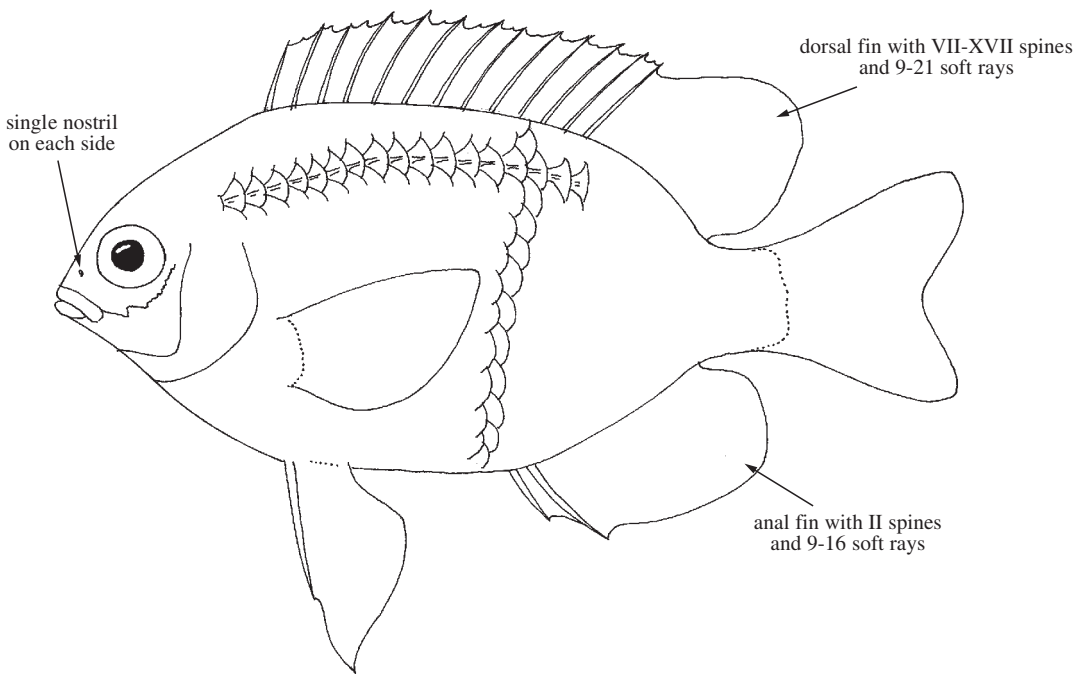
POMACENTRIDAE

Damselfishes (anemonefishes)

by J.R. Allen

Diagnostic characters: Small percoid fishes (to about 25 cm); body elongate to rounded in lateral profile, and laterally compressed. Eye medium sized, generally exceeding snout length. Mouth small, jaws equal and oblique. Small conical to incisiform teeth in jaws in a single row, sometimes with additional row of slender teeth behind primary row; teeth absent on vomer and palatines. Total gill rakers on first gill arch 10 to 85. **A single dorsal fin with VII to XVII stout spines and 9 to 21 soft rays; anal fin with II spines and 9 to 16 soft rays**; pelvic fins with I spine and 5 soft rays; pectoral fins with 14 to 22 rays; caudal fin rounded to forked. Scales comparatively large to moderate sized and ctenoid, extending onto head and median fins; cheeks and operculum scaly; lateral-line scales 12 to 60. **Colour:** extremely variable with many species exhibiting shades of brown, black, blue, green, red, and yellow; a dark spot or ocellus often present on dorsal fin, particularly in juvenile stages.

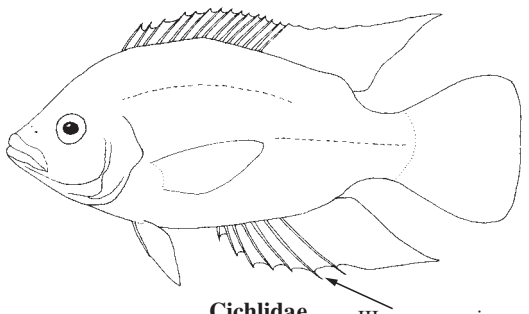


Habitat, biology, and fisheries: Coral and rocky reefs, but also common in other shallow-water habitats including sand and rubble patches, silty embayments, harbours, and seagrass beds. Pomacentrids are one of the most numerous groups occurring on tropical reefs, in terms of both number of species and number of individuals. Solitary fishes, pairs, or aggregations are equally common, depending on the species. Most species are highly territorial, especially during reproductive periods. Spawning occurs throughout most of the year at tropical localities. Eggs are deposited on the bottom and are guarded by the male. Hatching occurs within 2 to 7 days. Most species have a pelagic larval stage lasting between about 10 to 50 days, except *Acanthochromis polyacanthus*, which bypasses this stage entirely. Food habits are variable. The larger species tend to feed mainly on algae, many others are omnivorous, consuming algae and small invertebrates, while others rely on planktonic items. Larger species, such as various *Abudefduf*, are sometimes seen in markets, but the family is not considered as an important food fish. Their most important commercial use is as aquarium fishes. Anemone fishes are the leading group in this respect.

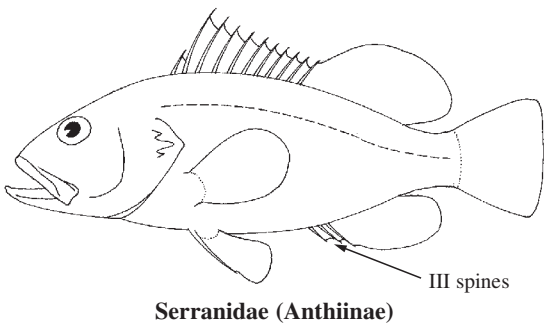
Similar families occurring in the area

Cichlidae: anal-fin spines III or more (only II in Pomacentridae); few species occurring in the area and all of these introduced fresh-water forms, commonly referred to as "tilapia".

Serranidae (genus *Pseudanthias*): generally resemble the pomacentrid genus *Chromis*, but easily distinguished by the presence of III anal-fin spines and smaller, more numerous scales (usually 35 to 65 lateral-line scales, compared to 12 to 20 in *Chromis*).



Cichlidae



Serranidae (Anthiinae)

Key to the genera of Pomacentridae occurring in the area

1a. Scales small, usually more than 50 in a longitudinal series from rear edge of opercle to base of caudal fin; all the opercles usually serrate . . . . . → *2*

1b. Scales comparatively large, usually less than 45 in a longitudinal series; all the opercles not serrate . . . . . → *3*

2a. Suborbital usually armed with a pair of elongate spines (Fig. 1); scales small, about 68 to 76 in a longitudinal series from upper edge of opercle to caudal-fin base . . . . . ***Premnas***

2b. Suborbital without pair of elongate spines, although serrae present (Fig. 2); scales moderate sized, about 50 to 60 in a longitudinal series . . . . . ***Amphiprion***

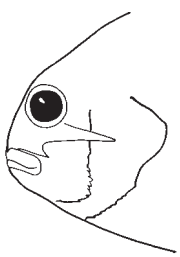


Fig. 1 ***Premnas***

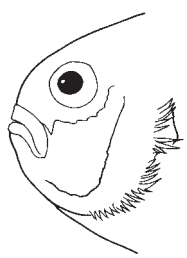


Fig. 2 ***Amphiprion***

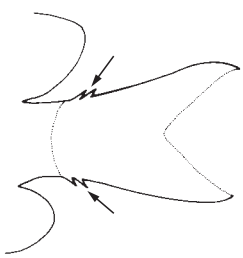
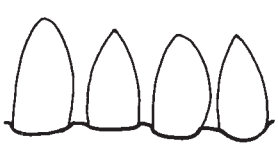


Fig. 3

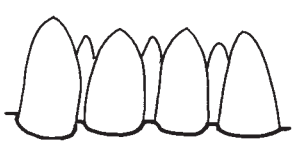
3a. Upper and lower edge of caudal-fin base usually with 2 or 3 projecting spiniform procurrent rays (Fig. 3); teeth conical, either biserial or multiserial; head usually entirely scaly except narrow region at front of snout and around each nostril . . . . . → *4*

3b. Upper and lower edge of caudal-fin base without projecting spiniform rays; teeth conical or incisiform, uniserial, or biserial (Fig. 4); head entirely scaly or frequently without scales on snout, preorbital, and suborbital . . . . . → *7*



a) conical teeth (uniserial)

b) incisiform teeth (uniserial)



c) biserial teeth

Fig. 4

4a. Dorsal-fin spines usually XVII . . . . . ***Acanthochromis***
4b. Dorsal-fin spines XII to XV . . . . . → ***5***

5a. Suborbital without scales (Fig. 5) . . . . . ***Altrichthys***
5b. Suborbital scaly . . . . . → ***6***

6a. Suborbital and hind margin of preopercle finely serrate (Fig. 6) . . . . . ***Dascyllus***
6b. Suborbital entire or covered with scales; hind margin of preopercle usually smooth (Fig. 7) . . . . . ***Chromis***

7a. Body very elongate, 2.9 to 3 times in standard length; small papilla-like structures on inner edge of posterior circumorbitals (Fig. 8) . . . . . ***Lepidozygus***
7b. Body orbiculate to elongate, usually 1.5 to 2.8 times in standard length; papilla-like structures on posterior circumorbitals absent . . . . . → ***8***

8a. Scales small, usually about 44 to 46 in longitudinal series from upper edge of gill opening to caudal-fin base; body depth 2.4 to 2.8 times in standard length . . . . . ***Teixeirichthys***
8b. Scales large, usually about 26 to 35 (except 30 to 46 in *Parma*, which has body depth 1.6 to 1.9 times in standard length) in longitudinal series from upper edge of gill opening to caudal-fin base; body depth 1.5 to 3 times in standard length . . . . . → ***9***

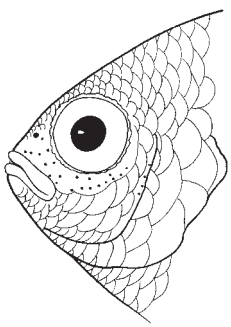


Fig. 5 ***Altrichthys***

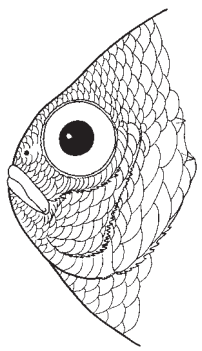


Fig. 6 ***Dascyllus***

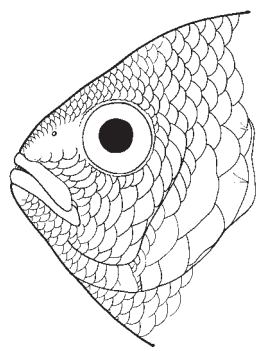


Fig. 7 ***Chromis***

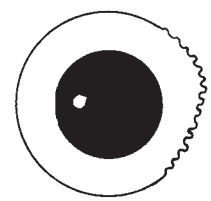
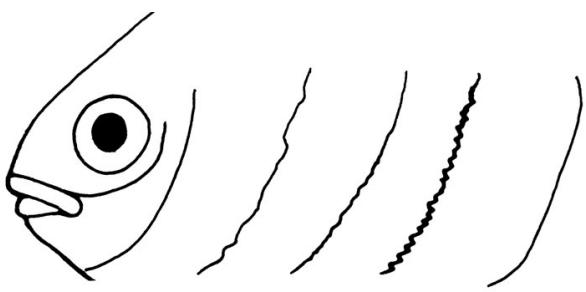


Fig. 8 eye of *Lepidozygus* showing papillae

9a. Hind margin of preopercle crenulate or weakly to strongly serrate (Fig. 9a-c) . . . . . → ***10***
9b. Hind margin of preopercle smooth (Fig. 9d) . . . . . → ***19***

10a. Lips greatly thickened, fimbriate, and curled back over snout (Fig. 10) . . . . . ***Cheiloprion***
10b. Lips normal, not greatly thickened and curled back over snout . . . . . → ***11***



a) crenulate b) weakly serrate c) strongly serrate d) smooth

Fig. 9 preopercular and suborbital serrations

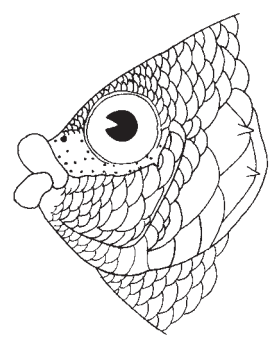


Fig. 10 ***Cheiloprion***

11a. Teeth of jaws uniserial (Fig. 11a) . . . . . → ***12***
11b. Teeth of jaws biserial (at least anteriorly) (Fig. 10b, c) . . . . . → ***15***

12a. Edge of subopercle serrate (Fig. 12) . . . . . ***Pristotis***
12b. Edge of subopercle smooth . . . . . → ***13***

13a. Suborbital scaly; snout scaly to about level of nostrils (Fig. 13) . . . . . ***Stegastes***
13b. Suborbital and snout scaleless; predorsal scales extending to about level of anterior edge of eyes . . . . . → ***14***

14a. Body elongate, its depth 2.6 to 2.9 times in standard length; dorsal-fin spines XIV (Fig. 14) . . . . . ***Pomachromis***
14b. Body ovate, its depth 2.1 to 2.3 times in standard length; dorsal-fin spines XIII . . . . . ***Amblypomacentrus***

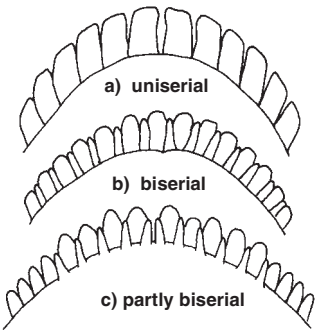


Fig. 11 pomacentrid dentition (viewed from inner side)

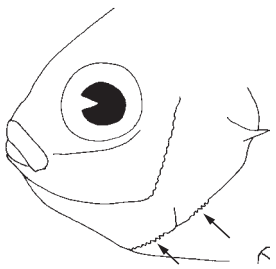


Fig. 12 ***Pristotis***

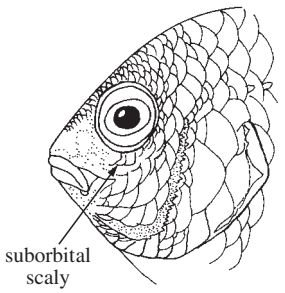


Fig. 13 ***Stegastes***

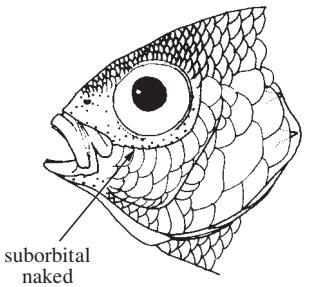


Fig. 14 ***Pomachromis***

15a. Suborbital scaleless (at least anteriorly); hind margin of preopercle distinctly serrate (Fig. 9c) . . . . . → ***16***
15b. Suborbital scaly; hind margin of preopercle crenulate or weakly serrate (Fig. 9a, b) . . . . . → ***17***

16a. Snout mostly scaleless, predorsal scales extending to about front of eyes or slightly beyond; notch between preorbital and suborbital absent (Fig. 15) . . . . . ***Dischistodus***
16b. Snout scaly to about level of nostrils or beyond; usually a prominent notch between preorbital and suborbital (much reduced in certain species) (Fig. 16) . . . . . ***Pomacentrus***

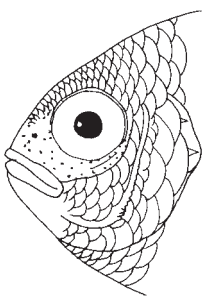


Fig. 15 ***Dischistodus***

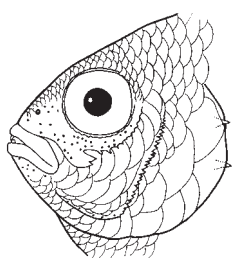


Fig. 16 ***Pomacentrus***

17a. Body more or less elongate, its depth usually 2.3 to 2.8 times in standard length; outer row of teeth with flattened tips (Fig. 17); caudal fin lunate or strongly emarginate; posterior portion of dorsal and anal fins and outer caudal-fin rays frequently produced into filaments . . . . . ***Neopomacentrus***
17b. Body ovate, its depth 2 to 2.3 times in standard length; outer row of teeth with conical tips; caudal fin rounded or slightly emarginate; outer caudal-fin rays and posterior portion of dorsal and anal fins not produced into filaments . . . . . → ***18***

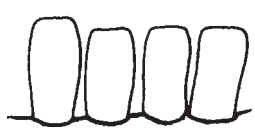


Fig. 17 teeth

18a. Dorsal- and anal-fin rays 14 or 15; pectoral-fin rays 18; slight notch between preorbital and suborbital (Fig. 18); spinous dorsal fin without dark spot at base of hindmost spines . . . ***Pomacentrus*** **(subgenus *Lepidopomacentrus*)**

18b. Dorsal- and anal-fin rays 11 to 13; pectoral-fin rays 15 or 16; notch between preorbital and suborbital absent; spinous dorsal fin with prominent dark spot at base of hindmost spine . . . . . ***Chrysiptera*** **(*C. traceyi* and *C. talboti* only)**

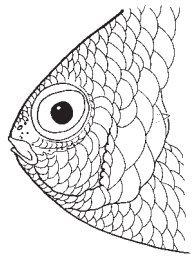


Fig. 18 ***Pomacentrus*** **(subgenus *Lepidopomacentrus*)**

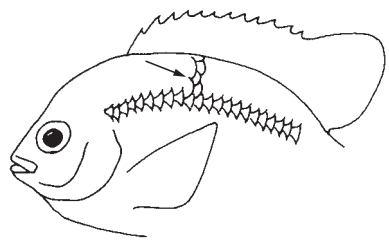


Fig. 19 ***Abudefduf***

19a. Vertical scale rows in excess of 30; horizontal scale rows below lateral line to origin of anal fin more than 14 . . . . . ***Parma***

19b. Vertical scale rows about 28; horizontal scale rows from lateral line to origin of anal fin less than 12 . . . . . → ***20***

20a. Horizontal scale rows above middle of lateral line usually 3 to 3 ½ (Fig. 19) . . . . . ***Abudefduf***

20b. Horizontal scale rows above middle of lateral line 1 to 2 ½ (usually 1 ½) . . . . . → ***21***

21a. Gill rakers on first gill arch approximately 65 to 85 . . . . . ***Hemiglyphidodon***

21b. Gill rakers on first gill arch 10 to 35 . . . . . → ***22***

22a. Dorsal-fin spines XII . . . . . ***Plectroglyphidodon***

22b. Dorsal-fin spines XIII or XIV . . . . . → ***23***

23a. Body somewhat orbiculate, its depth 1.5 to 1.8 times in standard length; longest dorsal-fin spine about equal in length to distance from tip of snout to uppermost portion of preopercle border (Fig. 20) . . . . ***Amblyglyphidodon***

23b. Body moderately deep to elongate, its depth usually 1.8 to 2.7 times in standard length; longest dorsal-fin spine usually much shorter than distance from tip of snout to uppermost portion of preopercle border . . . . . → ***24***

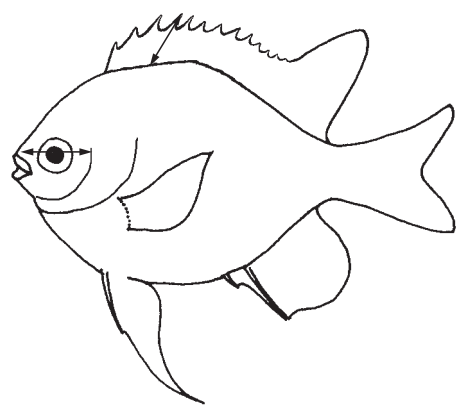


Fig. 20 ***Amblyglyphidodon***

24a. Body moderately deep, its depth usually less than 2 times in standard length; preorbital and suborbital scaly (Fig. 21); teeth biserial, at least anteriorly (Fig. 11c) . . ***Neoglyphidodon***

24b. Body ovate to elongate, its depth usually greater than 2 times in standard length; preorbital and suborbital scaly or naked (Fig. 22); teeth biserial or uniserial (Fig. 11a, b) . . . . . ***Chrysiptera***

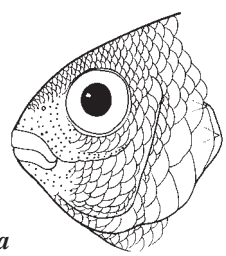


Fig. 21 ***Neoglyphidodon***

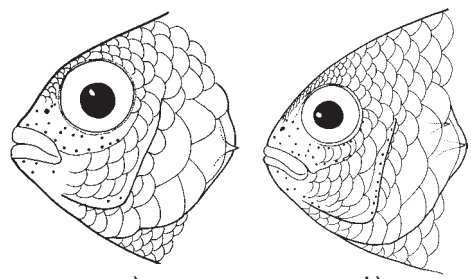


Fig. 22 ***Chrysiptera***

List of species occurring in the area

The symbol is given when species accounts are included.

Abudefduf bengalensis (Bloch, 1787)
Abudefduf lorenzi Hensley and Allen, 1977
Abudefduf notatus (Day, 1869)
Abudefduf septemfasciatus (Cuvier, 1830)
Abudefduf sexfasciatus Lacepède, 1802
Abudefduf sordidus Forsskål, 1775
Abudefduf vaigiensis (Quoy and Gaimard, 1825)
Abudefduf whitleyi Allen and Robertson, 1974

Acanthochromis polyacantha (Bleeker, 1855)

Altrichthys azurelineatus (Fowler and Bean, 1928)
Altrichthys curatus Allen, 1999

Amblyglyphidodon aureus (Cuvier, 1830)
Amblyglyphidodon azurelineatus (Fowler and Bean, 1928)
Amblyglyphidodon batunai Allen, 1995
Amblyglyphidodon curacao (Bloch, 1787)
Amblyglyphidodon leucogaster (Bleeker, 1847)
Amblyglyphidodon ternatensis (Bleeker, 1853)

Amblypomacentrus breviceps (Schlegel and Müller, 1839-44)
Amblypomacentrus clarus Allen and Adrim, 2000

Amphiprion akallopisos Bleeker, 1853
Amphiprion akindynos Allen, 1972
Amphiprion chrysopterus Cuvier, 1830
Amphiprion clarkii (Bennett, 1830)
Amphiprion ephippium (Bloch, 1790)
Amphiprion frenatus Brevoort, 1856
Amphiprion latezonatus Waite, 1900
Amphiprion leucokranos Allen, 1973
Amphiprion mccullochi Whitley, 1929
Amphiprion melanopus Bleeker, 1852
Amphiprion ocellaris (Cuvier, 1830)
Amphiprion percula (Lacepède, 1802)
Amphiprion perideraion Bleeker, 1855
Amphiprion polymnus (Linnaeus, 1758)
Amphiprion rubrocinctus Richardson, 1842
Amphiprion sandaracinos Allen, 1972
Amphiprion sebae Bleeker, 1853
Amphiprion thiellei Burgess, 1981
Amphiprion tricinctus Schultz and Welander, 1953

Cheiloprion labiatus (Day, 1877)

Chromis abyssicola Allen and Randall, 1985
Chromis acares Randall and Swerdloff, 1973
Chromis agilis Smith, 1960
Chromis alleni Randall, Ida, and Moyer, 1981
Chromis alpha Randall, 1988
Chromis amboinensis (Bleeker, 1873)
Chromis analis (Cuvier, 1830)
Chromis atripectoralis Welander and Schultz, 1951
Chromis atripes Fowler and Bean, 1928
Chromis axillaris (Bennett, 1831)
Chromis caudalis Randall, 1988
Chromis chrysura (Bliss, 1883)
Chromis cinerascens (Cuvier, 1830)
Chromis delta Randall, 1988

Chromis dimidiata (Klunzinger, 1871)
Chromis elerae Fowler and Bean, 1928
Chromis flavipectoralis Randall, 1989
Chromis flavomaculata Kamohara, 1960
Chromis fumea (Tanaka, 1917)
Chromis hypsilepis (Günther, 1867)
Chromis iomelas Jordan and Seale, 1906
Chromis klunzingeri Whitley, 1929
Chromis lepidolepis Bleeker, 1877
Chromis leucura Gilbert, 1905
Chromis lineata Fowler and Bean, 1928
Chromis margaritifer Fowler, 1946
Chromis mirationis Tanaka, 1917
Chromis nigrura Smith, 1960
Chromis nitida (Whitley, 1928)
Chromis opercularis (Günther, 1867)
Chromis ovatiformes Fowler, 1946
Chromis retrofasciata Weber, 1913
Chromis scotochiloptera Fowler, 1918
Chromis ternatensis (Bleeker, 1856)
Chromis vanderbilti Fowler, 1941
Chromis viridis (Cuvier, 1830)
Chromis weberi Fowler and Bean, 1928
Chromis westaustralis Allen, 1976
Chromis xanthochira (Bleeker, 1851)
Chromis xanthura (Bleeker, 1854)

Chrysiptera biocellata (Quoy and Gaimard, 1824)
Chrysiptera bleekeri (Fowler and Bean, 1928)
Chrysiptera brownriggii (Bennett, 1828) [formerly *C. leucopoma*]
Chrysiptera caruleolineata (Allen, 1973)
Chrysiptera cyanea (Quoy and Gaimard, 1824)
Chrysiptera cymatilis Allen, 1999
Chrysiptera flavipinnis (Allen and Robertson, 1974)
Chrysiptera galba (Allen and Randall, 1974)
Chrysiptera glauca (Cuvier, 1830)
Chrysiptera hemicyanea (Weber, 1913)
Chrysiptera niger (Allen, 1975)
Chrysiptera notialis (Allen, 1975)
Chrysiptera oxycephala (Bleeker, 1877)
Chrysiptera parasema (Fowler, 1918)
Chrysiptera pricei Allen and Adrim, 1992
Chrysiptera rex (Snyder, 1909)
Chrysiptera rollandi (Whitley, 1961)
Chrysiptera sinclairi Allen, 1987
Chrysiptera springeri Allen and Lubbock, 1976
Chrysiptera starcki (Allen, 1973)
Chrysiptera talboti (Allen, 1975)
Chrysiptera taupou (Jordan and Seale, 1906)
Chrysiptera trayceyi (Woods and Schultz, 1960)
Chrysiptera tricincta (Allen and Randall, 1974)
Chrysiptera unimaculata (Cuvier, 1830)

Dascyllus aruanus (Linnaeus, 1758)
Dascyllus carneus Fischer, 1885
Dascyllus flavicaudus Randall and Allen, 1974
Dascyllus melanurus Bleeker, 1854
Dascyllus reticulatus (Richardson, 1846)
Dascyllus strasburgi Klausewitz, 1960
Dascyllus trimaculatus (Rüppell, 1928)

Dischistodus chrysopoecilus (Schlegel and Müller, 1839)
Dischistodus fasciatus (Cuvier, 1830)
Dischistodus melanotus (Bleeker, 1858)
Dischistodus perspicillatus (Cuvier, 1830)
Dischistodus prosopotaenia (Bleeker, 1852)
Dischistodus pseudochrysopoecilus Allen and Robertson, 1974

Hemiglyphidodon plagiometopon (Bleeker, 1852)

Lepidozygus tapeinosoma (Bleeker, 1856)

Mecaenichthys immaculatus (Ogilby, 1885)

Neoglyphidodon bonang (Bleeker, 1853)
Neoglyphidodon carlsoni (Allen, 1975)
Neoglyphidodon crossi Allen, 1991
Neoglyphidodon melas (Cuvier, 1830)
Neoglyphidodon nigroris (Cuvier, 1830)
Neoglyphidodon oxyodon (Bleeker, 1857)
Neoglyphidodon polyacanthus (Ogilby, 1889)
Neoglyphidodon thoracotaeniatus (Fowler and Bean, 1928)

Neopomacentrus anabatoides (Bleeker, 1847)
Neopomacentrus azysron (Bleeker, 1877)
Neopomacentrus bankieri (Richardson, 1846)
Neopomacentrus cyanomos (Bleeker, 1856)
Neopomacentrus filamentosus (Macleay, 1833)
Neopomacentrus metallicus (Jordan and Seale, 1906)
Neopomacentrus nemurus (Bleeker, 1857)
Neopomacentrus taeniurus (Bleeker, 1856)
Neopomacentrus violascens (Bleeker, 1848)

Parma alboscapularis Allen and Hoese, 1975
Parma bicolor Allen and Larson, 1979
Parma mccullochi Whitley, 1929
Parma microlepis Günther, 1862
Parma occidentalis Allen and Hoese, 1975
Parma oligolepis Whitley, 1929
Parma polylepis Günther, 1862
Parma unifasciata (Steindachner, 1867)

Plectroglyphidodon dickii (Lienard, 1839)
Plectroglyphidodon flaviventris Allen and Randall, 1974
Plectroglyphidodon imparipennis (Vaillant and Sauvage, 1875)
Plectroglyphidodon johnstonianus Fowler and Ball, 1924
Plectroglyphidodon lacrymatus (Quoy and Gaimard, 1824)
Plectroglyphidodon leucozonus (Bleeker, 1859)
Plectroglyphidodon phoenixensis (Schultz, 1943)

Pomacentrus adelus Allen, 1991
Pomacentrus albimaculus Allen, 1975
Pomacentrus alexanderae Evermann and Seale, 1907
Pomacentrus alleni Burgess, 1981
Pomacentrus amboinensis Bleeker, 1868
Pomacentrus armillatus Allen, 1993
Pomacentrus auriventris Allen, 1991
Pomacentrus australis Allen and Robertson, 1973
Pomacentrus azuremaculatus Allen, 1991
Pomacentrus bankanensis Bleeker, 1853
Pomacentrur bintanensis Allen, 1999
Pomacentrus brachialis Cuvier, 1830
Pomacentrus burroughi Fowler, 1918
Pomacentrus chrysurus Cuvier, 1830

Pomacentrus coelestis Jordan and Starks, 1901
Pomacentrus colini Allen, 1991
Pomacentrus cuneatus Allen, 1991
Pomacentrus emarginatus Cuvier, 1830
Pomacentrus geminospilus Allen, 1993
Pomacentrus grammorhynchus Fowler, 1918
Pomacentrus imitator (Whitley, 1964)
Pomacentrus javanicus Allen, 1991
Pomacentrus komodoensis Allen, 1999
Pomacentrus lepidogenys Fowler and Bean, 1928
Pomacentrus limosus Allen, 1992
Pomacentrus littoralis Cuvier, 1830
Pomacentrus melanochir Bleeker, 1877
Pomacentrus milleri Taylor, 1964
Pomacentrus moluccensis Bleeker, 1853
Pomacentrus nagasakiensis Tanaka, 1917
Pomacentrus nigromanus Weber, 1913
Pomacentrus nigromarginatus Allen, 1973
Pomacentrus opisthostigma Fowler, 1918
Pomacentrus pavo (Bloch, 1878)
Pomacentrus philippinus Evermann and Seale, 1907
Pomacentrus polyspinus Allen, 1991
Pomacentrus reidi Fowler and Bean, 1928
Pomacentrus saksonoi Allen, 1995
Pomacentrus similis Allen, 1991
Pomacentrus simsiang Bleeker, 1856
Pomacentrus smithi Fowler and Bean, 1928
Pomacentrus stigma Fowler and Bean, 1928
Pomacentrus taeniometopon Bleeker, 1852
Pomacentrus tripunctatus Cuvier, 1830
Pomacentrus vaiuli Jordan and Seale, 1906
Pomacentrus wardi Whitley, 1927
Pomacentrus xanthosternus Allen, 1991

Pomachromis exilis Allen and Emery, 1972
Pomachromis fuscidorsalis Allen and Randall, 1974
Pomachromis guamensis Allen and Larson, 1975
Pomachromis richardsoni (Snyder, 1909)

Premnas biaculeatus (Bloch, 1790)

Pristotis jerdoni (Day, 1873)

Stegastes albifasciatus (Schlegel and Müller, 1839)
Stegastes apicalis (De Vis, 1885)
Stegastes aureus (Fowler, 1927)
Stegastes emeryi Allen and Randall, 1974
Stegastes fasciolatus (Ogilby, 1889)
Stegastes gascoynei (Whitley, 1964)
Stegastes insularis Allen and Emery, 1985
Stegastes lividus (Bloch and Schneider, 1801)
Stegastes nigricans (Lacepède, 1802)
Stegastes obreptus (Whitley, 1948)

References

Allen, G.R. 1972. *Anemonefishes*. Neptune City, New Jersey, T.F.H. Publications, Inc., 288 p.

Allen, G.R. 1975. *Damselfishes of the south seas*. Neptune City, New Jersey. T.F.H. Publications, Inc., 240 p.

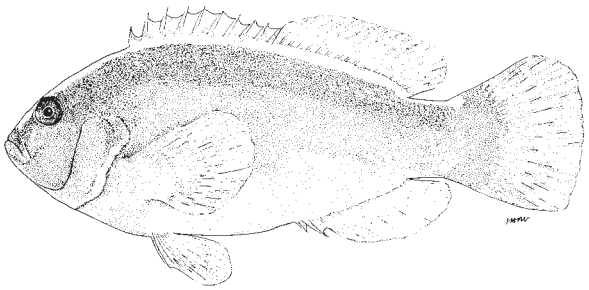
Allen, G.R. 1975. *The anemone fishes. Their classification and biology*. Second edition. Neptune City, New Jersey, T.F.H. Publications, Inc., 352 p.

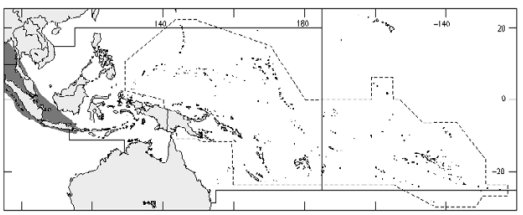
Allen, G.R. 1991. *Damselfishes of the world*. Melle, Germany, Mergus Publishers, 271 p.

Amphiprion akallopisos Bleeker, 1853

En - Skunk anemonefish.

Maximum total length about 10 to 11 cm. Coral reefs of lagoons and seaward reefs to depths of at least 15 m. Associated with the large anemones; frequently an adult pair and several smaller fishes with each anemone. Feeds on zooplankton, small benthic invertebrates, and algae. Used as an aquarium fish; caught with hand nets. Widespread in the Indian Ocean, ranging to the west coast of Thailand and southern coasts Sumatra and Java, also occurs in the Java Sea and around Bali.

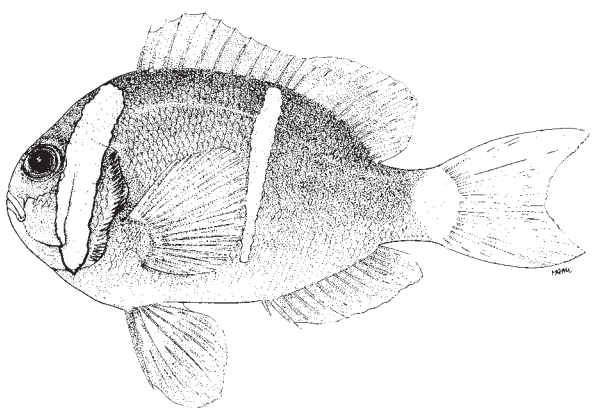


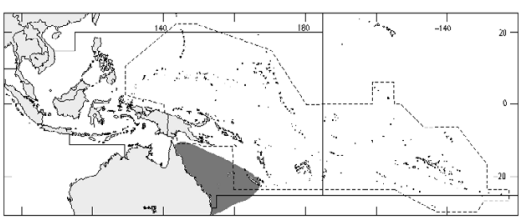


Amphiprion akindynos Allen, 1972

En - Barrier reef anemonefish.

Maximum total length about 12 to 13 cm. Coral reefs of lagoons and seaward reefs to depths of at least 20 m. Associated with the large anemones; frequently an adult pair and several smaller fish with each anemone. Feeds on zooplankton, small benthic invertebrates, and algae. Used as an aquarium fish; caught with hand nets. Great Barrier Reef and adjacent Coral Sea to New Caledonia and the Loyalty Islands.

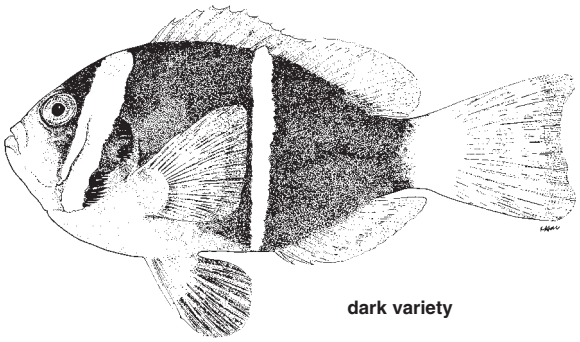




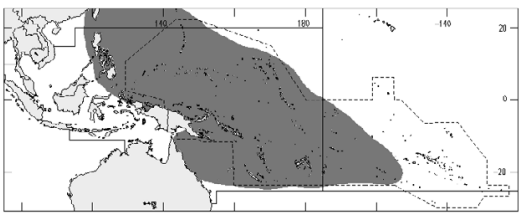
Amphiprion chrysopterus Cuvier, 1830

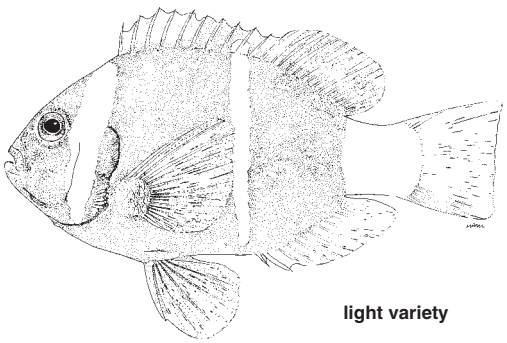
En - Orangefin anemonefish.

Maximum total length about 15 cm. Coral reefs of lagoons and seaward reefs to depths of at least 20 m. Associated with the large anemones; frequently an adult pair and several smaller fish with each anemone. Feeds on zooplankton, small benthic invertebrates, and algae. Used as an aquarium fish; caught with hand nets. Widespread in the western Pacific including New Guinea, Coral Sea, New Britain, Solomon Islands, Vanuatu, Fiji, Caroline Islands, Mariana Islands, Gilbert Islands, Samoa, Society Islands, and Tuamotu Islands.

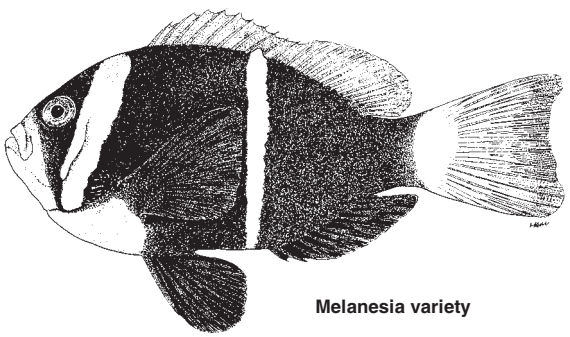


dark variety





light variety

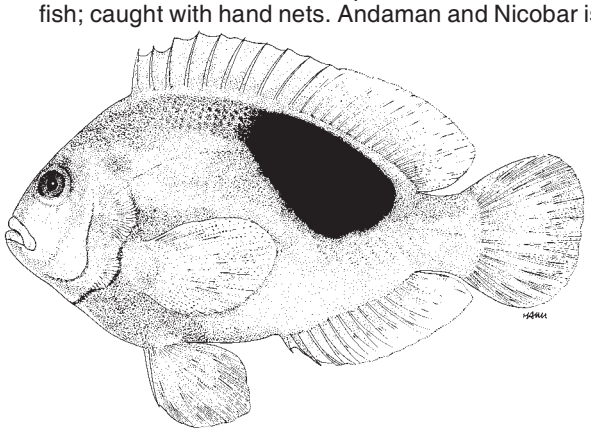


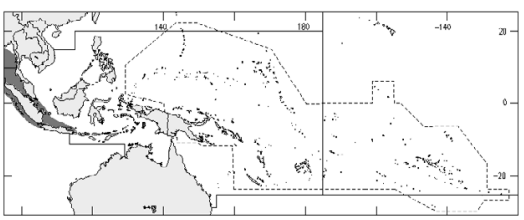
Melanesia variety

Amphiprion ephippium (Bloch, 1790)

En - Red saddleback anemonefish.

Maximum total length about 11 cm. Coral reefs of lagoons and seaward reefs to depths of at least 12 m. Associated with the large anemones; frequently an adult pair and several smaller fish with each anemone. Feeds on zooplankton, small benthic invertebrates, and algae. Used as an aquarium fish; caught with hand nets. Andaman and Nicobar islands, Thailand, Malaysia, Sumatra, and Java.

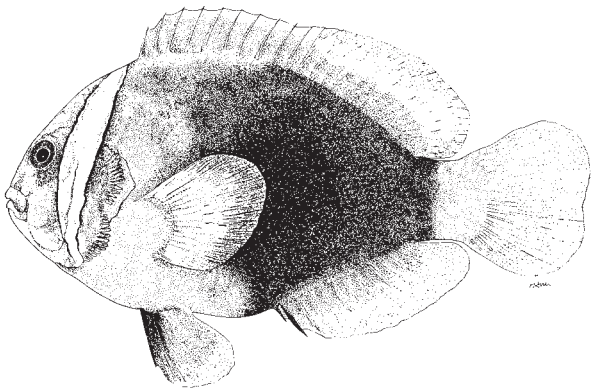


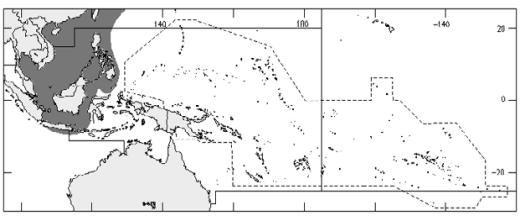


Amphiprion frenatus Brevoort, 1856

En - Tomato anemonefish.

Maximum total length about 14 cm. Coral reefs of lagoons and seaward reefs to depths of at least 12 m. Associated with the large anemones, frequently an adult pair and several smaller fish with each anemone, male partner is considerably smaller than the female. Feeds on zooplankton, small benthic invertebrates, and algae. An important aquarium fish; caught with hand nets. South China Sea and adjacent areas, north to Japan.

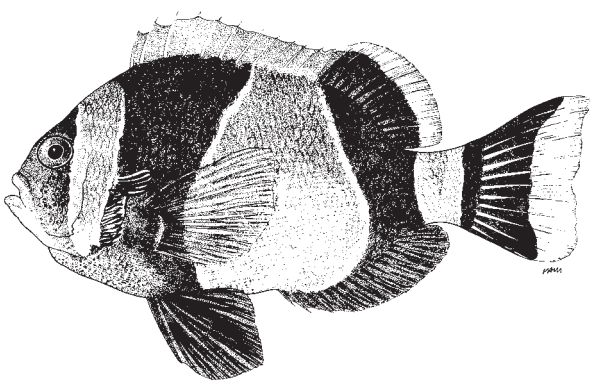


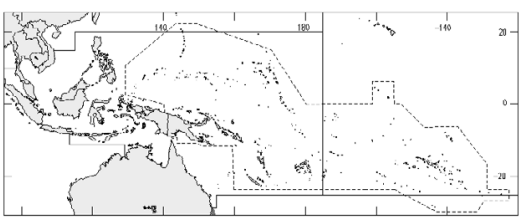


Amphiprion latezonatus Waite, 1900

En - Wideband anemonefish.

Maximum total length about 14 cm. Coral and rocky reefs to depths of at least 40 m. Associated with the large anemones, frequently an adult pair and several smaller fish with each anemone. Feeds on zooplankton, small benthic invertebrates, and algae. Used as an aquarium fish; caught with hand nets. Lord Howe Island off southeastern Australia and mainland reefs near the Queensland-New South Wales border.

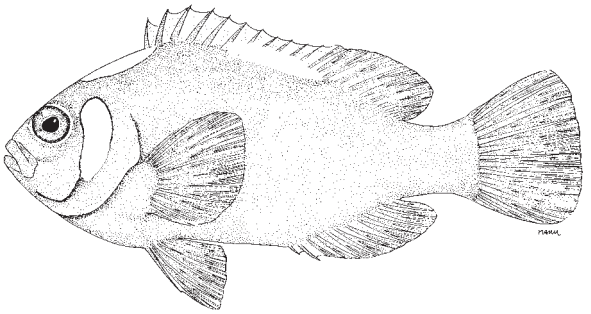


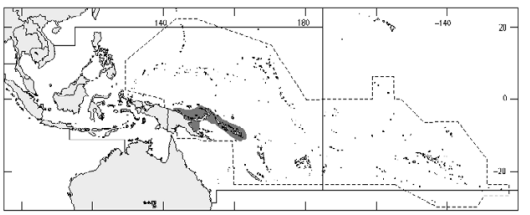


Amphiprion leucokranos Allen, 1973

En - Whitebonnet anemonefish.

Maximum total length about 9 cm. Coral reefs of lagoons and seaward reefs to depths of at least 15 m. Associated with the large anemones, usually solitary individuals or pairs. Feeds on zooplankton, small benthic invertebrates, and algae. Sometimes seen in the aquarium trade; caught with hand nets. Northern New Guinea, including Manus Island, also New Britain and the Solomon Islands.

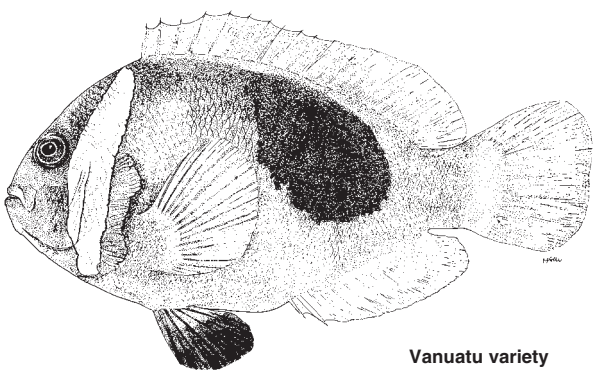




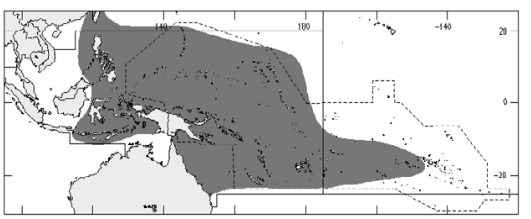
Amphiprion melanopus Bleeker, 1852

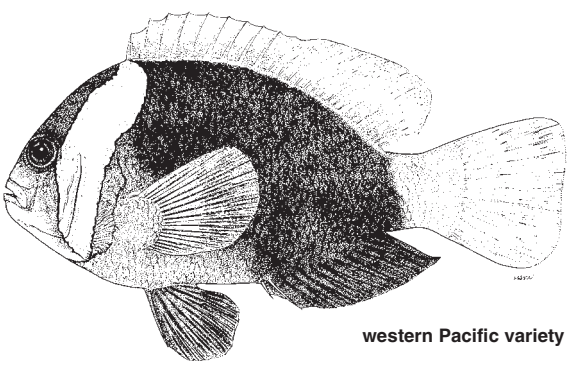
En - Red-and-black anemonefish.

Maximum total length about 12 cm. Coral reefs of lagoons and seaward reefs to depths of at least 10 m. Usually associated with colonial anemones, frequently an adult pair and several smaller fish with each cluster of anemones. Feeds on zooplankton, small benthic invertebrates, and algae. An important aquarium fish; caught with hand nets. Western Pacific, including Indonesia (Bali eastwards), Melanesia, Micronesia, southeastern Polynesia (to Society Islands), and from Great Barrier Reef to Coral Sea. Fish from Fiji to the Society Islands have a uniform red ground colour and some fish from the Coral Sea lack a white bar on the head.



Vanuatu variety





western Pacific variety