note: 1. The main stakeholders are identified 2. Most connections are two-way in direction | Disease Control Departments | Disease Control Depa

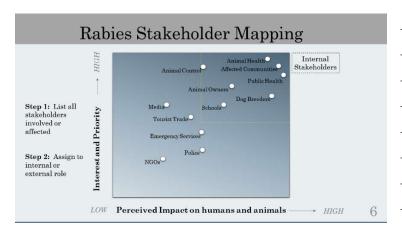
National One Human Communications Structure (Example: Uganda) Functional communication is needed within the One Health Communication Structure in order to achieve One Health goals and objectives Parmers and Other Conception Structure | Conception |

Reference: FAO

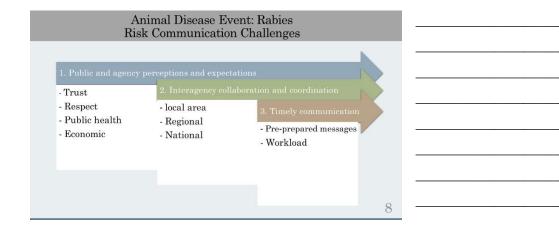
Animal Disease Event: Stakeholder Risk Communication (1) Scenario: A rabies epidemic is occurring in humans and in dogs in a small village near Nairobi, Kenya 1. What is risk communication? "an interactive process for exchanging information and opinions between risk evaluators, risk managers and other interested parties" Reference: OIE, 2004 https://www.oie.int/doc/ged/D1274.PDF 2. What is a stakeholder? "A stakeholder is a person, group or organisation involved in or affected by a course of action" Reference: Google images

5

Two types of Stakeholders: Internal versus External







Animal Disease Event: Best Practices for Stakeholder Risk Communication Communicate in ways that establish, maintain or restore trust Develop messages in advance and adapt for sharing with the public Information communicated should be complete, accurate and easily understood 4. Respect concerns from the community Community concerns are legitimate and must be considered 9

1. Build Trust With Each Stakeholder Group

Scenario: A rabies epidemic is occurring in humans and in dogs in a small village near Nairobi, Kenya

- Community trust
 Report only confirmed information about rabies cases and affected areas
 - Avoid creating undue fear
- To build, maintain or restore trust
- Provide regular updates on progress or challenges requiring community support about rabies . Protect patient confidentiality
- · Each stakeholder has different communication needs
- Dog owners require different messages compared to livestock farmers



Reference: Google images

2. Communicate Early

Scenario: A rabies epidemic is occurring in humans and in dogs in a small village near Nairobi, Kenya

- The most important communication is your first communication
- Provide timely alerts with accurate information about rabies cases in humans and animals
- Describe what measures are being taken by public health and animal health officials
- Discuss information that is likely to be wrong with each stakeholder
- Correct erroneous media messages immediately with correct information from public health and animal health agencies



Reference: Makerere University

3. Provide Transparency

Scenario: A rabies epidemic is occurring in humans and in dogs in a small village near

- Be candid and open: Admit errors or problems encountered
 Be understood easily: Avoid scientific terms about rabies
- ${\boldsymbol{\cdot}}$ Be accurate: Report only confirmed rabies cases
- Be complete: Give complete information about scope of rabies cases

• Transparency outcome = increased trust: Rabies and many diseases can create social distancing and stigma for the affected families

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4. Respect Concerns from the Community

Scenario: A rabies epidemic is occurring in humans and in dogs in a small village near Nairobi, Kenya

- Community concerns are legitimate
- Respect local customs when sharing information
- Rabies risk messages should be tailored for each audience:

 - Those at risk; warning
 The community: diligence
 Academia and researchers:
 accurate data is needed
 Government officials:
 cooperation
 Trade partners: Transparency
 Media: Accurate information



Reference: Google images

Animal Disease Event: Developing Your Risk Communication Messages



- · Creating clear and concise messages
- Use a message map which include up to three key messages with supporting materials
- Messages should be concise and
- Messages should be understandable
- Use positive terms. Avoid the terms

"no" or "do not"

Message Map Map Title: Stakeholder: Questions or concern: Overarching message: (Three sentences, up to 30 words at most, stated as simply as possible) Key Message 1 Key Message 2 Key Message 3 Most important message Supporting information Supporting information Supporting information Supporting information Add detail to overarching key messages (up to three statements) Add detail to overarching key messages (up to three statements) (up to three statements)

| Message Map: During a Rabies Outbreak in Village X near Nairobi, Kenya | | | | |
|---|---|--|---|----|
| | | | | |
| | Key Message 1 | Key Message 2 | Key Message 3 | |
| | Rabies is fatal disease and has been confirmed in the village | Dogs and other mammals transmit rabies virus when symptoms appear | Rabies can be prevented by regular vaccination of humans and animals | |
| | Confirmed positive cases 2 humans and 1 dog are reported | Infected persons or animals carry rabies virus from 10 days to over 12 months before signs of the disease appear | In rabies endemic countries, domestic animals and humans working with animals require vaccination. | |
| | Mortality in humans is 100% if not treated early There is no treatment for rabies in dogs Report all animal bites | Rabies virus is transmitted via saliva to open wounds or mucous membranes Report behavior changes in animals: dumb and furious forms | 2. Domestic animals in rabies endemic countries | 16 |

| Example: Befo | ore the LPAI H71 | N9 Outbreak | |
|---|---|--|----|
| | I H7N9 and why should we be prepare domestic and wild birds; 2) Humans c | | |
| Key Message 1 | Key Message 2 | Key Message 3 | |
| LPAI H7N9 is an influenza virus that may become established. | LPAI H7N9 poses a serious threat to human health. | Good market biosecurity measures can prevent infection | |
| The virus has been detected in a neighboring country. Brisk cross-border trade increases the chances of infected birds entering the country. | LPAI H7N9 poses a serious public health threat because infected poultry can pass the virus to human beings and cause serious disease and death. | The country has a good chance of preventing or mitigating the effects of an incursion by systematically planning and preparing its response. | |
| LPAI H7N9 spreads rapidly among chicken ducks without causing sickness Fear of LPAI H7N9 can cause panic and reducing demand for poultry, causing economic losses to the poultry industry. | Market managers should be trained to explain the facts about LPAI H7N9 to vendors, traders, suppliers, transporters and slaughterers. | Key poultry markets and collection points with poultry should be closed once or twice a month for cleaning and disinfection. | 17 |

Example: During the LPAI H7N9 Outbreak Map Title: Luwero Local Area During stakeholder: Market Managers, Vendors, Farmers questions or concern: How can you prevent LPAI H7N9 from spreading? Overarching message: Effective market management and disinfection are the key to controlling Key Message 1 LPAI H7N9 is disseminated through infected markets Prevention begins in poultry markets Report flu-like symptoms immediately. Since LPAI H7N9 caus In infected areas, LPAI H7N9 Poultry markets have been have been found in samples collected from poultry and the environment from wholesale and retail bird markets. identified as the locus of new infections, since birds from distant symptoms in birds, the first sign of LPAI H7N9 infection is areas arrive there ually a human infection. Cooperation of market managers is essential is essential Infected birds look 100% normal. All exposed birds will be culled from infected markets, and the market closed and disinfected. Humane culling and immediate disposal will be done. Poultry markets must be closed The first sign of LPAI H7N9 is a The first sign of LPAI H 1/No is a human infection. With early treatment, LPAI H7N9 can be cured. Seek immediate treatment for anyone with flu-like symptoms. deaned and disinfected. Market managers must be updated about the LPAI H7N9 threat, the government's contingency plan, and enhanced surveillance plan.

Example: After the LPAI H7N9 Outbreak Key Message 1 Key Message 2 Farm hygiene plays a key role in preventing future infections. This includes separation of species, hygiene, and prompt reporting of unexplained illnesses and deaths Culling is a humane activity. - Farmers are compensated adequately and can restart their activities. - $\begin{array}{ll} \mbox{Infected markets will be re-opened} \\ \mbox{for business after a period of} \\ \mbox{disinfection and closure} \\ \mbox{Farmers will be adequately} \end{array} \begin{array}{ll} - \mbox{LPAI H7N9 can evolve into HPAI} \\ \mbox{H7N9} \\ \mbox{Keep animal species separates to} \\ \mbox{minimize a new virus jumping} \\ \end{array}$ species Report any unexplained deaths or illness immediately compensated and can resume their livelihoods 19

Exercise 30: Stakeholder Communication Before, During and Following an Animal Disease Event

- This exercise will take 90 minutes. Map internal and external stakeholders in your Local Area using the mapping diagram illustrated in this lesson.
- Divide into groups of five participants.
 Group A: African swine fever
 Group B: Brucellosis
 Group C: Avian influenza
 Group D: Rift Valley fever
 Group E: Peste des petits ruminants
- Each group will develop three message maps for one stakeholder
 - group

 Communication before an outbreak

 During an outbreak

 After an outbreak
- Have one person from each group describe their stakeholder maps and message maps.

In Summary...

- •Internal and external stakeholder mapping is required before you develop risk communication messages.
- •Message maps are powerful tools to provide critical messages to different stakeholders during different stages of an animal outbreak event.
 - Communication before an animal disease event
 - -Communication during an animal disease event
 - Communication after an animal disease event

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Partners Contributors Contributors ISAVET Contributing Universities TEXAS A&M TEXAS A&M GRILIFE

Exercise 30 – Stakeholder Communication Before, During and Following an Animal Disease Event

Description of Exercise:

Develop a message map for a specific disease and identify a list of important stakeholders in your local area for communication of your messages. Should you have any questions over the exercise, please ask a trainer for clarification before during, and after the exercise.

Allotted Time: 1 hour and 30 minutes

Organisation of Group Work:

Divide into group of five participants -

- Group A: African swine fever
- Group B: Brucellosis
- Group C: Avian influenza
- Group D: Rift Valley fever
- Group E: Peste des petits ruminants

Exercise Objective(s):

- 1. Make a list of important stakeholders in your local area under the categories noted in the lecture.
- 2. Explain how communication differs before, during and following animal disease events.

Exercise Components and Structure:

- 1. Each group should develop three message maps for each stakeholder group.
- 2. Each message map should be tailored for each disease regarding communication before an outbreak, during an outbreak, and after an outbreak.
- 3. Have one person from each group describe their message maps to the entire group.

Materials, Data or Information:

- 1. MS Word
- 2. MS PowerPoint

Expected Outputs and Deliverables of Each Participant:

Inventory of local area stakeholders

 $1. Identify \ a \ list \ of \ important \ stakeholders \ in \ your \ local \ area$

| Category | Group or Name |
|----------------------------|---------------|
| Government personnel | |
| External audiences | |
| Collaborators and partners | |

| NGOs | |
|-------|--|
| Media | |

 $2.\ For\ your\ group,\ develop\ your\ message\ map\ over\ communication\ of\ your\ specific\ assigned\ disease\ topic\ \underline{before\ an\ outbreak}.$

| Message Map Title: | | | |
|------------------------|---------------------------|---------------------------|--|
| Key Message 1 | Key Message 2 | Key Message 3 | |
| | | | |
| Supporting information | Supporting Information | Supporting Information | |
| 1.1: | 2.1: | 3.1: | |
| 1.2: | 2.2: | 3.2: | |
| 1.3: | 2.3: | 3.3: | |

 $3.\ For\ your\ group,\ develop\ your\ message\ map\ over\ communication\ of\ your\ specific\ assigned\ disease\ topic\ during\ an\ outbreak.$

| Support Informat | Supporting Information | Supporting information |
|---------------------|---------------------------|------------------------|
| 3.1: | 1: | 1: |
| | | |
| 3.2: | 2: | 2: |
| | | |
| 3.3: | 3: | 3: |
| | | |

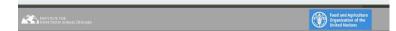
 $4.\ For\ your\ group,\ develop\ your\ message\ map\ over\ communication\ of\ your\ specific\ assigned\ disease\ topic\ \underline{after\ an\ outbreak}.$

| Key Message 1 | Key Message 2 | Key Message 3 |
|------------------------|---------------------------|---------------------------|
| | | |
| | | |
| | | |
| Supporting information | Supporting Information | Supporting Information |
| 1: | 2.1: | 3.1: |
| | | |
|): | 2.2: | 3.2: |
| | _ | _ |
| | | |

Lesson 26 – How to Plan and Prepare the Frontline ISAVET Field Study Proposal and Report

| Estimated Lesson and Exercise Time | 2 hours |
|------------------------------------|--|
| Participant Materials | ISAVET Lesson 26 Written and Oral Reports for Technical and Non-technical Audiences Participant Guide Version 4.PDF |
| | Computer and Microsoft Word |
| | Pen or Pencil |

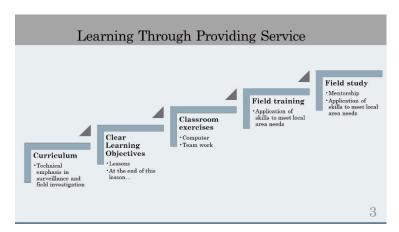
In Service Applied Veterinary Epidemiology Training (ISAVET) Lesson 26: How to Plan and Prepare the Frontline ISAVET Field Study Proposal and Report



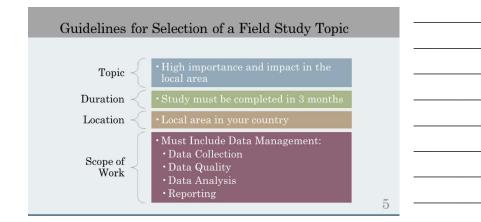
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|----------|-------|-------|
| Learning | Objec | tives |

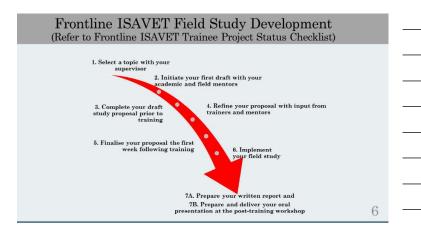
At the end of this lesson, you will be able to:

- Explain and follow the steps to prepare the Frontline ISAVET field study proposal.
- 2. Explain and follow the steps to prepare the Frontline ISAVET field study report and oral presentation.



| raining Course 4 Weeks | | | | | | | | ield W | | | | | |
|---------------------------|---|---|---|---|---|---|---|--------|---|----|----|----|---|
| | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | Expected Output |
| - Heens | | | - | _ | - | - | Ľ | | - | 20 | ** | | 10 Weekly surveillance reports |
| | | | | | | | - | | | | | | 3 Data quality audits |
| | - | • | - | | | | | | | | _ | | Brief Field Study (Max 10 pages) |
| | | | | | | | | _ | | | | | Project proposal |
| | | | | | | | - | | | | | _ | Data validation |
| | | | | | | | | | | | | | Useful high quality data for further analysis |
| | | | | | _ | | | _ | | | | | Results of descriptive data analysis |
| | | | | | | | | - | | | | | Finalize data analysis |
| | | | | | | | | | | _ | | - | Prepare oral and written Reports |
| | | - | 4 | - | | | | | | | | | |





Steps for Preparing the Frontline ISAVET Field Study Proposal

1. Select a field study topic that meets the local needs!

- Enter the topic on your Frontline ISAVET application form prior to the training.
- Consult with your work supervisor
- Contact both your academic and institutional
 mentors

1

Steps for Preparing the Frontline ISAVET Field Study Proposal

2. Initiate your first draft with your academic and field mentors

- *Share the proposed topic for your field study
- · Discuss your overall objective
- Discuss the type of study that is most appropriate
- Initiate your first draft and bring it to Frontline ISAVET training

Examples of Field Project Activities Possible field activities can include one of the following: · Secondary data analyses of existing local area field data or laboratory data · Report of a surveillance investigation · Report of a field (case) or an outbreak investigation ullet Develop a new or improve the existing local surveillance data collection and reporting $^{\circ}$ Analyses of the existing surveillance data in the local area · A disease survey or KAP (Knowledge, Attitudes, Practices) study Selection of the field activities should be based on local priorities, feasibility, practicality and availability of funds. 9 Steps for Preparing the Frontline ISAVET Field Study Proposal 3. Complete your draft study proposal prior to training · Complete a first draft of the Project Proposal for Frontline ISAVET Field Activities Template with input from your mentors Bring your first draft to Frontline ISAVET training 10 Steps for Preparing the Frontline ISAVET Field Study Proposal 4. Refine your proposal with input from trainers and mentors · Attend an appointment to discuss your field project proposal with a Frontline ISAVET $\boldsymbol{\cdot}$ Continue to refine the proposal and share each new draft with your mentors in your country

Frontline ISAVET Project Management and Work Plan TIMELINE Month 2 Month 3 ACTIVITIES OUTPUT Month 1 – Frontline ISAVET Finalize the detailed project proposa Conduct required field visits Garder required field visits Gather and prepare data from laboratory and/or field sources Compile descriptive data Compile descriptive analysis of data Report findings Data validation Useful high quality data for further analysis Results of descriptive data analysis Finalize data analysis Prepare oral and written Reports 12 Steps for Preparing the Frontline ISAVET Field Study Proposal 5. Finalise your proposal the first week following training · When you return home, discuss your field project proposal with both of your mentors and supervisor ${}^{\raisebox{-5pt}{\text{\circle*{1.5}}}}$ Complete your project proposal within one week upon your arrival home 13 Steps for Preparing the Frontline ISAVET Field Study Proposal 6. Implement your field study · Finalise your project field study work plan • Initiate data collection, quality control, analysis, display, conclusions and recommendations • Meet weekly to update and consult with your supervisor and mentors

Steps for Preparing the Frontline ISAVET Field Study Proposal 7. A. Prepare your written report ${}^{\textstyle \cdot} \, \underline{\text{Objective}} {}^{\textstyle \cdot} \, \text{Share clearly and simply what you have learned with your audience}$ · Expectation: *Write...re-write...re-write...until is meets the standards for publication ${\boldsymbol{\cdot}}$ Share each draft with your mentors and trainers to improve it over time ${\boldsymbol{\cdot}}$ Use feedback to further improve your skills in field epidemiology 15 Frontline ISAVET Field Report Structure Length: 10 Pages Maximum Title Page Sections Abstract Objectives Methods Results Conclusions and Limitations Recommendations References Acknowledgements 16 The Report Demonstrates Application of Technical Skills from the Frontline ISAVET Course Definitions: Case definition and Unit of interest Data collection Data quality Descriptive analysis - animal, place, time Data display - tables, graphs, maps Data interpretation Making recommendations for action Reporting and communicating findings Monitoring and evaluation (M&E) One Health - multi-disciplinary, multi-sectoral coordination 17

Steps for Preparing the Frontline ISAVET Field Study Proposal

7. B. Prepare your and oral presentation

- *Aim for 20 minutes in duration or 20 slides maximum
- * Follow guidelines in Lesson 22 for preparing PowerPoint Presentations

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Preparation of Oral Presentations: Prepare a 20-minute oral presentation using MS PowerPoint© consisting of a maximum of 20 slides

| SECTION | NO. SLIDES |
|-----------------------------|------------|
| Title Page | 1 |
| Background | 2 |
| Key Issue of Importance | 1. |
| Objectives | 1 |
| Methods | 4 |
| Results | 5 |
| Conclusions and Limitations | 3 |
| Recommendations | 2 |
| Acknowledgements | 1 |
| TOTAL | 20 |

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Delivery of Oral Presentation at the Post-Training Workshop

- Refer to Lesson 24 for guidance on preparing a Power Point presentation for further details
- Review the mentor and trainer review forms for the a) written report and b) oral presentation to understand how you will be evaluated
- Share your presentation with your mentors 2 to 3 weeks in advance of the post-training workshop
- $\boldsymbol{\cdot}$ Rehearse your presentation ahead of time with your mentors
- ${\boldsymbol{\cdot}}$ Be prepared in advance for questions from participants and have your mentors ask you questions that you can anticipate
- Apply recommendations from trainers and mentors to improve your written report and oral presentation within 1 to 2 weeks following the post-training workshop

| Guidelines for Sharing Technical Information with a Non- Technical Audience | |
|--|---|
| Technical Planetice | |
| 1 Don't assume anything | |
| 2 Don't try to do too much in a brief time | |
| 3 Level the playing field and avoid technical jargon | |
| 4 Link data directly to conclusions and recommendations | |
| 5 Engage the audience based on their needs | |
| 6 Ask for feedback | |
| 21 | |
| Exercise 31: Complete the Frontline ISAVET | |
| Trainee Project Status Checklist | |
| 1. This exercise will take 60 minutes. | |
| 2. Complete the Frontline ISAVET Trainee Project Status Checklist with a trainer or mentor. | |
| 3. Take action to complete the next steps for your project | |
| proposal. | |
| | |
| | |
| 22 | |
| | |
| References | |
| Blostein D . Technical writing and oral presentations. | |
| http://research.cs.queensu.ca/home/blostein/TechnicalCommunication.pdf Accessed 09/17/2019 | |
| Dianna Booher (2008). Presenting Technical Information to | |
| Nontechnical Audiences | |
| · Hannah Richardson (2017). The Fundamental Principles of | - |
| Report Writing | |

| ISAVE | T Contributing Universities | | |
|--------------|--|-----|--|
| Partners | TEXAS A&M GRILIFE | | |
| 6 | A STATE OF THE STA | | |
| Contributors | MAKERIER UNIVERSITY | 0.4 | |
| | | 24 | |

Exercise 31 – Frontline ISAVET Trainee Project Status Checklist

Description of Exercise:

Develop the introduction and objectives section for a technical report in MS Word and MS PowerPoint. Should you have any questions over the exercise, please ask a trainer for clarification before, during, and after the exercise.

Allotted Time: 60 minutes

Organisation of Group Work:

Work individually or in pairs.

Exercise Objective(s):

- 1. Complete the Frontline ISAVET Trainee Project Status Checklist with a trainer or mentor.
- 2. Take action to complete the next steps for your project proposal.

Exercise Components and Structure:

- 1. The trainer will introduce the objectives of the exercise and hand out a copy of the Frontline ISAVET Trainee Project Status Checklist (see below).
- 2. Complete the Frontline ISAVET Trainee Project Status Checklist with a trainer or mentor.
- 3. Take action to complete the next steps for your project proposal.
- 4. Complete the Frontline ISAVET Trainee Project Status Checklist with a trainer or mentor.

Take action to complete the next steps for your project proposal.

Materials, Data or Information:

- 1. MS Word
- 2. MS PowerPoint

Expected Outputs and Deliverables of Each Participant:

- 1. Update of the status of the project proposal.
- 2. An action plan to complete the project proposal.

Frontline ISAVET Trainee Project Status Checklist

Trainee Name: Country:

Date Survey Conducted with Trainee:

| Project Input | | | Status | Date | Follow up Action |
|---------------|-------------------------------|---|-----------------------------|------|------------------|
| | n : .nn | | | | Required |
| | Project Proposal Period | | | | |
| 1. | The subject of the ISAVET in- | | Not initiated | | |
| | country project is selected | | Initiated but not finalised | | |
| | by the trainee and the work | | Finalised | | |
| | supervisor prior to attending | | Unsure | | |
| | the training | | | | |
| 2. | The ISAVET trainee has been | | Yes | | |
| | contacted by the academic | | No | | |
| | mentor | | Unsure | | |
| 3. | The ISAVET trainee has been | | Yes | | |
| | contacted by the | | No | | |
| | institutional mentor | | Unsure | | |
| 4. | The ISAVET trainee is | | Yes | | |
| | familiar with the ISAVET | | No | | |
| | project protocol | | Unsure | | |
| 5. | The ISAVET trainee has | | Not initiated | | |
| | developed a first draft of | | Initiated but not finalised | | |
| | their project proposal | | Finalised | | |
| | according to the project | | Unsure | | |
| | protocol | | | | |
| 6. | A first draft of the project | | Not yet sent to both | | |
| | proposal has been sent to | | mentors | | |
| | the both academic and | | Sent to both mentors but | | |
| | institutional mentors and | | no feedback received by | | |
| | feedback has been received | | trainee | | |
| | by the trainee | | Trainee has received | | |
| | | | feedback from only one | | |
| | | _ | mentor | | |
| | | | Trainee has received | | |
| | | | feedback from both | | |
| | | _ | mentors | | |
| | | | Unsure | | |
| 7. | The trainee would like inputs | | Yes | | |
| | from the ISAVET trainers | | No | | |
| | before the end of the | | Unsure | | |
| _ | training course | | | | |
| ı Pro | ject Completion Period | | | | |

| | Project Input | Status | Date | Follow up Action Required |
|-----|--|---|------|------------------------------|
| 8. | The trainee has completed the collection of all data required to achieve the objectives of the project. | Yes No Partially completed Unsure | | |
| 9. | The trainee has produced at least one draft report of the project in MS Word according to the format provided by Frontline ISAVET. | Yes No Partially completed Unsure | | |
| 10. | A draft report from the trainee has been reviewed by both mentors or a trainer, and the trainee has received feedback. | Not yet sent to both mentors Sent to both mentors but no feedback received by trainee Trainee has received feedback from only one mentor Trainee has received | | |
| | | feedback from both mentors Trainee has received feedback from at least one trainer Unsure | | |
| 11. | The trainee has produced at least one draft report of the project in MS PowerPoint (20 slides maximum) according to the format provided by Frontline ISAVET. | Yes No Partially completed Unsure | | |
| 12. | The trainee would like inputs from the ISAVET trainers before presenting the final report and PowerPoint presentation at the follow up report meeting. | Yes No Unsure | | |

| 1. | Create a draft introduction and objectives statement for East Coast Fever using the information provided. |
|----|---|
| | Introduction: |
| | Objectives Statement: |
| 2. | Create a draft introduction and objectives statement for Brucellosis using the information provided. |
| | Introduction: |
| | Objectives Statement: |

Lesson 27 – FACE – Ethical Decision-Making Framework

| Estimated Lesson and Exercise Time | 2 hours and 30 minutes |
|------------------------------------|---|
| Participant Materials | Frontline ISAVET Lesson 27 FACE – Ethical Decision-Making Framework.PDF |
| | Computer and Microsoft Word |
| | Pen or Pencil |



Learning Objectives

At the end of this lesson, you will be able to:

- Describe the role of ethics in veterinary practice.
- Recognize common ethical issues in veterinary practice.
- Apply an approach to making ethical decisions in your work.
- Understand that ethics often is not about a single "right" answer, but about balancing competing moral and ethical claims.



ввс

The snipers trained to protect rhinos 7 February 2016

In Kenya's Borana nature reserve, drastic new measures are being taken to protect vulnerable rhinos from poachers who kill the animals for their horns.



"We lost 17 from a population of 90," says Borana's Michael Dyer. "We were outgunned and outwitted, so we had to up our game." Dyer recruited a former Special Forces instructor from the British army to train a hand-picked militia of local men, then gave them to right to use lethal force by enrolling them as Kenya police reservists.

The teams monitor 102 rhinos, but when they spot poachers, they don't arrest them. They don't even invite them to lay down their arms. Instead, they kill them - 19 so far, in split-second ambushes during which their victims probably never knew what hit them.

http://www.bbc.com/news/magazine-35503077

4

Protecting the Rhinos

- Is this intervention ethical?
- Why or why not?
- What are your assumptions?
- Whose interests are at stake?

What is Meant by "Ethics"?

- Living in accordance with our values.
- The goal of ethical deliberation is not absolute certainty but reliability in our behavior, choices, and character.

Joan Halifax (Living on the Edge, 2018)

 The hardest ethical decisions are not about "right" vs. "wrong"—they are about "right" vs. "right," or balancing competing ethical claims Rushworth Kidder (How Good People Make Tough Choices, 1955)

6

What is Meant by "Professional Ethics"?

- Codes for professionals that guide decisions and conduct
- Veterinarians have ethical obligations to:
 - Patients (animals)
 - Clients (humans who have the animals)
 - The health and safety of the public



nage: https://commons.wikimedia.org/wiki/File:Veterinary Outreach Hawaye Kebele Ethiopia.iog

7

Dealing with Conflicting Interests







Clients (animal owners)



Public health and safety

Have there been times when you felt the needs of these groups were in conflict?

integes:
Core https://www.independent.co.uk/infe-style/food-and-drisk/cous-beef-farming-reverse-climate-change-global-warming-ali202121.html
Goat famer- http://www.ipsrevs.net/2005/12/climate-change-spoot-famer-spain/
Public health: https://medium.com/the-cia/hyre-public-le-halth-gred-ev-wall-horts-a-draspug-76-bit/15/0id



Blue Local Area Scenario

Imagine that you are the local area veterinarian for Blue imagine that you are the local area veternarian for Bue local area. A local afferner named Isaach as requested permission to move his herd of cattle out of Blue local area in order to go to better grazing grounds in the neighboring Green local area. Before Isaac's request is approved, you need to certify that you have examined the animals and that the herd is healthy enough to travel.

You travel to Isaac's farm on the western boundary of Blue local area. When you arrive, Isaac says that he is very glad to see you today because he plans to move his herd next week, and he needs the certificate as soon as possible. He explains that his cousin is a private veterinarian, and she was able to take blood samples from 20 of Isaac's cows. Isaac shows you the lab results that he says were taken by his cousin, and they indicate that none of the animals tested positive for brucellosis. You know his cousin from past work experiences, and you like her and respect her work. When you ask Isaac'if his cousin came to draw blood samples from the cows or if he brought her he blood samples, he hesitates to answer. He finally says that he brought the blood samples to her, but only to save time because he needs to move his herd very soon.



10

Blue Local Area Scenario

What would you do?

Let's look at how ethical principles can guide decisionmaking.



Four Principles

Will this action provide benefits?

Will this action cause harm?
[Nonmaleficence]

Does this action respect the animals, your clients, and the general public? [Autonomy and Respect]

Is this action fair to the persons and animals involved? [Justice]

From Beauchamp & Childress (2012). Principles of Biomedical Ethics, 7th ed. New York: Oxford University Press.

12

Other Ethical Frameworks

- The four principles are a common way to approach ethical challenges because they are easy to remember and are relevant for many ethical dilemmas. However, there are other ethical approaches that are not based on the four principles.
- Ethics of Care
 - Taking care of others is a moral action.
 - The ethical course of action depends on the specific context and the relationships involved.
- Ubuntu
 - A term and concept in Bantu languages (Southern Africa).
 - Emphasis on interconnectedness of individuals and the community.

1

Beneficence: "Doing Good"

- In general, we all agree that it is good to be kind and to do good.
- In some circumstances, doing good for one person (or animal) may harm another.
- In some circumstances, doing good may not be possible.

In the Blue Local Area:

What does it mean to "do good" for the people and animals involved in the Blue Local Area scenario?

What is a way you could do good for Isaac?

For his animals

For other community members or animals?

Nonmaleficence: "Do No Harm"

- Hippocratic oath
 - "I will use treatment to help the sick according to my ability and judgment, but I will never use it to injure or wrong them."
 - Oath of Bourgelat (veterinary version) "First, do no harm"
- Are there situations where in order to achieve a greater good, you have to do harm?
- Harm can be intentional or unintentional.

In the Blue Local Area:

If you make Isaac wait to move the herd until you retest the animals, could this harm him or his animals? How?

If you do not retest the animals, could this harm other animals or people? How?

15

Autonomy and Respect

- Respect for animal patients, human clients and the general public.
 Allowing people to pursue their own desires is a form of respect.
- Are there situations in which it is difficult to act with respect for the needs of all involved?

In the Blue Local Area:

You want to respect Isaac's wish to move his herd, but you also have a responsibility to protect animal health and public health.

Which of these responsibilities is more important?

16

Justice: Fairness to All

- How do you balance fairness to the animal patient, the human client and the public?
- In a situation where the client's wellbeing and the public's health may be in opposition, we must ask what justice and fairness demand.

In the Blue Local Area:

Is it fair to ask Isaac to wait to move his herd when they have already been tested?

Is it fair to the public to use the herd's test results when you haven't verified them?

What conflicts of interest do you have in this case? If you didn't know Isaac, would your actions be different?

| 5 2 1 | | |
|---|----|--|
| Four Principles | | |
| • What do you think about these principles? | | |
| • How might you use them in your work? | | |
| How might you use them in your work? | | |
| Other questions or comments? | | |
| | | |
| | 18 | |
| | 10 | |
| | | |
| Common Ethical Challanges in Votaninam, Breating | | |
| Common Ethical Challenges in Veterinary Practic | e | |
| Situations characterized by fear, threat, and mistrust, | | |
| such as an infectious disease outbreakDivided loyalties between patients, clients, and public | | |
| health and safety (conflict of interest) | | |
| Moral Distress: when you realize you are causing harm or violating your core principles, but you can't avoid it | | |
| | | |
| | 19 | |
| | 13 | |
| | | |
| Law and Veterinary Ethics | | |
| • Decay your country house a veterinery accessistion? | | |
| Does your country have a veterinary association?Does the veterinary law in your country have bylaws or | | |
| regulations related to ethical behavior? • If you do have these laws, how much are they enforced? | | |
| What challenges are there with enforcement? | | |
| Are there mechanisms for reporting and discussing ethical issues? | | |
| | | |
| | 20 | |





Ethical Decision-Making in Global Health: A Proposed Approach

Let's look at a framework you can use when faced with an ethical dilemma.

Ethical Decision-Making in Global Health: A Proposed Approach Review the options and select one. Consider how each option can ensure respect and fairness. Corresponds to the principles of respect and justice Corresponds to the principles of respect and justice

Scenario about Conflicts of Interest:

Dilemmas for government veterinarians who also conduct private practice

25

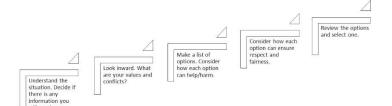
$\begin{tabular}{ll} A Scenario about Conflicts of Interest: Dilemmas for government veterinarians who also conduct private practice \end{tabular}$

You are sub-county Veterinary Officer in Local Area A, in East Africa. You receive an urgent call from a rural pastoralist farmer seeking your help for one of his prize cows, which is high-value (Frieslan pedigree). You quickly cancel your meetings for the day and drive to meet him. When you arrive, you find that the cow is recumbent, in advanced stages of pregnancy, with a ruptured amniotic sac, active uterine contractions, and severe vaginal prolapse with indications of dystocia. The animal has been showing signs of discomfort and restlessness for more than 6 hours. There are no other competent veterinary surgeons to handle the case in the area – the nearest being 75 km away.

surgeons to nandle the case in the area — the hearest being 7.5 km away. The situation is an emergency. The government pays your (modest) salary and expects you to be responsible for animal health, but doesn't provide supplies or materials to handle surgical and clinical cases except for special assignments on rare occasions. You happen to have the necessary supplies in your vehicle and are willing to use them in your capacity as a private veterinarian, but you must charge the farmer US \$40 for your time and materials (in part because you need the money to keep your two oldest children enrolled in school), which the farmer considers a lot of money.

ordest criticities enrolled in School), which the farmer considers a lot of money. Being an experienced Veterinary Officer, you know that African trypanosomiasis, which is a priority reportable zoonotic disease in your country, is associated with uterine or vaginal prolapse. The health regulations of your country regular you to report any clinically suspect case of trypanosomiasis to the Local Area veterinary Officer (DVO), and collect appropriate samples to send to the Local Area veterinary laboratory or a certified private laboratory. The government laboratory services are free. The government lab test results tend to take longer, but they are reliably reported to the national health authorities. In contrast, the certified private laboratory charges for tests; results are obtained much more quickly, but reporting positive results to the government authorities, as required by law, is less reliable. In either case, the farmer must pay the cost of specimen collection.

Ethical Decision-Making in Global Health: A Proposed Approach



27

In Summary...

- Veterinarians face a variety of ethical challenges in their day-to-day work.
- Professional codes of ethics for veterinarians describe responsibilities to three major stakeholders: patients (animals), clients, and the public.
- $\bullet\,$ The interests and ethical claims of these three groups may sometimes be in tension or in conflict with each other.
- Four principles of bioethics can be useful in weighing ethical claims of different groups. These principles are: beneficence; non-maleficence; autonomy (respect); and justice (fairness).
- $\bullet\,$ The 5-step approach to ethical decision-making can help the veterinarian to analyze ethical dilemmas, identify where additional information is needed, and weigh different courses of action.

28

Acknowledgements

- Food and Agriculture Organization (FAO) of the United Nations contributors:

 - Caryl Lockhart, FAO Rome
 Sam Okuthe, FAO Uganda
- Texas A&M University, Institute for Infectious Animal Diseases contributors:
 - David Castellan
 - · Heather Simmons
- The Task Force for Global Health contributors:
 - David Addiss, Focus Area for Compassion and Ethics (FACE)
 - Angela Hilmers, Training Programs in Epidemiology and Public Health Interventions Network (TEPHINET)
 - Jessica Hill, Focus Area for Compassion and Ethics (FACE)
 - Emma Cooke, Emory University School of Medicine

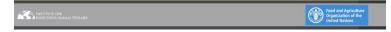
Case Study 3: Practical Ethics for Veterinarians

Your instructor will provide an additional resource after group discussions conclude.

Lesson 28 – Professionalism in the Practice of Veterinary Field Epidemiology

| Estimated Lesson and Exercise Time | 2 hours |
|------------------------------------|------------------------------------|
| Participant Materials | Frontline ISAVET Lesson 28 |
| | Professionalism in the Practice of |
| | Veterinary Field Epidemiology |
| | Participant GuidePDF |
| | Computer and Microsoft Word |

In Service Applied Veterinary Epidemiology Training (ISAVET) Lesson 28: Professionalism in the Practice of Veterinary Field Epidemiology

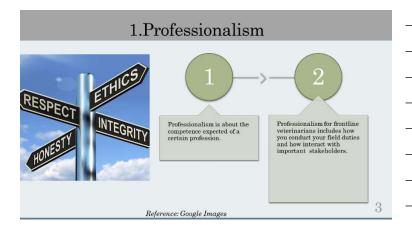


Learning Objectives

At the end of this lesson, you will be able to:

- 1. Define professionalism;
- 2. Describe modalities of professional best practices:
 - a) community relations;
 - b) data integrity and confidentiality;
 - c) information sharing;
 - d) protecting the population at risk;
 - e) animal welfare; and
- $_{\mbox{\scriptsize 3.}}$ Be an effective team member and adopt the role assigned to you.

| 0 | | |
|---|--|--|
| 2 | | |



| Veterinary Statutory Bodies | | |
|------------------------------|--|--|
| Туре | Role | Example |
| Professional associations | Oversee the quality and competence of veterinarians in a country (OIE) | American Veterinary Medical Association promotes continuing education and accredits Veterinary Colleges |
| Professional regulatory body | Assure legal compliance with veterinary legislation and regulations | California Veterinary Medical Board issues veterinary licenses based on qualifications and enforces standards of professional conduct |

Objectives of Veterinary Statutory Bodies



- To promote and safeguard the interests, welfare, terms and conditions of service for veterinary professionals.
- Enhance recognition of their crucial role in promoting animal health, food safety, animal welfare and public health.
- Encourage members to maintain the honor, dignity, independence, integrity, tradition of the profession, professional standards, discipline, conduct and etiquette.

Reference: Google Images

Professionalism in Frontline Veterinary Field Epidemiology

- -Frontline Veterinary Field Epidemiology embraces the One Health approach:
- It means communicating, collaborating, cooperating and working together to achieve better health for humans, animals and the environment.
- The health of people is connected to the health of animals and the environment.



References: Google Image

6

a) Community Relations for Frontline Veterinarians

- Establish & maintain a mutually beneficial relationships with communities and important stakeholders.
- $\cdot \ Work\ with\ the\ leaders\ to\ enlist\ community\ and\ stakeholder\ support$
- Foster two way communication among frontline veterinarians, the community and important stakeholders.
- ${\bf stakeholders}. \\ {\bf \cdot Communicate \ risk \ regularly \ with \ communities \ and \ stakeholders \ (see \ Lesson \ 24 \ for \ details)}$
- Utilise community support & cooperation in order to better prevent and control the disease. Cultivate trust with key informants and leaders who can assist with case finding
- Inform the community and important stakeholders of you role to investigate, identify the cause
 of the problem, provide interventions to stop transmission and protect the population at risk.
- of the problem, provide interventions to stop transmission and protect the population at risk. Enlist community leaders and stakeholders to take part in disease prevention and control activities



Reference: Google Images

Relationship Building Building Ingredients - Transparency - Trust - Commitment - Social bonding - Adaptation - Professionalism - Cooperation - Collaboration - Engagement - Synergy - Word of mouth

Best Practices for Community Relations 1 Recognition of cultural institutions and elders or opinion leaders 2 Collaboration with local leaders and institutions 3 Dress appropriately for the community of interest 4 Determine correct language for community of interest 5 Avoid mannerisms that are disruptive or destructive 6 Show respect to the community members b) Best Practices for Data Integrity and Confidentiality Capture data Use a reliable means and ensure accuracy DATA INTEGRITY Hold daily team meetings to cross check completeness Reference: Google Images Store appropriately to avoid loss Encourage confidentiality 10 c) Best Practices for Information Sharing 1 Share information with those who need it 2 Release to media only information meant for them 3 Only release information that is cleared by the team leader 4 Appoint one spokesperson for each field investigation 5 Provide regular updates to the general public 6 Package information in different forms to suit different audiences

d) and e) Best Practices For Protecting the Population at Risk & **Ensuring Animal Welfare** Maintain biosafety and biosecurity during investigations Use correct disinfection or change PPE between farms Use appropriate restraint and handling methods Practice humane euthanasia when required Affected animals health during an investigation Avoid overcrowding Discomfort and distress to animals during an investigation Humane Euthanasia and Disposal \cdot Humane euthanasia is a standard every local veterinarian must practice • "Do no Harm!" $\dot{}$ Standards for each species of animal are available but sometimes difficult maintain under field conditions – DO YOUR BEST! · Promotes goodwill with animal owners and the community • Prevent negative publicity in the media and on the internet! $\boldsymbol{\cdot}$ Immediate, proper disposal method is a professional duty to stop further disease transmission $^{\bullet}$ Know the advantages and disadvantages of each disposal method * Example: Burial is challenging in the rainy season when the water table is high 13 3. Be an Effective Team Member

Exercise 32: Professionalism During Veterinary Field Investigations

- 1. This exercise should take 90 minutes.
- Form into six (6) groups
- List and describe best practices for veterinary field epidemiologists in your local area for the following issues and add to this list:
- List of additional best practices:
 Build rapport with the community leaders as soon as possible.
 - Recognise the socio-cultural setting of the affected community.
 - Community using a language that is well understood. The team should dress appropriately.

 - Respect privacy of the community members.
 - Cross cultural skills so as interpret the cultural practices appropriately and respect the culture of the affected community. Don't despise the important aspects of the community.

15

In Summary...

- 1. If the veterinary field epidemiologists-community relations are not handled well, it can lead to suspicion, mistrust, lack of transparency and accountability leading to sabotage of the field investigations by the community members.
- This would undermine the overall success thus hindering appropriate intervention.
- To sum it up, for successful veterinary field epidemiology investigations good relations that provide mutual benefits and foster interdependence between the team and the community members is required.

ISAVET Contributing Universities

Partners

Contributors



Exercise 32 – Professionalism During Veterinary Field Investigations

Description of Exercise:

List and describe best practices for veterinary field epidemiologists in your country. Should you have any questions over the exercise, please ask a trainer for clarification before during, and after the exercise.

Allotted Time: 90 minutes

Organisation of Group Work:

Form into six (6) groups.

Exercise Objective(s):

- 1. Define professionalism.
- 2. Describe modalities of professional best practices: Community relations; data integrity and confidentiality; information sharing; protecting the population at risk and animal welfare.
- 3. Be an effective team member and adopt the role assigned to you.

Exercise Components and Structure:

- 1. List and describe best practices for veterinary field epidemiologists in your country for the following issues and add to this list:
 - List of additional best practices:
 - Build rapport with the community leaders as soon as possible.
 - Recognise the socio-cultural setting of the affected community.
 - Community using a language that is well understood.
 - The team should dress appropriately.
 - Respect privacy of the community members.
 - Cross cultural skills so as interpret the cultural practices appropriately and respect the culture of the affected community. Don't despise the important aspects of the community.

Materials, Data or Information:

- 1. Microsoft Word
- 2. Microsoft PowerPoint

Expected Outputs and Deliverables of Each Participant:

1. List and description of best practices for field epidemiologists.

List and describe best practices for the following:

| • | Community relations |
|---|------------------------------------|
| | |
| | |
| | |
| | |
| | |
| | |
| | |
| • | Data integrity and confidentiality |
| • | Data integrity and confidentiality |
| | |
| | |
| | |
| | |
| | |
| | |
| | |
| • | Information sharing |
| | |
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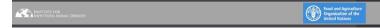
• Protection of the population at risk and animal welfare

Lesson 29 – Emergency Preparedness and Response Planning for Animal Health Emergencies at the Local Level: Part 1

| Estimated Lesson and Exercise Time | 3 hours |
|------------------------------------|---|
| Participant Materials | Frontline ISAVET Lesson 29 Emergency Preparedness and Response Planning for Animal Health Emergencies at the Local Level: Part 1 Participant Guide.PDF |
| | Computer and Microsoft Word Pen or Pencil |

In Service Applied Veterinary Epidemiology Training (ISAVET)

Lesson 29: Emergency Preparedness and Response Planning for Animal Health Emergencies at the Local Level: Part 1

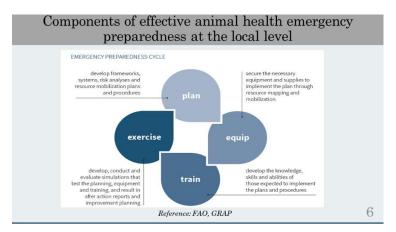


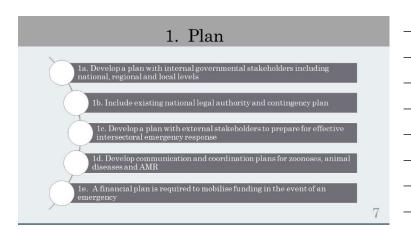
Learning Objectives

At the end of this lesson, you will be able to:

- Define animal health emergency, emergency preparedness and emergency planning;
- 2. Describe the goals of developing an animal health emergency plan for preparedness and response at the local level;
- Explain the components of effective animal health emergency preparedness and response at the local level;
- 4. Describe who the stakeholders are in developing and implementing the preparedness plan at the local level.

| Tronomic carrioaram Taronipani man | atti |
|---|-------------|
| Definition of Animal Disease Emergency | - |
| (OIE, 2010) | |
| Transboundary animal disease (TAD), defined as an epidemic; | |
| Disease which is highly contagious or transmissible; | |
| The potential for very rapid spread, irrespective of national borders; | |
| | |
| Cause serious socio-economic and possibly public health consequences. | |
| | |
| | |
| Definitions (OIE, 2018): | |
| Emergency Preparedness and Emergency Response | |
| | |
| PREPAREDNESS RESPONSE | |
| · Is based on an emergency preparedness plan • Is based on a contingency plan: | |
| • The plan outlines what a government needs to do before an outbreak of a • The response plan details what a government will do | |
| disease happens in order to be prepared (i.e. getting ready). in the event of an incursion of a disease, from the point when a suspect case is | |
| reported (i.e. responding). | |
| 4 | |
| | |
| Goals of an Animal Health Emergency Plan | |
| (Adapted: FAO, GEMP) | |
| | |
| Reduce the impact of animal health emergencies, | |
| whether caused by natural occurrence, accidental or deliberate introduction, by enhancing | |
| [local] preparedness and response capacity. | |
| | |
| | |
| | |



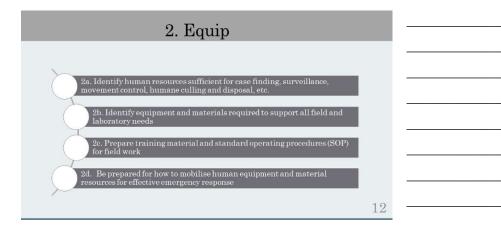


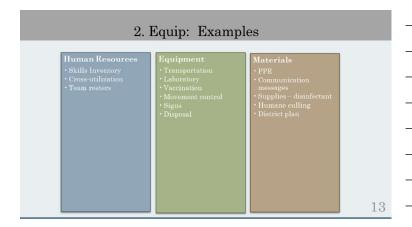
Animal Health Emergency Preparedness is a Partnership at Every Level! Planning depends on partnerships coordinated by the Government Define roles and expectations for: Government Internal National District (Local) Sub-district Private Sector Understand and acknowledge their importance! Academia National planning provides structure for district participants during an outbreak

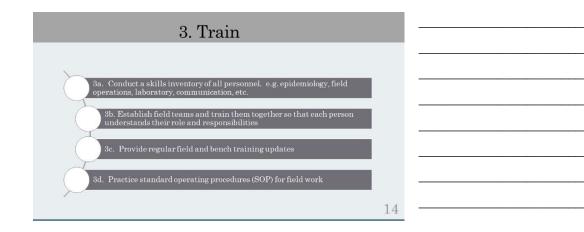
Government Preparedness Is an animal health emergency defined within your National Contingency Plan? If so, does the contingency plan contain: Adequate authority, rules and regulations Contains a chain of command structure Procedures for resource mobilization and release A compensation scheme granting authority A list of priority disease and risks Strategies and standard operating procedures for priority diseases Partnerships with: Academia Veterinary statutory bodies Non-governmental organisations and the private sector e.g. ILRI, producer cooperatives and associations



| External Stakeholders Planning and Preparedness | |
|---|--|
| National Disaster Management Authorities • Animal Health • Public Health • Wildlife • National Emergency Management Agency | |
| Livestock Industry Groups Veterinary Statutory Bodies | |
| Non-governmental Organisations (NGOs) Local Farmers | |
| Livestock Market Participants – owners, traders, vendors, public Laboratories – government, academic, private | |







| 3. Train: Examples | |
|---|---|
| 5. 11am. Examples | |
| Human Resources * Skills inventory of personnel Participate in regular Participate in regular Training Models * Contingency plan Training curriculum | |
| personnel Cross-utilization Train and update regularly Participate in regular Training curriculum and materials Standard operating procedures (SOP) | |
| • Table top • Field simulation | |
| * Joint animal health public health field training | |
| | |
| 15 | |
| | |
| | |
| 4. Exercise | , |
| 4. Exercise | , |
| | |
| 4a. Organise field teams according to disease surveillance, disease diagnosis and disease control functions | |
| 4b. Provide as many opportunities as possible for staff to | |
| participate in outbreaks in neighbouring districts | |
| 4c. Conduct short regular tabletop and field exercises based on each components of the emergency response plan | |
| 16 | |
| | |
| | |
| 4. Exercise: Examples | |
| | |
| Planning A simulation exercise is a test of how the advance of conducting improve the simulation Plan 3-6 months in advance of conducting improve the simulation | |
| contingency plan works in the real world, at the training exercises training exercises • Evaluate the outcome of | |
| • Establish a simulation exercise planning committee • Include public and • Establish realistic goals contingency plan • Establish realistic goals and deliverables for the exercise • Update training | |
| Establish a simulation exercise planning committee Include public and private sectors Part of the exercise and deliverables for the exercise and deliverables for the exercise and update the contingency plan accordingly Update training curriculum and materials and standard operating procedures (SOP) | |
| (SOP) | |
| | |

Exercise 33: District Level Planning Matrix

- 1. This exercise will take 120 minutes.
- 2. Divide into two groups of roughly equal size.
- Groups should elect a leader to facilitate discussion.
- Groups should elect a scribe to record information.
- Check and record the emergency preparedness elements that you can achieve in your local area to achieve the following:
 - · PLAN
 - · EQUIP · TRAIN

 - EXERCISE
- Discussion will follow related to implementation and inclusion in existing plans when the students return home.

In Summary...

- This module introduces preparedness planning concepts for animal health emergencies with a focus on district implementation;
- · Local veterinary offices need to plan, equip, train and exercise to prepare for emergencies;
- Use the output from Exercise 33 to further improve your local area emergency preparedness.

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ISAVET Contributing Universities

Partners



Contributors

Exercise 33 - Local Area Level Planning Matrix

Description of Exercise:

Develop a local-level planning matrix in preparation for an animal disease outbreak. Use avian influenza as the disease. Should you have any questions over the exercise, please ask a trainer for clarification.

Allotted Time: 120 minutes

Organisation of Group Work:

Form yourselves into groups of roughly equal size.

Exercise Objective(s):

1. Check and record the emergency preparedness elements that you can achieve in your local area.

Exercise Components and Structure:

- 1. Check and record the emergency preparedness elements that you can achieve in your local area to achieve the following:
 - PLAN
 - EQUIP
 - TRAIN
 - EXERCISE
- 2. Develop a local level planning matrix.

Materials, Data or Information:

- 1. Flip chart
- 2. Markers
- 3. Computer

Expected Outputs and Deliverables of Each Participant:

1. Complete matrix of local preparedness.

Local Preparedness Matrix

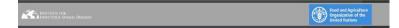
| Preparedness Step | Details of Local Preparedness |
|-------------------|-------------------------------|
| 1. Plan | - |
| 2. Equip | • |
| 3. Train | - |
| 4. Exercise | • |

Lesson 30 – Emergency Preparedness and Response Planning for Animal Health Emergencies at the Local Level: Part 2

| Estimated Lesson and Exercise Time | 3 hours |
|------------------------------------|--|
| Participant Materials | ISAVET Lesson 30 Emergency Preparedness and Response Planning for Animal Health Emergencies at the Local Level - Part 2 Participant Guide.doc |
| | Computer and Microsoft Word Pen or Pencil |

In Service Applied Veterinary Epidemiology Training (ISAVET)

Lesson 30: Emergency Preparedness and Response Planning for Animal Health Emergencies at the Local Level: Part 2



Learning Objectives

At the end of this lesson, you will be able to...

1. Map value chains with stakeholders during peace time.

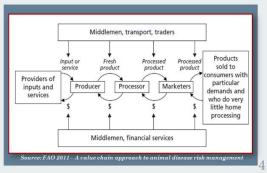
3

Value Chain Maps are part of Preparing for an Emergency Emergency Value Chain mapping is an important part of emergency preparedness: 1. It informs risk based surveillance 2. It informs rapid field response for prevention and control of disease transmission through marketing channels

What are Animal Value Chains?

Reference: FAO, GRAP

- Value chains are the marketing channels for movement of animals, animal products and animal by products
- animal by products
 • Value chains include inputs, processes and outputs

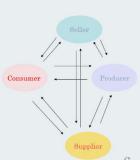


What are Animal Value Chains? INPUTS > PROCESSES > OUTPUTS Treat White MANUEL STATE AND ANTONS Services Front State Animal Value Chain approach to animal disease risk management

The Livestock Sector

- · Who are involved in livestock value chains?
- Initial thoughts:
- People who eat consumers
- People who own and produce ${\bf producers}$
- People who supply suppliers
- People who sell sellers
- People who buy consumers
- · Each player has a unique motivation for their actions within the value chains.
 - · Profit: Producers, sellers and Suppliers
 - Nourishment: Consumers

Source: FAO 2011 – A value chain approach to animal disease risk management 6



Value Chains Operate Both Inside and Outside of the Local Area



- Food is produced through a chain of processes for each type of animal production system- Farm production
 Transport
 Marketing

- Consumption
- Each type of livestock production system is composed of the following: Inputs

- Outputs
 People who influence the movement of animal products

Value Chains Operate Across Border Lines

- · Animal products move from production areas to consumption
- · Movement occurs through movement corridors which contain market points along the way.
- · Animal traders and marketers are key players that control the system.



Benefits of District Value Chain Mapping • Conduct Act quickly Assess risk points before and risk-based surveillance anticipate conducting with limited surveillance where disease can human and material be spreading resources in advance 9

How do we Develop an Understanding of Value Chains at the Local Level?

- Value chain mapping is the first step to developing a preventive strategy for early disease detection based on the trading behaviour of farmers, middlemen and marketers.
- Build value chain maps in peace time (before there are outbreaks.
- Bring together the local stakeholders:
 - Producers
 - Suppliers
 - Sellers
- Consumers

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Steps in Developing a District Value Chain Map 1. Select an animal production system to map Dairy, beef, sheep, goat, pigs and/or poultry 2. Assemble the risk mapping team Farmers, breeders, district veterinarians, traders, marketers and suppliers 3. Discuss the objectives and benefits of value chain mapping Targeted risk-based surveillance and early detection 4. Draw a district map Show boundaries, roads, rivers, lakes and railways 5. Build the map layers and nodes Location of breeders, producers, processors, markets, feed and medication sources

Example: Value Chain Map of Poultry Value Chain



Reference: Frontline ISAVET

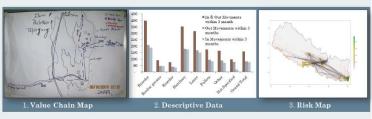
12

Outcomes From Value Chain Mapping

- · Link risk-based surveillance to available resources:
 - Human resources
 - Laboratory resources
 - Financial resources
- Develop risk-based criteria for prevention and control:
 - Identify high-risk nodes for surveillance
 - Establish the basis for descriptive value chain analysis
- Establish the basis for advanced studies such as social network analysis

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Value Chain Outputs



Reference: FAO, 2014

FAO Value Chain Studies

- · Guidelines on designing livestock value chain studies for disease risk assessment and control purposes - Acknowledgement: Jan Hinrichs



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Exercise 34: Preparedness

- · This exercise should take 120 minutes.
- · Participate in the Value Chain Workshop demonstration.
- · Local veterinarians, farmers, dealers and market owners will develop a value chain map for a local area animal production system (cattle, sheep, goats, poultry, etc.).

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In Summary...

- Value chain mapping should be conducted as part of preparedness activities ${\tt BEFORE}$ an outbreak occurs
- Value chain mapping at the district level offers the following benefits:
 - Risk Assessment: Assess risk points before conducting surveillance
 - Disease Prevention: Conduct risk-based surveillance with limited human and material resources
 - ${\it Disease~Control:}$ Act quickly and anticipate where disease can be spreading in advance

| ISAVE | T Contributing Universities | | |
|--------------|-------------------------------|----|--|
| Partners | TEXAS A&M TEXAS A&M GRILIFE | | |
| | TEXAS ARM. | | |
| | AND DE COLOR | | |
| Contributors | | | |
| | MAKERERE UNIVERSITY | 18 | |
| | | 10 | |

Exercise 34 - Preparedness

Description of Exercise:

Develop a local-level value chain may in preparation for an animal disease outbreak. Should you have any questions over the exercise, please ask a trainer for clarification.

Allotted Time: 120 minutes

Organisation of Group Work:

Participate in the value chain workshop.

Exercise Objective(s):

1. Local veterinarians, farmers, dealers and market owners will develop a value chain map for a local production system with assistance of the local veterinarians using the following method.

Exercise Components and Structure:

1. Develop a value chain map.

Materials, Data or Information:

- 1. Flip chart
- 2. Markers
- 3. Computer

Expected Outputs and Deliverables of Each Participant:

1.Experience of doing value chain mapping in preparation for animal disease outbreak events.

The process for developing local value chain maps for each animal production system is as follows:

- 1. Select an animal production system to map
- 2. Assemble the risk mapping team
- 3. Discuss the objectives and benefits of value chain mapping
- 4. Draw a local map using a large piece of white paper that is taped to the wall
- 5. Build the map layers and nodes

Lesson 31 – Emergency Preparedness and Response Planning at the Local Level: Part 3

| Estimated Lesson and Exercise Time | 3 hours |
|------------------------------------|--|
| Participant Materials | ISAVET Lesson 30 Emergency Preparedness and Response Planning at the Local Level - Part 3 Participant Guide.doc |
| | Computer and Microsoft Word Pen or Pencil |

In Service Applied Veterinary Epidemiology Programme (ISAVET)

Lesson 31: Emergency Disease Preparedness and Response Planning at the Local Level: Part 3



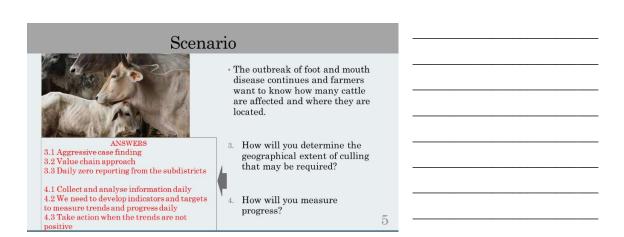
Learning Objectives

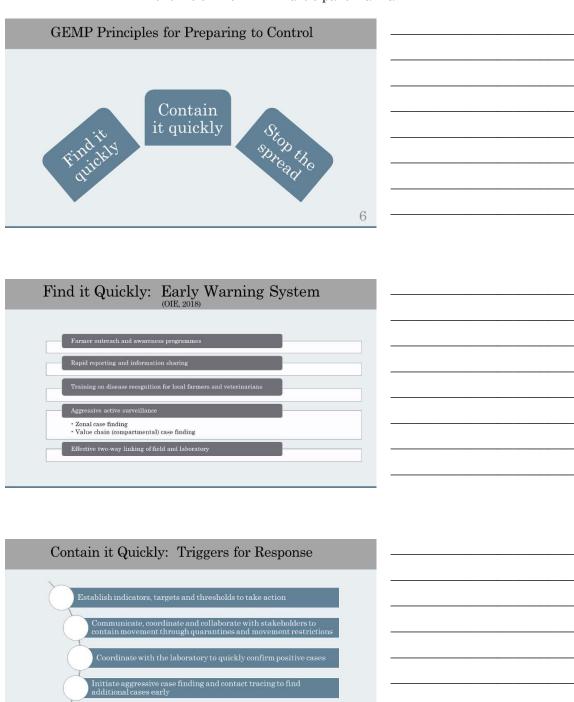
At the end of this lesson you will be able to:

- 1. Apply GEMP principles to improve outbreak response:
 - · Find it quickly
- · Contain it quickly
- · Stop the spread
- 2. Develop indicators and targets to measure and improve daily disease response!

Health Intervention Model for Disease Prevention and Control (Adapted: FAO, 2006) **Enhance Host** Prevent Prevent **Immunity** Exposure Transmission • Vaccination · Humane culling • Movement control • Biosecurity • Surveillance · Biosecurity • Quarantine · Surveillance Biosecurity • Seromonitoring Surveillance Compensation, Communication, Public-Private Collaboration 3

Scenario There has been an outbreak of foot and mouth disease (FMD) in your area that has affected many diary and beef farms. You live in a cattle exporting country. Assume that the official policy is humane culling of affected animals. 1. How will you begin to assess the size (magnitude) of this outbreak? 2. How will you measure progress? 2. How will you measure progress? 2. I We need to develop indicators and targets to measure trends and progress daily 2.2 Take action if disease incidence increases





| Stop the Spread: Control and Verify | |
|---|--|
| Humane Culling and Safe Disposal Reduce and contain virus transmission from affected sites | |
| Quarantine Prevent movement from suspect and confirmed cases until action is taken | |
| Biosecurity Train farmers and veterinarians to follow "Biosecurity Best Practices" | |
| Active Surveillance - Verify Look in the local area and along the value chain to confirm control | |

Find it Quickly: Daily Outbreak Report Metrics

| Subdistrict Name | Ratio of positive to negative suspect or confirmed cases | % of suspect or confirmed positive cases detected by active surveillance | No. days from onset of clinical signs to farmer reporting (days) | No. of surveillance events reported today |
|------------------|---|--|---|--|
| Subdistrict A | | | | |
| Subdistrict B | | | | |
| Subdistrict C | | | | |
| Subdistrict D | | | | |
| Subdistrict E | | | | |
| Subdistrict F | | | | |
| Subdistrict G | | | | |
| Subdistrict H | | | | |
| Total | i | % | | |

Contain it Quickly: Daily Outbreak Report Metrics

| Subdistrict Name | No. days from reporting to quarantine | No. days from reporting to culling confirmed cases | No. days from culling to burial or burning | No. days from burial to cleaning and disinfection |
|------------------|---|---|--|---|
| Subdistrict A | | | | |
| Subdistrict B | | | | |
| Subdistrict C | | | | |
| Subdistrict D | | | | |
| Subdistrict E | | | | |
| Subdistrict F | | | | |
| Subdistrict G | | | | |
| Subdistrict H | | | | |
| Total | | | | |

Stop the Spread: Daily Outbreak Report Metrics

| Subdistrict Name | No. of affected premises today (daily incidence) | Ratio of affected premises today versus previous day | Point prevalence today | Weekly prevalence estimate |
|------------------|--|---|------------------------|----------------------------------|
| Subdistrict A | | | | |
| Subdistrict B | | | | |
| Subdistrict C | | | | |
| Subdistrict D | | | | |
| Subdistrict E | | | | |
| Subdistrict F | | | | |
| Subdistrict G | | | | |
| Subdistrict H | | | | |
| Total | | | | |

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Daily Surveillance Reports by Animal-Place-Time

| Species or Type | Class | Total No. at Risk | No. Sick | No. Dead | Disease(S) Suspected (S), Probable (P) or Confirmed (C) | Notes: (Age groups, breed and sex most affected, etc) | Active (A) Passive (P) |
|--------------------|---------------------|----------------------------------|-----------------------------------|---------------|---|---|-------------------------------------|
| | Pla | ıce | | | Ti | me | |
| Region | Subdistrict Name | GPS Coordinates (Latitude) | GPS Coordinates (Longitude) | Date of Visit | Date Symptoms First Observed | Date of First Death | Date of Laboratory Submission |

13

How do we measure the effectiveness of disease control at the district level?

- 1. Define indicators and targets
- 2. Take daily action



$Frontline\ curriculum-Participant\ manual$ Indicators and Targets (Adapted, Frontline FETP) **Target** Indicator · Desired level of Statement to achievement measure achievement of an activity objective • Example: 80% of disease outbreak investigations are done within 4 hours • Example: Is disease investigation done on time? of reporting 15 Surveillance Indicators You can use in Your Local Area

| | Local A | ica. | | | | |
|--|---|--|--|---|--|--|
| District Response | District and Laboratory Response | Completeness of Subdistrict Reporting | Use of Active Surveillance | | | |
| Time to respond to farmer request for field investigation | Time from sample collection to laboratory reporting | Percentage of daily reports received from subdistrict offices | % of surveillance samples collected actively each day | _ | | |
| Percentage of district field investigations with a complete report | Time from laboratory report received by district until farmer notification | Percentage of complete disease data from each subdistrict | Number of secondary field investigations generated from a primary field investigation | | | |

Exercise 35

- 1. Form into four groups
- Develop indicators and targets to measure the effectiveness of disease control using the template provided in the next slide
- 3. Propose any changes or modifications that would be useful in your district

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Appropriate Indicators for Your Local Area: Group into country teams and fill in the cells

| District Response | Target | District and Laboratory Response | Target | Completeness of Subdistrict Reporting | Target | Use of Active Surveillance | Target |
|---|--------|--|--------|---|--------|--|--------|
| Time to respond to farmer request for field investigation | | Time from sample collection to laboratory reporting | | Percentage of daily reports received from subdistrict offices | | % of surveillance samples collected actively daily | |
| Percentage of district field investigations with a complete report | | Time from laboratory report received by district until farmer notification | | Percentage of complete disease data from each subdistrict | | Percentage of secondary field investigations generated from a primary field investigation | |

In Summary...

- *Apply GEMP principles to improve outbreak response:
 - 1. Find it quickly
 - 2. Contain it quickly
 - 3. Stop the spread
- Develop indicators and targets to measure and improve daily disease response!

| ISAVE | T Contributing Universities | | |
|--------------|-----------------------------|----|--|
| Partners | TEXAS A&M GRILIFE | | |
| Contributors | MAKERBE UNIVERSITY | 21 | |

Exercise 35 – Develop Indicators and Targets to Measure the Effectiveness of Disease Response and Control

Description of Exercise:

Prepare to measure and improve disease outbreak response in your local area

Time: 120 minutes

• Group work: 60 minutes

• Group reports and discussion: 60 minutes

Organization:

The lesson includes theory from FAO GEMP principles, a scenario exercise and a metrics exercise.

Exercise Objective(s):

- 1. Explain the health interventions model.
- 2. Explain specific principles that are used to prepare for and improve disease response following an outbreak.

Exercise Components and Structure:

- Form into four groups
- Develop indicators and targets to measure the effectiveness of disease control using the template provided in the next slide
- Propose any changes or modifications that would be useful in your local area
- Deliver group reports and participate in plenary discussion

| District Response | Target | District and Laboratory Response | Target | Completeness of Subdistrict Reporting | Target | Use of Active Surveillance | Target |
|---|--------|--|--------|---|--------|--|--------|
| Time to respond to farmer request for field investigation | | Time from sample collection to laboratory reporting | | Percentage of daily reports received from subdistrict offices | | % of surveillance samples collected actively daily | |
| Percentage of district field investigations with a complete report | | Time from laboratory report received by district until farmer notification | | Percentage of complete disease data from each subdistrict | | Percentage of secondary field investigations generated from a primary field investigation | |

Expected Outputs and Deliverables of Each Participant:

- 1. Active listening and participation in asking questions, providing targets related to response indicators to improve outbreak response.
- 2. Justify your indicators and target values you have chosen.

Lesson 32 – Characteristics of a Functional Disease Prevention and Control Programme at the Local Level

| Estimated Lesson and Exercise Time | 3 hours |
|------------------------------------|--|
| Participant Materials | ISAVET Lesson 30 Emergency Preparedness and Response Planning at the Local Level - Part 3 Participant Guide.doc |
| | Computer and Microsoft Word Pen or Pencil |

In Service Applied Veterinary Epidemiology Programme (ISAVET)

Lesson 32: Characteristics of a Functional Disease Prevention and Control Programme at the Local Level



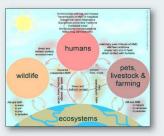
Learning Objectives

At the end of this lesson, you will be able to:

- 1. Explain the health interventions model
- 2. Explain specific principles that are used to prepare for and improve disease response following an outbreak

Current Challenges

- Population growth, consumerism and increasing protein demand
- · Increasing global trade and animal
- · Urban encroachment and land use
- · Livestock intensification
- Spill over of disease at the animalhuman-wildlife and environmental interface
- · Antimicrobial use and resistance
- · Climate change



3

Role of Frontline Veterinarians for the Prevention and Control of Animal Diseases



- Veterinary extension
- extension
 Animal production

Prevent disease introduction

- Rapid detectionProtect susceptible
- Protect susceptible populations

Contain and control disease

- Surveillance
- \bullet Field investigation



1

Responsibilities of Frontline Veterinarians for the Prevention and Control of Animal Diseases

1. Veterinary extension

2. Partnerships

o. Communication

5. Movement control

6. Humane culling

7. Safe disposal

8. Compensation

9. Vaccination and sero-monitoring

10. Disease surveillance and investigation



Veterinary Extension: Setting the Stage for Cooperation and Collaboration Benefits of Veterinary Extension Extension Services Provides preventive health promotion services to farmers to improve production Entry point to ensure cooperation and collaboration with farmers

Creates positive relationships based on mutual benefit

Measures baseline production levels and permits data collection

Improve efficiency, profits and the adoption of prevention practices

Positive working relationship is crucial for effective prevention and control of disease



Source: Google images

6

Public-Private Partnership (PPP)

- Mutual benefit for both public and private sectors including:
 - 1. Product and export certification programmes
 - Compartmentalisation
 - Food safety quality assurance programmes
 - 2. Vaccination programmes
 - FMD, PPR, Rabies, Anthrax
- PPP are often developed through extension training and education programmes at the local level



Source: Google images

Frontline curriculum – Participant manual Communication Communication channels important at the local level Chain of command (regular timetable) Laboratory and district communication Vertical Communication: Public-private sectors (farmers, producer associations) Horizontal Communication: Office of emergency services Research and education institutions 9 Veterinary Biosecurity • Personal protective actions and equipment - Hand hygiene - Disinfectants and sterilants $% \left(-1\right) =-1$ - Personal protective equipment - Prevention of bites and other animal-related injuries · Protective actions during veterinary procedures - Examination of animals - Post mortem investigations - Diagnostic specimen handling - Environmental infection control - Cleaning and disinfection of equipment - Veterinary waste disposal 10

| | | aı | 1S | | |
|---|-----|----|------------|--|--|
| Biosecurity for Veterina | Yes | | o tomastir | | |
| The veterinarian does not own or manage his/her own poultry flock. [essential] The veterinarian only visits a maximum of one poultry farm each day. [essential] | - | | | | |
| The veterinarian wears clean, dedicated clothing and footwear for each farm visit [essential] | | | | | |
| The veterinarian does not visit farms or poultry markets outside of the poultry production zone (PPZ) on the same day that he/she is visiting farms within the PPZ. Jessential! | П | | | | |
| The veterinarian does not visit farms with different poultry species on the same day. [essential] | | | | | |
| The veterinarian does not buy or sell poultry or poultry products. [essential] The veterinarian attended at least one biosecurity training course in the previous year. | - | | | | |
| The veterinaria has delived at least one observating training course in me piervous year. The veterinarian has delivered bioscentry training to farmers in the previous year. Is it true that bioscentry principles include isolation, traffic centrel, cleaning and disinfection and waste discosal? | | | | | |
| 10. On each farm visit: the veterinarian removes rings and watches, does not use his/her mobile phone. | | | | | |
| 11. The veterinarian always visits younger poultry first, before visiting older poultry. | | | | | |
| The veterinarian maintains separate "clean" and "dirty" areas in his/her vehicle. | | | | | |
| 13. The veterinarian washes his/her hands and feet and removes protective clothing after leaving a poultry house and before entering his/her vehicle. | | | | | |
| 14. Dirty clothing and footwear are placed in sealed bags and stored in the "dirty" area of the vehicle until they are washed at home. | | | | | |
| 15. The veterinarian cleans and disinfects all equipment before leaving the poultry farm. | | | | | |
| 16. The veterinarian cleans and disinfects the outside of his vehicle following each farm call. | | | | | |
| 17. The veterinarian signs the farmer's visitor logbook during each visit. | | | | | |
| 18. The veterinarian changes into clean footwear when they enter the farm block. | | | | | |
| 19. The veterinarian carries a pail, brush and disinfectant in his vehicle. | | | | | |
| The veterinarian parks his vehicle outside of the poultry farm before entering the farm. Subtotals: | | | 11 | | |

Movement Control

- Critical during an outbreak event but may also be used preventively before and following an outbreak
- The modes of transmission of disease vary and disease spread due to the movement of live animals and animal products can be controlled by movement restrictions
- · Such movement restrictions need to be well-supported by legislation
- · Livestock keepers should understand the need for restrictions
- $\boldsymbol{\cdot}$ Difficult to implement in pastoral areas and in areas where borders and boundaries are porous
- · For this reason, risk-based surveillance along the value chains that cross these boundaries is absolutely critical

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Quarantine

- $\boldsymbol{\cdot}$ Is an official extension of the biosecurity principle of isolation
- Quarantining of infected or potentially infected farms or areas
- Restrictions are placed on the movement of susceptible species animals into or out of the quarantined area until infection is considered to have been removed
- Restrictions may also be placed on the movement of people, potentially contaminated animal products and other materials
- · Enforcement require appropriate legal framework

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Humane Culling and Safe Disposal

- Prevention includes safe culling to rapidly remove sources of infectious materials
- For most animal disease emergencies, some degree of culling is likely to be necessary
- Include animal wastes and feathers
- · Involves cleansing and disinfection of properties
- \cdot Culling must be carried out in a humane manner
- · Method used varies from situation to situation

Compensation

- $\boldsymbol{\cdot}$ A compensation policy is needed for killing of animals or the destruction of property
- Should be seen as mostly an incentive to encourage rapid reporting of disease
- Not as compensation for all losses
- Inadequate or too generous can encourage behaviours that are damaging to the control efforts
- Poor compensation might encourage owners to hide or move their animals to avoid culling
- · Generous compensation encourage risky behavior

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Vaccination and Seromonitoring

- \cdot The objective of vaccination should be specified in the vaccination policy (prevention, control or eradication)
- ${\boldsymbol \cdot}$ Seromonitoring before and following vaccination is important to define prior exposure and immune response, respectively
- · The vaccination policy includes the following measures
 - Preventive vaccination
 - Emergency vaccination
- Routine vaccination
- $\boldsymbol{\cdot}$ Vaccination in the face of an outbreak is a difficult, resource intensive and expensive task
- $\boldsymbol{\cdot}$ The properties of the vaccines must be well-understood
- It is rare that vaccination alone will eradicate infection

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Animal Disease Prevention and Control Strategies (Adapted: FAO, 2006) Exposure Host Immunity Movement control Biosecurity Biosecurity Surveillance Compensation, Communication, Public-Private Collaboration Host Immunity Vaccination Biosecurity Surveillance Seromonitoring

Surveillance Before an Animal Disease Outbreak

- \cdot Goal: detect and respond to suspicious disease events as rapidly as possible.
- $\hbox{-} Surveillance options include:} \\$
 - Targeted surveillance at markets and abattoirs
 - Risk-based surveillance along value chains
 - Passive surveillance from farmers and sub-district offices
 - Active surveillance and surveys
 - Other

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Surveillance and Outbreak Investigation

Surveillance



Outbreak Investigation

Systematic ongoing collection of health and disease data that requires <u>Reporting</u> and a <u>Response ACTION</u> Systematic method to investigate suspected disease events including <u>Reporting</u> and a <u>Response ACTION</u>

Requires clear case definition

Requires clear case definition

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Exercise 36

- List the roles, responsibilities and time dedicated to:
- 1. Health promotion/extension;
- 2. Disease prevention;
- 3. Disease control.
- Provide suggestions for improving each of these activities in your district

Exercise 36: District Preparedness

- Summarise the roles and responsibilities of stakeholders in your district related to preparedness and response to animal disease emergencies.

 $Expected\ Output: Profile\ of\ district\ veterinary\ activities\ and\ suggestions\ for\ improvement.$

| Stakeholder | Area | Responsibility | Suggestion for improvement |
|-------------|------------|----------------|----------------------------|
| | Extension | | |
| | Prevention | | |
| | Control | | |
| | | | |

9.1

In Summary...

Roles and responsibilities of veterinarian:

- Health promotion (Extension)
- · Disease prevention and control
 - Surveillance
 - Biosecurity
 - Quarantine
 - Movement control
 - Humane culling
 - Vaccination and seromonitoring

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Partners Contributors Contributors TEXAS A&M TEXAS A&

Exercise 36 – Roles, Responsibilities and Time Dedicated to: Health Promotion/Extension, Disease Prevention and Disease Control at the Local Level.

Description of Exercise:

The roles and responsibilities of frontline veterinarians and various stakeholders in health promotion as well as disease prevention and control described, implementation gaps and opportunities for enhancing this collaboration will be discussed with ISAVET trainees.

Time: 2 hours

Organization:

There will be a lecture on Characteristic of a Functional Disease Prevention and Control Programme at Local area Level followed by questions posed by the facilitator and ISAVET trainees.

Exercise Objective(s):

1. Explain the roles and responsibilities of frontline veterinarians and various stakeholders in health promotion as well as disease prevention and control.

Exercise Components and Structure:

- C. Facilitator guiding questions related to the participating countries to identify the role and responsibilities of the various stakeholders in extension, disease prevention and control efforts in their respective countries. These includes:
 - List the roles, responsibilities and time dedicated to:
 - 1. Health promotion extension:
 - 2. Disease prevention:
 - 3. Disease control.
 - 4. Provide suggestions for improvement of each activities in your local area
- D. Open the floor to questions, answers and further discussion

Expected Outputs and Deliverables of Each Participant:

1. Active listening and participation in asking questions, providing examples related to the role and responsibilities of the various stakeholders at local area with regard to extension, disease prevention and control at the local area level.

Section IV: Week 4 – Field Work Training

Lesson 33 – Local Area Profile to Prepare for Field Training

Lesson 34 - Disease Profile to Prepare for Field Training

Lesson 35 – Role of Wildlife Related to the Priority Disease

Lesson 36 – Preparing for Field Work

This Manual provides the curricular contents of the Frontline In-Service Applied Veterinary Epidemiology Training (ISAVET) programme. The intended audience of the Manual are participants in Frontline (ISAVET) at national-level. The manual will serve as an FAO Global resource for National capacity development of Veterinary Services to detect and respond to emerging infectious animal diseases including transboundary animal diseases and zoonotic diseases.