



Food and Agriculture
Organization of the
United Nations



International Commission on
Poplars and Other Fast-Growing
Trees Sustaining People and the
Environment

Activities Related to the Cultivation and Utilization of Poplars, Willows, and Other Fast-Growing Trees in the United States, 2020-2023

UNITED STATES REPORT TO THE 27TH SESSION OF THE INTERNATIONAL COMMISSION ON POPLARS AND OTHER FAST-GROWING TREES SUSTAINING PEOPLE AND THE ENVIRONMENT (IPC) OF THE FOOD AND AGRICULTURE ORGANIZATION OF THE UNITED NATIONS (UN FAO)

August 2024



Activities Related to the Cultivation and Utilization of Poplars, Willows, and Other Fast-Growing Trees in the United States, 2020-2023

August 2024

United States Report to the
International Commission on Poplars and Other Fast-Growing Trees Sustaining People and the
Environment (IPC) of the Food and Agriculture Organization of the United Nations (UN FAO)

27th Session
Bordeaux, France
21-25 October 2024

Prepared by:

Ronald S. Zalesny Jr.

United States National Representative to the IPC
Supervisory Research Plant Geneticist
USDA Forest Service | Northern Research Station
Rhineland, Wisconsin, USA
ronald.zalesny@usda.gov

Elizabeth R. Rogers

Pathways Intern
USDA Forest Service | Northern Research Station
Rhineland, Wisconsin, USA
elizabeth.r.rogers@usda.gov



ACKNOWLEDGEMENTS

This report was developed by Ron Zalesny Jr. and Liz Rogers as part of their official duties as research employees of the USDA Forest Service. The findings and conclusions in this report are those of the authors and should not be construed to represent any official USDA or United States Government determination or policy. Along with others¹, the authors presented portions of this information at the [2024 Short Rotation Woody Crops International Conference](#); 13-16 May 2024; Columbia, Missouri, USA. Ron Zalesny Jr. is grateful to Emile Gardiner, previous United States National Representative to the IPC (2012-2022), for his insight about the IPC, technical knowledge of fast-growing trees, and mentorship. The authors thank: 1) Sonja Oswalt for data analysis and generating the summary statistics in section IV, 2) Dana Mitchell and Carlos Rodriguez Franco for providing policy-related information, 3) Nathaniel Rogers and Ryan Vinhal for technical support of the literature database, 4) Deahn Donner, Adam Fullerton, Beth Larry, Dana Nelson, and Ryan Vinhal for reviewing earlier versions of the report, and 5) the following partners and their organizations for contributing information.

Partner	Organization
J-A. Adorno	University of Connecticut, Storrs, Connecticut, USA
C. Babor	University of Connecticut, Storrs, Connecticut, USA
S.L. Doty	University of Washington, Seattle, Washington, USA
J. DuPlissis	University of Minnesota Duluth NRRI, Duluth, Minnesota, USA
A. Fullerton	USDA Forest Service, Washington, District of Columbia, USA
B. Garzon	University of Connecticut, Storrs, Connecticut, USA
S.B. Ghezehei	North Carolina State University, Raleigh, North Carolina, USA
A. Groover	USDA Forest Service, Burlington, Vermont, USA
D.W. Hazel	North Carolina State University, Raleigh, North Carolina, USA
A. Himes	Mississippi State University, Mississippi State, Mississippi, USA
J.G. Isebrands	Environmental Forestry Consultants, New London, Wisconsin, USA
J. Jackson	University of Minnesota Duluth NRRI, Duluth, Minnesota, USA
J. Kuzovkina	University of Connecticut, Storrs, Connecticut, USA
W.F. Lazarus	University of Minnesota, St. Paul, Minnesota, USA
P.N. McGovern	Open4st, Grand Rapids, Michigan, USA
D. Mitchell	US Department of Energy, Auburn, Alabama, USA
E.G. Nichols	North Carolina State University, Raleigh, North Carolina, USA
C. O'Dea	USDA Forest Service, Washington, District of Columbia, USA
H.J. Renninger	Mississippi State University, Mississippi State, Mississippi, USA
D.L. Rockwood	Florida FGT LLC, Orlando, Florida, USA
C. Rodriguez Franco	USDA Forest Service, Washington, District of Columbia, USA
M. Schillberg	University of Connecticut, Storrs, Connecticut, USA
C.M. Siegert	Mississippi State University, Mississippi State, Mississippi, USA
L.B. Smart	Cornell University, Geneva, New York, USA
N.F. Sonti	USDA Forest Service, Baltimore, Maryland, USA
S.H. Strauss	Oregon State University, Corvallis, Oregon, USA
T.J. Tschaplinski	Oak Ridge National Laboratory, Oak Ridge, Tennessee, USA
G.A. Tuskan	Oak Ridge National Laboratory, Oak Ridge, Tennessee, USA
L. Urban	University of Connecticut, Storrs, Connecticut, USA
R.A. Vinhal	USDA Forest Service, Rhinelander, Wisconsin, USA
K. Zobrist	Washington State University, Snohomish, Washington, USA

¹ Zalesny, R.S. Jr., Rogers, E.R., Vinhal, R.A., Adorno, J-A., Babor, C., Doty, S.L., Garzon, B., Ghezehei, S.B., Groover, A., Hazel, D.W., Himes, A., Kuzovkina, J., Nichols, E.G., Renninger, H.J., Rockwood, D.L., Schillberg, M., Siegert, C.M., Strauss, S.H., Tschaplinski, T.J., Tuskan, G.A., & Urban, L. (2024). Research and development of fast-growing trees in the United States from 2020 to 2023: a report from the International Commission on Poplars and Other Fast-Growing Trees Sustaining People and the Environment (IPC) of the Food and Agriculture Organization of the United Nations (UN FAO). In: 14th Biennial Short Rotation Woody Crops Operations Working Group Conference: 2024 Short Rotation Woody Crops International Conference, The Future is Green: Integrating Short Rotation Woody Crops (SRWC), Agroforestry, and Ecosystem Services for Sustainable, Productive Landscapes; 13-16 May 2024; Columbia, Missouri, USA.

TABLE OF CONTENTS

I. POLICY AND LEGAL FRAMEWORK

1. Introduction	4
2. National Laws, Policies, and Regulations	5

II. TECHNICAL INFORMATION

1. Taxonomy, Nomenclature, and Registration	
a) <i>Populus</i>	7
b) <i>Salix</i>	7
c) <i>Eucalyptus</i>	8
2. Domestication, Breeding, and Selection of Genetic Resources	
a) <i>Populus</i>	9
b) <i>Salix</i>	11
c) <i>Eucalyptus</i>	12
3. Plant Health, Resilience to Threats and Climate Change	
a) Biotic Factors	14
b) Abiotic Factors	15
c) Resilience to Threats and Climate Change	16
4. Production Systems for the Bioeconomy	
a) Nursery Practices and Propagation Techniques	16
b) Planted Forests	17
c) Naturally Regenerating Forest	20
d) Agroforestry and Trees Outside Forest	21
e) Harvesting	22
f) Utilization for Wood Products	22
g) Utilization for Bioenergy	23
5. Environmental and Ecosystem Services	
a) Site and Landscape Improvement	24
b) Phytoremediation and Associated Phytotechnologies	25
c) Environmental Applications for Ecosystem Services	26

III. GENERAL INFORMATION

1. Administration and Operation of the National Poplar Commission or Equivalent	
a) USDA Forest Service as an “Other Suitable National Body”	27
b) Meetings, Congresses, and Study Tours Sponsored by IPC-Affiliated Organizations	28
2. Relations with Other Countries	29
3. Innovations not Included in Other Sections	30
4. Literature	38

IV. SUMMARY STATISTICS, 2020-2023	128
---	-----

I. POLICY AND LEGAL FRAMEWORK

1. Introduction

Substantial investments in major economic, policy, and technological development affected activities related to the cultivation and utilization of poplars, willows, and eucalypts in the United States (US) from 2020-2023. Funding for research on the development and application of fast-growing trees came from private and public sectors, and significant shifts in funding priorities occurred across both sectors throughout the reporting period. The [National Science Board \(NSB\)](#), which governs the National Science Foundation (NSF), develops science and engineering indicators, and advises the President and Congress on science and engineering policy, research, and education issues. The NSB publishes a suite of reports related to Science and Engineering Indicators. In particular, the information presented in the following bullets is quoted verbatim from the 2024 report¹, [Research and Development: U.S. Trends and International Comparisons](#).

- ☛ In 2022, the United States performed an estimated \$885.6 billion in research and development (R&D) in current U.S. dollars. This is an increase from 2021 of 12% in current (nominal) dollars and a 5% increase in constant (inflation-adjusted) dollars.
- ☛ The business sector is by far the largest performer of U.S. R&D. In 2022, this sector performed an estimated \$692.7 billion in domestic R&D (current U.S. dollars), or 78% of U.S. R&D, a 14% increase from the \$608.6 billion performed in 2021 (6% increase in constant dollars).
- ☛ The second-largest performing sector in 2022 was higher education, with \$91.4 billion (or 10% of the U.S. R&D total). This represented a 7% increase from 2021 in current dollars, but performance stagnated in constant dollars (-0.4% change). In 2022, the federal government performed \$73.3 billion, for an 8% share of U.S. R&D, compared with \$66.8 billion in 2021, for a 10% increase (3% in constant dollars).
- ☛ In addition to being the largest performer, the business sector is also the largest R&D funder in the United States. In 2022, the sector funded \$672.9 billion, or 76% of total U.S. R&D, up from 69% in 2000 and 61% in 2010.
- ☛ The federal government funded 18% of U.S. R&D (\$159.8 billion dollars) in 2022 as the second-largest source. The federal government funds the largest proportion of U.S. basic research performance (40%).
- ☛ Across all agencies in FY 2022, 24% of federal R&D obligations were devoted to basic research (\$45.4 billion), 25% to applied research (\$48.4 billion), and 51% to experimental development (\$96.6 billion).

Three federal agencies exhibiting the greatest potential relevance to the cultivation of poplars, willows, and eucalypts in natural forests, planted forests, and agroforestry throughout the country had federal obligations for R&D that increased throughout the 2020, 2021, and 2022 fiscal years¹. Specifically, federal R&D obligations for the US Department of Energy (DOE) increased nearly 13.9% from FY2020 to FY2022, while that for the US Department of the Interior (US DOI) and US Environmental Protection Agency (US EPA) increased 12.2% and 7.7%, respectively. Federal R&D obligations for the US Department of Agriculture (USDA), within which the USDA Forest Service resides, were 3.4% less in FY2022 than FY2020.

2. National Laws, Policies, and Regulations

In the US, there were substantial developments throughout the reporting period (2020-2023) for longstanding national policies, as well as new enactments that have or may affect the cultivation of poplars, willows, and eucalypts in natural forests, planted forests, and agroforestry throughout the country. These national policies focus on the production of renewable fuels, domestic energy security, resilience to climate change, restoration of degraded lands and waters, and rural economic development. Of particular note is the ***Energy Independence and Security Act (EISA) of 2007*** ([Public Law 110-140](#); 19 December 2007). Enacted long before the reporting period, the EISA aimed to: 1) increase US energy security, 2) improve vehicle fuel economy, and 3) **develop renewable energy**. For the latter, EISA contained a National Renewable Fuel Standard (RFS) requiring fuel producers to use at least 36 billion gallons of biofuels by 2022, with 21 billion gallons being cellulosic biofuels. Estimates for this RFS were informed by an interdepartmental partnership between the US DOE and USDA that resulted in the release of a [2005 US Billion-Ton Report](#) that demonstrated how agriculture and forest resources could potentially produce an annual sustainable output of one billion dry tons of biomass to displace approximately 30% of national petroleum consumption¹. In 2011, the US DOE released a [2011 US Billion-Ton Update Report](#) that provided more rigorous assessment of availability, prices, and resource sustainability in the context of implementation of the revised RFS and other policy initiatives². The *2011 US Billion-Ton Update Report* greatly increased the potential of energy crops, including poplars, willows, and other fast-growing trees, for meeting projected feedstock demands necessary to support future supplies of renewable fuels. In 2016, the US DOE published the [2016 US Billion-Ton Report](#) which supported earlier conclusions that the US could sustain production of sufficient renewable biomass to achieve the RFS target established for 2022, and the report included a more in-depth analysis of the role that poplars, willows, eucalypts, and other fast-growing trees could play in reaching renewable energy goals out to 2040³. Just after the current reporting period (2020-2023), in May 2024, the US DOE Bioenergy Technologies Office (BETO) released its fourth iteration of the billion-ton reports, the [2023 Billion-Ton Report](#). Among its key outcomes, the US can produce 1.1 to 1.5 billion tons of biomass annually in a future mature market, with the largest source of biomass being from purpose-grown energy crops⁴.

Similarly, the ***Energy Act of 2020*** that was incorporated into the ***Consolidated Appropriations Act of 2021*** ([Public Law 116-260](#); 27 December 2020) aims to modernize US energy policies to encourage innovation in technologies that are important for energy, national security, and environmental protection. In addition, the *Energy Act of 2020* authorizes spending on research and development for a wide variety of energy technologies, including growing and utilizing poplars, willows, eucalypts, and other fast-growing trees for biofuels.

The ***Agriculture Improvement Act of 2018*** ([Public Law 115-334](#); 20 December 2018) (i.e., the 2018 Farm Bill) is also noteworthy with respect to the potential cultivation of poplars, willows, and eucalypts. The act reauthorized and modified many USDA programs, including those related to forestry, energy, and horticulture, as well as conservation, research, and extension activities. The 2018 Farm Bill was extended on 16 November 2023 through 30 September 2024, allowing programs to continue to the end of the 2024 fiscal year. Three such programs are pertinent to the production of fast-growing trees. First, the [Conservation Reserve Program \(CRP\)](#) pays a yearly rental payment in exchange for farmers removing environmentally sensitive land from agricultural production and planting species that will improve environmental quality. General CRP includes a Climate-Smart Practice Incentive to help increase carbon sequestration and reduce greenhouse gas emissions by helping producers and landowners establish

trees and perennial grasses, enhance wildlife habitat, and restore wetlands. Second, the [Conservation Reserve Enhancement Program \(CREP\)](#) is a voluntary land retirement program that helps agricultural producers protect environmentally sensitive land, decrease erosion, restore wildlife habitat, and safeguard ground and surface water. Objectives of the CREP include: 1) protecting the Nation's ability to produce food and fiber, 2) reducing soil erosion and sedimentation, 3) improving water quality, and 4) creating or enhancing habitat for wildlife. Third, the [Biomass Crop Assistance Program \(BCAP\)](#) is a program that helps farmers grow crops for bioenergy production. The USDA's Farm Service Agency (FSA) administers the BCAP. Objectives of the BCAP include: 1) promoting energy-efficient bioenergy crops, 2) developing new crops and cropping systems that preserve natural resources, and 3) connecting farmers with energy companies that use the feedstocks.

The ***Inflation Reduction Act (IRA) of 2022*** ([Public Law 117-169](#); 16 August 2022) also has implications for the cultivation of fast-growing trees in the US. The IRA is the largest investment in reducing carbon pollution in the history of the US. The IRA includes funding, incentives, and programs to help the country transition to a clean energy economy. Provisions of the IRA can help reduce climate damage and increase economic output in five ways, with the first three being directly relevant to growing poplar, willow, and eucalypt biomass feedstock production systems for biofuels, bioenergy, and bioproducts: 1) **mitigating greenhouse gases**, 2) **adapting to climate change**, 3) **reducing local pollution**, 4) supporting electric vehicle (EV) charging, and 5) supporting power infrastructure. The IRA has a section dedicated to the advancement of Sustainable Aviation Fuels (SAF), with those achieving a 50% or greater reduction in lifecycle greenhouse gas emissions under the most recent Carbon Offsetting and Reduction Scheme for International Aviation standards continuing to qualify for tax credits under US EPA and Internal Revenue Service (IRS) guidance. Similarly, these activities are a high priority for the US DOE that along with the USDA and other agencies recently released a SAF Grand Challenge, a government-wide approach to reduce the cost, enhance the sustainability, and expand the production of domestic SAF to meet 100% of the aviation fuel demand by 2050, which also advances the Biden Administration's goal to achieve carbon pollution-free electricity by 2035 ([Executive Order 14008](#), Tackling the Climate Crisis at Home and Abroad; 27 January 2021). Funding announcements from the US DOE in spring 2024 supported high-impact R&D on mobilizing low carbon intensity, purpose-grown energy crops necessary to support the production goals of the SAF Grand Challenge.

It is worth noting that some provisions of the IRA dated back to the Biden Administration's American Jobs Plan that were redirected to the ***Infrastructure Investment and Jobs Act of 2021*** (IIJA) (a.k.a., Bipartisan Infrastructure Law, BIL) ([Public Law 117-58](#); 15 November 2021). Key goals of the American Jobs Plan that carried forward to the enacted IIJA included: 1) **making infrastructure more resilient** through maximizing the resilience of land and water resources to protect communities and the environment, 2) **investing in R&D and future technologies** to establish the US as a leader in climate science, innovation, and R&D, 3) **reenergizing America's power infrastructure** to conserve public lands and waters, bolster community resilience, and advance environmental justice, and 4) **delivering the largest investment in tackling legacy pollution in American history** by cleaning up Superfund and brownfield sites, reclaiming abandoned mines, and capping orphaned oil and gas wells. For the latter, in 2023, the USDA Forest Service invested millions of dollars to remediate and restore mined lands, reducing the threat to human health and the environment [Provision 40804(b)(8) - Mitigate Environmental Hazards on Mined Lands]. These efforts involve nature-based solutions, such as utilizing poplars and willows for phytoremediation and associated phytotechnologies. Additional USDA programs supporting reforestation include: 1) USDA conservation programs such as the CRP (see above), [Environmental Quality Incentives Program \(EQIP\)](#), and [Agricultural Conservation Easement Program](#)

(ACEP), which can incentivize private landowners to plant trees and maintain forests, 2) USDA Forest Service, State and Private Forestry programs, including Urban and Community Forestry, Landscape Scale Restoration, and Forest Health Management, which can accelerate tree restoration on non-federal lands, and 3) USDA Forest Service National Forest programs such as Vegetation and Watershed Management and Collaborative Forest Landscape Restoration, which can help restore forest ecosystems while restocking federal forests affected by pests, diseases, or other disturbances.

¹ National Science Board, National Science Foundation. (2024). Research and Development: U.S. Trends and International Comparisons. Science and Engineering Indicators 2024. NSB-2024-6. Alexandria, VA. 51 p. <https://ncses.nsf.gov/pubs/nsb20246/> | ² Perlack, R.D., Wright, L.L., Turhollow, A.F., Graham, R.L., Stokes, B.J., & Erblich, D.C. (2005). Biomass as Feedstock for a Bioenergy and Bioproducts Industry: The Technical Feasibility of a Billion-Ton Annual Supply. DOE/GO-102995-112135, US Department of Energy; ORNL/TM-2005/66, Oak Ridge National Laboratory. Oak Ridge National Laboratory, Oak Ridge, TN. 59 p. <https://doi.org/10.2172/885984> | ³ US Department of Energy. (2011). U.S. Billion-Ton Update: Biomass Supply for a Bioenergy and Bioproducts Industry. R.D. Perlack and B.J. Stokes (Leads), ORNL/TM-2011/224. Oak Ridge National Laboratory, Oak Ridge, TN. 227 p. <https://doi.org/10.2172/1023318> | ⁴ US Department of Energy. (2016). 2016 Billion-Ton Report: Advancing Domestic Resources for a Thriving Bioeconomy, Volume 1: Economic Availability of Feedstocks. M.H. Langholtz, B.J. Stokes, and L.M. Eaton (Leads), ORNL/TM-2016/160. Oak Ridge National Laboratory, Oak Ridge, TN. 448 p. <https://doi.org/10.2172/1271651> | ⁵ US Department of Energy. (2024). 2023 Billion-Ton Report: An Assessment of U.S. Renewable Carbon Resources. M.H. Langholtz (Lead), ORNL/SPR-2024/3103. Oak Ridge National Laboratory, Oak Ridge, TN. 268 p. <https://doi.org/10.23720/BT2023/2316165>

II. TECHNICAL INFORMATION

The information presented in this section is provided as a summary of 114 peer-reviewed manuscripts highlighting selected activities related to each area of work conducted in the US and/or involving coauthorship from US-based researchers, and, therefore, is not meant to represent a full synthesis of all 941 peer-reviewed manuscripts listed below (Section III.4. General Information: Literature).

1. Taxonomy, Nomenclature, and Registration

a) *Populus*

There was limited information published from 2020-2023 on the Taxonomy, Nomenclature, and Registration of poplars in the US. Please see the records below for recent advancements in the cultivation and utilization of poplars in the US. Previously, Isebrands and Richardson (2014) provided a comprehensive summary of the Taxonomy, Nomenclature, and Registration of poplars.

Isebrands, J.G., & Richardson, J. (eds). (2014). Poplars and Willows: Trees for Society and the Environment. Food and Agriculture Organization of the United Nations (UN FAO) and CAB International (CABI); Rome, Italy. ISBN 978-1-78064-108-9 (CABI); ISBN 978-92-5-107185-4 (FAO). 650 p. <https://doi.org/10.1079/9781780641089.0000>

b) *Salix*

Similarly, as for poplars, Isebrands and Richardson (2014) provided a comprehensive summary of the Taxonomy, Nomenclature, and Registration of willows.

Isebrands, J.G., & Richardson, J. (eds). (2014). Poplars and Willows: Trees for Society and the Environment. Food and Agriculture Organization of the United Nations (UN FAO) and CAB International (CABI); Rome, Italy. ISBN 978-1-78064-108-9 (CABI); ISBN 978-92-5-107185-4 (FAO). 650 p. <https://doi.org/10.1079/9781780641089.0000>

Throughout the reporting period (2020-2023), there was an extensive amount of research focusing on the taxonomic classification and identification of different willow species and hybrids¹⁻⁷. From 2016-2020, with the IPC as the International Cultivar Registration Authority, a total of 854 cultivar epithets were included in the checklist for cultivars of *Salix*, providing a baseline for the formal registration of new cultivars¹. Since 2016, 26 new cultivar epithets have been registered and included in the International

Register of Cultivars of *Salix*, providing detailed information about each registered cultivar, including traits such as foliage color, stem color, and growth habit¹. Complementing this work, McGovern et al. (2021) discussed a proof-of-concept for an IPC *Salix* Cultivar Database that stores and manages information about *Salix* tree and shrub cultivars, including integrating information about epithets, families, and pedigrees⁷.

In addition to classification, extensive efforts have been devoted to identification of willows, focusing on various morphological characteristics. For example, ovule number was studied to differentiate among willow species and hybrids³⁻⁵. Here, Marchenko and Kuzovkina (2021a, 2021b, 2023) verified the taxonomic designation of *Salix vitellina* L. using ovule number, comparing values with related *Salix* species³⁻⁵. They analyzed several specimens of *S. vitellina* from Europe and North America and revealed that *S. vitellina* specimens have an ovule index of 6-10, with most valves having four and five ovules. With this ovule distribution being similar to that of *Salix alba* L. specimens, their results suggested that *S. vitellina* belongs to *S. alba*. Based on their results, the number of ovules present in the ovaries of the willow flower can be a useful tool for species identification, with their data indicating that these differences in ovule number can be used as diagnostic characters to distinguish among species³. Furthermore, to aid in species and hybrid verification, a method for calculating ovule number in the genus *Salix* was described⁴, as well as data showing morphological comparisons to support identification and classification of species hybrids that distinguish them from their parent species⁵. Lastly, Marchenko and Kuzovkina (2022) evaluated the recent replacements of *Salix fragilis* L. with *Salix euxina* I.V. Belyaeva and *Salix* × *rubens* Schrank with *Salix* × *fragilis* L., proposing to continue the use of *S. fragilis* as the name for a glabrous crack willow and *S.* × *rubens* for its hybrid with *S. alba*⁶.

In conclusion, these results provide valuable insights into the taxonomic classification, identification, and nomenclature of different species and hybrids within the genus *Salix*, as well as detailing the importance of databases for storing and managing cultivar information. Overall, these findings contribute to a better understanding of the taxonomy, classification, and archiving of willows.

¹ Kuzovkina, Y. A., & Vietto, L. (2021). International registration of cultivar names for *Salix* L. (willow) in 2016–20. *HortScience*, 56(1), 106–107. <https://doi.org/10.21273/HORTSCI15356-20> | ² Marchenko, A., & Kuzovkina, Y. (2023). Notes on the Taxonomy of *Salix vitellina* (Salicaceae). *Plants*, 12(14), 2610. <https://doi.org/10.3390/plants12142610> | ³ Marchenko, A. M., & Kuzovkina, Y. A. (2023). The ovule number variation provides new insights into taxa delimitation in willows (*Salix* subgen. *Salix*; salicaceae). *Plants*, 12(3), Article 3. <https://doi.org/10.3390/plants12030497> | ⁴ Marchenko, A. M., & Kuzovkina, Y. A. (2021a). Calculation of the ovule number in the genus *Salix*: A method for taxa differentiation. *Applications in Plant Sciences*, 9(11–12), e11450. <https://doi.org/10.1002/aps3.11450> | ⁵ Marchenko, A. M., & Kuzovkina, Y. A. (2021b). Identification of hybrid formulae of a few willows (*Salix*) using ovule numbers. *Silvae Genetica*, 70(1), 75–83. <https://doi.org/10.2478/sg-2021-0006> | ⁶ Marchenko, A. M., & Kuzovkina, Y. A. (2022). Notes on the nomenclature and taxonomy of *Salix fragilis* (Salicaceae). *TAXON*, 71(4), 721–732. <https://doi.org/10.1002/tax.12685> | ⁷ McGovern, P. N., Kuzovkina, Y. A., & Soolanayakanahally, R. Y. (2021). Short communication: IPC *Salix* cultivar database proof-of-concept. *Forests*, 12(5), 631. <https://doi.org/10.3390/f12050631>

c) *Eucalyptus*

There was limited information published from 2020-2023 on the Taxonomy, Nomenclature, and Registration of eucalypts in the US. Please see the records below in Section III.4. (General Information: Literature) for recent advancements in the cultivation and utilization of eucalypts in the US.

2. Domestication, Breeding, and Selection of Genetic Resources

Sanderson et al. (2021) provided a comprehensive summary of the evolutionary history and diversification patterns of *Populus* (poplars, cottonwoods, and aspens) and *Salix* (willows). Their findings highlighted the role of hybridization and incomplete lineage sorting in generating phylogenetic conflict and shaping the diversity of these genera. They used targeted sequence capture and whole genome

sequencing techniques to obtain genomic data from 165 samples representing various subgenera and sections within *Populus* and *Salix* for phylogenetic analysis using two approaches. First, they used the ASTRAL method to estimate gene trees for each gene and then inferred the best species tree by minimizing quartet distances among gene trees. Second, they concatenated all genes and used maximum likelihood analysis to estimate the most likely concatenated tree. In addition, they performed dating analyses to estimate the timing of diversification events. Their results revealed extensive phylogenetic conflict among genes, indicating the presence of hybridization and incomplete lineage sorting. Their results also highlighted the significant role of hybridization in the diversification and diversity of both genera. They discussed the challenges posed by hybridization in reconstructing phylogenetic histories of these genera. Overall, Sanderson et al. (2021) contributed to our knowledge of the factors influencing species diversity and phylogenetic relationships in willows, poplars, and their close relatives.

Sanderson, B. J., Gambhir, D., Feng, G., Hu, N., Cronk, Q. C., Percy, D. M., Freaner, F. M., Johnson, M. G., Smart, L. B., Keefover-Ring, K., Yin, T., Ma, T., DiFazio, S. P., Liu, J., & Olson, M. S. (2023). Phylogenomics reveals patterns of ancient hybridization and differential diversification that contribute to phylogenetic conflict in willows, poplars, and close relatives. *Systematic Biology*, 72(6), 1220–1232. <https://doi.org/10.1093/sysbio/syad042>

a) *Populus*

i. Sections

Isebrands and Richardson (2014) provided a comprehensive summary of the Sections of poplars.

Isebrands, J.G., & Richardson, J. (eds). (2014). *Poplars and Willows: Trees for Society and the Environment*. Food and Agriculture Organization of the United Nations (UN FAO) and CAB International (CABI); Rome, Italy. ISBN 978-1-78064-108-9 (CABI); ISBN 978-92-5-107185-4 (FAO). 650 p. <https://doi.org/10.1079/9781780641089.0000>

ii. Breeding and Testing

Two selected studies provided contrasting yet valuable information on the cultural history of the Lombardy poplar¹ and the efficiency of genotype testing in hybrid poplar breeding². Collectively, these findings can be useful for those interested in the cultural and historical aspects of poplar culture, as well as researchers, breeders, and horticulturists working with poplar trees. First, Beamish (2022) discussed how Lombardy poplar went from being greatly admired to reviled, facing hostility and negative labels such as odious and worthless. The shunning of Lombardy poplar was purported to be influenced by xenophobia, nativist movements, changes in horticultural fashion, and a growing sense of pride in native trees¹. Second, Nelson et al. (2021) provided a detailed account of various ways to increase efficiency of hybrid poplar tree improvement, focusing on the testing process. They reported that early selection, elimination of certain nursery-based progeny trial measurements, and providing clone performance estimates through family field trials could potentially increase efficiency in genotype testing². In addition, by examining age-age correlations, they found that selection for growth traits in hybrid poplars can be effective as early as one fourth of the rotation age. This is important given that previous studies have shown that age-age correlations can be predictive of performance at later ages, with the strength of such correlations dependent on factors such as site, clonal composition, region, and disease resistance².

¹ Beamish, A. (2022). A much-abused tree: The rise and fall of the Lombardy poplar. *Studies in the History of Gardens & Designed Landscapes*, 42(2), 120–137. <https://doi.org/10.1080/14601176.2022.2059287> | ² Nelson, N. D., Berguson, W. E., McMahon, B. G., Cai, M., & Buchman, D. J. (2021). Vectors of efficiency in hybrid poplar genotype testing. *Silvae Genetica*, 70(1), 39–56. <https://doi.org/10.2478/sg-2021-0004>

iii. Genomics

Throughout the reporting period (2020-2023), there was an extensive amount of research focusing on various aspects of poplar genomics, including gene dosage variation, lignin structure, wood anatomical and morphological traits, adventitious root development, and sex determination¹⁻⁸. Bastiaanse et al. (2021) investigated the effect of gene dosage variation on leaf morphology in *Populus*¹. They used a systems genetics approach and a large population of *Populus* lines and reported that gene dosage had significant impacts on leaf shape, size, stomatal patterning, and serration characteristics. In addition, they identified dosage-sensitive genomic regions that influenced specific morphological traits¹. Brunner et al. (2021) provided a collection of abstracts showcasing current research and advancements in plant tissue culture, genetic engineering, and gene editing². Bryant et al. (2023) focused on lignin structure in *Populus* and its genetic basis³. They conducted a genome-wide association study (GWAS) to identify candidate genes associated with lignin phenotypes and found several promising candidate genes involved in lignin biosynthesis³. Complementary work was also reported for transgenic poplar designed for biofuels⁴. Furthermore, Chhetri et al. (2020) conducted a GWAS using a large population of *Populus* genotypes and identified candidate genes associated with wood anatomical and morphological traits in *Populus trichocarpa* Torr. & Gray, thus providing insights into the genetic control of such traits⁵. Similarly, Nagle et al. (2023) conducted a GWAS to identify regulators of adventitious root development and found hundreds of candidate genes involved in various functions such as cell division and structure, hormone signaling, reactive oxygen species (ROS) signaling, and post-translational modifications (PTMs)⁶. Wang et al. (2020) examined the phylogenetic relationships and evolutionary diversification of the genus *Populus*, with results highlighting the complex patterns of gene flow and ancient polymorphisms within the genus. Their findings have important implications for understanding the evolutionary history and adaptive potential of poplars⁷. Lastly, Zhou et al. (2020) investigated the sex determination region (SDR) in *P. trichocarpa* and identified sex-associated markers and male-specific sequences within the SDR, confirming male heterogamety in the species⁸. Overall, these findings contributed to our understanding of the genetic basis of biologically- and economically-important traits in *Populus*, and the results have implications for genetic improvement and biofuel production in poplars. In particular, the studies highlighted the importance of considering multiple data types, such as gene dosage, gene expression, and co-expression networks, in understanding complex traits. Further research is needed to fully understand the genetic mechanisms underlying these traits and to optimize genetic modifications for desired outcomes in poplar breeding and biofuel production.

¹ Bastiaanse, H., Henry, I. M., Tsai, H., Lieberman, M., Canning, C., Comai, L., & Groover, A. (2021). A systems genetics approach to deciphering the effect of dosage variation on leaf morphology in *Populus*. *The Plant Cell*, 33(4), 940–960. <https://doi.org/10.1093/plcell/koaa016> | ² Brunner, A. M., Beers, E. P., Sheng, X., Petzold, H. E., & Teixeira, R. (2021). From *Populus* molecular networks to transgenic field phenotypes. *In Vitro Cellular & Developmental Biology - Animal*, 57(S1), 20–31. <https://doi.org/10.1007/s11626-021-00576-4> | ³ Bryant, N., Zhang, J., Feng, K., Shu, M., Ployet, R., Chen, J.-G., Muchero, W., Yoo, C. G., Tschaplinski, T. J., Pu, Y., & Ragauskas, A. J. (2023). Novel candidate genes for lignin structure identified through genome-wide association study of naturally varying *Populus trichocarpa*. *Frontiers in Plant Science*, 14, 1153113. <https://doi.org/10.3389/fpls.2023.1153113> | ⁴ Bryant, N. D., Pu, Y., Tschaplinski, T. J., Tuskan, G. A., Muchero, W., Kalluri, U. C., Yoo, C. G., & Ragauskas, A. J. (2020). Transgenic poplar designed for biofuels. *Trends in Plant Science*, 25(9), 881–896. <https://doi.org/10.1016/j.tplants.2020.03.008> | ⁵ Chhetri, H. B., Furches, A., Macaya-Sanz, D., Walker, A. R., Kainer, D., Jones, P., Harman-Ware, A. E., Tschaplinski, T. J., Jacobson, D., Tuskan, G. A., & DiFazio, S. P. (2020). Genome-wide association study of wood anatomical and morphological traits in populus trichocarpa. *Frontiers in Plant Science*, 11, 545748. <https://doi.org/10.3389/fpls.2020.545748> | ⁶ Nagle, M. F., Yuan, J., Kaur, D., Ma, C., Peremylova, E., Jiang, Y., Zuhl, B., Niño de Rivera, A., Muchero, W., Fuxin, L., & Strauss, S. H. (2023). GWAS identifies candidate genes controlling adventitious rooting in *Populus trichocarpa*. *Horticulture Research*, 10(8), uhad125. <https://doi.org/10.1093/hr/uhad125> | ⁷ Wang, M., Zhang, L., Zhang, Z., Li, M., Wang, D., Zhang, X., Xi, Z., Keefover-Ring, K., Smart, L. B., DiFazio, S. P., Olson, M. S., Yin, T., Liu, J., & Ma, T. (2020). Phylogenomics of the genus *Populus* reveals extensive interspecific gene flow and balancing selection. *New Phytologist*, 225(3), 1370–1382. <https://doi.org/10.1111/nph.16215> | ⁸ Zhou, R., Macaya-Sanz, D., Schmutz, J., Jenkins, J. W., Tuskan, G. A., & DiFazio, S. P. (2020). Sequencing and analysis of the sex determination region of *Populus trichocarpa*. *Genes*, 11(8), 843. <https://doi.org/10.3390/genes11080843>

b) *Salix*

i. Sections

Isebrands and Richardson (2014) provided a comprehensive summary of the Sections of willows.

Isebrands, J.G., & Richardson, J. (eds). (2014). *Poplars and Willows: Trees for Society and the Environment*. Food and Agriculture Organization of the United Nations (UN FAO) and CAB International (CABI); Rome, Italy. ISBN 978-1-78064-108-9 (CABI); ISBN 978-92-5-107185-4 (FAO). 650 p.
<https://doi.org/10.1079/9781780641089.0000>

ii. Breeding and Testing

Carlson and Smart (2022) presented findings from a detailed study on biomass yield and growth traits in shrub willow hybrids and concluded that triploids produced greater levels of heterosis for biomass yield compared to diploids¹. They focused on diploid, triploid, and tetraploid genotypes of willow and aimed to understand their performance, inheritance patterns, and heterosis effects on biomass yield. Their results showed that triploid genotypes exhibited significantly greater aboveground biomass (AGB) compared to diploid and tetraploid genotypes; however, biomass allocation ratios (i.e., ratios of leaf dry mass to stem dry mass, and root dry mass to stem dry mass) were significantly greater in tetraploid genotypes compared to triploids and diploids, and diploids had significantly lower biomass allocation ratios compared to both triploids and tetraploids¹. In addition to biomass yield, Carlson et al. (2022) studied the inheritance patterns of gene expression on triploid F₁ progeny and their parents². The results of their transcriptome analysis revealed that gene expression inheritance in the triploid F₁ progeny was mostly conserved and additive. However, a percentage of genes showed nonadditive inheritance patterns, ranging from 27% to 60% across the families. Transgressively expressed genes, which exhibited under- and over-dominant expression, accounted for a small percentage. The majority of the non-additive only genes showed dominance in the direction of the tetraploid parent². Overall, findings from these studies contributed to our understanding of heterosis and can inform the development of improved willow bioenergy crops.

¹ Carlson, C. H., & Smart, L. B. (2022). Heterosis for biomass-related traits in interspecific triploid hybrids of willow (*Salix* spp.). *BioEnergy Research*, 15(2), 1042–1056. <https://doi.org/10.1007/s12155-021-10305-0> | ² Carlson, C. H., Choi, Y., Chan, A. P., Town, C. D., & Smart, L. B. (2022). Nonadditive gene expression is correlated with nonadditive phenotypic expression in interspecific triploid hybrids of willow (*Salix* spp.). *G3 Genes/Genomes/Genetics*, 12(3), jkab436. <https://doi.org/10.1093/g3journal/jkab436>

iii. Genomics

Throughout the reporting period (2020-2023), there was an extensive amount of research focusing on the evolution, genetic basis, and pathways of sex determination in willows¹⁻⁶. In particular, researchers identified candidate genes, explored genetic changes during sex chromosome transitions, and highlighted the importance of comparative studies in understanding sex determination mechanisms. In addition, studies were conducted investigating the impact of specific genes on protein abundance and identifying variations in gene copy numbers and expression between genotypes. Overall, the collective findings contributed to our understanding of the genetic pathways and gene expression patterns associated with sex determination in plants. Hu et al. (2023) investigated the transition from an XY system to a ZW system in willows and reported that the origins of sex chromosomes during homologous transitions may be more flexible than previously considered¹. Feng et al. (2020) explored the pathways to sex determination in plants and the parallel evolution of molecular pathways underlying unisexual flowers. They reported that dioecy has independently evolved thousands of times in angiosperms, and

that hormone response pathways, particularly cytokinin and ethylene response pathways, play a role in controlling unisexuality². Hyden et al. (2021; 2023a; 2023b) investigated the role of specific genes in sex determination in willows³⁻⁵. They identified candidate genes involved in sex determination, such as AGO4, DRB1, GATA15, and ARR17. These genes showed differential expression between males and females and are expected to be present only in females. The expression of these genes results in differential abundance of proteins involved in floral development and sex determination processes³⁻⁵. Zhou et al. (2020) evaluated the gene content of palindromes in the female sex determination region of *Salix purpurea* L.⁶. Palindromes contain duplicated copies of genes involved in sex determination, such as the RR gene.

¹ Hu, N., Sanderson, B. J., Guo, M., Feng, G., Gambhir, D., Hale, H., Wang, D., Hyden, B., Liu, J., Smart, L. B., DiFazio, S. P., Ma, T., & Olson, M. S. (2023). Evolution of a ZW sex chromosome system in willows. *Nature Communications*, 14(1), 7144. <https://doi.org/10.1038/s41467-023-42880-5> | ² Feng, G., Sanderson, B. J., Keefover-Ring, K., Liu, J., Ma, T., Yin, T., Smart, L. B., DiFazio, S. P., & Olson, M. S. (2020). Pathways to sex determination in plants: How many roads lead to Rome? *Current Opinion in Plant Biology*, 54, 61–68. <https://doi.org/10.1016/j.pbi.2020.01.004> | ³ Hyden, B., Carper, D. L., Abraham, P. E., Yuan, G., Yao, T., Baumgart, L., Zhang, Y., Chen, C., O'Malley, R., Chen, J., Yang, X., Hettich, R. L., Tuskan, G. A., & Smart, L. B. (2023a). Functional analysis of *Salix purpurea* genes support roles for ARR17 and GATA15 as master regulators of sex determination. *Plant Direct*, 7(11), e546. <https://doi.org/10.1002/pld3.546> | ⁴ Hyden, B., Feng, K., Yates, T. B., Jawdy, S., Cereghino, C., Smart, L. B., & Muchero, W. (2023b). De Novo Assembly and annotation of 11 diverse shrub willow (*Salix*) genomes reveals novel gene organization in sex-linked regions. *International Journal of Molecular Sciences*, 24(3), 2904. <https://doi.org/10.3390/ijms24032904> | ⁵ Hyden, B., Carlson, C. H., Gouker, F. E., Schmutz, J., Barry, K., Lipzen, A., Sharma, A., Sandor, L., Tuskan, G. A., Feng, G., Olson, M. S., DiFazio, S. P., & Smart, L. B. (2021). Integrative genomics reveals paths to sex dimorphism in *Salix purpurea* L. *Horticulture Research*, 8, 170. <https://doi.org/10.1038/s41438-021-00606-y> | ⁶ Zhou, R., Macaya-Sanz, D., Carlson, C. H., Schmutz, J., Jenkins, J. W., Kudrna, D., Sharma, A., Sandor, L., Shu, S., Barry, K., Tuskan, G. A., Ma, T., Liu, J., Olson, M., Smart, L. B., & DiFazio, S. P. (2020). A willow sex chromosome reveals convergent evolution of complex palindromic repeats. *Genome Biology*, 21(1), 38. <https://doi.org/10.1186/s13059-020-1952-4>

c) *Eucalyptus*

i. Sections

Yost et al. (2021) aimed to determine the genetic variation and native origin of *Eucalyptus globulus* Labill. in California and to explore the factors influencing its naturalization in the region. They reported that *E. globulus* in California primarily originated from eastern Tasmania, with the genetic diversity of the Californian landrace being lower than that of the native Australian population. In addition, neither specific introduced genotype nor native origin influenced naturalization of *E. globulus* in California; instead, naturalization was influenced by local climate and disturbance factors. Overall, these findings contributed to our understanding of the naturalization patterns of *E. globulus* and highlight the importance of considering environmental factors along with genetics in invasion biology.

Yost, J. M., Wise, S. L., Love, N. L. R., Steane, D. A., Jones, R. C., Ritter, M. K., & Potts, B. M. (2021). Origins, diversity and naturalization of *Eucalyptus globulus* (Myrtaceae) in California. *Forests*, 12(8), 1129. <https://doi.org/10.3390/f12081129>

ii. Breeding and Testing

Castro et al. (2021; 2022) highlighted the results of using top grafting as a technique to induce early flowering in *Eucalyptus* trees¹⁻². They conducted grafting experiments at different time periods before natural flowering and evaluated the survival rate, canopy area growth, and flowering capacity of the top grafts. In addition, the influence of paclobutrazol (PBZ), a growth regulator, on these traits was also examined. Overall, they concluded: 1) the top grafting technique was found to be viable for inducing early flowering in *Eucalyptus* trees, 2) the number of flower buds and fruits produced was satisfactory in both grafting periods (albeit higher for the grafts performed three months before natural flowering), and 3) the application of PBZ increased flowering but did not significantly affect survival or canopy area growth. Their information provided valuable insights into the use of top grafting as a technique for inducing early flowering in *Eucalyptus* trees. As such, their findings can be applied in tree improvement programs to obtain superior clones through self-pollination cycles¹⁻². Furthermore, Ibarra et al. (2023)

provided results from field tests comparing the growth and wood properties of a hybrid species (GloNi) derived from *Eucalyptus nitens* H. Deane & Maiden (NIT) and *E. globulus* (GLO) in two breeding zones, Arauco and Valdivia³. Their results provided valuable information for selecting suitable species for different breeding zones and understanding the performance of hybrid species in terms of growth and wood properties³. Rockwood et al. (2022) discussed the potential uses of *Eucalyptus amplifolia* Naudin and *Corymbia torelliana* (F. Muell.) K.D. Hill in Florida, USA, focusing on market opportunities for these woods that are currently limited but have potential for future expansion⁴. In particular, they highlighted specialty pulps, medium density fiberboard (MDF), wood-cement boards, plywood, oriented strand board, biochar feedstocks, biofuel and bioenergy production, windbreaks, dendroremediation, and weed suppression as potential opportunities⁴.

¹ Castro, C. A. D. O., Santos, G. A. D., Takahashi, E. K., Nunes, A. C. P., Souza, G. A. D., Resende, M. D. V. D., & Zanuncio, J. C. (2022). Top grafting to accelerate self-pollination in *Eucalyptus* breeding. *Revista Árvore*, 46, e4630. <https://doi.org/10.1590/1806-908820220000030> | ² Castro, C. A. de O., dos Santos, G. A., Takahashi, E. K., Nunes, A. C. P., Souza, G. A., & de Resende, M. D. V. (2021). Accelerating *Eucalyptus* breeding strategies through top grafting applied to young seedlings. *Industrial Crops and Products*, 171, 113906. <https://doi.org/10.1016/j.indcrop.2021.113906> | ³ Ibarra, L., Hodge, G., & Acosta, J. J. (2023). Quantitative genetics of a hybrid population of *Eucalyptus nitens* × *Eucalyptus globulus*: Estimation of genetic parameters and implications for breeding strategies. *Forests*, 14(2), 381. <https://doi.org/10.3390/f14020381> | ⁴ Rockwood, D. L., Huber, D. A., Crawford, M. A., Rucks, P. C., Lamb, E., Fabbro, K. W., Minogue, P. J., Jump, R., & Hodge, G. R. (2022). *Eucalyptus amplifolia* and *Corymbia torelliana* in the southeastern USA: Genetic improvement and potential uses. *Forests*, 13(1), 75. <https://doi.org/10.3390/f13010075>

iii. Genomics

Throughout the reporting period (2020-2023), there was an extensive amount of research focusing on the genomics of *Eucalyptus*, including population structure, genomic predictions, genetic containment, structural variation, adaptive introgression, and genomic selection¹⁻⁸. Overall, this information contributed to our understanding of the genetic basis of important traits in *Eucalyptus* and has implications for breeding programs and the genetic improvement of *Eucalyptus* populations. Callister et al. (2022) explored the impact of accounting for population structure in genomic predictions of *E. globulus*¹. They used the single-step genomic BLUP method to compare breeding value predictions from models that either excluded base population effects or included fixed genetic groups or marker-derived proxies called metafounders. They found that the inclusion of metafounders improved accuracy and stability in breeding value predictions¹. Elorriaga et al. (2021) evaluated the use of CRISPR technology to disrupt the function of the LEAFY gene in *Eucalyptus* trees². Their objective was to achieve genetic containment in *Eucalyptus* trees while retaining desirable vegetative growth traits, and they found that CRISPR disruption of the LEAFY gene resulted in sterile indeterminate inflorescences without significant adverse effects on juvenile vegetative growth or leaf morphology². Similar results were reported by Nagle et al. (2023)⁷. Lötter et al. (2023) investigated the structural variation in *Eucalyptus* interspecific hybrids using long-read DNA sequencing data to assemble the haplogenomes of *Eucalyptus grandis* Hill & Maiden and *Eucalyptus urophylla* S.T. Blake hybrids, and they identified structural variants between the two haplogenomes, providing insights into genome structural rearrangement in these species³. Mostert-O'Neill et al. (2021) conducted a study on *E. grandis* to investigate the potential for adaptive genetic variation through hybridization with other species⁴. They reported that adaptive introgression may play a significant role in the ability of *E. grandis* to adapt to rapid climate change, identifying environmental factors that contributed to population differentiation patterns in *E. grandis*⁴. Mostert-O'Neill et al. (2022) studied the genomic consequences of artificial selection in *E. grandis* breeding and wild populations. Their aim was to understand the genetic differentiation between breeding and wild populations and identify the genomic changes that have occurred during domestication. They discovered that the breeding and wild populations were genetically distinct, indicating that artificial selection has led to genomic changes in *E. grandis*⁵. In addition to evaluating the efficiency of genomic selection compared to traditional breeding cycles, Mphahlele et al. (2020) conducted a study on genetic

parameters, genomic selection accuracy, and genetic gains in *E. grandis*⁶. They reported that the use of a genomic relationship matrix improved breeding value prediction accuracy and genetic gains in *E. grandis*⁶. Rocha et al. (2022) conducted a GWAS to identify quantitative trait loci (QTLs) related to growth and wood quality traits in *E. grandis*, and they identified significant associations for both growth and wood quality traits and highlighted the importance of considering population structure and using multi-trait models in GWAS⁸.

¹ Callister, A. N., Bermann, M., Elms, S., Bradshaw, B. P., Lourenco, D., & Brawner, J. T. (2022). Accounting for population structure in genomic predictions of *Eucalyptus globulus*. *G3 Genes/Genomes/Genetics*, 12(9), jkac180. <https://doi.org/10.1093/g3journal/jkac180> | ² Elorriaga, E., Klocko, A. L., Ma, C., Du Plessis, M., An, X., Myburg, A. A., & Strauss, S. H. (2021). Genetic containment in vegetatively propagated forest trees: CRISPR disruption of *LEAFY* function in *Eucalyptus* gives sterile indeterminate inflorescences and normal juvenile development. *Plant Biotechnology Journal*, 19(9), 1743–1755. <https://doi.org/10.1111/pbi.13588> | ³ Lötter, A., Duong, T. A., Candotti, J., Mizrachi, E., Wegrzyn, J. L., & Myburg, A. A. (2023). Haplogenome assembly reveals structural variation in *Eucalyptus* interspecific hybrids. *GigaScience*, 12, giad064. <https://doi.org/10.1093/gigascience/giad064> | ⁴ Mostert-O'Neill, M. M., Reynolds, S. M., Acosta, J. J., Lee, D. J., Borevitz, J. O., & Myburg, A. A. (2021). Genomic evidence of introgression and adaptation in a model subtropical tree species, *Eucalyptus grandis*. *Molecular Ecology*, 30(3), 625–638. <https://doi.org/10.1111/mec.15615> | ⁵ Mostert-O'Neill, M. M., Tate, H., Reynolds, S. M., Mphahlele, M. M., Van Den Berg, G., Verry, S. D., Acosta, J. J., Borevitz, J. O., & Myburg, A. A. (2022). Genomic consequences of artificial selection during early domestication of a wood fibre crop. *New Phytologist*, 235(5), 1944–1956. <https://doi.org/10.1111/nph.18297> | ⁶ Mphahlele, M. M., Isik, F., Mostert-O'Neill, M. M., Reynolds, S. M., Hodge, G. R., & Myburg, A. A. (2020). Expected benefits of genomic selection for growth and wood quality traits in *Eucalyptus grandis*. *Tree Genetics & Genomes*, 16(4), 49. <https://doi.org/10.1007/s11295-020-01443-1> | ⁷ Nagle, M. F., Nahata, S. S., Zahl, B., de Rivera, A. N., Tacker, X. V., Elorriaga, E., Ma, C., Goraloglia, G. S., Klocko, A. L., Gordon, M., Joshi, S., & Strauss, S. H. (2023). Knockout of floral and meiosis genes using CRISPR/Cas9 produces male-sterility in *Eucalyptus* without impacts on vegetative growth. *Plant Direct*, 7(7), e507. <https://doi.org/10.1002/pld3.507> | ⁸ Rocha, L. F., Benatti, T. R., de Siqueira, L., de Souza, I. C. G., Bianchin, I., de Souza, A. J., Fernandes, A. C. M., Oda, S., Stape, J. L., Yassue, R. M., Carvalho, H. F., Müller, N. A., Fladung, M., Acosta, J. J., Fritsche-Neto, R., & Tamarussi, E. V. (2022). Quantitative trait loci related to growth and wood quality traits in *Eucalyptus grandis* W. Hill identified through single- and multi-trait genome-wide association studies. *Tree Genetics & Genomes*, 18(6), 38. <https://doi.org/10.1007/s11295-022-01570-x>

3. Plant Health, Resilience to Threats and Climate Change

a) Biotic Factors

Throughout the reporting period (2020–2023), there was an extensive amount of research focusing on various aspects of the interactions of poplars and willows with endophytes, pathogens, and environmental stressors^{1–7}. Banan et al. (2024) contributed to the understanding of how endophytes can influence water use efficiency (WUE) in *P. trichocarpa* under water-deficit conditions¹. They reported that endophyte inoculation can lead decreased stomatal conductance and increased intrinsic WUE (iWUE), but the impact on aboveground productivity may be limited. They highlighted the potential of endophytes as a tool for improving plant resilience and resource use efficiency in bioenergy feedstock systems¹. Crowell et al. (2020a; 2020b; 2022) studied the susceptibility of different *Salix* species to two species of *Melampsora* rust, *Melampsora americana* and *Melampsora paradoxa*, and they identified *M. americana* as the most prevalent rust species in the northeastern United States^{2–4}. They reported the presence of both vertical and horizontal resistance among willow genotypes, and they suggested that stomatal and trichome densities may contribute to rust resistance in *Salix*. Their findings have important implications for rust management in shrub willow cultivation and can guide future breeding efforts to develop resistant cultivars^{2–4}. Doty et al. (2022) explored the potential of endophytes from wild *Populus* trees to act as biocontrol agents against plant pathogens⁵. They investigated the inhibitory mechanisms employed by these endophytes and their effectiveness against various plant pathogens. Their genomic analysis revealed the presence of gene clusters associated with antimicrobial activities in different endophyte strains, suggesting that the wild poplar tree microbiome could be a valuable source of beneficial endophyte strains for biocontrol applications against various pathogens⁵. Lenz et al. (2021) used metabolomics techniques to identify and quantify metabolites involved in signaling and cell wall remodeling in resistant and susceptible *P. trichocarpa* genotypes after inoculation with the fungal pathogen *Sphaerulina musiva*, the causal agent of Septoria canker disease⁶. They reported that resistant genotypes employed systemic acquired resistance, cell wall apposition, and lignin deposition as modes of resistance to *S. musiva*. In addition, susceptible genotypes lacked a robust defense response and

showed depletion of metabolites involved in defense mechanisms. Their results provided useful information about the metabolic pathways and mechanisms associated with resistance and can inform breeding efforts to develop resistant poplar varieties⁶. Søndreli et al. (2020) identified *S. musiva* as the causal agent of an outbreak of Septoria canker on *P. trichocarpa* trees in Eastern Oregon, USA⁷. Their results demonstrated the varying susceptibility of different *P. trichocarpa* genotypes to *S. musiva*, as well as contributing to the importance of monitoring and managing this pathogen to protect tree health⁷.

¹ Banan, D., Sher, A., Doty, S. L., & Kim, S.-H. (2024). Endophyte mediated *Populus trichocarpa* water use efficiency is dependent on time of day and plant water status. *Phytobiomes Journal*, PBIOMES-11-22-0077-R. <https://doi.org/10.1094/PBIOMES-11-22-0077-R> | ² Crowell, C. R., Bekauri, M. M., Cala, A. R., McMullen, P., Smart, L. B., & Smart, C. D. (2020a). Differential susceptibility of diverse *Salix* spp. to *Melampsora americana* and *Melampsora paradoxa*. *Plant Disease*, 104(11), 2949–2957. <https://doi.org/10.1094/PDIS-04-20-0718-RE> | ³ Crowell, C. R., Wilkerson, D. G., Beckauri, M., Cala, A. R., McMullen, P. W., Mondo, S., Andreopoulos, W., Lipzen, A., Lail, K., Yan, M., Ng, V., Grigoriev, I. V., Smart, L. B., & Smart, C. D. (2022). The *Melampsora americana* population on *Salix purpurea* in the Great Lakes region is highly diverse with a contributory influence of clonality. *Phytopathology*, 112(4), 907–916. <https://doi.org/10.1094/PHYTO-05-21-0201-R> | ⁴ Crowell, C. R., Wilkerson, D. G., McMullen, P., Cala, A. R., Smart, L. B., & Smart, C. D. (2020b). Population biology of *Melampsora americana* and eQTL mapping for rust resistance in *Salix purpurea* shrub willow. *Plant Health* 2020. APS Annual Meeting. | ⁵ Doty, S. L., Joubert, P. M., Firrincieli, A., Sher, A. W., Tournay, R., Kill, C., Parikh, S. S., & Okubara, P. (2022). Potential biocontrol activities of *Populus* endophytes against several plant pathogens using different inhibitory mechanisms. *Pathogens*, 12(1), 13. <https://doi.org/10.3390/pathogens12010013> | ⁶ Lenz, R. R., Louie, K. B., Søndreli, K. L., Galanie, S. S., Chen, J.-G., Muchero, W., Bowen, B. P., Northen, T. R., & LeBoldus, J. M. (2021). Metabolomic patterns of Septoria canker resistant and susceptible *Populus trichocarpa* genotypes 24 hours postinoculation. *Phytopathology*, 111(11), 2052–2066. <https://doi.org/10.1094/PHYTO-02-21-0053-R> | ⁷ Søndreli, K. L., Keriö, S., Frost, K., Muchero, W., Chen, J.-G., Haiby, K., Gantz, C., Tuskan, G., & LeBoldus, J. M. (2020). Outbreak of Septoria canker caused by *Sphaerulina musiva* on *Populus trichocarpa* in eastern Oregon. *Plant Disease*, 104(12), 3266–3266. <https://doi.org/10.1094/PDIS-03-20-0494-PDN>

b) Abiotic Factors

There were several studies highlighting the importance of considering temperature, water stress, and genetic traits in forest management and bioengineering projects¹⁻⁴. The results of these studies contributed to a better understanding of the physiological responses of different tree species to environmental stressors. Binkley et al. (2020) described the Tolerance of *Eucalyptus* Clones to Hydric, Thermal and Biotic Stresses (TECHS) project that focused on clonal plantations of *Eucalyptus* and examined the influence of temperature and water stress on tree growth across a 3,500 km gradient from the Amazon to Uruguay¹. They reported that the mean annual increment (MAI) of stemwood production exhibited a humped pattern in relation to temperature, with the highest MAI observed at mean annual temperatures of 20 °C, and that whole-rotation MAI declined with increasing temperature and decreasing rainfall¹. Chen et al. (2020) reported that drought resulted in a significant loss of stem xylem hydraulic conductance, leading to hydraulic failure and potential mortality in *E. urophylla* under drought stress². In addition, decreased whole-plant hydraulic conductivity, sap flow density, and conversion of starch to soluble sugar were observed under drought conditions, thus highlighting the importance of considering both water and carbon dynamics in understanding drought-induced tree mortality². Himes et al. (2021) identified hybrid poplar leaf traits that can serve as indicators of drought resistance³. They reported that carbon isotope discrimination ($\Delta^{13}\text{C}$) was not an effective indicator of drought resistance in hybrid poplar varieties, although abaxial stomatal density (ABAX) showed a significant correlation with varietal performance under drought stress. Overall, the best indicator of drought stress was a composite of multiple traits³. Keita et al. (2021) investigated the growth response of three willow species (*Salix discolor* Muhl., *Salix eriocephala* Michx., and *Salix interior* Rowlee) to different soil moisture treatments and its implications for riverbank soil bioengineering⁴. Their results highlighted the importance of considering species-specific responses to water stress when designing riverbank soil bioengineering projects. By selecting the appropriate willow species based on their tolerance to water stress, the success of these projects can be greatly improved⁴.

¹ Binkley, D., Campoe, O. C., Alvares, C. A., Carneiro, R. L., & Stape, J. L. (2020). Variation in whole-rotation yield among *Eucalyptus* genotypes in response to water and heat stresses: The TECHS project. *Forest Ecology and Management*, 462, 117953. <https://doi.org/10.1016/j.foreco.2020.117953> | ² Chen, X., Zhao, P., Ouyang, L., Zhu, L., Ni, G., & Schäfer, K. V. R. (2020). Whole-plant water hydraulic integrity to predict drought-induced *Eucalyptus urophylla* mortality under drought stress. *Forest Ecology and Management*, 468, 118179. <https://doi.org/10.1016/j.foreco.2020.118179> | ³ Himes, A., Emerson, P., McClung, R., Renninger, H., Rosenstiel, T., & Stanton, B. (2021). Leaf traits indicative of drought resistance in hybrid poplar. *Agricultural Water Management*, 246, 106676. <https://doi.org/10.1016/j.agwat.2020.106676> | ⁴ Keita, N., Bourgeois, B., Evette, A., Tisserant, M., González, E., Breton, V., Goulet, C., & Poulin, M. (2021). Growth

c) Resilience to Threats and Climate Change

Many studies contributed to our understanding of plant adaptation, climate change mitigation, and sustainable land use practices. For example, several studies highlighted the importance of understanding the factors influencing leaf trait variation, the scale and sustainability of bioenergy expansion, the effects of eCO₂ on fine root dynamics, and the growth and response of *Eucalyptus* plantations to different treatments and environmental conditions¹⁻⁴. Asao et al. (2020) tested the variation in leaf functional traits of *Eucalyptus camaldulensis* Dehnh. genotypes from different climates and reported that genotypic differences contributed significantly to the variation in aboveground biomass, indicating that some genotypes are better adapted to their specific environment in terms of growth¹. There was no significant relationship between the climate of genotype provenance and leaf or performance traits, indicating that genotypes of *E. camaldulensis* are not specifically adapted to their climate of origin¹. Calvin et al. (2021) discussed the importance of bioenergy for global climate change mitigation². They described bioenergy with carbon capture and storage (BECCS), as well as integrated assessment models (IAMs) that link biophysical models of biosphere and atmosphere processes with socio-economic models to explore future scenarios of global energy, land, economy, and climate. Their key take home messages were that these IAMs provided valuable insights into the potential impacts of bioenergy deployment, but caution should be applied in interpreting their results due to uncertainties and limitations. In addition, alternative assessment methods can provide complementary insights and help inform sustainable bioenergy policies and practices². Piñeiro et al (2020) studied the effects of elevated carbon dioxide (eCO₂) on fine root dynamics in a *Eucalyptus* woodland ecosystem and reported that the response of fine roots to eCO₂ was influenced by soil water availability and depth³. Under well-watered conditions, eCO₂ had limited effects on fine root biomass and production, while eCO₂ stimulated fine root growth and production under water-limited conditions³. Ryan et al. (2020) tested the growth and response of *Eucalyptus* plantations to different treatments and environmental conditions in southeastern Brazil⁴. They studied the effects of fertilization and water input on plantation growth and analyzed the relationships between growth and prior weather conditions, and the findings of this study were compared to the results of the TECHS Project described above⁴.

¹ Asao, S., Hayes, L., Aspinwall, M. J., Rymer, P. D., Blackman, C., Bryant, C. J., Cullerne, D., Egerton, J. J. G., Fan, Y., Innes, P., Millar, A. H., Tucker, J., Shah, S., Wright, I. J., Yvon-Durocher, G., Tissue, D., & Atkin, O. K. (2020). Leaf trait variation is similar among genotypes of *Eucalyptus camaldulensis* from differing climates and arises in plastic responses to the seasons rather than water availability. *New Phytologist*, 227(3), 780–793. <https://doi.org/10.1111/nph.16579> | ² Calvin, K., Cowie, A., Berndes, G., Arneeth, A., Cherubini, F., Portugal-Pereira, J., Grassi, G., House, J., Johnson, F. X., Popp, A., Rounsevell, M., Slade, R., & Smith, P. (2021). Bioenergy for climate change mitigation: Scale and sustainability. *GCB Bioenergy*, 13(9), 1346–1371. <https://doi.org/10.1111/gcbb.12863> | ³ Piñeiro, J., Ochoa-Hueso, R., Drake, J. E., Tjoelker, M. G., & Power, S. A. (2020). Water availability drives fine root dynamics in a *Eucalyptus* woodland under elevated atmospheric CO₂ concentration. *Functional Ecology*, 34(11), 2389–2402. <https://doi.org/10.1111/1365-2435.13660> | ⁴ Ryan, M. G., Stape, J. L., Binkley, D., & Alvares, C. A. (2020). Cross-site patterns in the response of *Eucalyptus* plantations to irrigation, climate and intra-annual weather variation. *Forest Ecology and Management*, 475, 118444. <https://doi.org/10.1016/j.foreco.2020.118444>

4. Production Systems for the Bioeconomy

a) Nursery Practices and Propagation Techniques

Two studies provided insights into the growth and development of *Eucalyptus* seedlings¹⁻². Acevedo et al. (2021) highlighted the importance of optimal N concentration for promoting *E. globulus* seedling growth, as well as the use of a SPAD meter for monitoring N status¹. They showed that 300 mg N L⁻¹ was optimum, with seedlings exhibiting maximum growth, enhanced root growth, and higher tissue nutrient

concentrations at this rate. In addition, they found the use of a SPAD meter, which measures chlorophyll content, could be an effective tool for monitoring N status in forest nurseries¹. Chu et al. (2023) emphasized the potential benefits of using bagasse and zeolite in sewage sludge compost to reduce the concentration and bioavailability of potentially toxic elements (PTEs) in soil². The addition of composts to the soil promoted growth of *Eucalyptus urophyllus* S.T. Blake seedlings, but the addition of sewage sludge compost increased concentrations of PTEs in the seedlings. Therefore, combined addition of bagasse and zeolite were shown to be an effective strategy for reducing the environmental risks associated with sewage sludge compost application². Overall, these findings contributed to our understanding of seedling growth and provided practical recommendations for nutrient management in forest nurseries and the safe use of compost in soil.

¹ Acevedo, M., Rubilar, R., Dumroese, R. K., Ovalle, J. F., Sandoval, S., & Chassin-Trubert, R. (2021). Nitrogen loading of *Eucalyptus globulus* seedlings: Nutritional dynamics and influence on morphology and root growth potential. *New Forests*, 52(1), 31–46. <https://doi.org/10.1007/s11056-020-09778-2> | ² Chu, S., Xian, L., Zhao, N., Lai, C., Yang, W., Wang, J., Long, M., Liao, D., Ouyang, J., Wang, Z., Jacobs, D. F., & Zeng, S. (2023). Combined addition of bagasse and zeolite stabilizes potentially toxic elements in sewage sludge compost and improves *Eucalyptus urophylla* seedling growth. *Forest Ecology and Management*, 539, 121003. <https://doi.org/10.1016/j.foreco.2023.121003>

b) Planted Forests

i. *Populus*

Throughout the reporting period (2020-2023), there was an extensive amount of research focusing on the cultivation, productivity, and profitability of hybrid poplar trees for bioenergy, bioproducts, and biofuels¹⁻⁸. Studies focused on factors that influence growth and performance of poplar trees, including pests and diseases, leaf area index (LAI), foliar nitrogen (N), stand density, fertilization, clone selection, and site suitability. Overall, the findings can be used to inform decision-making processes related to the development of poplar-based environmental systems and the sustainable production of feedstocks for the bioeconomy. Ghezehei et al. (2021) tested the early growth and productivity of poplar trees in North Carolina, USA¹. Their study focused on the influence of pests and diseases, crown-related traits such as leaf area index (LAI), foliar nitrogen (N), and stand density, and silvicultural aspects such as fertilization, clone selection, and site suitability. Also, they highlighted the importance of considering real-world conditions when estimating the productivity of poplar plantations based on small-scale trials¹. Ghezehei et al. (2020) examined the survival, growth, and biomass of 89 poplar clones in the coastal southeastern USA at four- and eight-year rotations². They recommended selecting site-suitable clones for productivity-focused stands based on productivity first, then narrowing down the selection based on survival and rotation length. Changes were observed in the most site-suitable clones between longer and shorter rotations². Pilipović et al. (2022) quantified water use efficiency (WUE) of hybrid poplar clones at three different sites in the Midwestern United States, with results showing significant variation in WUE and biomass production among different clones and sites³. As such, they highlighted the importance of considering site-specific factors when selecting poplar clones for biomass production. For example, there is a need to match poplar genotypes to site conditions for successful bioenergy deployment, particularly in water-limited environments³. Renninger et al. (2023) defined water use strategies of different genotypes of *Populus deltoides* and its hybrids grown for biomass production and ecosystem services in the Lower Mississippi Alluvial Valley, USA⁴. In particular, *P. deltoides* × *P. maximowiczii* 'DM' hybrids had lower WUE and higher leaf area loss compared to other genotypes, while *P. deltoides* × *P. deltoides* 'DD' genotypes had higher WUE and larger leaf areas. Their findings have implications for selecting genotypes with desirable water use strategies for sustainable tree growth and productivity, with 'DD' genotypes exhibiting more efficient water use strategies, while 'DM' hybrids being more susceptible to water stress⁴. Similarly, Renninger et al. (2022) compared the physiological functioning and productivity of *P.*

deltoides and three hybrid clones in the southeastern US⁵. Genotypes were tested under different site conditions and endophyte inoculations, and their results provided valuable insights into the performance and potential of these genotypes for biomass production and environmental remediation purposes⁵. Renninger et al. (2021) provided similar results, with genotype × environment interactions governing biomass, growth, and physiology traits⁶. Furthermore, Rogers et al. (2023) conducted a literature review detailing the methodologies used to measure poplar water use, as well as intrinsic and extrinsic factors influencing poplar water use⁷. They concluded that performance varied among different hybrid types and tree age classes, which can affect water use. Similarly, planting density also had a significant impact on hybrid poplar water use, with different densities leading to variations in growth, productivity, and morphology. Their findings are useful to guide decision-making in the design and management of poplar-based production systems⁷. Stanton et al. (2021) discussed the practice and economics of hybrid poplar biomass production for biofuels and bioproducts in the Pacific Northwest, USA⁸. In addition to the chemical composition of hybrid poplar biomass and production costs over a 20-year rotation, they evaluated management practices, yields, and harvesting methods. Their information can be used to inform decision-making processes related to the development of a biorefining industry in the region and the sustainable production of biofuels and bioproducts⁸.

¹ Ghezehei, S. B., Ewald, A. L., Hazel, D. W., Zalesny, R. S., & Nichols, E. G. (2021). Productivity and profitability of poplars on fertile and marginal sandy soils under different density and fertilization treatments. *Forests*, 12(7), 869. <https://doi.org/10.3390/f12070869> | ² Ghezehei, S. B., Wright, J., Zalesny, R. S., Nichols, E. G., & Hazel, D. W. (2020). Matching site-suitable poplars to rotation length for optimized productivity. *Forest Ecology and Management*, 457, 117670. <https://doi.org/10.1016/j.foreco.2019.117670> | ³ Pilipović, A., Headlee, W. L., Zalesny Jr., R. S., Pekeč, S., & Bauer, E. O. (2022). Water use efficiency of poplars grown for biomass production in the Midwestern United States. *GCB Bioenergy*, 14(3), 287–306. <https://doi.org/10.1111/gcbb.12887> | ⁴ Renninger, H. J., Pitts, J. J., & Rousseau, R. J. (2023). Comparisons of biomass, water use efficiency and water use strategies across five genomic groups of *Populus* and its hybrids. *GCB Bioenergy*, 15(1), 99–112. <https://doi.org/10.1111/gcbb.13014> | ⁵ Renninger, H. J., Stewart, L. F., Freeman, J. L., & Rousseau, R. J. (2022). Physiological functioning and productivity in eastern cottonwood and hybrid poplars on contrasting sites in the Southeastern US. *BioEnergy Research*, 15(2), 1057–1070. <https://doi.org/10.1007/s12155-021-10377-y> | ⁶ Renninger, H. J., Stewart, L. F., & Rousseau, R. J. (2021). Water use, efficiency, and stomatal sensitivity in eastern cottonwood and hybrid poplar varieties on contrasting sites in the southeastern United States. *Frontiers in Forests and Global Change*, 4, 704799. <https://doi.org/10.3389/ffgc.2021.704799> | ⁷ Rogers, E. R., Zalesny, R. S., Lin, C.-H., & Vinhal, R. A. (2023). Intrinsic and extrinsic factors influencing *Populus* water use: A literature review. *Journal of Environmental Management*, 348, 119180. <https://doi.org/10.1016/j.jenvman.2023.119180> | ⁸ Stanton, B. J., Bourque, A., Coleman, M., Eisenbies, M., Emerson, R. M., Espinoza, J., Gantz, C., Himes, A., Rodstrom, A., Shuren, R., Stonex, R., Volk, T., & Zepa, J. (2021). The practice and economics of hybrid poplar biomass production for biofuels and bioproducts in the Pacific Northwest. *BioEnergy Research*, 14(2), 543–560. <https://doi.org/10.1007/s12155-020-10164-1>

ii. *Salix*

Three studies provided insights into the sustainable production of willow^{1–3}. Information obtained from these studies can be used to guide the selection, management, and optimization of willow genotypes and plantations for maximizing productivity and ecosystem services. Gouker et al. (2021) highlighted the importance of selecting appropriate genotypes and planting designs for maximizing biomass yield¹. Conducted in Geneva, New York, USA, results included differences in budbreak rates, biomass traits, wood composition, and calorific value among genotypes and between two planting designs. Their information can be valuable for selecting and managing willow genotypes for biomass production¹. Montes et al. (2021) estimated the potential and water-limited yield of six elite willow cultivars grown in a commercial-scale plantation in Centre County, Pennsylvania, USA². They found that planting density had a significant impact on biomass yield, and the information obtained can be valuable for optimizing the cultivation of willow for biomass production². Muklada et al. (2022) discussed favorable morphological characteristics and nutritional value of willow, making it a promising crop for forage production and bioenergy purposes³. Their results highlighted the potential of willow as a sustainable and productive crop for various agricultural applications³.

¹ Gouker, F. E., Fabio, E. S., Serapiglia, M. J., & Smart, L. B. (2021). Yield and biomass quality of shrub willow hybrids in differing rotation lengths and spacing designs. *Biomass and Bioenergy*, 146, 105977. <https://doi.org/10.1016/j.biombioe.2021.105977> | ² Montes, F., Fabio, E. S., Smart, L. B., Richard, T. L., Añó, R. M., & Kemanian, A. R. (2021). A semi-commercial case study of willow biomass production in the northeastern United States. *Agronomy Journal*, 113(2), 1287–1302. <https://doi.org/10.1002/agj2.20603> | ³ Muklada, H., Fabio, E. S., & Smart, L. B. (2022). Growth, nitrogen uptake, and nutritional value of a diverse panel of shrub willow (*Salix* spp.) genotypes in response to nitrogen fertilization. *Agronomy*, 12(11), 2678. <https://doi.org/10.3390/agronomy12112678>

iii. *Eucalyptus*

Research on *Eucalyptus* plantations has provided valuable insights into the factors influencing growth, including climate, genotype, and spacing. For example, several studies highlighted the importance of proper management practices for maximizing productivity, such as selecting appropriate genotypes, optimizing spacing, and managing competition¹⁻⁶. Overall, these results contributed to the understanding of *Eucalyptus* growth and adaptation and can inform management practices and genotype selection for specific environments. In particular, Campoe and Binkley (2020) described the TECHS Project, a comprehensive research initiative that aims to understand the factors influencing the growth of *Eucalyptus* plantations¹. Including intensive experimentation across diverse geography, their findings have provided valuable insights contributing to the development of improved management practices for *Eucalyptus* plantations¹. Similarly, Campoe et al. (2020) studied the effects of climate and genotype on carbon fluxes and partitioning in *Eucalyptus* plantations². They asserted that understanding how carbon budgets differ across sites and why clones can differ in wood production was crucial for improving productivity in stressful environments. In addition, they highlighted the need for further research on genotype × environment interactions to develop more process-based production methods and recommend genotypes with higher tolerance to different climatic regions². Conti et al. (2020) investigated the climatic characterization and physiological parameters of *Eucalyptus* clones at three sites within the TECHS research platform³. Leaf turgor loss point and leaf water potential measurements showed variations among seasons and clones at each site, contributing to the conclusions that there were differences in drought tolerance among the *Eucalyptus* clones, but these differences were not solely determined by the leaf turgor loss point. Overall, the study provided valuable insights into the climatic characterization and physiological parameters of *Eucalyptus* clones, which can contribute to the understanding of their adaptation to different environmental conditions³. Rubilar et al. (2020) studied the effects of climate and water availability on the growth of different *Eucalyptus* genotypes and reported irrigation increased cumulative growth and leaf area index for most taxa⁴. They emphasized the need for strategic allocation of genotypes to contrasting drought risk environments and the importance of understanding genotype × environment interactions for predicting and maximizing plantation productivity in the face of climate change⁴. Hall et al. (2019) used various equations and models to analyze the growth and productivity data of *Eucalyptus benthamii* in the southeastern US and Brazil⁵. They developed the growth and yield models based on factors such as age, dominant height, and site index. Their conclusions provided valuable insights into the growth, productivity, and management of *Eucalyptus* plantations, including the impact on water resources, the potential for bioenergy production, and the development of modeling techniques to improve predictions⁵. Stape et al. (2022) reported that spacing and geometric layout had significant effects on the growth and productivity of *Eucalyptus grandis* × *Eucalyptus urophylla* clones in Brazil⁶. Square planting designs with high tree densities were found to be most productive, while wide spacing between rows may have cost-saving benefits or allow for intercropping. These findings provided valuable insights for forest managers and researchers in optimizing productivity of *Eucalyptus* plantations⁶.

¹ Campoe, O. C., & Binkley, D. (2020). Environmental and genetic influences on growth in *Eucalyptus* plantations: The TECHS special issue. *Forest Ecology and Management*, 476, 118464. <https://doi.org/10.1016/j.foreco.2020.118464> | ² Campoe, O. C., Alvares, C. A., Carneiro, R. L., Binkley, D., Ryan, M. G., Hubbard, R. M., Stahl, J., Moreira, G., Moraes, L. F., & Stape, J. L. (2020). Climate and genotype influences on carbon fluxes and partitioning in *Eucalyptus* plantations. *Forest Ecology and Management*, 475, 118445. <https://doi.org/10.1016/j.foreco.2020.118445> | ³ Conti Junior, J. L. F., De Araujo, M. J., De Paula, R. C., Queiroz, T. B., Hakamada, R. E., & Hubbard, R. M. (2020). Quantifying turgor loss point and leaf water potential across contrasting *Eucalyptus* clones and sites within the TECHS research platform. *Forest Ecology and Management*, 475, 118454. <https://doi.org/10.1016/j.foreco.2020.118454> | ⁴ Rubilar, R., Hubbard, R., Emhart, V., Mardones, O., Quiroga, J. J., Medina, A., Valenzuela, H., Espinoza, J., Burgos, Y., & Bozo, D. (2020). Climate and water availability impacts on early growth and growth efficiency of *Eucalyptus* genotypes: The importance of G×E interactions. *Forest Ecology and Management*, 458, 117763. <https://doi.org/10.1016/j.foreco.2019.117763> | ⁵ Hall, K. B., Stape, J., Bullock, B. P., Frederick, D., Wright, J., Scolforo, H. F., & Cook, R. (2019). A growth and yield model for *Eucalyptus benthamii* in the southeastern United States. *Forest Science*, 66(1), 25–37. <https://doi.org/10.1093/forsci/fxz061> | ⁶ Stape, J. L., Silva, C. R., & Binkley, D. (2022). Spacing and geometric layout effects on the productivity of clonal *Eucalyptus* plantations. *Trees, Forests and People*, 8, 100235. <https://doi.org/10.1016/j.tfp.2022.100235>

c) Naturally Regenerating Forest

Throughout the reporting period (2020-2023), there was an extensive amount of research focusing on naturally regenerating forests of *Populus tremuloides* Michx. (quaking aspen)¹⁻⁸. These studies investigated various aspects of the species, including its genetic structure, population history, range dynamics, phenology, trait variation, ecological importance, and management strategies. The studies provided valuable insights into the evolutionary history, ecological dynamics, and conservation of quaking aspen. Bagley et al. (2020) explored the genetic structure, population history, and range dynamics of quaking aspen in North America¹. They identified three distinct genetic clusters within the species range and provided evidence for stable-edge dynamics in the southwestern portions of the range¹. Blonder et al. (2023) investigated the spatiotemporal patterns and drivers of canopy phenology in quaking aspen². They reported that phenological traits vary across space and time and are influenced by genetic factors, such as cytotype and sex, as well as environmental factors, such as temperature, snowmelt date, and soil moisture. A primary emphasis of their results was the importance of considering lagged environmental effects and genetic factors in understanding phenological variation in quaking aspen². Cole et al. (2021) examined the levels of trait variation, heritabilities, and ontogenetic trajectories of various traits in aspen trees³. They reported high levels of genotypic variation in growth, defense, and reproductive traits, and they identified trade-offs between growth and defense. They stressed the importance of considering ontogenetic patterns and genetic variability in understanding trait variation in aspen trees³. Crouch et al. (2023) provided a systematic review of aspen ecology and management in the southwest US⁴. They identified factors influencing aspen forest dynamics, such as climate, fire, ungulates, insects, and diseases, as well as management strategies to promote aspen ecosystem resilience and adaptive capacity. Their review emphasized the importance of considering fire, ungulate browse, and climate in shaping aspen populations⁴. DeRose et al. (2022) evaluated the relationship between polyploidy, growth, and defense mechanisms in natural populations of quaking aspen⁵. They found that polyploidy influences the allocation of resources and trade-offs between growth and chemical defense in aspen populations. Overall, their study provided insights into the genetic basis of trait variation in aspen trees and the ecological dynamics of plant populations⁵. Ding et al. (2020) tested the performance of aspen trees in different breeding regions and assessed the phenotypic traits of the offspring⁶. They reported high genetic variation in growth and adaptive traits among aspen genotypes and provided valuable information for tree improvement programs in a changing climate⁶. Kruger et al. (2020) quantified the costs of allocation to chemical defenses in aspen trees⁷. Allocation to chemical defenses is costly for aspen trees under conditions of intense intraspecific competition and resource limitation, and the study highlighted the importance of considering the independent roles of chemical defenses and other growth co-determinants in understanding the costs of allocation to defense⁷. Lindroth et al. (2023) tested the phytochemical variation in trembling aspen in the Intermountain West region of North America⁸. They reported high genetic variation in foliar chemistry among aspen genotypes and ontogenetic changes in chemical expression as ramets age. In addition, the phytochemical profiles of western and Great Lakes populations of aspen were compared, highlighting the ecological implications of phytochemical variation⁸.

¹ Bagley, J. C., Heming, N. M., Gutiérrez, E. E., Devisetty, U. K., Mock, K. E., Eckert, A. J., & Strauss, S. H. (2020). Genotyping-by-sequencing and ecological niche modeling illuminate phylogeography, admixture, and pleistocene range dynamics in quaking aspen (*Populus tremuloides*). *Ecology and Evolution*, 10(11), 4609–4629. <https://doi.org/10.1002/ece3.6214> | ² Blonder, B. W., Brodrick, P. G., Chadwick, K. D., Carroll, E., Cruz-de Hoyos, R. M., Expósito-Alonso, M., Hateley, S., Moon, M., Ray, C. A., Tran, H., & Walton, J. A. (2023). Climate lags and genetics determine phenology in quaking aspen (*Populus tremuloides*). *New Phytologist*, 238(6), 2313–2328. <https://doi.org/10.1111/nph.18850> | ³ Cole, C. T., Morrow, C. J., Barker, H. L., Rubert-Nason, K. F., Riehl, J. F. L., Köllner, T. G., Lackus, N. D., & Lindroth, R. L. (2021). Growing up aspen: Ontogeny and trade-offs shape growth, defence and reproduction in a foundation species. *Annals of Botany*, 127(4), 505–517. <https://doi.org/10.1093/aob/mcaa070> | ⁴ Crouch, C. D., Rogers, P. C., Moore, M. M., & Waring, K. M. (2023). Building ecosystem resilience and adaptive capacity: A systematic review of aspen ecology and management in the Southwest. *Forest Science*, 69(3), 334–354. <https://doi.org/10.1093/forsci/foxad004> | ⁵ DeRose, R. J., Gardner, R. S., Lindroth, R. L., & Mock, K. E. (2022). Polyploidy and growth—Defense tradeoffs in natural populations of western quaking aspen. *Journal of Chemical*

Ecology, 48(4), 431–440. <https://doi.org/10.1007/s10886-022-01355-5> | ⁶ Ding, C., Hamann, A., Yang, R.-C., & Brouard, J. S. (2020). Genetic parameters of growth and adaptive traits in aspen (*Populus tremuloides*): Implications for tree breeding in a warming world. *PLOS ONE*, 15(3), e0229225. <https://doi.org/10.1371/journal.pone.0229225> | ⁷ Kruger, E. L., Keefover-Ring, K., Holeski, L. M., & Lindroth, R. L. (2020). To compete or defend: Linking functional trait variation with life-history tradeoffs in a foundation tree species. *Oecologia*, 192(4), 893–907. <https://doi.org/10.1007/s00442-020-04622-y> | ⁸ Lindroth, R. L., Wooley, S. C., Donaldson, J. R., Rubert-Nason, K. F., Morrow, C. J., & Mock, K. E. (2023). Phenotypic variation in phytochemical defense of trembling aspen in western North America: Genetics, development, and geography. *Journal of Chemical Ecology*, 49(5–6), 235–250. <https://doi.org/10.1007/s10886-023-01409-2>

d) Agroforestry and Trees Outside Forest

There are many potential benefits of incorporating short rotation woody crops (SRWCs) into conventional agricultural practices and agroforestry systems. Although these purpose-grown trees have the potential to enhance biomass productivity, carbon sequestration, reduction of GHG, and nutrient accrual, the adoption of SRWCs for agroforestry requires support in terms of information, resources, and networking opportunities for farmers and growers. Several studies have addressed the biological, ecological, and social aspects of integrating SRWCs with agroforestry¹⁻⁵. Gamble et al. (2020) described a study investigating the effects of different alley cropping systems on soil organic carbon (SOC) sequestration and nutrient accrual in Minnesota, USA¹. Their results provided valuable insights into the potential of these systems for agroforestry and climate change mitigation¹. Ile et al. (2022) reviewed the efficacy of producing SRWCs in the southeastern US to optimize biomass productivity, aboveground carbon sequestration, and belowground carbon sequestration in agroforestry systems². They concluded that using SRWCs for bioenergy production can lead to more efficient use of forest resources, improved forest management, and the growth of a sustainable green economy, while the establishment of SRWCs on degraded agricultural lands can help restore soil health and enhance soil carbon storage². Ile et al. (2023) documented a case study evaluating farmers' motivations and perceptions regarding the adoption of SRWCs in North Carolina³. They identified four key factors strongly influencing the adoption process: 1) market availability, 2) education awareness, 3) funding, and 4) social networking. Tailored information and support, including the establishment of farmers' cooperatives and online training courses for extension agents, were recommended to facilitate the adoption of SRWCs. Overall, farmers between the ages of 40 to 50 years old with larger land sizes were most likely to integrate SRWCs into their traditional agricultural systems³. Furthermore, from a biophysical perspective, Salceda et al. (2023) compared the removal efficiencies of total nitrogen (TN) and dissolved nitrogen (DN) in shallow groundwater by grass-only buffers and tree + grass buffers on grazing watersheds with deep loess soils⁴. They reported that both types of buffers were effective in reducing the concentrations of DN and TN in shallow groundwater, and that the establishment of sufficiently wide buffers can help mitigate groundwater contamination by nutrients and protect water quality⁴. Shults et al. (2020) measured the productivity and nutrient return of cover crops, plant-available soil nitrogen, and poplar leaf nutrients in different spacing treatments⁵. They reported that red clover cover crops can enhance nutrient availability and biomass production, but the effect on soil nitrogen and poplar leaf nutrients may vary depending on the spacing and vegetation treatments⁵.

¹ Gamble, J. D., Johnson, G. A., Current, D. A., Wyse, D. L., Zamora, D. S., & Sheaffer, C. C. (2020). Alley cropping affects perennial bioenergy crop root distribution, carbon, and nutrient stocks. *Agronomy Journal*, 112(5), 3718–3732. <https://doi.org/10.1002/agg2.20350> | ² Ile, O. J., McCormick, H., Skrabacz, S., Bhattacharya, S., Aguilos, M., Carvalho, H. D. R., Idassi, J., Baker, J., Heitman, J. L., & King, J. S. (2022). Integrating short rotation woody crops into conventional agricultural practices in the southeastern United States: A review. *Land*, 12(1), 10. <https://doi.org/10.3390/land12010010> | ³ Ile, O., Typhina, E., Brannum, K., Parajuli, R., Bardon, R., & King, J. (2023). Toward the adoption of new farming systems among farmers: A case study of short rotation woody crops in North Carolina. *Journal of Extension*, 61(1). <https://doi.org/10.34068/joe.61.01.09> | ⁴ Salceda, M., Udawatta, R. P., Anderson, S. H., Mendis, S. S., & Liu, F. (2023). Agroforestry buffers on nitrogen reduction in groundwater on a grazed hillslope. *Agrosystems, Geosciences & Environment*, 6(2), e20370. <https://doi.org/10.1002/agg2.20370> | ⁵ Shults, P., Nzokou, P., & Koc, I. (2020). Nitrogen contributions of alley cropped *Trifolium pratense* may sustain short rotation woody crop yields on marginal lands. *Nutrient Cycling in Agroecosystems*, 117(2), 261–272. <https://doi.org/10.1007/s10705-020-10068-8>

e) Harvesting

Several studies provided valuable insights into harvesting of poplars, willows, and eucalypts in the US, with key areas of study including understanding the impact of harvest timing, cultivar selection, field drying, and logistics optimization on biomass production, quality, and cost optimization¹⁻⁵. Overall, the findings of these studies may inform decision-making and policy development in the bioenergy sector, contributing to the development of more efficient and sustainable biomass production and utilization practices. De Souza et al. (2022) investigated the effects of harvest timing on biomass production and nutrient removal in shrub willow crops¹. They found that late-season harvests yielded more biomass compared to early-season harvests, and that second rotation yield was also higher than first rotation yield. Harvest timing affected nutrient removal, with nitrogen removal being highest in January and lowest in April¹. Eisenbies et al. (2021) summarized the results of a study on the performance of a single-pass cut-and-chip harvester in commercial-scale short-rotation poplar crops². Their objective was to understand the dynamics of material capacity and fuel consumption in poplar biomass crops, which can improve economic and environmental assessments of these systems. They showed that material capacity is influenced by standing biomass and field conditions, with material capacity increasing steadily with standing biomass, and that fuel consumption increases with tree height and is more pronounced in wetter weather conditions². Eisenbies et al. (2020) provided information on a study for collecting harvester performance data from commercial-scale willow harvests on several sites in New York, USA³. Overall, they evaluated the performance of the harvesters based on material capacity and fuel consumption, and they analyzed the impact of independent variables such as standing biomass, rainfall, and season³. Rockwood et al. (2023) focused on field drying of *Eucalyptus* biomass in Florida, USA, and its potential to enhance biomass quality⁴. They concluded that field drying effectively reduced wood moisture content and enhanced biomass quality. The duration of field drying affected drying, but species, tree size, and tree form did not have a significant impact. Their findings have implications for the energy production industry, as reducing the moisture content of woody biomass can improve the efficiency of conversion processes and reduce energy costs⁴. Wang et al. (2020) discussed the optimization of biomass supply chains and the delivered cost of multiple biomass feedstocks in the northeastern US⁵. They identified the most cost-efficient logistics for biomass transportation and analyzed the factors that affect the delivered cost. Overall, supplying feedstocks to local and relatively small to medium-sized facilities may reduce transportation costs and make the overall economics feasible. These and other findings of the study may be used to inform decision-making and policy development in the bioenergy sector⁵.

¹ De Souza, D. P., Eisenbies, M. H., & Volk, T. A. (2022). Growing season harvests of shrub willow (*Salix* spp.) have higher nutrient removals and lower yields compared to dormant-season harvests. *Forests*, 13(11), 1936. <https://doi.org/10.3390/f13111936> | ² Eisenbies, M. H., Volk, T. A., DeSouza, D., Hallen, K., Stanton, B., Espinoza, J., Himes, A., Shuren, R., Stonex, R., Summers, B., & Zepa, J. (2021). An assessment of the harvesting and fuel performance of a single-pass cut-and-chip harvester in commercial-scale short-rotation poplar crops as influenced by crop and weather conditions. *Biomass and Bioenergy*, 149, 106075. <https://doi.org/10.1016/j.biombioe.2021.106075> | ³ Eisenbies, M. H., Volk, T. A., De Souza, D. P. L., & Hallen, K. W. (2020). Cut-and-chip harvester material capacity and fuel performance on commercial-scale willow fields for varying ground and crop conditions. *GCB Bioenergy*, 12(6), 380–395. <https://doi.org/10.1111/gcbb.12679> | ⁴ Rockwood, D. L., Tamang, B., & Ellis, M. F. (2023). Field drying for enhancing biomass quality of *Eucalyptus* logs and trees in Florida, USA. *Forests*, 14(5), 899. <https://doi.org/10.3390/f14050899> | ⁵ Wang, Y., Wang, J., Schuler, J., Hartley, D., Volk, T., & Eisenbies, M. (2020). Optimization of harvest and logistics for multiple lignocellulosic biomass feedstocks in the northeastern United States. *Energy*, 197, 117260. <https://doi.org/10.1016/j.energy.2020.117260>

f) Utilization for Wood Products

Several research studies contributed to the development of sustainable lignocellulosic fibers from renewable resources using green chemistry. The findings have implications for the production of pellets, medium density fiberboard (MDF), packaging, bioplastics, sanitary towels, wipes, and other products in the bio-based and circular economy¹⁻⁵. Overall, these studies highlighted the importance of sustainable

utilization of bioresources and the need for alternative pulping methods and raw materials to reduce ecological damage and promote efficient recycling of lignocellulosic fibers. Elniski et al. (2023) discussed the use of lignin as an additive to enhance the properties of willow pellets, which are used as a renewable energy source¹. While lignin interacts with cellulose and hemicelluloses to promote bonding during pellet production, its effect on bulk density and ash content of pellets is inconsistent. For example, lignin generally increases the energy content of pellets, but its impurities can reduce energy content¹. Ghezehei and Saloni (2023) evaluated the suitability of poplar stem-wood samples as feedstocks for pellet production, based on feedstock source location/region, age of stands, poplar clones, and tree stem sections². They identified the best-performing poplar clones for each region and examined the effects of stem position on feedstock quality, comparing their results to pellet quality threshold values². Himes et al. (2021) described the variation in wood properties [i.e., specific gravity, dimensional stability, hardness, screw withdrawal, and bending strength and stiffness {modulus of rupture (MOR) and modulus of elasticity (MOE), respectively}] of hybrid poplar grown in Boardman, Oregon, USA³. They assessed radial and vertical stem positions and determined wind influenced significant differences in some traits between different sides of the trees. In addition, boards milled closer to the bark had greater MOE and MOR than those from interior stem sections. Their results may be used to target specific stem positions for different applications³. Rockwood et al. (2022) highlighted the suitability of different poplar and eucalypt wood species and genotypes for MDF production in Florida, USA⁴. They analyzed the physical and mechanical properties of MDF panels, and tested effects from species, genotype, and log. Their results provided valuable insights for the selection of wood materials for MDF manufacturing⁴. Salem et al. (2021) explored the dynamics of fiber production from wood, non-wood, and agro-residues using carbonate hydrolysis and a mild kraft process without bleaching agents⁵. Their objective was to identify a sustainable alternative to synthetic polymer feedstocks for the production of packaging, bioplastics, sanitary towels, wipes, and related products. Based on their results, carbonate hydrolysis and mild kraft pulping can produce higher value fibers with sustainable properties⁵.

¹ Elniski, A., Dongre, P., & Bujanovic, B. M. (2023). Lignin use in enhancing the properties of willow pellets. *Forests*, 14(10), 2041. <https://doi.org/10.3390/f14102041> | ² Ghezehei, S. B., & Saloni, D. (2023). Assessment of feedstock quality of poplars (*Populus* L.) using selected pellet-quality parameters. *BioEnergy Research*, 17(2), 993–1003. <https://doi.org/10.1007/s12155-023-10711-6> | ³ Himes, A., Leavengood, S., & Polinko, A. (2021). Variation in wood properties of hybrid poplar lumber by radial and vertical position in stem: A case study from Boardman, Oregon. *Wood and Fiber Science*, 53(3), 161–177. <https://doi.org/10.22382/wfs-2021-17> | ⁴ Rockwood, D. L., Winandy, J. E., & Gribbins, N. R. (2022). Processing and wood factors influence medium density fiberboard production from young *Eucalyptus grandis*, *E. amplifolia*, *Corymbia torelliana*, and cottonwood grown in Florida USA. *Forests*, 13(2), 266. <https://doi.org/10.3390/f13020266> | ⁵ Salem, K. S., Naithani, V., Jameel, H., Lucia, L., & Pal, L. (2021). Lignocellulosic fibers from renewable resources using green chemistry for a circular economy. *Global Challenges*, 5(2), 2000065. <https://doi.org/10.1002/gch2.202000065>

g) Utilization for Bioenergy

Throughout the reporting period (2020-2023), there was an extensive amount of research focusing on the potential of using feedstocks from SRWCs for bioenergy, including bio-jet fuel and bioethanol production¹⁻⁶. For example, Budsberg et al. (2022) highlighted the environmental and economic benefits of switching to poplar-based biofuels, including reduced fuel use, lower greenhouse gas emissions, and potential economic viability¹. Buell et al. (2023) summarized the potential of plant biology, genetics, and biotechnology in addressing various challenges in agriculture, renewable energy, and climate change – as they relate to feedstocks for renewable bioenergy sources². In particular, their review focused on plant architecture, gene expression and regulation, lignin biosynthesis, trichome development, CRISPR-Cas9 technology, biofuel production, and climate change and plant growth². Davis et al. (2020) modeled the economic potential for *Eucalyptus* as feedstocks for jet fuel additives in the US and found that the genus had potential to fulfill a portion of the demand for these additives³. They discussed production potential, distribution, market opportunities, and co-product utilization as they relate to challenges and

uncertainties of producing *Eucalyptus*³. Jeong et al. (2020) evaluated the economic feasibility and environmental impact of using short rotation coppice (SRC) poplar as a feedstock for a biopower plant in Indiana, USA⁴. Based on a sensitivity analysis, they discussed different land types and the carbon effects of the biopower plant and concluded the plant operation is currently not economically feasible but could be viable with a higher carbon tax and stringent environmental policy targets⁴. Kim et al. (2021) assessed the tradeoffs between yield, disease incidence, and conversion efficiency of hybrid poplar genotypes used as bioenergy feedstocks⁵. They contributed information about wood chemistry (e.g., carbohydrate and lignin content) that may inform decisions for biomass utilization and bioenergy production⁵. Morales-Vera et al. (2022) emphasized the importance of conducting comprehensive life cycle assessments (LCA) to evaluate the environmental performance of biofuel production in the Pacific Northwest, USA⁶. Their results highlighted the environmental impacts of different stages of production and provided insights into areas for improvement. For example, future studies may focus on environmentally safe ways to produce ethanol in biorefineries and explore additional approaches to assess both the environmental and cost aspects of ethanol life cycles⁶.

¹ Budsberg, E., Parker, N., Bandaru, V., Bura, R., & Gustafson, R. (2022). Hydrocarbon bio-jet fuel from bioconversion of poplar biomass: Life cycle assessment of site-specific impacts. *Forests*, 13(4), 549. <https://doi.org/10.3390/f13040549> | ² Buell, C. R., Dardick, C., Parrott, W., Schmitz, R. J., Shih, P. M., Tsai, C.-J., & Urbanowicz, B. (2023). Engineering custom morpho- and chemotypes of *Populus* for sustainable production of biofuels, bioproducts, and biomaterials. *Frontiers in Plant Science*, 14, 1288826. <https://doi.org/10.3389/fpls.2023.1288826> | ³ Davis, M. R., Kainer, D., Tuskan, G. A., Langholtz, M. H., Hellwinckel, C. M., Shedden, M., & Eaton, L. (2020). Modeled economic potential for *Eucalyptus* spp. Production for jet fuel additives in the United States. *Biomass and Bioenergy*, 143, 105807. <https://doi.org/10.1016/j.biombioe.2020.105807> | ⁴ Jeong, D., Tyner, W. E., Meilan, R., Brown, T. R., & Doering, O. C. (2020). Stochastic techno-economic analysis of electricity produced from poplar plantations in Indiana. *Renewable Energy*, 149, 189–197. <https://doi.org/10.1016/j.renene.2019.11.061> | ⁵ Kim, K., Voothuluru, P., Hamilton, C., McCord, J., Tamang, B., Cunningham, M., Eberhardt, T. L., Rials, T., & Labbé, N. (2021). Tradeoffs between yield, disease incidence and conversion efficiency for selection of hybrid poplar genotypes as bioenergy feedstocks. *Biomass and Bioenergy*, 154, 106259. <https://doi.org/10.1016/j.biombioe.2021.106259> | ⁶ Morales-Vera, R., Vázquez-Ibarra, L., Scott, F., Puettmann, M., & Gustafson, R. (2022). Life cycle assessment of bioethanol production: A case study from poplar biomass growth in the U.S. Pacific Northwest. *Fermentation*, 8(12), 734. <https://doi.org/10.3390/fermentation8120734>

5. Environmental and Ecosystem Services

a) Site and Landscape Improvement

There were several studies highlighting the importance of using different materials such as biochar, sewage sludge, and fibercake residuals as soil amendments for unique restoration techniques¹⁻⁵. These studies aimed to improve soil fertility, nutrient availability, and carbon sequestration in different types of soils and ecosystems across varying landscapes. Domingues et al. (2020) showed the addition of different biochars significantly increased soil pH and cation exchange capacity (CEC)¹. They reported that high-ash biochars produced at low pyrolysis temperatures were effective in increasing CEC without improving soil fertility, while low-ash biochars increased carbon storage without causing alkalization. Their study provided valuable insights for tailoring and selecting biochars for specific agronomic and environmental applications, such as improving soil CEC¹. Guerrini et al. (2021) investigated the potential of sewage sludge as a pedotechnomaterial (PTM) for soil recovery in degraded tropical soils². Their results showed that applying sewage sludge led to improvements in soil fertility, nutrient availability, and plant growth, highlighting the potential of sewage sludge as a sustainable alternative for soil and ecosystem restoration². Rockwood et al. (2022) discussed the carbon sequestration potential of *Eucalyptus* trees in Florida, USA using biochar as a soil amendment that enhanced carbon sequestration of the intensively managed short rotation woody crop systems³. In addition, they provided cost estimates for using biochar as a soil amendment, which are lower compared to previous estimates for carbon sequestration in forestry³. Rogers et al. (2021) tested the use of fibercake residuals from paper and containerboard production as soil amendments for hybrid poplar clones⁴. While specific effects varied depending on clone and type of fibercake used, their results showed the use of such residuals can positively impact the

early growth and establishment of poplar trees⁴. Vinhal et al. (2022) described the design, implementation, and testing of specialized willow restoration mats using the DeValix technique at phytoremediation sites⁵. They identified generalist and specialist clones based on performance across sites in the Lake Superior and Lake Michigan watersheds of the US, and they showed DeValix mats can successfully establish willow clones and promote their growth for site and landscape improvement⁵.

¹ Domingues, R. R., Sánchez-Monedero, M. A., Spokas, K. A., Melo, L. C. A., Trugilho, P. F., Valenciano, M. N., & Silva, C. A. (2020). Enhancing cation exchange capacity of weathered soils using biochar: Feedstock, pyrolysis conditions and addition rate. *Agronomy*, 10(6), 824. <https://doi.org/10.3390/agronomy10060824> | ² Guerrini, I. A., Sampaio, T. F., Bogiani, J. C., Backes, C., Harrison, R. B., Oliveira, F. C., Gava, J. L., Traballi, R. C., Garuba De Menezes Mota, R., Roder, L. R., Grilli, E., Ganga, A., James, J. N., & Capra, G. F. (2021). Sewage sludge as a pedotechnomaterial for the recovery of soils compacted by heavy machinery on *Eucalyptus* commercial plantation. *Journal of Cleaner Production*, 325, 129320. <https://doi.org/10.1016/j.jclepro.2021.129320> | ³ Rockwood, D. L., Ellis, M. F., & Fabbro, K. W. (2022). Economic potential for carbon sequestration by short rotation eucalypts using biochar in Florida, USA. *Trees, Forests and People*, 7, 100187. <https://doi.org/10.1016/j.tfp.2021.100187> | ⁴ Rogers, E. R., Zalesny, R. S., Benzel, T., & Wiese, A. H. (2021). Establishment of poplars in soils amended with fibercake residuals from paper and containerboard production. *Agroforestry Systems*, 95(8), 1719–1733. <https://doi.org/10.1007/s10457-021-00680-0> | ⁵ Vinhal, R. A., Zalesny, R. S., DeBauche, B. S., Rogers, E. R., Pilipović, A., Soolanayakanahally, R. Y., & Wiese, A. H. (2022). Establishment of willows using the novel DeValix technique: Ecological restoration mats designed for phytotechnologies. *International Journal of Phytoremediation*, 24(7), 730–743. <https://doi.org/10.1080/15226514.2021.1970102>

b) Phytoremediation and Associated Phytotechnologies

Throughout the reporting period (2020-2023), there was an extensive amount of research focusing on phytoremediation and associated phytotechnologies as sustainable, nature-based approaches utilizing plants to remove and/or degrade pollutants from soil, water, and air¹⁻⁶. Overall, the studies demonstrated the potential of poplars and other short rotation woody crops (SRWCs) for environmental solutions in various contaminated environments. The findings highlighted the importance of selecting appropriate clones based on specific contamination and environmental conditions. Overall, the studies contributed to our understanding of the long-term benefits and ecosystem services provided by SRWCs and emphasized the need for further research and the development of best management practices to maximize the potential of phytoremediation in ecological restoration efforts. For example, Landmeyer et al. (2020) tested salt-tolerant hybrid poplar trees for phytoremediation of slightly brackish groundwater contaminated with polycyclic aromatic hydrocarbons (PAHs) at a Superfund site in the Central Valley of California, USA¹. They found that all salt-tolerant genotypes grew well and effectively removed naphthalene, a PAH compound, from the groundwater, highlighting the potential of using these trees for phytoremediation in these and other contaminated sites with deep groundwater¹. Pilipović et al. (2020) investigated the growth and physiological responses of three poplar clones grown on soils artificially contaminated with heavy metals, diesel fuel, and herbicides². They reported that growth and physiological responses were influenced by the interactions of clones with different contaminant types (i.e., genotype × environment interactions), as well as emphasizing the importance of physiological traits in poplar breeding and subsequent phytoremediation testing². Zalesny et al. (2021) described the use of phyto-recurrent selection for the establishment of a regional phytotechnologies network consisting of sixteen phytoremediation buffer systems (i.e., phyto buffers) in the Great Lakes Basin, USA³. They examined genotype × environment interactions and evaluated the health, biomass, and growth of twelve poplar clones across the sixteen phyto buffers. Certain clones exhibited specialist or generalist responses in terms of biomass and growth, emphasizing the importance of considering genotype × environment interactions in phytoremediation projects and while developing phytotechnology best management practices³. Building on the work of Zalesny et al. (2021), Pilipović et al. (2020) compared the growth and physiology of three clone groups tested across the phyto buffers: 1) NRRI clones, 2) Experimental clones, and 3) Common clones⁴. The NRRI clones were new genotypes produced by the University of Minnesota Duluth, Natural Resources Research Institute. The Experimental clones had been tested broadly in the region but had not reached commercial status. The Common clones had been used in decades of testing and deployment in the Midwestern United States. Overall, they showed the NRRI clones exhibited the

greatest potential for improving water quality and reducing nutrient runoff at landfills and similar liability sites in the region⁴. Salehi et al. (2023) tested the growth, biomass, nutrition, and heavy-metal accumulation of three poplar clones and one willow clone following irrigation with uncontaminated tap water versus wastewater⁵. They reported that eight months of irrigation with heavy-metal contaminated wastewater increased height, leaf area, tissue biomass, and phytoextraction and phytoaccumulation of macro-/micro-nutrients and heavy metals in tree tissues relative to the tap water control, demonstrating that these genotypes have potential for uses in systems with soils affected by similarly contaminated urban wastewaters⁵. Zalesny et al. (2020) evaluated the phytoremediation potential, biomass production, and biofuel recalcitrance of two hybrid poplar clones, 'DN34' and 'NM6', grown at a former landfill site in Rhineland, Wisconsin, USA⁶. They highlighted that while both genotypes were effective in phytoaccumulation and phytoextraction of contaminants, 'NM6' outperformed 'DN34' for biomass and carbon-related traits while 'DN34' exhibited higher uptake levels for most elements despite 'NM6' having higher concentrations of certain elements in specific tissues⁶. Overall, their results contributed to the understanding of the long-term benefits and ecosystem services provided by poplar clones in landfill phytoremediation.

¹ Landmeyer, J. E., Rock, S., Freeman, J. L., Nagle, G., Samolis, M., Levine, H., Cook, A., & O'Neill, H. (2020). Phytoremediation of slightly brackish, polycyclic aromatic hydrocarbon-contaminated groundwater from 250 ft below land surface: A pilot-scale study using salt-tolerant, endophyte-enhanced hybrid poplar trees at a superfund site in the central valley of California, April–November 2019. *Remediation Journal*, 31(1), 73–89. <https://doi.org/10.1002/rem.21664> | ² Pilipović, A., Zalesny Jr., R. S., Orlović, S., Drekić, M., Pekeć, S., Katanić, M., & Poljaković-Pajnik, L. (2020). Growth and physiological responses of three poplar clones grown on soils artificially contaminated with heavy metals, diesel fuel, and herbicides. *International Journal of Phytoremediation*, 22(4), 436–450. <https://doi.org/10.1080/15226514.2019.1670616> | ³ Zalesny, R. S., Pilipović, A., Rogers, E. R., Burken, J. G., Hallett, R. A., Lin, C.-H., McMahon, B. G., Nelson, N. D., Wiese, A. H., Bauer, E. O., Buechel, L., DeBauche, B. S., Peterson, M., Seegers, R., & Vinhal, R. A. (2021). Establishment of regional phytoremediation buffer systems for ecological restoration in the Great Lakes Basin, USA. I. Genotype × Environment Interactions. *Forests*, 12(4), 430. <https://doi.org/10.3390/f12040430> | ⁴ Pilipović, A., Zalesny Jr., R. S., Rogers, E. R., McMahon, B. G., Nelson, N. D., Burken, J. G., Hallett, R. A., & Lin, C.-H. (2021). Establishment of regional phytoremediation buffer systems for ecological restoration in the Great Lakes Basin, USA. II. New clones show exceptional promise. *Forests*, 12(4), 474. <https://doi.org/10.3390/f12040474> | ⁵ Salehi, A., Zalesny, R. S., & Calagari, M. (2023). Effects of urban wastewater application on growth, biomass, nutrition, and heavy-metal accumulation of *Populus nigra* L. “62/154,” *P. alba* L. “20/45,” *P. euramericana* (Dode) Guinier “92/40,” and *Salix excelsa* S.G. Gmel grown in heavy-metal contaminated soil. *International Journal of Phytoremediation*, 25(10), 1371–1383. <https://doi.org/10.1080/15226514.2022.2158783> | ⁶ Zalesny Jr., R. S., Zhu, J. Y., Headlee, W. L., Gleisner, R., Pilipović, A., Acker, J. V., Bauer, E. O., Birr, B. A., & Wiese, A. H. (2020). Ecosystem services, physiology, and biofuels recalcitrance of poplars grown for landfill phytoremediation. *Plants*, 9(10), 1357. <https://doi.org/10.3390/plants9101357>

c) Environmental Applications for Ecosystem Services

Studies highlighting the potential benefits of using shrub willow buffers for bioenergy production and land management exemplify environmental applications for ecosystem services¹⁻⁵. These buffers reduce greenhouse gas (GHG) emissions, provide ecosystem services, and improve soil health. Strategic placement of buffers and consideration of factors such as yield, fertilizer application rate, and transportation distance are important for optimizing the benefits and minimizing the impacts of shrub willow production. Overall, the findings contributed information on the potential development of a sustainable bioeconomy and provided insights for decision-making in the establishment of shrub willow biomass crops¹⁻⁵. Canter et al. (2022) showcased the importance of strategic placement of shrub willow buffers to optimize GHG emissions and transportation distance¹. They reported that shrub willow production on marginal land has the potential to contribute to a sustainable bioeconomy with reduced GHG emissions¹. Englund et al. (2020) conducted a global assessment of the performance and progress of multifunctional perennial production systems for bioenergy and presented several case studies that illustrate the potential to meet bioenergy demands². Also, they described the importance of identifying suitable locations for the strategic establishment of perennial crops. They emphasized the need to understand the specific landscape characteristics and environmental impacts in order to direct implementation efforts to areas where the benefits of perennialization could be maximized². Jager et al. (2023) focused on the Mid-Atlantic region of the US and examined the potential value of ecosystem services provided by multi-zone riparian buffers consisting of native, perennial crops such as switchgrass and willow³. Their results suggested that strategic placement and choice of low-input perennial

feedstocks are important for providing ecosystem services and reducing the carbon footprint of biofuels³. Yang et al. (2020) tested the production of willow biomass crops in upstate New York, USA and reported that willow crops were carbon-negative when grown on former cropland or pasture⁴. They highlighted the importance of considering multiple factors, such as land cover, transportation distance, and yield, in assessing GHG emissions and energy ratios. Their results may be used to inform policy-makers, land managers, and scientists in making informed decisions about the establishment of willow biomass crops and reducing GHG emissions in the bioenergy sector⁴. Zumpf et al. (2021) demonstrated that strategically integrated shrub willow buffers in an Illinois corn-soybean field had positive effects on soil health and microbial community composition⁵. Overall, their study contributed to the understanding of the relationship between crop management, soil health, and microbial communities, and provided insights into the potential benefits of perennial bioenergy crop production for sustainable land management⁵.

¹ Canter, C. E., Zolton, K., Cacho, J. F., Negri, M. C., Zumpf, C. R., & Quinn, J. J. (2022). Impact of landscape design on the greenhouse gas emissions of shrub willow bioenergy buffers in a US midwest corn-production landscape. *Biofuels, Bioproducts and Biorefining*, 16(3), 629–639. <https://doi.org/10.1002/bbb.2345> | ² Englund, O., Dimitriou, I., Dale, V. H., Kline, K. L., Mola-Yudego, B., Murphy, F., English, B., McGrath, J., Busch, G., Negri, M. C., Brown, M., Goss, K., Jackson, S., Parish, E. S., Cacho, J., Zumpf, C., Quinn, J., & Mishra, S. K. (2020). Multifunctional perennial production systems for bioenergy: Performance and progress. *WIREs Energy and Environment*, 9(5), e375. <https://doi.org/10.1002/wene.375> | ³ Jager, H. I., Nair, S. S., Efroymson, R. A., DeRolph, C. R., Parish, E. S., & Wang, G. (2023). Ecosystem services from partially harvested riparian buffers can offset biomass production costs. *Science of The Total Environment*, 889, 164199. <https://doi.org/10.1016/j.scitotenv.2023.164199> | ⁴ Yang, S., Volk, T., & Fortier, M.-O. (2020). Willow biomass crops are a carbon negative or low-carbon feedstock depending on prior land use and transportation distances to end users. *Energies*, 13(16), 4251. <https://doi.org/10.3390/en13164251> | ⁵ Zumpf, C., Cacho, J., Grasse, N., Quinn, J., Hampton-Marcell, J., Armstrong, A., Campbell, P., Negri, M. C., & Lee, D. K. (2021). Influence of shrub willow buffers strategically integrated in an Illinois corn-soybean field on soil health and microbial community composition. *Science of The Total Environment*, 772, 145674. <https://doi.org/10.1016/j.scitotenv.2021.145674>

III. GENERAL INFORMATION

1. Administration and Operation of the National Poplar Commission or Equivalent

a) USDA Forest Service as an “Other Suitable National Body”

The US Poplar Council, which was started in 1964, strongly advocated for the US to join the International Poplar Commission (IPC). Article IV of the original IPC Convention, which came into force in 1961, established that member countries had to designate a National Poplar Commission. As a private organization, the US Poplar Council could not be sanctioned by the US to act as the National Poplar Commission in fulfillment of the requirement to join the IPC Convention. Recognizing that the National Poplar Commission requirement was preventing the US (and other nations) from joining the IPC Convention, the IPC Executive Committee amended the Convention in 1967 with the adopted text “either for the establishment of a National Poplar Commission, or if not possible, for the designation of some other suitable National body”.

In July of 1970, Secretary of State William P. Rogers wrote Secretary of Agriculture Clifford M. Hardin to inform him the Circular 175 procedure for the IPC had been completed. The Instrument of Acceptance of the Convention was transmitted to the FAO Director-General A.H. Boerma in August 1970, at which time he wrote the Secretary of State to confirm receipt. In October 1970, USDA Forest Service Chief Edward Cliff informed the Poplar Council of the acceptance of US membership into the IPC, and he thanked them for their support. In this letter, Chief Cliff noted that the USDA Forest Service had been designated as the US member agency — thus, **it was established that the USDA Forest Service fulfilled the IPC Convention requirement of “some other suitable National body”** in lieu of a National Poplar Commission. The USDA

Forest Service has maintained this role since 1970, and the national delegate to the IPC has been under the appointment of the USDA Forest Service Chief (or Deputy Chief for Research and Development).

Historically, there have been two IPC-affiliated organizations from the US. First, the US Poplar Council was active from 1964 to 2017. Second, the Short Rotation Woody Crops Operations Working Group (SRWCOWG) was formed in 1995 and remains active today. Given the substantially reduced number of US experts on poplars and other fast-growing trees throughout the 2000s, the US Poplar Council disbanded in 2017, with active members at that time joining the SRWCOWG. As a result, the SRWCOWG remains the sole organization dedicated to the cultivation and utilization of poplars, willows, eucalypts, and other fast-growing species in the US. Throughout time, USDA Forest Service researchers, including the US National Representative to the IPC, have held leadership roles in both groups, and this continues at present. The current connection between the IPC and SRWCOWG remains strong, and additional partnerships with the Poplar and Willow Council of Canada (PWCC) have resulted in substantial advancements in the development of short rotation woody crops throughout North America. These accomplishments are reflected in the activities, relations with other countries, innovations, and publications listed below.

b) Meetings, Congresses, and Study Tours Sponsored by IPC-Affiliated Organizations

- ✦ 2nd World Forum on Urban Forests: Greener, Healthier, and Happier Cities for All; 16-20 October 2023; Washington, District of Columbia, USA.
- ✦ 2023 International Phytoremediation Training Academy, USDA Forest Service International Programs; 14-18 August 2023; Menomonee Falls, Wisconsin, USA.
- ✦ 17th International Phytotechnologies Conference; 23-26 May 2023; Chicago, Illinois, USA.
- ✦ 8th International Poplar Symposium (IPS VIII): Poplars and willows in the era of global change: agroforestry, environmental improvement, and ecosystem services to enhance livelihoods; IUFRO Working Group 2.08.04 (Physiology and Genetics of Poplars and Willows); 4-6 October 2022; Novi Sad, Serbia (virtual).
- ✦ Joint Workshop of the Food and Agriculture Organization of the United Nations (UN FAO), International Commission on Poplars and Other Fast-Growing Trees Sustaining People and the Environment (IPC), and TreeDivNet: Management of Mixed and Pure Planted Forests – Stock-taking of Science and Practice; 30 September 2022; Rome, Italy.
- ✦ International Commission on Poplars and Other Fast-Growing Trees Sustaining People and the Environment (IPC) of the Food and Agriculture Organization of the United Nations (UN FAO); 29 September 2022; Rome, Italy.
- ✦ 9th Bioenergy Week of the Global Bioenergy Partnership (GBEP) of the Food and Agriculture Organization of the United Nations (UN FAO) (from Rhinelander, WI, USA); 26-29 September 2022; Asunción, Paraguay (virtual).
- ✦ Phyto Day: Enhancing Ecosystem Services Using Phytotechnologies for a Sustainable World; 14 June 2022; Mequon, Wisconsin, USA.
- ✦ 13th Biennial Short Rotation Woody Crops Operations Working Group Conference: 2022 International Short Rotation Woody Crops Conference; 2-4 May 2022; Mills River, North Carolina, USA.

- ☛ 26th Session of the International Commission on Poplars and Other Fast-Growing Trees Sustaining People and the Environment (IPC), Food and Agriculture Organization (FAO) of the United Nations; 5-8 October 2021; Rome, Italy (virtual).
- ☛ Short Rotation Woody Crops Regional Updates Webinar Series (this is different than the webinar series listed below); hosted monthly by the University of Minnesota Duluth, Natural Resources Research Institute (NRRI); August-December 2020; Duluth, Minnesota, USA (virtual).
- ☛ Short Rotation Woody Crops Webinar Series (this took place in lieu of the 2020 Woody Crops International Conference due to the COVID-19 pandemic and is different than the regional updates webinar series listed above); hosted monthly by the University of Minnesota Duluth, Natural Resources Research Institute (NRRI); August-December 2020; Duluth, Minnesota, USA (virtual).

2. Relations with Other Countries

- ☛ Following all national phytosanitary requirements, there are active exchanges of poplar, willow, and eucalypt plant material from the US with many countries around the globe, including Brazil, Canada, China, India, Italy, Sweden, and the United Kingdom. US scientists that work with poplar, willow, and eucalypt collaborate all over the world through IPC and IUFRO activities in their area of expertise.
- ☛ Ron Zalesny of the USDA Forest Service in Rhinelander, Wisconsin USA hosted a former scientist from the University of Novi Sad, Institute of Lowland Forestry and Environment, in Novi Sad, Serbia (Andrej Pilipović, who is currently with the Natural Resources Research Institute, University of Minnesota Duluth, Duluth, Minnesota, USA) for a long-term (i.e., 20-month, 2020-2021) scientific exchange conducting phytotechnologies research at the USDA Forest Service, Northern Research Station. The overarching outcome was a set of phytotechnologies best management practices (i.e., Phyto BMPs) that are locally adapted within the Great Lakes Basin yet globally relevant. The scientific exchange was administratively supported by USDA Forest Service International Programs.
- ☛ Ron Zalesny, Liz Rogers, and Ryan Vinhal of the USDA Forest Service in Rhinelander, Wisconsin USA, along with Liza Paqueo of the USDA Forest Service International Programs office in Washington, District of Columbia, USA, developed and hosted the International Phytoremediation Training Academy (2023-current). The overarching objective of the academy is to bring together global researchers and practitioners to promote the successful planning, design, and establishment of phytoremediation systems through a combination of hands-on experiences, engagement with phytotechnologies professionals, and ready-to-use resources. Participants in 2023 were from the Democratic Republic of the Congo, Bangladesh, Lebanon, and India. Currently, participants for 2024 are being selected. The academy is funded by USDA Forest Service International Programs. Other instructors include Chung-Ho Lin (University of Missouri, Columbia, Missouri, USA) and Akshat Tyagi (Give Me Trees Trust, Delhi, India).
- ☛ From 2019-2020, Andrew Groover of the USDA Forest Service in Burlington, Vermont, USA hosted a Ph.D. student (Aleksandra Słupianek) from Poland in support of her developing laser capture methods in poplar wood (visit ended by pandemic).

- ✦ From 2018-2020, Andrew Groover of the USDA Forest Service in Burlington, Vermont, USA hosted an Associate Professor from the Chinese Academy of Forestry (Lijuan Wang) for training in genomics (visit ended by pandemic).
- ✦ Don Rockwood of Florida FGT, LLC in Orlando, Florida, USA provided eucalypt seed to North Carolina State University's Camcore program for evaluation by its members in several wet subtropic countries.
- ✦ The University of Minnesota Duluth, Natural Resources Research Institute (NRRI) (Duluth, Minnesota, USA) shipped hybrid poplar cuttings to a phytoremediation company in Canada to begin a trial in 2023.
- ✦ The University of Minnesota Duluth, Natural Resources Research Institute (NRRI) (Duluth, Minnesota, USA) collaborated with Euromediana in Lithuania that established a nursery bed of NRRI hybrid poplar clones. In 2023, Euromediana shared the NRRI cuttings with a wood products company in Europe and the Swedish University of Agricultural Sciences.

3. Innovations not Included in Other Sections

a) Notable Publications from Sources Outside Peer-Reviewed Journals

1. Buchman, D. Jackson, J., Berguson, W.E., McMahon, B.G., Nelson, N.D.; DuPlissis, J., & Host, G.E. (2020). Grower's Guide for Hybrid Poplar Plantations for Biomass Production. Natural Resources Research Institute, University of Minnesota Duluth. Retrieved from the University of Minnesota Digital Conservancy; Minneapolis, Minnesota, USA. 15 p. <http://hdl.handle.net/11299/213759>
2. Ghezehei, S., & Saloni, D. (2023). *Populus*-based Production of Solid Biofuel. Final Technical Report prepared for the North Carolina Department of Agriculture and Consumer Services Bioenergy Research Initiative. North Carolina State University; Raleigh, North Carolina, USA.
3. Ghezehei, S. (2023). The Silviculture of *Populus* for Veneer. Final Technical Report prepared for the North Carolina Department of Agriculture and Consumer Services New and Emerging Crops Program. North Carolina State University; Raleigh, North Carolina, USA.
4. Ghezehei, S., & Hazel, D. (2022). Intercropping *Populus* for Bioenergy and Veneer. Final Technical Report prepared for the North Carolina Department of Agriculture and Consumer Services Bioenergy Research Initiative. North Carolina State University; Raleigh, North Carolina, USA.
5. Ghezehei, S., Kurkalova, L.A., Hazel, D., & Nichols, E.G. (2021). Predicting SRWC Productivity and Economic Feasibility. Final Technical Report prepared for the North Carolina Department of Agriculture and Consumer Services Bioenergy Research Initiative. North Carolina State University; Raleigh, North Carolina, USA.
6. Ghezehei, S., & Hazel, D. (2020). Western North Carolina Bioenergy Forest Demonstration. Final Technical Report prepared for the North Carolina Department of Agriculture and Consumer Services Bioenergy Research Initiative. North Carolina State University; Raleigh, North Carolina, USA.
7. Hazel, D.W., Ghezehei, S.B., & Nichols, E.G. (2021). Growing Hardwood Veneer in 20 Years or Less on Your Land? National Woodlands, a Magazine for the National Woodland Owners Association. Autumn, 2021, 44(1). <https://nationalwoodlands.com/publications/>

8. Heppenstall, M., Himes, A., Haider, N., & Hart, N. (2023). Harvesting Hybrid Poplar for Bioenergy. In: Growing Hybrid Poplar for Bioenergy in the PNW. Zobrist, K., Haider, N., Stanton, B.J., & Stonex, R. (eds.). Washington State University Extension; Pullman, Washington, USA. 115 p. <https://doi.org/10.7273/000005545>
9. Jackson, J., & Reichenbach, M. (2023). A Collaborative Learning Process from a Distance. Natural Resources Research Institute, University of Minnesota Duluth. Retrieved from the University of Minnesota Digital Conservancy; Minneapolis, Minnesota, USA. 15 p. <https://hdl.handle.net/11299/256906>
10. Jackson, J., DuPlissis, J., & Meyer, N. (2022). Next-Gen Poplars Project: Extension Program Evaluation Report. University of Minnesota Extension and Natural Resources Research Institute University of Minnesota Duluth. Retrieved from the University of Minnesota Digital Conservancy; Minneapolis, Minnesota, USA. 5 p. <https://hdl.handle.net/11299/241234>
11. Kolka, R., Haight, R.G., Chun, C.L., Berens, M.J., Rogers, E.R., Vinhal, R.A., Zalesny, R.S. Jr., Nislow, K.H., Perry, C.H., & Connolly, S.J. (2022). Mercury Sulfur Initiative: Suggested Program of Research for the Upper Great Lakes States. General Technical Report, NRS-206. Madison, WI: U.S. Department of Agriculture, Forest Service, Northern Research Station. 28 p. <https://doi.org/10.2737/NRS-GTR-206>
12. Lazarus, W.F., Nelson, N.D., Jackson, J., Berguson, W.E., McMahon, B.G., Buchman, D., & Cai, M. (2021). Comparison of Hybrid Poplar Wood Breakeven Prices as Affected by Current and Improved Genetics. Natural Resources Research Institute, University of Minnesota Duluth, Technical Report NRRI/TR-2021/16. Retrieved from the University of Minnesota Digital Conservancy; Duluth, Minnesota, USA. 20 p. <https://hdl.handle.net/11299/219608>
13. McGovern, P.N., Rogers, E.R., Vinhal, R.A., & Zalesny, R.S. Jr. (2023). Leveraging *Populus* Field Trials to Manage Environmental and Human-Induced Change. Technical Paper of the International Union of Forest Research Organizations (IUFRO) Working Party 2.08.04: Poplars and Willows; Vienna, Austria. 27 p. <https://www.iufro.org/science/divisions/division-2/20000/20800/20804/publications/>
14. Mitchell, D., & Jansen, A. (2024). Deploying Purpose-Grown Energy Crops for Sustainable Aviation Fuel Workshop Summary Report. US Department of Energy, Office of Energy Efficiency and Renewable Energy, Technical Report DOE-EE 2808. 41 p. <https://www.energy.gov/eere/bioenergy/articles/deploying-purpose-grown-energy-crops-sustainable-aviation-fuel-workshop>
15. Nelson, N.D., Berguson, W.E., McMahon, B.G., Jackson, J., Buchman, D., DuPlissis, J., & White, T.W. (2021). Intellectual Property in the NRRI Hybrid Poplar Program – Inventory, Commercialization Plan, and Progress Report. Natural Resources Research Institute, University of Minnesota Duluth, Technical Report NRRI/TR-2021/07. Retrieved from the University of Minnesota Digital Conservancy; Minneapolis, Minnesota, USA. 13 p. <https://hdl.handle.net/11299/218837>
16. Nelson, N.D., Buchman, D., Cai, M., McMahon, B.G., & Berguson, W.E. (2022). Inter-Tree Competition Effects in Hybrid Poplar Genotype Testing. Natural Resources Research Institute, University of Minnesota Duluth, Technical Report NRRI/TR-2022/02. Retrieved from the University of Minnesota Digital Conservancy; Minneapolis, Minnesota, USA. 21 p. <https://hdl.handle.net/11299/226513>

17. Nichols, E.G., Hazel, D.W., Ghezehei, S.B., & Bowen, L. (2021). Opportunities and Guide for Short-Rotation Forestry in North Carolina. Online handbook. North Carolina State University; Raleigh, North Carolina, USA. 39 p. <https://sites.google.com/view/shortrotationforestry/home/guide-to-s-r-forestry>
18. Rockwood, D.L., Fabbro, K.W., Ellis, M.F., Ozores-Hampton, M., & Varshovi, A. (2022). Carbon Sequestration by Eucalypts in Florida, USA: Management Options Including Biochar and Associated Economics. In: Carbon Sequestration. Sarvajayakesavalu, S., & Karthikeyan. K. (eds.). IntechOpen. <https://doi.org/10.5772/intechopen.104923>
19. Rockwood, D.L., Ellis, M.F., Liu, R., Zhao, F., Fabbro, K.W., He, Z., & Derbowka, D.R. (2020). Forest Trees for Biochar and Carbon Sequestration: Production and Benefits. In: Applications of Biochar for Environmental Safety. Abdelhafez, A.A., & Abbas, M.H.H. (eds.). IntechOpen. <https://doi.org/10.5772/intechopen.92377>
20. Rogers, E.R., Zalesny, R.S. Jr., Vinhal, R.A., Weger, J., Van Acker, J., Soolanayakanahally, R.Y., Pajel, P., Springgay, E., Zoveda, R., Chauhan, S., Mclvor, I., Paris, P., Singh, H., Thevs, N., & Worms, P. (2023). How Can Fast-Growing Trees Optimize Agroforestry Benefits? The role of the International Commission on Poplars and Other Fast-Growing Trees Sustaining People and the Environment. Technical Brief of the Food and Agriculture Organization (FAO) of the United Nations. Rome, Italy. 6 p. <https://www.fao.org/3/cc8646en/cc8646en.pdf>
21. Zalesny, R.S. Jr., & Pilipović, A. (eds). (2022). Growth and Development of Short Rotation Woody Crops for Rural and Urban Applications. Printed edition of the special issue published in Forests. MDPI Publishers; Basel, Switzerland. ISBN 978-3-0365-4502-8. 390 p. <https://doi.org/10.3390/books978-3-0365-4501-1>
22. Zalesny, R.S. Jr., Casler, M.D., Hallett, R.A., Lin, C-H., & Pilipović, A. (2021). Bioremediation and Soils. Chapter 9, pages 237-273. In: Soils and Landscape Restoration. Stanturf, J.A., & Callahan, M.A. (eds). Elsevier Academic Press. 440 p. <https://doi.org/10.1016/B978-0-12-813193-0.00009-6>
23. Zalesny, R.S. Jr., Headlee, W.L., Soolanayakanahally, R.Y., & Richardson, J. (eds). (2020). Short Rotation Woody Crop Production Systems for Ecosystem Services and Phytotechnologies. Printed edition of the special issue published in Forests. MDPI Publishers; Basel, Switzerland. ISBN 978-3-03921-509-6. 303 p. <https://doi.org/10.3390/books978-3-03921-510-2>
24. Zobrist, K., Haider, N., Stanton, B.J., & Stonex, R. (2023). Growing Hybrid Poplar for Bioenergy in the PNW. Washington State University Extension; Pullman, Washington, USA. 115 p. <https://doi.org/10.7273/000005545>

b) Patents, Trademarks, Commercializations

- ☛ Doty, S. & Sher, A. (21 April 2022). Nitrogen fixing bacteria and an activity-enhancing microbial partner closely associated with wild poplar roots. University of Washington Record of Innovation Number 49511.
- ☛ Doty, S., & Tournay, R. (26 October 2021). Endophytes for improved phytoremediation of arsenic. University of Washington Record of Innovation Number 49368.

- University of Minnesota Duluth, Natural Resources Research Institute (NRRI) (Duluth, Minnesota, USA) commercialized their InnovaTree™ hybrid poplar (*Populus × euramericana*) which was registered by the IPC in 2021. In 2023, they have a US plant patent pending, as well as two commercial tree nurseries that are selling it and several other nurseries evaluating InnovaTree™ for sale.

c) Additional Innovations Not Listed Elsewhere

- United Nations Endorsement of Phyto-Recurrent Selection**

In 2003, USDA Forest Service researchers developed **phyto-recurrent selection** as a methodology to choose favorable tree varieties for phytoremediation and other phytotechnology applications. Since then, USDA Forest Service researchers and their partners have conducted extensive greenhouse, nursery, and field testing to refine and validate the method. As a result of these efforts, phyto-recurrent selection was endorsed as a “Good Practice” by the [UN Decade on Ecosystem Restoration Task Force on Best Practices](#) in 2023. Good Practices are evaluated by the Task Force according to their adaptability, scalability, and broad applicability for implementation in restoration efforts worldwide. Phyto-recurrent selection was determined to meet these criteria, and is now available online within the UN Framework for Ecosystem Restoration Monitoring (FERM) Registry: [Phyto-Recurrent Selection to Enhance Ecosystem Services and Livelihoods in Rural and Urban Communities](#).

Phyto-recurrent selection provides a sustainable solution to many diverse ecological challenges through the greening of degraded lands. As a selection practice, phyto-recurrent selection can be used to select the most appropriate trees for remediating contamination in soils, groundwater, and surface waters. Selected trees can then be used to reforest the contaminated or otherwise degraded lands, which in turn contributes to improving the quality of soils and waters. This technique can also be leveraged to address the effects of climate change through the selection of varieties with enhanced potential for carbon sequestration, climate adaptability, and climate change mitigation. Additionally, superior genotypes with optimal growth can be selected in order to maximize the production of bioenergy and bioproducts. For instance, varieties can be selected for the provisioning of fuelwood, thereby improving grower livelihoods. Finally, native species and genetically appropriate germplasm can serve as the base population for phyto-recurrent selection activities, which will help ensure the sustainability of restoration practices.

For more information about phyto-recurrent selection, please contact: Ron Zalesny Jr. (ronald.zalesny@usda.gov), Liz Rogers (elizabeth.r.rogers@usda.gov), and/or Ryan Vinhal (ryan.vinhal@usda.gov).

- Fast-Growing Trees Networks**

A team at the University of Connecticut, led by Julia Kuzovkina, recently initiated a project to construct a network of researchers, practitioners, and institutions involved with 20 fast-growing tree genera to facilitate information exchange and collaborative projects. The first steps included the analyses of references and scientists who have published their work related to fast-growing species over the past 7 years. The following databases were analyzed: Google Scholar, Academic Search Premiere, BIOSIS Citation Index, Cab Abstracts, EBSCO, Scopus, JSTOR, CRIS. <https://tinyurl.com/yp3f77ym>

For more information about the fast-growing trees networks, please contact: Julia Kuzovkina (jkuzovkina@uconn.edu).

- ✎ Blumenfeld, S., Nichols, E.G., & Ghezehei, S.B. (2023) A productivity and economic assessment tool for poplar production for veneer in Western North Carolina. North Carolina State University.
- ✎ Groover, A. (2011-present). Developed poplar germplasm resource for dissecting complex traits based on gene dosage. USDA Forest Service. Burlington, Vermont, USA.
- ✎ Groover, A. (2014-present). Demonstrated gene copy number variation as a fundamental source of variation contributing to ecological and economically important traits in poplar. USDA Forest Service. Burlington, Vermont, USA.
- ✎ McGovern, P.N. (2023). Produced ten hybrid aspen families using established Open4st project parents. The families were comprised of three F₁ and five F₃ backcrosses with three females, four males and two wind pollinated families.
- ✎ North Carolina State University identified nine *Populus* clones suitable for veneer.
- ✎ North Carolina State University identified three *Populus* clones suitable for veneer and plywood.

d) Poplar, Willow, and Eucalypt Experts in the US

- ✎ Gary Banuelos, USDA Agricultural Research Service, gary.banuelos@ars.usda.gov
- ✎ Amy Brunner, Virginia Technological University, abrunner@vt.edu
- ✎ Joel Burken, Missouri University of Science and Technology, burken@mst.edu
- ✎ Victor Busov, Michigan Technological University, vbusov@mtu.edu
- ✎ Mark Coleman, University of Idaho, mcoleman@uidaho.edu
- ✎ Maggie Davis, Oak Ridge National Laboratory, davismr@ornl.gov
- ✎ Steve DiFazio, West Virginia University, spdifazio@mail.wvu.edu
- ✎ Sharon Doty, University of Washington, sldoty@uw.edu
- ✎ John DuPlissis, University of Minnesota Duluth, jdupliss@d.umn.edu
- ✎ Mark Eisenbies, State University of New York, mheisenb@esf.edu
- ✎ John Freeman, Intrinsyx Environmental, jfreeman@intrinsyx.com
- ✎ Esteban Galeano, Mississippi State University, egg113@msstate.edu
- ✎ Emile Gardiner, USDA Forest Service, emile.gardiner@usda.gov
- ✎ Solomon Ghezehei, North Carolina State University, sbghezeh@ncsu.edu
- ✎ Andrew Groover, USDA Forest Service, andrew.t.groover@usda.gov
- ✎ Richard Gustafson, University of Washington, pulp@uw.edu
- ✎ Elizabeth Guthrie-Nichols, North Carolina State University, egnichol@ncsu.edu

- ✦ Edo Gutman, Intrinsyx Environmental, edo@intrinsyx.com
- ✦ Rich Hallett, USDA Forest Service, richard.hallett@usda.gov
- ✦ Dennis Hazel, North Carolina State University, hazeld@ncsu.edu
- ✦ Austin Himes, Mississippi State University, austin.himes@msstate.edu
- ✦ Gary Hodge, North Carolina State University, grh@ncsu.edu
- ✦ Jud Isebrands, Environmental Forestry Consultants, efcllc@athenet.net
- ✦ Jeff Jackson, University of Minnesota Duluth, JeffJ@d.umn.edu
- ✦ Matias Kirst, University of Florida, mkirst@ufl.edu
- ✦ Julia Kuzovkina, University of Connecticut, jkuzovkina@uconn.edu
- ✦ Jim Landmeyer, US Geological Survey, jlandmey@usgs.gov
- ✦ Matt Langholtz, Oak Ridge National Laboratory, langholtzmh@ornl.gov
- ✦ Bill Lazarus, University of Minnesota, wlazarus@umn.edu
- ✦ Jared LeBoldus, Oregon State University, jared.leboldus@science.oregonstate.edu
- ✦ Lou Licht, Ecolotree, lou-licht@ecolotree.com
- ✦ Chung-Ho Lin, University of Missouri, linchu@missouri.edu
- ✦ Chris Maier, USDA Forest Service, christopher.maier@usda.gov
- ✦ Jessica McCord, University of Tennessee, jfox16@tennessee.edu
- ✦ Pat McGovern, Open4st, pmcgover@gmail.com
- ✦ Dana Mitchell, US Department of Energy, dana.mitchell@ee.doe.gov
- ✦ Renee Murphy, Intrinsyx Environmental, rmurphy@intrinsyx.com
- ✦ Lee Newman, State University of New York, lanewman@esf.edu
- ✦ Galen O'Toole, Intrinsyx Environmental, galen@intrinsyx.com
- ✦ Max Piana, USDA Forest Service, max.piana@usda.gov
- ✦ Andrej Pilipović, University of Minnesota Duluth, pilip015@d.umn.edu
- ✦ John Quinn, Argonne National Laboratory, quinnj@anl.gov
- ✦ Drew Rakers, Sand County Environmental, drew.rakers@sandcountyeniv.com
- ✦ Heidi Renninger, Mississippi State University, heidi.renninger@msstate.edu
- ✦ Tim Rials, University of Tennessee, trials@utk.edu
- ✦ Don Rockwood, Florida FGT LLC, dlock@ufl.edu
- ✦ Levi Rhody, Sand County Environmental, levi.rhody@sandcountyeniv.com

- ✎ Liz Rogers, USDA Forest Service, elizabeth.r.rogers@usda.gov
- ✎ Erik Schilling, National Council for Air and Stream Improvement (NCASI), eschilling@ncasi.org
- ✎ Jamie Schuler, West Virginia University, jamie.schuler@mail.wvu.edu
- ✎ Bart Sexton, Sand County Environmental, bart.sexton@sandcountyeniv.com
- ✎ Courtney Siegert, Mississippi State University, courtney.siegert@msstate.edu
- ✎ Reid Simmer, University of Iowa, reid-simmer@uiowa.edu
- ✎ Larry Smart, Cornell University, lbs33@cornell.edu
- ✎ Nancy Sonti, USDA Forest Service, nancy.f.sonti@usda.gov
- ✎ Brian Stanton, Poplar Innovations, brian.stanton@poplarinnovations.com
- ✎ Rick Stonex, Poplar Innovations, rick.stonex@poplarinnovations.com
- ✎ Steve Strauss, Oregon State University, Steve.strauss@oregonstate.edu
- ✎ Tim Tschaplinski, Oak Ridge National Laboratory, tschaplinstj@ornl.gov
- ✎ Jerry Tuskan, Oak Ridge National Laboratory, tuskanga@ornl.gov
- ✎ Ranjith Udawatta, University of Missouri, UdawattaR@missouri.edu
- ✎ Ryan Vinhal, USDA Forest Service, ryan.vinhal@usda.gov
- ✎ Tim Volk, State University of New York, tavolk@esf.edu
- ✎ Hairong Wei, Michigan Technological University, hairong@mtu.edu
- ✎ Ron Zalesny Jr., USDA Forest Service, ronald.zalesny@usda.gov
- ✎ JY Zhu, USDA Forest Service, junyong.zhu@usda.gov
- ✎ Kevin Zobrist, Washington State University, kevin.zobrist@wsu.edu
- ✎ Colleen Zumpf, Argonne National Laboratory, czumpf@anl.gov

e) Organizations and Programs Relevant to Poplars, Willows, and Eucalypts in the US

- ✎ **International Commission on Poplars and Other Fast-Growing Trees Sustaining People and the Environment**, <https://www.fao.org/ipc/en/>
- ✎ Advanced Hardwood Biofuels Northwest, <https://hardwoodbiofuels.org/>
- ✎ Argonne National Laboratory, Environmental Science Division, <https://www.anl.gov/evs/departments>
- ✎ American Farmland Trust, <https://farmland.org/>
- ✎ Association for Temperate Agroforestry, <https://www.aftaweb.org/>
- ✎ Beyond Trees Network, <https://www.beyondtrees.global/>
- ✎ Cornell University, Willowpedia, <https://willow.cals.cornell.edu/>

- ✎ Ecolotree, <https://www.ecolotree.com/>
- ✎ eXtension Farm Energy, <https://farm-energy.extension.org/>
- ✎ Florida FGT, LLC, <https://www.floridafgt.com/>
- ✎ Green Trees, <https://www.green-trees.com/>
- ✎ IEA Bioenergy Task 43: Biomass, <https://task43.ieabioenergy.com/>
- ✎ International Phytotechnology Society, <https://phytosociety.org/>
- ✎ Intrinsyx Environmental, <https://www.intrinsyxenvironmental.com/>
- ✎ IUFRO 1.03.00 Short Rotation Forestry, <https://www.iufro.org/science/divisions/division-1/10000/10300/>
- ✎ IUFRO 1.06.01 Phytotechnologies, <https://www.iufro.org/science/divisions/division-1/10000/10600/10601/>
- ✎ IUFRO 2.08.04 Poplars and Willows, <https://www.iufro.org/science/divisions/division-2/20000/20800/20804/>
- ✎ Leachate Management Specialists, <https://www.leachate.us/>
- ✎ Michigan State University, Forest Biomass Innovation Center, <https://www.canr.msu.edu/fbic>
- ✎ North Carolina State University, Camcore, <https://camcore.cnr.ncsu.edu/>
- ✎ North Carolina State University, Forest Productivity Cooperative, <https://www.forestproductivity.org/>
- ✎ Oak Ridge National Laboratory, Center for Bioenergy Innovation (CBI), <https://cbi.ornl.gov/>
- ✎ Oak Ridge National Laboratory, National Transportation Research Center, <https://www.ornl.gov/facility/ntrc/research-areas/>
- ✎ Oak Ridge National Laboratory, Plant Microbe Interfaces (PMI), <https://pmiweb.ornl.gov/>
- ✎ Poplar and Willow Council of Canada, <http://www.poplar.ca/>
- ✎ Sand County Environmental, <https://www.sandcountyenvironment.com/>
- ✎ Savanna Institute, <https://www.savannainstitute.org/>
- ✎ Short Rotation Woody Crops Operations Working Group, <https://woodycrops.tennessee.edu/>
- ✎ UN Decade on Ecosystem Restoration, Best Practices Task Force, <https://www.decadeonrestoration.org/task-forces/best-practice>
- ✎ University of Minnesota, CINRAM, <https://cinram.umn.edu/>
- ✎ University of Minnesota Duluth, Natural Resources Research Institute, <https://nrri.umn.edu/>
- ✎ University of Missouri, Center for Agroforestry, <https://centerforagroforestry.org/>
- ✎ University of Tennessee, Center for Renewable Carbon, <https://crc.tennessee.edu/>
- ✎ US DOE, Bioenergy Technologies Office, <https://www.energy.gov/eere/bioenergy/bioenergy-technologies-office>
- ✎ US DOE, Joint BioEnergy Institute, <https://www.jbei.org/>
- ✎ US DOE, National Renewable Energy Laboratory, <https://www.nrel.gov/bioenergy/index.html>
- ✎ USDA Forest Service, Forest Products Laboratory, <https://www.fs.usda.gov/research/fpl>

- ✎ USDA Forest Service, International Programs, <https://www.fs.usda.gov/about-agency/international-programs/about-us>
- ✎ USDA Forest Service, National Agroforestry Center, <https://www.fs.usda.gov/nac/>
- ✎ USDA Forest Service, Northern Research Station, <https://www.fs.usda.gov/research/nrs>
- ✎ USDA Forest Service, Pacific Northwest Research Station, <https://www.fs.usda.gov/research/pnw>
- ✎ USDA Forest Service, Pacific Southwest Research Station, <https://www.fs.usda.gov/research/psw>
- ✎ USDA Forest Service, Research and Development, <https://www.fs.usda.gov/research/about>
- ✎ USDA Forest Service, Rocky Mountain Research Station, <https://www.fs.usda.gov/research/rmrs>
- ✎ USDA Forest Service, Southern Research Station, <https://www.fs.usda.gov/research/srs>
- ✎ USDA Forest Service, Urban and Community Forestry, <https://www.fs.usda.gov/managing-land/urban-forests/ucf>
- ✎ Willow Biomass, State University of New York, <https://www.esf.edu/willow/>

4. Literature

In addition to receiving papers from experts, a comprehensive literature review was performed to identify records related to the cultivation and utilization of poplars, willows, eucalypts, and other fast-growing trees (a.k.a., short rotation woody crops, SRWC) in the United States from 2020-2023. Following the methods of Rogers et al. (2023), the literature review was conducted using the Web of Science database (<https://clarivate.com/webofsciencelibrary/solutions/web-of-science/>). Search logic included:

Populus:

(TS=(*Populus* OR poplar)) AND CU=(United States OR USA OR United States of America)

Salix:

(TS=(*Salix* OR willow)) AND CU=(United States OR USA OR United States of America)

Eucalyptus:

(TS=(*Eucalyptus* OR eucalypt)) AND CU=(United States OR USA OR United States of America)

SRWCs:

(TS=((SRWC AND rotation) OR (SRC AND rotation) OR (FGTs AND trees) OR ("short rotation" AND trees) OR "short rotation woody crops" OR "short rotation coppice" OR "fast growing trees")) AND CU=(United States OR USA OR United States of America)

where: TS = searches for topic terms in the following fields within a record: Title, Abstract, Author, Keywords, Keywords Plus®; and CU = searches for countries/regions in the Addresses field within a record. The timespan (i.e., publication date) for all searches ranged from 1 January 2020 to 31 December 2023. Only peer-reviewed manuscripts are included below. Notable publications from sources other than peer-reviewed journals are listed above [Section III.3.a)] Notable Publications from Sources Outside Peer-Reviewed Journals].

In total, 2,561 records were screened, with 199 papers received from experts (*Populus* = 156; *Salix* = 27; *Eucalyptus* = 9; SRWCs = 7) and 2,362 manuscripts identified from Web of Science (*Populus* = 1,174; *Salix* = 477; *Eucalyptus* = 618; SRWC = 93) (Figure 1). Each record was reviewed based on title, abstract, and content, and 1,679 records were excluded for lack of relevancy and/or redundancy among

sources/searches. Therefore, there are 882 total unique records. Of these, 59 records are cross-listed among *Populus*, *Salix*, *Eucalyptus*, and/or SRWC, resulting in a total of 941 records reported below. Of those, 66% are for *Populus* (n = 621), followed by 20% for *Eucalyptus* (n = 185), 12% for *Salix* (n = 110), and 2% for SRWCs (n = 25) (Figure 1). The distribution of the five areas of work summarized in Section II above [Technical Information. 1) Taxonomy, Nomenclature, and Registration; 2) Domestication, Breeding, and Selection of Genetic Resources; 3) Plant Health, Resilience to Threats and Climate Change, 4) Production Systems for the Bioeconomy; 5) Environmental and Ecosystem Services] are illustrated in Figure 2. Across genera and SRWCs, the number of records was greatest for Production Systems for the Bioeconomy and least for Taxonomy, Nomenclature, and Registration. The number of records among the three remaining areas of work are dependent on specific category (i.e., *Populus*, *Salix*, *Eucalyptus*, SRWC) (Figure 2).

Rogers, E. R., Zalesny, R. S., Lin, C.-H., & Vinhal, R. A. (2023). Intrinsic and extrinsic factors influencing *Populus* water use: A literature review. *Journal of Environmental Management*, 348, 119180. <https://doi.org/10.1016/j.jenvman.2023.119180>

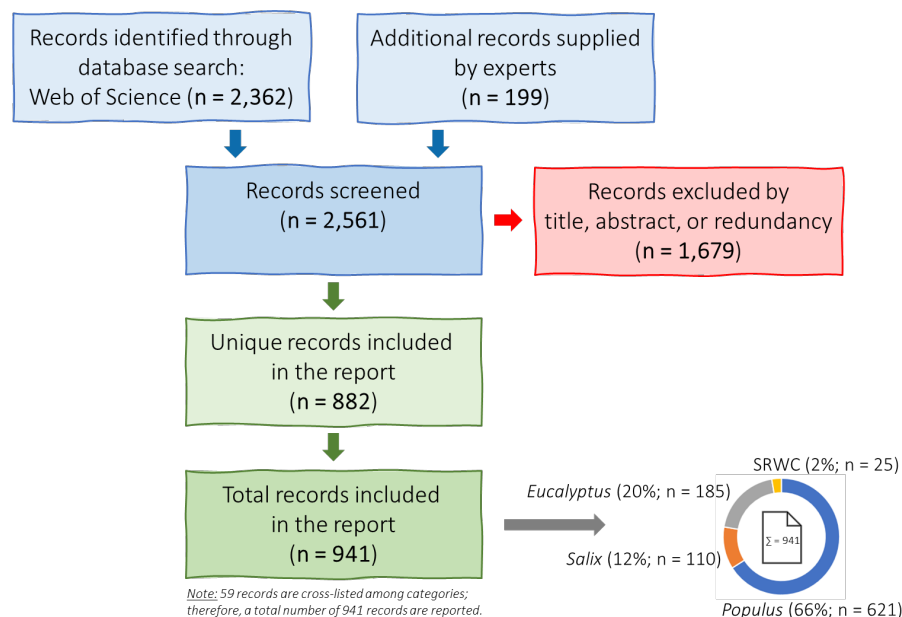


Figure 1. Flow chart of literature review process for selection of peer-reviewed manuscripts related to the cultivation and utilization of poplars, willows, eucalypts, and other fast-growing trees (a.k.a., short rotation woody crops, SRWCs) in the United States from 2020-2023. Methods were adapted from Rogers et al. (2023).

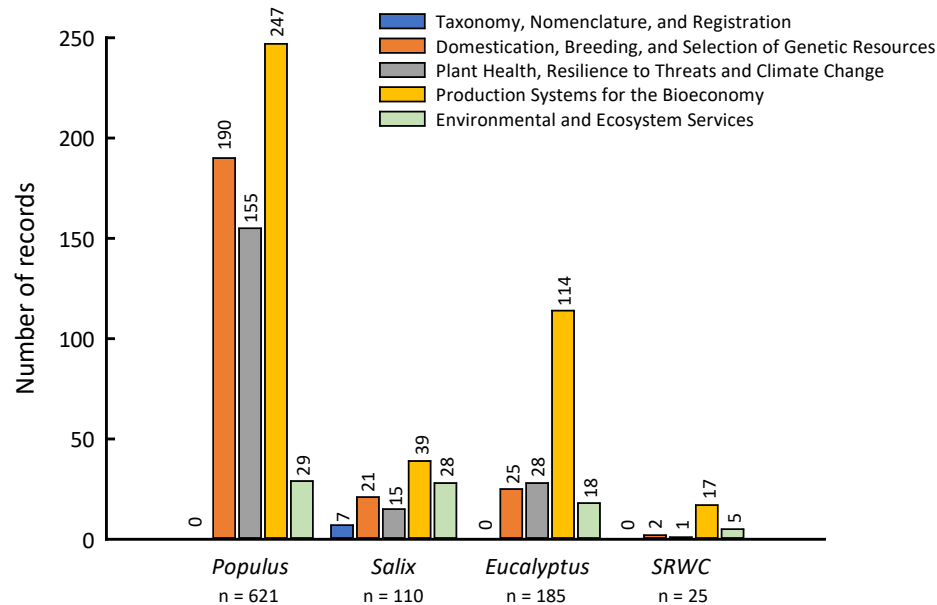


Figure 2. Number of records for each of five areas of work for *Populus*, *Salix*, *Eucalyptus*, and SRWC resulting from a literature review of peer-reviewed manuscripts related to the cultivation and utilization of poplars, willows, eucalypts, and other fast-growing trees (a.k.a., short rotation woody crops, SRWCs) in the United States from 2020-2023.

Populus

Taxonomy, Nomenclature, and Registration

None reported.

Domestication, Breeding, and Selection of Genetic Resources

1. Abbas, M., Peszlen, I., Shi, R., Kim, H., Katahira, R., Kafle, K., Xiang, Z., Huang, X., Min, D., Mohamadamin, M., Yang, C., Dai, X., Yan, X., Park, S., Li, Y., Kim, S. H., Davis, M., Ralph, J., Sederoff, R. R., ... Li, Q. (2020). Involvement of *Cesa4*, *Cesa7-A/B* and *Cesa8-A/B* in secondary wall formation in *Populus trichocarpa* wood. *Tree Physiology*, 40(1), 73–89. <https://doi.org/10.1093/treephys/tpz020>
2. Abeyratne, C. R., Macaya-Sanz, D., Zhou, R., Barry, K. W., Daum, C., Haiby, K., Lipzen, A., Stanton, B., Yoshinaga, Y., Zane, M., Tuskan, G. A., & DiFazio, S. P. (2023). High-resolution mapping reveals hotspots and sex-biased recombination in *Populus trichocarpa*. *G3*, 13(1), jkac269. <https://doi.org/10.1093/g3journal/jkac269>
3. Abraham, P. E., Matthiadis, A., Hettich, R. L., & Kalluri, U. C. (2020). Molecular remodeling in *Populus PdkOR* RNAi roots profiled using LC-MS/MS proteomics. *Proteomics*, 20(24), 2000067. <https://doi.org/10.1002/pmic.202000067>

4. Ahlawat, Y. K., Nookaraju, A., Harman-Ware, A. E., Doeppke, C., Biswal, A. K., & Joshi, C. P. (2021). Genetic modification of *KNAT7* transcription factor expression enhances saccharification and reduces recalcitrance of woody biomass in poplars. *Frontiers in Plant Science*, 12, 762067. <https://doi.org/10.3389/fpls.2021.762067>
5. Amdoun, R., Sahli, F., Hamadache, K., Ouzzane, A.-H., Khelifi-Slaoui, M., Moustafa, K., Hefferon, K., Makhzoum, A., & Khelifi, L. (2020). Optimization of caulogenesis in *Populus nigra* under lead (Pb) stress via response surface methodology (RSM) and desirability function analysis. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 142(1), Article 1. <https://doi.org/10.1007/s11240-020-01827-0>
6. An, X., Gao, K., Chen, Z., Li, J., Yang, X., Yang, X., Zhou, J., Guo, T., Zhao, T., Huang, S., Miao, D., Ullah Khan, W., Rao, P., Ye, M., Lei, B., Liao, W., Wang, J., Ji, L., Li, Y., ... Strauss, S. H. (2022). High quality haplotype-resolved genome assemblies of *Populus tomentosa* Carr., a stabilized interspecific hybrid species widespread in Asia. *Molecular Ecology Resources*, 22(2), 786–802. <https://doi.org/10.1111/1755-0998.13507>
7. An, X., Gao, K., Chen, Z., Li, J., Yang, X., Yang, X., Zhou, J., Guo, T., Zhao, T., Huang, S., Miao, D., Khan, W. U., Rao, P., Ye, M., Lei, B., Liao, W., Wang, J., Ji, L., Li, Y., ... Strauss, S. H. (2020). *Hybrid origin of Populus tomentosa* Carr. Identified through genome sequencing and phylogenomic analysis. <https://doi.org/10.1101/2020.04.07.030692>
8. Appidi, M. R., Bible, A. N., Carper, D. L., Jawdy, S. S., Giannone, R. J., Hettich, R. L., Morrell-Falvey, J., & Abraham, P. E. (2022). Development of an experimental approach to achieve spatially resolved plant root-associated metaproteomics using an agar-plate system. *Molecular Plant-Microbe Interactions*®, 35(8), 639–649. <https://doi.org/10.1094/MPMI-01-22-0011-TA>
9. Azeez, A., & Busov, V. (2021). CRISPR/Cas9-mediated single and biallelic knockout of poplar *STERILE APETALA (PopSAP)* leads to complete reproductive sterility. *Plant Biotechnology Journal*, 19(1), 23–25. <https://doi.org/10.1111/pbi.13451>
10. Azeez, A., Zhao, Y. C., Singh, R. K., Yordanov, Y. S., Dash, M., Miskolczi, P., Stojkovič, K., Strauss, S. H., Bhalerao, R. P., & Busov, V. B. (2021). *EARLY BUD-BREAK 1* and *EARLY BUD-BREAK 3* control resumption of poplar growth after winter dormancy. *Nature Communications*, 12(1), 1123. <https://doi.org/10.1038/s41467-021-21449-0>
11. Azizpor, P., Sullivan, L., Lim, A., & Groover, A. (2022). Facile labeling of sieve element phloem-protein bodies using the reciprocal oligosaccharide probe OGA488. *Frontiers in Plant Science*, 13, 809923. <https://doi.org/10.3389/fpls.2022.809923>
12. Bagley, J. C., Heming, N. M., Gutiérrez, E. E., Devisetty, U. K., Mock, K. E., Eckert, A. J., & Strauss, S. H. (2020). Genotyping-by-sequencing and ecological niche modeling illuminate phylogeography, admixture, and pleistocene range dynamics in quaking aspen (*Populus tremuloides*). *Ecology and Evolution*, 10(11), 4609–4629. <https://doi.org/10.1002/ece3.6214>
13. Bai, X., Wang, W., Ji, J., Jiang, J., Yu, Q., Yang, C., & Liu, G. (2023). *PabCIPK24* plays an important role in the response of hybrid poplar to salt stress. *Industrial Crops and Products*, 205, 117452. <https://doi.org/10.1016/j.indcrop.2023.117452>
14. Bai, X., Ji, J., Wang, W., Gu, C., Yu, Q., Jiang, J., Yang, C., & Liu, G. (2022). Characterization of CBL-interacting protein kinases' gene family and expression pattern reveal their important

roles in response to salt stress in poplar. *Forests*, 13(9), 1353.

<https://doi.org/10.3390/f13091353>

15. Balasubramanian, V. K., Rivas-Ubach, A., Winkler, T., Mitchell, H., Moran, J., & Ahkami, A. H. (2023). Modulation of polar auxin transport identifies the molecular determinants of source–sink carbon relationships and sink strength in poplar. *Tree Physiology*, tpad073. <https://doi.org/10.1093/treephys/tpad073>
16. Barge, E. G., Leopold, D. R., Rojas, A., Vilgalys, R., & Busby, P. E. (2022). Phylogenetic conservatism of mycoparasitism and its contribution to pathogen antagonism. *Molecular Ecology*, 31(10), 3018–3030. <https://doi.org/10.1111/mec.16436>
17. Bastiaanse, H., Henry, I. M., Tsai, H., Lieberman, M., Canning, C., Comai, L., & Groover, A. (2021). A systems genetics approach to deciphering the effect of dosage variation on leaf morphology in *Populus*. *The Plant Cell*, 33(4), 940–960. <https://doi.org/10.1093/plcell/koaa016>
18. Beamish, A. (2022). A much-abused tree: The rise and fall of the Lombardy poplar. *Studies in the History of Gardens & Designed Landscapes*, 42(2), 120–137. <https://doi.org/10.1080/14601176.2022.2059287>
19. Bewg, W. P., Harding, S. A., Engle, N. L., Vaidya, B. N., Zhou, R., Reeves, J., Horn, T. W., Joshee, N., Jenkins, J. W., Shu, S., Barry, K. W., Yoshinaga, Y., Grimwood, J., Schmitz, R. J., Schmutz, J., Tschaplinski, T. J., & Tsai, C.-J. (2022). Multiplex knockout of trichome-regulating MYB duplicates in hybrid poplar using a single gRNA. *Plant Physiology*, 189(2), 516–526. <https://doi.org/10.1093/plphys/kiac128>
20. Bible, A. N., Chang, M., & Morrell-Falvey, J. L. (2021). Identification of a diguanylate cyclase expressed in the presence of plants and its application for discovering candidate gene products involved in plant colonization by *Pantoea* sp. YR343. *PLOS ONE*, 16(7), e0248607. <https://doi.org/10.1371/journal.pone.0248607>
21. Brooks, E. G., Elorriaga, E., Liu, Y., Duduit, J. R., Yuan, G., Tsai, C.-J., Tuskan, G. A., Ranney, T. G., Yang, X., & Liu, W. (2023). Plant promoters and terminators for high-precision bioengineering. *BioDesign Research*, 5, 0013. <https://doi.org/10.34133/bdr.0013>
22. Brunner, A. M., Beers, E. P., Sheng, X., Petzold, H. E., & Teixeira, R. (2021). From *Populus* molecular networks to transgenic field phenotypes. *In Vitro Cellular & Developmental Biology - Animal*, 57(S1), 20–31. <https://doi.org/10.1007/s11626-021-00576-4>
23. Bryant, N., Zhang, J., Feng, K., Shu, M., Ployet, R., Chen, J.-G., Muchero, W., Yoo, C. G., Tschaplinski, T. J., Pu, Y., & Ragauskas, A. J. (2023). Novel candidate genes for lignin structure identified through genome-wide association study of naturally varying *Populus trichocarpa*. *Frontiers in Plant Science*, 14, 1153113. <https://doi.org/10.3389/fpls.2023.1153113>
24. Bryant, N. D., Pu, Y., Tschaplinski, T. J., Tuskan, G. A., Muchero, W., Kalluri, U. C., Yoo, C. G., & Ragauskas, A. J. (2020). Transgenic poplar designed for biofuels. *Trends in Plant Science*, 25(9), 881–896. <https://doi.org/10.1016/j.tplants.2020.03.008>
25. Çelik, E. N. Y., Baloglu, M. C., & Ayan, S. (2021). Gene expression profiles of Hsp family members in different poplar taxa under cadmium stress. *Turkish Journal of Agriculture and Forestry*, 45(1). <https://doi.org/10.3906/tar-2003-83>

26. Chang, Y., Xu, X., Zheng, H., Xie, H., Li, B., Chen, S., Li, Y., & Dai, S. (2023). Genome-wide identification and characterization of ACBP gene family in *Populus* reveal salinity alkali-responsive profiles. *Journal of Forestry Research*, 34(2), 481–496. <https://doi.org/10.1007/s11676-022-01485-2>
27. Chao, N., Qi, Q., Li, S., Ruan, B., Jiang, X., & Gai, Y. (2021). Characterization and functional analysis of the hydroxycinnamoyl-CoA: Shikimate hydroxycinnamoyl transferase (HTC) gene family in poplar. *PeerJ*, 9, e10741. <https://doi.org/10.7717/peerj.10741>
28. Chen, S., Yu, Y., Wang, X., Wang, S., Zhang, T., Zhou, Y., He, R., Meng, N., Wang, Y., Liu, W., Liu, Z., Liu, J., Guo, Q., Huang, H., Sederoff, R. R., Wang, G., Qu, G., & Chen, S. (2023). Chromosome-level genome assembly of a triploid poplar *Populus alba* 'Berolinensis'. *Molecular Ecology Resources*, 23(5), 1092–1107. <https://doi.org/10.1111/1755-0998.13770>
29. Chen, Y.-H., Sharma, S., Bewg, W. P., Xue, L.-J., Giselbach, C. R., & Tsai, C.-J. (2023). Multiplex editing of the *Nucleoredoxin1* tandem array in poplar: From small indels to translocations and complex inversions. *The CRISPR Journal*, 6(4), 339–349. <https://doi.org/10.1089/crispr.2022.0096>
30. Chhetri, H. B., Furches, A., Macaya-Sanz, D., Walker, A. R., Kainer, D., Jones, P., Harman-Ware, A. E., Tschaplinski, T. J., Jacobson, D., Tuskan, G. A., & DiFazio, S. P. (2020). Genome-wide association study of wood anatomical and morphological traits in populus trichocarpa. *Frontiers in Plant Science*, 11, 545748. <https://doi.org/10.3389/fpls.2020.545748>
31. Chiatante, D., Montagnoli, A., Trupiano, D., Sferra, G., Bryant, J., Rost, T. L., & Scippa, G. S. (2021). Meristematic connectome: A cellular coordinator of plant responses to environmental signals? *Cells*, 10(10), 2544. <https://doi.org/10.3390/cells10102544>
32. Cole, C. T., Morrow, C. J., Barker, H. L., Rubert-Nason, K. F., Riehl, J. F. L., Köllner, T. G., Lackus, N. D., & Lindroth, R. L. (2021). Growing up aspen: Ontogeny and trade-offs shape growth, defence and reproduction in a foundation species. *Annals of Botany*, 127(4), 505–517. <https://doi.org/10.1093/aob/mcaa070>
33. Conde, D., Triozzi, P. M., Balmant, K. M., Doty, A. L., Miranda, M., Boullosa, A., Schmidt, H. W., Pereira, W. J., Dervinis, C., & Kirst, M. (2021). A robust method of nuclei isolation for single-cell RNA sequencing of solid tissues from the plant genus *Populus*. *PLOS ONE*, 16(5), e0251149. <https://doi.org/10.1371/journal.pone.0251149>
34. Cope, O. L., Keefover-Ring, K., Kruger, E. L., & Lindroth, R. L. (2021). Growth–defense trade-offs shape population genetic composition in an iconic forest tree species. *Proceedings of the National Academy of Sciences*, 118(37), e2103162118. <https://doi.org/10.1073/pnas.2103162118>
35. Cope, O. L., Lindroth, R. L., Helm, A., Keefover-Ring, K., & Kruger, E. L. (2021). Trait plasticity and trade-offs shape intra-specific variation in competitive response in a foundation tree species. *New Phytologist*, 230(2), 710–719. <https://doi.org/10.1111/nph.17166>
36. Daguerre, Y., Basso, V., Hartmann-Wittulski, S., Schellenberger, R., Meyer, L., Bailly, J., Kohler, A., Plett, J. M., Martin, F., & Veneault-Fourrey, C. (2020). The mutualism effector MiSSP7 of *Laccaria bicolor* alters the interactions between the poplar JAZ6 protein and its associated proteins. *Scientific Reports*, 10(1), 20362. <https://doi.org/10.1038/s41598-020-76832-6>

37. Dahal, S., Hurst, G., Chourey, K., Engle, N., Burdick, L., Morrell-Falvey, J., Tschaplinski, T., Doktycz, M., & Pelletier, D. (2023). Mechanism for utilization of the *Populus*-Derived Metabolite salicin by a *Pseudomonas*—*Rahnella* Co-Culture. *Metabolites*, 13(2), 140. <https://doi.org/10.3390/metabo13020140>
38. Dai, H., Wei, S., & Noori, A. (2020). The mechanism of chelator improved the tolerance and accumulation of poplar to Cd explored through differential expression protein based on iTRAQ. *Journal of Hazardous Materials*, 393, 122370. <https://doi.org/10.1016/j.jhazmat.2020.122370>
39. Dai, X., Zhai, R., Lin, J., Wang, Z., Meng, D., Li, M., Mao, Y., Gao, B., Ma, H., Zhang, B., Sun, Y., Li, S., Zhou, C., Lin, Y.-C. J., Wang, J. P., Chiang, V. L., & Li, W. (2023). Cell-type-specific PtrWOX4a and PtrVCS2 form a regulatory nexus with a histone modification system for stem cambium development in *Populus trichocarpa*. *Nature Plants*, 9(1), 96–111. <https://doi.org/10.1038/s41477-022-01315-7>
40. De Vries, L., MacKay, H. A., Smith, R. A., Mottiar, Y., Karlen, S. D., Unda, F., Muirragui, E., Bingman, C., Vander Meulen, K., Beebe, E. T., Fox, B. G., Ralph, J., & Mansfield, S. D. (2022). PHBMT1, a BAHD-family monolignol acyltransferase, mediates lignin acylation in poplar. *Plant Physiology*, 188(2), 1014–1027. <https://doi.org/10.1093/plphys/kiab546>
41. De Vries, L., Brouckaert, M., Chanoca, A., Kim, H., Regner, M. R., Timokhin, V. I., Sun, Y., De Meester, B., Van Doorselaere, J., Goeminne, G., Chiang, V. L., Wang, J. P., Ralph, J., Morreel, K., Vanholme, R., & Boerjan, W. (2021). CRISPR-Cas9 editing of CAFFEYOYL SHIKIMATE ESTERASE 1 and 2 shows their importance and partial redundancy in lignification in *Populus tremula* × *P. alba*. *Plant Biotechnology Journal*, 19(11), 2221–2234. <https://doi.org/10.1111/pbi.13651>
42. Ding, C., & Brouard, J. S. (2022). Assisted migration is plausible for a boreal tree species under climate change: A quantitative and population genetics study of trembling aspen (*Populus tremuloides* Michx.) in western Canada. *Ecology and Evolution*, 12(10), e9384. <https://doi.org/10.1002/ece3.9384>
43. Feng, G., Sanderson, B. J., Keefover-Ring, K., Liu, J., Ma, T., Yin, T., Smart, L. B., DiFazio, S. P., & Olson, M. S. (2020). Pathways to sex determination in plants: How many roads lead to Rome? *Current Opinion in Plant Biology*, 54, 61–68. (cross-listed with *Salix*) <https://doi.org/10.1016/j.pbi.2020.01.004>
44. Fitzpatrick, M. C., Chhatre, V. E., Soolanayakanahally, R. Y., & Keller, S. R. (2021). Experimental support for genomic prediction of climate maladaptation using the machine learning approach gradient forests. *Molecular Ecology Resources*, 21(8), 2749–2765. <https://doi.org/10.1111/1755-0998.13374>
45. Fu, Y., Li, F., Mu, S., Jiang, L., Ye, M., & Wu, R. (2021). Heterophylly quantitative trait loci respond to salt stress in the desert tree *Populus euphratica*. *Frontiers in Plant Science*, 12, 692494. <https://doi.org/10.3389/fpls.2021.692494>
46. Gao, Y., Liu, X., Jin, Y., Wu, J., Li, S., Li, Y., Chen, B., Zhang, Y., Wei, L., Li, W., Li, R., Lin, C., Reddy, A. S. N., Jaiswal, P., & Gu, L. (2022). Drought induces epitranscriptome and proteome changes in stem-differentiating xylem of *Populus trichocarpa*. *Plant Physiology*, 190(1), 459–479. <https://doi.org/10.1093/plphys/kiac272>

47. Gao, Y., Liu, X., Wu, B., Wang, H., Xi, F., Kohnen, M. V., Reddy, A. S. N., & Gu, L. (2021). Quantitative profiling of N6-methyladenosine at single-base resolution in stem-differentiating xylem of *Populus trichocarpa* using nanopore direct RNA sequencing. *Genome Biology*, 22(1), 22. <https://doi.org/10.1186/s13059-020-02241-7>
48. Goralogia, G. S., Howe, G. T., Brunner, A. M., Helliwell, E., Nagle, M. F., Ma, C., Lu, H., Goddard, A. L., Magnuson, A. C., Klocko, A. L., & Strauss, S. H. (2021). Overexpression of *SHORT VEGETATIVE PHASE-LIKE (SVL)* in *Populus* delays onset and reduces abundance of flowering in field-grown trees. *Horticulture Research*, 8(1), 167. <https://doi.org/10.1038/s41438-021-00600-4>
49. Gougherty, A. V., Chhatre, V. E., Keller, S. R., & Fitzpatrick, M. C. (2020). Contemporary range position predicts the range-wide pattern of genetic diversity in balsam poplar (*Populus balsamifera* L.). *Journal of Biogeography*, 47(6), 1246–1257. <https://doi.org/10.1111/jbi.13811>
50. Han, Y., Lou, X., Zhang, W., Xu, T., & Tang, M. (2022). Arbuscular mycorrhizal fungi enhanced drought resistance of *Populus cathayana* by regulating the 14-3-3 family protein genes. *Microbiology Spectrum*, 10(3), e02456-21. <https://doi.org/10.1128/spectrum.02456-21>
51. Harding, S. A., Tuma, T. T., Aulakh, K., Ortega, M. A., Ci, D., Ou, Y., & Tsai, C.-J. (2022). Tonoplast sucrose trafficking modulates starch utilization and water deficit behavior in Poplar leaves. *Plant and Cell Physiology*, 63(8), 1117–1129. <https://doi.org/10.1093/pcp/pcac087>
52. Harding, S. A., Frost, C. J., & Tsai, C. (2020). Defoliation-induced compensatory transpiration is compromised in *SUT4*-RNAi *Populus*. *Plant Direct*, 4(9), e00268. <https://doi.org/10.1002/pld3.268>
53. Harman-Ware, A. E., Martin, M. Z., Engle, N. L., Doeppke, C., & Tschaplinski, T. J. (2023). Rapid screening of secondary aromatic metabolites in *Populus trichocarpa* leaves. *Biotechnology for Biofuels and Bioproducts*, 16(1), 41. <https://doi.org/10.1186/s13068-023-02287-2>
54. Harman-Ware, A. E., Happs, R. M., Macaya-Sanz, D., Doeppke, C., Muchero, W., & DiFazio, S. P. (2022). Abundance of major cell wall components in natural variants and pedigrees of *Populus trichocarpa*. *Frontiers in Plant Science*, 13, 757810. <https://doi.org/10.3389/fpls.2022.757810>
55. Harman-Ware, A. E., Macaya-Sanz, D., Abeyratne, C. R., Doeppke, C., Haiby, K., Tuskan, G. A., Stanton, B., DiFazio, S. P., & Davis, M. F. (2021). Accurate determination of genotypic variance of cell wall characteristics of a *Populus trichocarpa* pedigree using high-throughput pyrolysis-molecular beam mass spectrometry. *Biotechnology for Biofuels*, 14(1), 59. <https://doi.org/10.1186/s13068-021-01908-y>
56. Hassan, M. M., Martin, S., Feng, K., Yates, T. B., Yuan, G., Martin, M. Z., Martin, S., Muchero, W., Griffiths, N. A., Weston, D. J., & Yang, X. (2023). Genome-wide identification and functional prediction of silicon (Si) transporters in poplar (*Populus trichocarpa*). *Plant Biotechnology Reports*, 17(2), 285–302. <https://doi.org/10.1007/s11816-022-00788-4>
57. Hofmeister, B. T., Denkena, J., Colomé-Tatché, M., Shahryary, Y., Hazarika, R., Grimwood, J., Mamidi, S., Jenkins, J., Grabowski, P. P., Sreedasyam, A., Shu, S., Barry, K., Lail, K., Adam, C., Lipzen, A., Sorek, R., Kudrna, D., Talag, J., Wing, R., ... Schmitz, R. J. (2020). A genome assembly and the somatic genetic and epigenetic mutation rate in a wild long-lived perennial *Populus trichocarpa*. *Genome Biology*, 21(1), 259. <https://doi.org/10.1186/s13059-020-02162-5>

58. Hong, J., Gunasekara, C., He, C., Liu, S., Huang, J., & Wei, H. (2021). Identification of biological pathway and process regulators using sparse partial least squares and triple-gene mutual interaction. *Scientific Reports*, 11(1), 13174. <https://doi.org/10.1038/s41598-021-92610-4>
59. Hu, J., Su, H., Cao, H., Wei, H., Fu, X., Jiang, X., Song, Q., He, X., Xu, C., & Luo, K. (2022). AUXIN RESPONSE FACTOR7 integrates gibberellin and auxin signaling via interactions between DELLA and AUX/IAA proteins to regulate cambial activity in poplar. *The Plant Cell*, 34(7), 2688–2707. <https://doi.org/10.1093/plcell/koac107>
60. Hu, R., Zhang, J., Jawdy, S., Sreedasyam, A., Lipzen, A., Wang, M., Ng, V., Daum, C., Keymanesh, K., Liu, D., Lu, H., Ranjan, P., Chen, J.-G., Muchero, W., Tschaplinski, T. J., Tuskan, G. A., Schmutz, J., & Yang, X. (2022). Comparative genomics analysis of drought response between obligate CAM and C3 photosynthesis plants. *Journal of Plant Physiology*, 277, 153791. <https://doi.org/10.1016/j.jplph.2022.153791>
61. Hu, X.-L., Lu, H., Hassan, M. M., Zhang, J., Yuan, G., Abraham, P. E., Shrestha, H. K., Solis, M. I. V., Chen, J.-G., Tschaplinski, T. J., Doktycz, M. J., Tuskan, G. A., Cheng, Z.-M., & Yang, X. (2021). Advances and perspectives in discovery and functional analysis of small secreted proteins in plants. *Horticulture Research*, 8(1), 130. <https://doi.org/10.1038/s41438-021-00570-7>
62. Jia, H., Liu, G., Li, J., Zhang, J., Sun, P., Zhao, S., Zhou, X., Lu, M., & Hu, J. (2020). Genome resequencing reveals demographic history and genetic architecture of seed salinity tolerance in *Populus euphratica*. *Journal of Experimental Botany*, 71(14), 4308–4320. <https://doi.org/10.1093/jxb/eraa172>
63. John, C., Muchero, W., & Emrich, S. (2020). Extreme phenotype sampling improves LASSO and random forest marker selection for complex traits. *2020 IEEE International Conference on Bioinformatics and Biomedicine (BIBM)*, 2771–2778. <https://doi.org/10.1109/BIBM49941.2020.9313524>
64. Kaya, Z., Taşkıran, B., Uluğ, A., Güvendiren, A. D., Wegrzyn, J. L., & Neale, D. B. (2023). Association genetics of wood traits in European black poplar (*Populus nigra* L.). *Tree Genetics & Genomes*, 19(4), 38. <https://doi.org/10.1007/s11295-023-01612-y>
65. Kim, M.-H., Cho, J.-S., Park, E.-J., Lee, H., Choi, Y.-I., Bae, E.-K., Han, K.-H., & Ko, J.-H. (2020). Overexpression of a poplar RING-H2 zinc finger, *Ptxerico*, confers enhanced drought tolerance via reduced water loss and ion leakage in *Populus*. *International Journal of Molecular Sciences*, 21(24), 9454. <https://doi.org/10.3390/ijms21249454>
66. Klocko, A. L., Elorriaga, E., Ma, C., & Strauss, S. H. (2023). Variation in floral form of CRISPR knock-outs of the poplar homologs of *LEAFY* and *AGAMOUS* after FT heat-induced early flowering. *Horticulture Research*, 10(8), Article 8. <https://doi.org/10.1093/hr/uhad132>
67. Klocko, A. L., Goddard, A. L., Jacobson, J. R., Magnuson, A. C., & Strauss, S. H. (2021). RNAi suppression of *LEAFY* gives stable floral sterility, and reduced growth rate and leaf size, in field-grown poplars. *Plants*, 10(8), 1594. <https://doi.org/10.3390/plants10081594>
68. Kuchma, O., Janz, D., Leinemann, L., Polle, A., Krutovsky, K., & Gailing, O. (2020). Hybrid and environmental effects on gene expression in poplar clones in pure and mixed with black locust stands. *Forests*, 11(10), 1075. <https://doi.org/10.3390/f11101075>

69. Lagergren, J., Pavicic, M., Chhetri, H. B., York, L. M., Hyatt, D., Kainer, D., Rutter, E. M., Flores, K., Bailey-Bale, J., Klein, M., Taylor, G., Jacobson, D., & Streich, J. (2023). Few-shot learning enables population-scale analysis of leaf traits in *Populus trichocarpa*. *Plant Phenomics*, 5, 0072. <https://doi.org/10.34133/plantphenomics.0072>
70. Lawrence, E. H., Springer, C. J., Helliker, B. R., & Poethig, R. S. (2022). The carbon economics of vegetative phase change. *Plant, Cell & Environment*, 45(4), 1286–1297. <https://doi.org/10.1111/pce.14281>
71. Lawrence, E. H., Leichty, A. R., Doody, E. E., Ma, C., Strauss, S. H., & Poethig, R. S. (2021). Vegetative phase change in *Populus tremula* × *alba*. *New Phytologist*, 231(1), 351–364. <https://doi.org/10.1111/nph.17316>
72. Lawrence, E. H., Springer, C. J., Helliker, B. R., & Poethig, R. S. (2021). MicroRNA156-mediated changes in leaf composition lead to altered photosynthetic traits during vegetative phase change. *New Phytologist*, 231(3), 1008–1022. <https://doi.org/10.1111/nph.17007>
73. Li, H., Dai, X., Huang, X., Xu, M., Wang, Q., Yan, X., Sederoff, R. R., & Li, Q. (2021). Single-cell RNA sequencing reveals a high-resolution cell atlas of xylem in *Populus*. *Journal of Integrative Plant Biology*, 63(11), 1906–1921. <https://doi.org/10.1111/jipb.13159>
74. Li, M., Dong, H., Li, J., Dai, X., Lin, J., Li, S., Zhou, C., Chiang, V. L., & Li, W. (2023). *PtrVCS2* regulates drought resistance by changing vessel morphology and stomatal closure in *Populus trichocarpa*. *International Journal of Molecular Sciences*, 24(5), 4458. <https://doi.org/10.3390/ijms24054458>
75. Li, S., Zhang, Y., Xin, X., Ding, C., Lv, F., Mo, W., Xia, Y., Wang, S., Cai, J., Sun, L., Du, M., Dong, C., Gao, X., Dai, X., Zhang, J., & Sun, J. (2020). The osmotin-like protein gene *PdOLP1* is involved in secondary cell wall biosynthesis during wood formation in poplar. *International Journal of Molecular Sciences*, 21(11), 3993. <https://doi.org/10.3390/ijms21113993>
76. Li, Y., Bako, C. M., Saktrakulka, P., Lehmler, H.-J., Hornbuckle, K. C., & Schnoor, J. L. (2021). Interconversion between methoxylated, hydroxylated and sulfated metabolites of PCB 3 in whole poplar plants. *Science of The Total Environment*, 785, 147341. <https://doi.org/10.1016/j.scitotenv.2021.147341>
77. Lin, C.-Y., Sun, Y., Song, J., Chen, H.-C., Shi, R., Yang, C., Liu, J., Tunlaya-Anukit, S., Liu, B., Loziuk, P. L., Williams, C. M., Muddiman, D. C., Lin, Y.-C. J., Sederoff, R. R., Wang, J. P., & Chiang, V. L. (2021). Enzyme complexes of Ptr4CL and PtrHCT modulate Co-enzyme A ligation of Hydroxycinnamic acids for Monolignol Biosynthesis in *Populus trichocarpa*. *Frontiers in Plant Science*, 12, 727932. <https://doi.org/10.3389/fpls.2021.727932>
78. Liu, B., Sanchez, M., Truong, J., Ford, P. C., & Abu-Omar, M. M. (2022). Catalytic conversion of high S-lignin to a sustainable tri-epoxide polymer precursor. *Green Chemistry*, 24(12), 4958–4968. <https://doi.org/10.1039/D2GC01278B>
79. Liu, B., Liu, J., Yu, J., Wang, Z., Sun, Y., Li, S., Lin, Y.-C. J., Chiang, V. L., Li, W., & Wang, J. P. (2021). Transcriptional reprogramming of xylem cell wall biosynthesis in tension wood. *Plant Physiology*, 186(1), 250–269. <https://doi.org/10.1093/plphys/kiab038>
80. Liu, C., Wang, S., Liu, Y., Wang, M., Fan, E., Liu, C., Zhang, S., Yang, C., Wang, J., Sederoff, H. W., You, X., Chiang, V. L., Chen, S., Sederoff, R. R., & Qu, G. (2023). Exceptionally high genetic

variance of the doubled haploid (DH) population of poplar. *Journal of Forestry Research*, 34(6), 1941–1950. <https://doi.org/10.1007/s11676-023-01612-7>

81. Liu, C., Li, K., Wang, M., Fan, E., Yang, C., Wang, J., Fu, P., Ge, X., Sederoff, H. W., Sederoff, R. R., Chiang, V. L., Wang, S., & Qu, G. (2021). Qu-2, a robust poplar suspension cell line for molecular biology. *Journal of Forestry Research*, 32(2), 733–740. <https://doi.org/10.1007/s11676-020-01266-9>
82. Liu, D., Tang, D., Xie, M., Zhang, J., Zhai, L., Mao, J., Luo, C., Lipzen, A., Zhang, Y., Savage, E., Yuan, G., Guo, H.-B., Tadesse, D., Hu, R., Jawdy, S., Cheng, H., Li, L., Yer, H., Clark, M. M., ... Yang, X. (2023). *Agave REVEILLE1* regulates the onset and release of seasonal dormancy in *Populus*. *Plant Physiology*, 191(3), 1492–1504. <https://doi.org/10.1093/plphys/kiac588>
83. Liu, H., Gao, J., Sun, J., Li, S., Zhang, B., Wang, Z., Zhou, C., Sulis, D. B., Wang, J. P., Chiang, V. L., & Li, W. (2022). Dimerization of PtrMYB074 and PtrWRKY19 mediates transcriptional activation of *Ptrbhlh186* for secondary xylem development in *Populus trichocarpa*. *New Phytologist*, 234(3), 918–933. <https://doi.org/10.1111/nph.18028>
84. Liu, X., Gao, Y., Liao, J., Miao, M., Chen, K., Xi, F., Wei, W., Wang, H., Wang, Y., Xu, X., Reddy, A. S. N., & Gu, L. (2021). Genome-wide profiling of circular RNAs, alternative splicing, and R-loops in stem-differentiating xylem of *Populus trichocarpa*. *Journal of Integrative Plant Biology*, 63(7), 1294–1308. <https://doi.org/10.1111/jipb.13081>
85. Liu, Y., Tao, H., Wang, Y., Fang, Y., Xiang, Y., Liu, G., Guo, Y., Liu, J., Yin, Y., Cai, Y., & Jiang, G. (2021). Gaseous elemental mercury [Hg(0)] oxidation in poplar leaves through a Two-Step single-electron transfer process. *Environmental Science & Technology Letters*, 8(12), 1098–1103. <https://doi.org/10.1021/acs.estlett.1c00735>
86. Liu, Y., Wang, R., Yu, J., Huang, S., Zhang, Y., Wei, H., & Wei, Z. (2023). Genome-wide identification and characterization of auxin response factor (ARF) gene family involved in wood formation and response to exogenous hormone treatment in *Populus trichocarpa*. *International Journal of Molecular Sciences*, 24(1), 740. <https://doi.org/10.3390/ijms24010740>
87. Lu, H., Gordon, M. I., Amarasinghe, V., & Strauss, S. H. (2020). Extensive transcriptome changes during seasonal leaf senescence in field-grown black cottonwood (*Populus trichocarpa* Nisqually-1). *Scientific Reports*, 10(1), 6581. <https://doi.org/10.1038/s41598-020-63372-2>
88. Luo, S., Coutinho, B. G., Dadhwal, P., Oda, Y., Ren, J., Schaefer, A. L., Greenberg, E. P., Harwood, C. S., & Tong, L. (2021). Structural basis for a bacterial pip system plant effector recognition protein. *Proceedings of the National Academy of Sciences*, 118(10), e2019462118. <https://doi.org/10.1073/pnas.2019462118>
89. Ma, C., Goddard, A., Peremyslova, E., Duan, C., Jiang, Y., Nagle, M., & Strauss, S. H. (2022). Factors affecting in vitro regeneration in the model tree *Populus trichocarpa* I. Medium, environment, and hormone controls on organogenesis. *In Vitro Cellular & Developmental Biology - Plant*, 58(6), 837–852. <https://doi.org/10.1007/s11627-022-10301-9>
90. Ma, C., Duan, C., Jiang, Y., Nagle, M., Peremyslova, E., Goddard, A., & Strauss, S. H. (2022). Factors affecting in vitro regeneration in the model tree *Populus trichocarpa*: II. Heritability estimates, correlations among explant types, and genetic interactions with treatments among

wild genotypes. *In Vitro Cellular & Developmental Biology - Plant*, 58(6), 853–864.
<https://doi.org/10.1007/s11627-022-10302-8>

91. Madina, M. H., Rahman, M. S., Huang, X., Zhang, Y., Zheng, H., & Germain, H. (2020). A poplar rust effector protein associates with protein disulfide isomerase and enhances plant susceptibility. *Biology*, 9(9), 294. <https://doi.org/10.3390/biology9090294>
92. Mahon, E. L., de Vries, L., Jang, S.-K., Middar, S., Kim, H., Unda, F., Ralph, J., & Mansfield, S. D. (2022). Exogenous chalcone synthase expression in developing poplar xylem incorporates naringenin into lignins. *Plant Physiology*, 188(2), 984–996.
<https://doi.org/10.1093/plphys/kiab499>
93. Matthews, M. L., Wang, J. P., Sederoff, R., Chiang, V. L., & Williams, C. M. (2021). A multiscale model of lignin biosynthesis for predicting bioenergy traits in *Populus trichocarpa*. *Computational and Structural Biotechnology Journal*, 19, 168–182.
<https://doi.org/10.1016/j.csbj.2020.11.046>
94. Matthews, M. L., Wang, J. P., Sederoff, R., Chiang, V. L., & Williams, C. M. (2020). Modeling cross-regulatory influences on monolignol transcripts and proteins under single and combinatorial gene knockdowns in *Populus trichocarpa*. *PLOS Computational Biology*, 16(4), e1007197. <https://doi.org/10.1371/journal.pcbi.1007197>
95. Mottiar, Y., Karlen, S. D., Goacher, R. E., Ralph, J., & Mansfield, S. D. (2023). Metabolic engineering of *p*-hydroxybenzoate in poplar lignin. *Plant Biotechnology Journal*, 21(1), 176–188. <https://doi.org/10.1111/pbi.13935>
96. Movahedi, A., Wei, H., Kadkhodaei, S., Sun, W., Zhuge, Q., Yang, L., & Xu, C. (2023). CRISPR-mediated genome editing in poplar issued by efficient transformation. *Frontiers in Plant Science*, 14, 1159615. <https://doi.org/10.3389/fpls.2023.1159615>
97. Movahedi, A., Wei, H., Zhou, X., Fountain, J. C., Chen, Z.-H., Mu, Z., Sun, W., Zhang, J., Li, D., Guo, B., Varshney, R. K., Yang, L., & Zhuge, Q. (2022). Precise exogenous insertion and sequence replacements in poplar by simultaneous HDR overexpression and NHEJ suppression using CRISPR-Cas9. *Horticulture Research*, 9, uhac154. <https://doi.org/10.1093/hr/uhac154>
98. Nagle, M. F., Yuan, J., Kaur, D., Ma, C., Peremyslova, E., Jiang, Y., Zahl, B., Niño de Rivera, A., Muchero, W., Fuxin, L., & Strauss, S. H. (2023). GWAS identifies candidate genes controlling adventitious rooting in *Populus trichocarpa*. *Horticulture Research*, 10(8), uhad125.
<https://doi.org/10.1093/hr/uhad125>
99. Nagle, M. F., Yuan, J., Kaur, D., Ma, C., Peremyslova, E., Jiang, Y., De Rivera, A. N., Jawdy, S., Chen, J.-G., Feng, K., Yates, T. B., Tuskan, G. A., Muchero, W., Fuxin, L., & Strauss, S. H. (2022). GWAS identifies candidate regulators of in planta regeneration in *Populus trichocarpa*. <https://doi.org/10.1101/2022.06.08.495082>
100. Nagle, M., Ma, C., Peremyslova, E., & Strauss, S. (2020). Transformation improvement in wild black cottonwood (*Populus trichocarpa*): Effects of developmental regulators, surfactants, and other factors. *In Vitro Cellular & Developmental Biology-Animal*, 56.
101. Nelson, N. D., Berguson, W. E., McMahon, B. G., Cai, M., & Buchman, D. J. (2021). Vectors of efficiency in hybrid poplar genotype testing. *Silvae Genetica*, 70(1), 39–56.
<https://doi.org/10.2478/sg-2021-0004>

102. Noble, J. D., Balmant, K. M., Dervinis, C., De Los Campos, G., Resende, M. F. R., Kirst, M., & Barbazuk, W. B. (2020). The genetic regulation of alternative splicing in *Populus deltoides*. *Frontiers in Plant Science*, 11, 590. <https://doi.org/10.3389/fpls.2020.00590>
103. Ortega, M. A., Zhou, R., Chen, M. S. S., Bewg, W. P., Simon, B., & Tsai, C. (2023). *In vitro* floral development in poplar: Insights into seed trichome regulation and trimonoecy. *New Phytologist*, 237(4), 1078–1081. <https://doi.org/10.1111/nph.18624>
104. Qiao, Z., Yates, T. B., Shrestha, H. K., Engle, N. L., Flanagan, A., Morrell-Falvey, J. L., Sun, Y., Tschaplinski, T. J., Abraham, P. E., Labbé, J., Wang, Z., Hettich, R. L., Tuskan, G. A., Muchero, W., & Chen, J. (2021). Towards engineering ectomycorrhization into switchgrass bioenergy crops via a lectin receptor-like kinase. *Plant Biotechnology Journal*, 19(12), 2454–2468. <https://doi.org/10.1111/pbi.13671>
105. Payyavula, R. S., Badmi, R., Jawdy, S. S., Rodriguez, M., Gunter, L., Sykes, R. W., Winkeler, K. A., Collins, C. M., Rottmann, W. H., Chen, J., Yang, X., Tuskan, G. A., & Kalluri, U. C. (2022). Biomass formation and sugar release efficiency of *Populus* modified by altered expression of a NAC transcription factor. *Plant Direct*, 6(8), e419. <https://doi.org/10.1002/pld3.419>
106. Pelletier, D. A., Burdick, L. H., Podar, M., Schadt, C. W., & Kalluri, U. C. (2020). Draft genome sequence of *Larkinella* sp. Strain BK230, isolated from *Populus deltoides* roots. *Microbiology Resource Announcements*, 9(12), e00159-20. <https://doi.org/10.1128/MRA.00159-20>
107. Peng, X., Tong, B., Lee, J., Wang, K., Yu, X., Huang, X., Wen, J., Makarem, M., Pang, H., Hinjan, S., Yan, X., Yao, S., Lu, F., Wang, B., Peng, F., Ralph, J., Kim, S. H., Sederoff, R. R., & Li, Q. (2023). Overexpression of a gibberellin 20-oxidase gene in poplar xylem led to an increase in the size of nanocellulose fibrils and improved paper properties. *Carbohydrate Polymers*, 314, 120959. <https://doi.org/10.1016/j.carbpol.2023.120959>
108. Podar, M., Turner, J., Burdick, L. H., & Pelletier, D. A. (2020a). Complete genome sequence of *Starkeya* sp. Strain ORNL1, a soil alphaproteobacterium isolated from the rhizosphere of *Populus deltoides*. *Microbiology Resource Announcements*, 9(27), e00644-20. <https://doi.org/10.1128/MRA.00644-20>
109. Podar, M., Turner, J., Burdick, L. H., & Pelletier, D. A. (2020b). Complete genome sequence of the novel *Roseimicrobium* sp. Strain ORNL1, a verrucomicrobium isolated from the *Populus deltoides* rhizosphere. *Microbiology Resource Announcements*, 9(27), e00617-20. <https://doi.org/10.1128/MRA.00617-20>
110. Reeves, P. A., Tetreault, H. M., & Richards, C. M. (2020). Bioinformatic extraction of functional genetic diversity from heterogeneous germplasm collections for crop improvement. *Agronomy*, 10(4), 593. <https://doi.org/10.3390/agronomy10040593>
111. Ren, M., Zhang, Y., Wang, R., Liu, Y., Li, M., Wang, X., Chen, X., Luan, X., Zhang, H., Wei, H., Yang, C., & Wei, Z. (2022). *PtrHAT22*, as a higher hierarchy regulator, coordinately regulates secondary cell wall component biosynthesis in *Populus trichocarpa*. *Plant Science*, 316, 111170. <https://doi.org/10.1016/j.plantsci.2021.111170>
112. Ren, M., Zhang, Y., Liu, C., Liu, Y., Tian, S., Cheng, H., Zhang, H., Wei, H., & Wei, Z. (2021). Characterization of a high hierarchical regulator, *PtrGATA12*, functioning in differentially

- regulating secondary wall component biosynthesis in *Populus trichocarpa*. *Frontiers in Plant Science*, 12, 657787. <https://doi.org/10.3389/fpls.2021.657787>
113. Rho, H., Van Epps, V., Kim, S.-H., & Doty, S. L. (2020). Endophytes increased fruit quality with higher soluble sugar production in honeycrisp apple (*Malus pumila*). *Microorganisms*, 8(5), 699. <https://doi.org/10.3390/microorganisms8050699>
 114. Rodriguez-Zaccaro, F. D., Henry, I. M., & Groover, A. (2021). Genetic regulation of vessel morphology in *Populus*. *Frontiers in Plant Science*, 12, 705596. <https://doi.org/10.3389/fpls.2021.705596>
 115. Saint-Vincent, P. M. B., Furches, A., Galanie, S., Teixeira Prates, E., Aldridge, J. L., Labbe, A., Zhao, N., Martin, M. Z., Ranjan, P., Jones, P., Kainer, D., Kalluri, U. C., Chen, J.-G., Muchero, W., Jacobson, D. A., & Tschaplinski, T. J. (2023). Validation of a metabolite–GWAS network for *Populus trichocarpa* family 1 UDP-glycosyltransferases. *Frontiers in Plant Science*, 14, 1210146. <https://doi.org/10.3389/fpls.2023.1210146>
 116. Saint-Vincent, P. M., Ridout, M., Engle, N. L., Lawrence, T. J., Yearly, M. L., Tschaplinski, T. J., Newcombe, G., & Pelletier, D. A. (2020). Isolation, characterization, and pathogenicity of two *Pseudomonas syringae* pathovars from *Populus trichocarpa* seeds. *Microorganisms*, 8(8), 1137. <https://doi.org/10.3390/microorganisms8081137>
 117. Sanderson, B. J., Gambhir, D., Feng, G., Hu, N., Cronk, Q. C., Percy, D. M., Frenner, F. M., Johnson, M. G., Smart, L. B., Keefover-Ring, K., Yin, T., Ma, T., DiFazio, S. P., Liu, J., & Olson, M. S. (2023). Phylogenomics reveals patterns of ancient hybridization and differential diversification that contribute to phylogenetic conflict in willows, poplars, and close relatives. *Systematic Biology*, 72(6), 1220–1232. (cross-listed with *Salix*) <https://doi.org/10.1093/sysbio/syad042>
 118. Sanderson, B. J., DiFazio, S. P., Cronk, Q. C. B., Ma, T., & Olson, M. S. (2020). A targeted sequence capture array for phylogenetics and population genomics in the Salicaceae. *Applications in Plant Sciences*, 8(10), e11394. (cross-listed with *Salix*) <https://doi.org/10.1002/aps3.11394>
 119. Sankey, T., Hultine, K., Blasini, D., Koepke, D., Bransky, N., Grady, K., Cooper, H., Gehring, C., & Allan, G. (2021). UAV thermal image detects genetic trait differences among populations and genotypes of fremont cottonwood (*Populus fremontii* , salicaceae). *Remote Sensing in Ecology and Conservation*, 7(2), 245–258. <https://doi.org/10.1002/rse2.185>
 120. Sarkar, D., & Maranas, C. D. (2020). SNPeffect: Identifying functional roles of SNPs using metabolic networks. *The Plant Journal*, 103(2), 512–531. <https://doi.org/10.1111/tpj.14746>
 121. Shen, C., Li, Q., An, Y., Zhou, Y., Zhang, Y., He, F., Chen, L., Liu, C., Mao, W., Wang, X., Liang, H., Yin, W., & Xia, X. (2022). The transcription factor GNC optimizes nitrogen use efficiency and growth by up-regulating the expression of nitrate uptake and assimilation genes in poplar. *Journal of Experimental Botany*, 73(14), 4778–4792. <https://doi.org/10.1093/jxb/erac190>
 122. Shrestha, H. K., Solis, M. I. V., Jawdy, S. S., Tuskan, G. A., Yang, X., & Abraham, P. E. (2021). Temporal dynamics of protein and post-translational modification abundances in *Populus* leaf across a diurnal period. *PROTEOMICS*, 21(20), 2100127. <https://doi.org/10.1002/pmic.202100127>

123. Sow, M. D., Le Gac, A., Fichot, R., Lanciano, S., Delaunay, A., Le Jan, I., Lesage-Descauses, M., Citerne, S., Caius, J., Brunaud, V., Soubigou-Taconnat, L., Cochard, H., Segura, V., Chaparro, C., Grunau, C., Daviaud, C., Tost, J., Brignolas, F., Strauss, S. H., ... Maury, S. (2021). RNAi suppression of DNA methylation affects the drought stress response and genome integrity in transgenic poplar. *New Phytologist*, 232(1), 80–97. <https://doi.org/10.1111/nph.17555>
124. Sreedasyam, A., Plott, C., Hossain, M. S., Lovell, J. T., Grimwood, J., Jenkins, J. W., Daum, C., Barry, K., Carlson, J., Shu, S., Phillips, J., Amirebrahimi, M., Zane, M., Wang, M., Goodstein, D., Haas, F. B., Hiss, M., Perroud, P.-F., Jawdy, S. S., ... Schmutz, J. (2023). JGI plant gene atlas: An updateable transcriptome resource to improve functional gene descriptions across the plant kingdom. *Nucleic Acids Research*, 51(16), 8383–8401. <https://doi.org/10.1093/nar/gkad616>
125. Sun, P., Jia, H., Cheng, X., Zhang, Y., Li, J., Zhang, L., Lu, M., Zhang, J., & Hu, J. (2020). Genetic architecture of leaf morphological and physiological traits in a *Populus deltoides* ‘Danhong’ × *P. simonii* ‘Tongliao1’ pedigree revealed by quantitative trait locus analysis. *Tree Genetics & Genomes*, 16(3), 45. <https://doi.org/10.1007/s11295-020-01438-y>
126. Sun, P., Jia, H., Yue, Z., Li, J., Li, J., Zhang, J., Lu, M., & Hu, J. (2020). Genetic identification of 91 poplar cultivars based on SSR markers. *Nordic Journal of Botany*, 38(6), njb.02504. <https://doi.org/10.1111/njb.02504>
127. Sun, Y., Qiao, Z., Muchero, W., & Chen, J.-G. (2020). Lectin receptor-like kinases: The sensor and mediator at the plant cell surface. *Frontiers in Plant Science*, 11, 596301. (cross-listed with *Eucalyptus*) <https://doi.org/10.3389/fpls.2020.596301>
128. Tang, X., Wang, C., Chai, G., Wang, D., Xu, H., Liu, Y., He, G., Liu, S., Zhang, Y., Kong, Y., Li, S., Lu, M., Sederoff, R. R., Li, Q., & Zhou, G. (2022). Ubiquitinated DA1 negatively regulates vascular cambium activity through modulating the stability of WOX4 in *Populus*. *The Plant Cell*, 34(9), 3364–3382. <https://doi.org/10.1093/plcell/koac178>
129. Tao, Y., Chiu, L.-W., Hoyle, J. W., Dewhurst, R. A., Richey, C., Rasmussen, K., Du, J., Mellor, P., Kuiper, J., Tucker, D., Crites, A., Orr, G. A., Heckert, M. J., Godinez-Vidal, D., Orozco-Cardenas, M. L., & Hall, M. E. (2023). Enhanced photosynthetic efficiency for increased carbon assimilation and woody biomass production in engineered hybrid poplar. *Forests*, 14(4), 827. <https://doi.org/10.3390/f14040827>
130. Triozzi, P. M., Schmidt, H. W., Dervinis, C., Kirst, M., & Conde, D. (2021). Simple, efficient and open-source CRISPR/Cas9 strategy for multi-site genome editing in *Populus tremula* × *alba*. *Tree Physiology*, 41(11), 2216–2227. <https://doi.org/10.1093/treephys/tpab066>
131. Tsai, C.-J., Xu, P., Xue, L.-J., Hu, H., Nyamdari, B., Naran, R., Zhou, X., Goeminne, G., Gao, R., Gjersing, E., Dahlen, J., Pattathil, S., Hahn, M. G., Davis, M. F., Ralph, J., Boerjan, W., & Harding, S. A. (2020). Compensatory guaiacyl lignin biosynthesis at the expense of syringyl lignin in 4CL1-knockout poplar. *Plant Physiology*, 183(1), 123–136. <https://doi.org/10.1104/pp.19.01550>
132. Umana, G. E., Perez, J. M., Unda, F., Lin, C.-Y., Sener, C., Karlen, S. D., Mansfield, S. D., Eudes, A., Ralph, J., Donohue, T. J., & Noguera, D. R. (2022). Biological funneling of phenolics from transgenic plants engineered to express the bacterial 3-dehydroshikimate dehydratase (*qsuB*) gene. *Frontiers in Chemical Engineering*, 4, 1036084. <https://doi.org/10.3389/fceng.2022.1036084>

133. Unda, F., Mottiar, Y., Mahon, E. L., Karlen, S. D., Kim, K. H., Loqué, D., Eudes, A., Ralph, J., & Mansfield, S. D. (2022). A new approach to zip-lignin: 3,4-dihydroxybenzoate is compatible with lignification. *New Phytologist*, 235(1), 234–246. <https://doi.org/10.1111/nph.18136>
134. Vidyagina, E. O., Subbotina, N. M., Belyi, V. A., Lebedev, V. G., Krutovsky, K. V., & Shestibratov, K. A. (2020). Various effects of the expression of the xyloglucanase gene from *Penicillium canescens* in transgenic aspen under semi-natural conditions. *BMC Plant Biology*, 20(1), 251. <https://doi.org/10.1186/s12870-020-02469-2>
135. Wang, H., Chen, J.-G., & Chang, Y. (2021). Identification, expression, and interaction analysis of ovate family proteins in *Populus trichocarpa* reveals a role of PtOFP1 regulating drought stress response. *Frontiers in Plant Science*, 12, 650109. <https://doi.org/10.3389/fpls.2021.650109>
136. Wang, H., Liu, S., Dai, X., Yang, Y., Luo, Y., Gao, Y., Liu, X., Wei, W., Wang, H., Xu, X., Reddy, A. S. N., Jaiswal, P., Li, W., Liu, B., & Gu, L. (2021). PSDX: A comprehensive multi-omics association database of *Populus trichocarpa* with a focus on the secondary growth in response to stresses. *Frontiers in Plant Science*, 12, 655565. <https://doi.org/10.3389/fpls.2021.655565>
137. Wang, H., Pak, S., Yang, J., Wu, Y., Li, W., Feng, H., Yang, J., Wei, H., & Li, C. (2022). Two high hierarchical regulators, PuMYB40 and PuWRKY75, control the low phosphorus driven adventitious root formation in *Populus ussuriensis*. *Plant Biotechnology Journal*, 20(8), 1561–1577. <https://doi.org/10.1111/pbi.13833>
138. Wang, M., Zhang, L., Zhang, Z., Li, M., Wang, D., Zhang, X., Xi, Z., Keefover-Ring, K., Smart, L. B., DiFazio, S. P., Olson, M. S., Yin, T., Liu, J., & Ma, T. (2020). Phylogenomics of the genus *Populus* reveals extensive interspecific gene flow and balancing selection. *New Phytologist*, 225(3), 1370–1382. <https://doi.org/10.1111/nph.16215>
139. Wang, Q., Dai, X., Pang, H., Cheng, Y., Huang, X., Li, H., Yan, X., Lu, F., Wei, H., Sederoff, R. R., & Li, Q. (2021). BEL1-like homeodomain protein BLH6a is a negative regulator of *Cald5H2* in sinapyl alcohol monolignol biosynthesis in poplar. *Frontiers in Plant Science*, 12, 695223. <https://doi.org/10.3389/fpls.2021.695223>
140. Wang, R., Reng, M., Tian, S., Liu, C., Cheng, H., Liu, Y., Zhang, H., Saqib, M., Wei, H., & Wei, Z. (2021). Transcriptome-wide identification and characterization of micrnas in diverse phases of wood formation in *Populus trichocarpa*. *G3 Genes/Genomes/Genetics*, 11(8), jkab195. <https://doi.org/10.1093/g3journal/jkab195>
141. Wang, Y., Yang, Y., Wang, F., Wang, G., Wang, C., Wang, W., Chen, K., Gu, C., Yu, Q., & Jiang, J. (2021). Growth adaptability and foreign gene stability of TaLEA transgenic *Populus simonii* × *nigra*. *Annals of Forest Science*, 78(2), Article 2. <https://doi.org/10.1007/s13595-021-01038-3>
142. Wang, Z., Mao, Y., Guo, Y., Gao, J., Liu, X., Li, S., Lin, Y.-C. J., Chen, H., Wang, J. P., Chiang, V. L., & Li, W. (2020). MYB transcription factor161 mediates feedback regulation of *secondary wall-associated NAC-domain1* family genes for wood formation. *Plant Physiology*, 184(3), 1389–1406. <https://doi.org/10.1104/pp.20.01033>
143. Wei, M., Zhang, M., Sun, J., Zhao, Y., Pak, S., Ma, M., Chen, Y., Lu, H., Yang, J., Wei, H., Li, Y., & Li, C. (2023). PuHox52 promotes coordinated uptake of nitrate, phosphate, and iron under nitrogen deficiency in *Populus ussuriensis*. *Journal of Integrative Plant Biology*, 65(3), 791–809. <https://doi.org/10.1111/jipb.13389>

144. Wei, M., Liu, Q., Wang, Z., Yang, J., Li, W., Chen, Y., Lu, H., Nie, J., Liu, B., Lv, K., Mao, X., Chen, S., Sanders, J., Wei, H., & Li, C. (2020). PuHox52-mediated hierarchical multilayered gene regulatory network promotes adventitious root formation in *Populus ussuriensis*. *New Phytologist*, 228(4), 1369–1385. <https://doi.org/10.1111/nph.16778>
145. Wu, W., Li, J., Wang, Q., Lv, K., Du, K., Zhang, W., Li, Q., Kang, X., & Wei, H. (2021). Growth-regulating factor 5 (GRF5)-mediated gene regulatory network promotes leaf growth and expansion in poplar. *New Phytologist*, 230(2), Article 2. <https://doi.org/10.1111/nph.17179>
146. Wu, W., Liao, T., Du, K., Wei, H., & Kang, X. (2021). Transcriptome comparison of different ploidy reveals the mechanism of photosynthetic efficiency superiority of triploid poplar. *Genomics*, 113(4), 2211–2220. <https://doi.org/10.1016/j.ygeno.2021.05.009>
147. Xiao, Y., Poovaiah, C., Unda, F., Ritchie, L., Dombrov, M., Phalen, C., Argyros, A., & Coleman, H. (2020). Expression of the *Trichoderma reesei* expansin-like protein, swollenin, in poplar results in biomass with improved sugar release by enzymatic hydrolysis. *Biomass and Bioenergy*, 134, 105473. <https://doi.org/10.1016/j.biombioe.2020.105473>
148. Xie, M., Zhang, J., Singan, V. R., McGranahan, M. J., LaFayette, P. R., Jawdy, S. S., Engle, N., Doeppke, C., Tschaplinski, T. J., Davis, M. F., Lindquist, E., Barry, K., Schmutz, J., Parrott, W. A., Chen, F., Tuskan, G. A., Chen, J., & Muchero, W. (2020). Identification of functional single nucleotide polymorphism of *Populus trichocarpa* *PtrEPSP-TF* and determination of its transcriptional effect. *Plant Direct*, 4(1), e00178. <https://doi.org/10.1002/pld3.178>
149. Xue, L., Wu, H., Chen, Y., Li, X., Hou, J., Lu, J., Wei, S., Dai, X., Olson, M. S., Liu, J., Wang, M., Charlesworth, D., & Yin, T. (2020). Evidences for a role of two Y-specific genes in sex determination in *Populus deltoides*. *Nature Communications*, 11(1), 5893. <https://doi.org/10.1038/s41467-020-19559-2>
150. Yang, X., Liu, D., Lu, H., Weston, D. J., Chen, J.-G., Muchero, W., Martin, S., Liu, Y., Hassan, M. M., Yuan, G., Kalluri, U. C., Tschaplinski, T. J., Mitchell, J. C., Wulschleger, S. D., & Tuskan, G. A. (2021). Biological parts for plant biodesign to enhance land-based carbon dioxide removal. *BioDesign Research*, 2021, 9798714. <https://doi.org/10.34133/2021/9798714>
151. Yang, X., Medford, J. I., Markel, K., Shih, P. M., De Paoli, H. C., Trinh, C. T., McCormick, A. J., Ployet, R., Hussey, S. G., Myburg, A. A., Jensen, P. E., Hassan, M. M., Zhang, J., Muchero, W., Kalluri, U. C., Yin, H., Zhuo, R., Abraham, P. E., Chen, J.-G., ... Tuskan, G. A. (2020). Plant biosystems design research roadmap 1.0. *BioDesign Research*, 2020, 8051764. (cross-listed with *Eucalyptus*) <https://doi.org/10.34133/2020/8051764>
152. Yang, W., Wang, D., Li, Y., Zhang, Z., Tong, S., Li, M., Zhang, X., Zhang, L., Ren, L., Ma, X., Zhou, R., Sanderson, B. J., Keefover-Ring, K., Yin, T., Smart, L. B., Liu, J., DiFazio, S. P., Olson, M., & Ma, T. (2021). A general model to explain repeated turnovers of sex determination in the salicaceae. *Molecular Biology and Evolution*, 38(3), 968–980. <https://doi.org/10.1093/molbev/msaa261>
153. Yang, Y., Shao, Y., Chaffin, T. A., Lee, J. H., Poindexter, M. R., Ahkami, A. H., Blumwald, E., & Stewart, C. N. (2022). Performance of abiotic stress-inducible synthetic promoters in genetically engineered hybrid poplar (*Populus tremula* × *Populus alba*). *Frontiers in Plant Science*, 13, 1011939. <https://doi.org/10.3389/fpls.2022.1011939>

154. Yang, Y., Lee, J. H., Poindexter, M. R., Shao, Y., Liu, W., Lenaghan, S. C., Ahkami, A. H., Blumwald, E., & Stewart, C. N. (2021). Rational design and testing of abiotic stress-inducible synthetic promoters from poplar *cis*-regulatory elements. *Plant Biotechnology Journal*, 19(7), 1354–1369. <https://doi.org/10.1111/pbi.13550>
155. Yao, T., Yuan, G., Lu, H., Liu, Y., Zhang, J., Tuskan, G. A., Muchero, W., Chen, J.-G., & Yang, X. (2023). CRISPR/Cas9-based gene activation and base editing in *Populus*. *Horticulture Research*, 10(6), uhad085. <https://doi.org/10.1093/hr/uhad085>
156. Yao, T., Zhang, J., Yates, T. B., Shrestha, H. K., Engle, N. L., Ployet, R., John, C., Feng, K., Bewg, W. P., Chen, M. S. S., Lu, H., Harding, S. A., Qiao, Z., Jawdy, S. S., Shu, M., Yuan, W., Mozaffari, K., Harman-Ware, A. E., Happs, R. M., ... Muchero, W. (2023). Expression quantitative trait loci mapping identified *PtrXB38* as a key hub gene in adventitious root development in *Populus*. *New Phytologist*, 239(6), 2248–2264. <https://doi.org/10.1111/nph.19126>
157. Yao, T., Zhang, J., Xie, M., Yuan, G., Tschaplinski, T. J., Muchero, W., & Chen, J.-G. (2021). Transcriptional regulation of drought response in *Arabidopsis* and woody plants. *Frontiers in Plant Science*, 11, 572137. <https://doi.org/10.3389/fpls.2020.572137>
158. Yao, T., Feng, K., Xie, M., Barros, J., Tschaplinski, T. J., Tuskan, G. A., Muchero, W., & Chen, J.-G. (2021). Phylogenetic occurrence of the phenylpropanoid pathway and lignin biosynthesis in plants. *Frontiers in Plant Science*, 12, 704697. <https://doi.org/10.3389/fpls.2021.704697>
159. Yao, W., Li, C., Lin, S., Wang, J., Zhou, B., & Jiang, T. (2020). Transcriptome analysis of salt-responsive and wood-associated NACs in *Populus simonii* × *Populus nigra*. *BMC Plant Biology*, 20(1), 317. <https://doi.org/10.1186/s12870-020-02507-z>
160. Yao, W., Zhang, D., Zhou, B., Wang, J., Li, R., & Jiang, T. (2020). Over-expression of poplar *NAC15* gene enhances wood formation in transgenic tobacco. *BMC Plant Biology*, 20(1), 12. <https://doi.org/10.1186/s12870-019-2191-2>
161. Yates, T. B., Feng, K., Zhang, J., Singan, V., Jawdy, S. S., Ranjan, P., Abraham, P. E., Barry, K., Lipzen, A., Pan, C., Schmutz, J., Chen, J.-G., Tuskan, G. A., & Muchero, W. (2021). The ancient Salicoid genome duplication event: A platform for reconstruction of de novo gene evolution in *populus trichocarpa*. *Genome Biology and Evolution*, 13(9), evab198. <https://doi.org/10.1093/gbe/evab198>
162. Yu, J., Zhou, C., Li, D., Li, S., Jimmy Lin, Y.-C., Wang, J. P., Chiang, V. L., & Li, W. (2022). A PtrLBD39-mediated transcriptional network regulates tension wood formation in *Populus trichocarpa*. *Plant Communications*, 3(1), 100250. <https://doi.org/10.1016/j.xplc.2021.100250>
163. Yuan, G., Tuskan, G. A., & Yang, X. (2023). Use of fluorescent protein reporters for assessing and detecting genome editing reagents and transgene expression in plants. In B. Yang, W. Harwood, & Q. Que (Eds.), *Plant Genome Engineering* (Vol. 2653, pp. 115–127). Springer US. https://link.springer.com/10.1007/978-1-0716-3131-7_8
164. Yuan, G., Martin, S., Hassan, M. M., Tuskan, G. A., & Yang, X. (2022). PARA: A new platform for the rapid assembly of gRNA arrays for multiplexed CRISPR technologies. *Cells*, 11(16), 2467. <https://doi.org/10.3390/cells11162467>
165. Yuan, G., Hassan, Md. M., Yao, T., Lu, H., Vergara, M. M., Labbé, J. L., Muchero, W., Pan, C., Chen, J.-G., Tuskan, G. A., Qi, Y., Abraham, P. E., & Yang, X. (2021). Plant-based biosensors for

- detecting CRISPR-mediated genome engineering. *ACS Synthetic Biology*, 10(12), 3600–3603. <https://doi.org/10.1021/acssynbio.1c00455>
166. Yuan, G., Liu, Y., Yao, T., Muchero, W., Chen, J.-G., Tuskan, G. A., & Yang, X. (2023). eYGFPuv-Assisted transgenic selection in *Populus deltoides* WV94 and multiplex genome editing in protoplasts of *P. trichocarpa* × *P. deltoides* clone '52-225.' *Plants*, 12(8), 1657. <https://doi.org/10.3390/plants12081657>
 167. Yuan, G., Lu, H., De, K., Hassan, M. M., Liu, Y., Islam, Md. T., Muchero, W., Tuskan, G. A., & Yang, X. (2023). Split selectable marker systems utilizing inteins facilitate gene stacking in plants. *Communications Biology*, 6(1), 567. <https://doi.org/10.1038/s42003-023-04950-8>
 168. Yuan, G., Lu, H., Tang, D., Hassan, M. M., Li, Y., Chen, J.-G., Tuskan, G. A., & Yang, X. (2021). Expanding the application of a UV-visible reporter for transient gene expression and stable transformation in plants. *Horticulture Research*, 8(1), 234. <https://doi.org/10.1038/s41438-021-00663-3>
 169. Yuan, J., Kaur, D., Zhou, Z., Nagle, M., Kiddle, N. G., Doshi, N. A., Behnoudfar, A., Peremyslova, E., Ma, C., Strauss, S. H., & Li, F. (2022). Robust high-throughput phenotyping with deep segmentation enabled by a web-based annotator. *Plant Phenomics*, 2022, 2022/9893639. <https://doi.org/10.34133/2022/9893639>
 170. Zhang, B., Wang, Z., Dai, X., Gao, J., Zhao, J., Ma, R., Chen, Y., Sun, Y., Ma, H., Li, S., Zhou, C., Wang, J. P., & Li, W. (2024). A COMPASS histone H3K4 trimethyltransferase pentamer transactivates drought tolerance and growth/biomass production in *Populus trichocarpa*. *New Phytologist*, 241(5), 1950–1972. <https://doi.org/10.1111/nph.19481>
 171. Zhang, L., Liu, M., Liu, Y., Wang, J., Lei, Y., Chen, Z., & Yan, L. (2023). Evaluation of the colour parameters on hygroscopicity and dimensional stability of thermally modified *Populus tomentosa* Carr. *Industrial Crops and Products*, 205, 117506. <https://doi.org/10.1016/j.indcrop.2023.117506>
 172. Zhang, L., Zhao, J., Bi, H., Yang, X., Zhang, Z., Su, Y., Li, Z., Zhang, L., Sanderson, B. J., Liu, J., & Ma, T. (2021). Bioinformatic analysis of chromatin organization and biased expression of duplicated genes between two poplars with a common whole-genome duplication. *Horticulture Research*, 8(1), 62. <https://doi.org/10.1038/s41438-021-00494-2>
 173. Zhang, L., Liu, B., Zhang, J., & Hu, J. (2020). Insights of molecular mechanism of xylem development in five black poplar cultivars. *Frontiers in Plant Science*, 11, 620. <https://doi.org/10.3389/fpls.2020.00620>
 174. Zhang, Q., Su, Z., Guo, Y., Zhang, S., Jiang, L., & Wu, R. (2020). Genome-wide association studies of callus differentiation for the desert tree, *Populus euphratica*. *Tree Physiology*, 40(12), 1762–1777. <https://doi.org/10.1093/treephys/tpaa098>
 175. Zhang, S., Wang, B., Li, Q., Hui, W., Yang, L., Wang, Z., Zhang, W., Yue, F., Liu, N., Li, H., Lu, F., Zhang, K., Zeng, Q., & Wu, A.-M. (2023). CRISPR/CAS9 mutated *p*-coumaroyl shikimate 3'-hydroxylase 3 gene in *Populus tomentosa* reveals lignin functioning on supporting tree upright. *International Journal of Biological Macromolecules*, 253, 126762. <https://doi.org/10.1016/j.ijbiomac.2023.126762>

176. Zhang, X.-Y., Huang, Z., Su, X., Siu, A., Song, Y., Zhang, D., & Fang, Q. (2020). Machine learning models for net photosynthetic rate prediction using poplar leaf phenotype data. *PLOS ONE*, 15(2), e0228645. <https://doi.org/10.1371/journal.pone.0228645>
177. Zhang, Y., Liu, Y., Wang, X., Wang, R., Chen, X., Wang, S., Wei, H., & Wei, Z. (2022). *PtrWOX13A* promotes wood formation and bioactive gibberellins biosynthesis in *Populus trichocarpa*. *Frontiers in Plant Science*, 13, 835035. <https://doi.org/10.3389/fpls.2022.835035>
178. Zhang, Y., Liu, C., Cheng, H., Tian, S., Liu, Y., Wang, S., Zhang, H., Saqib, M., Wei, H., & Wei, Z. (2020). DNA methylation and its effects on gene expression during primary to secondary growth in poplar stems. *BMC Genomics*, 21(1), 498. <https://doi.org/10.1186/s12864-020-06902-6>
179. Zhang, Y., Xu, S., Ji, F., Hu, Y., Gu, Z., & Xu, B. (2020). Plant cell wall hydrolysis process reveals structure–activity relationships. *Plant Methods*, 16(1), 147. <https://doi.org/10.1186/s13007-020-00691-5>
180. Zhang, Z., Chen, Y., Zhang, J., Ma, X., Li, Y., Li, M., Wang, D., Kang, M., Wu, H., Yang, Y., Olson, M. S., DiFazio, S. P., Wan, D., Liu, J., & Ma, T. (2020). Improved genome assembly provides new insights into genome evolution in a desert poplar (*Populus euphratica*). *Molecular Ecology Resources*, 20(3), 781–794. <https://doi.org/10.1111/1755-0998.13142>
181. Zhao, Y., Guo, Q., Cao, S., Tian, Y., Han, K., Sun, Y., Li, J., Yang, Q., Ji, Q., Sederoff, R., & Li, Y. (2022). Genome-wide identification of the *AlkB* homologs gene family, *PagALKBH9B* and *PagALKBH10B* regulated salt stress response in *Populus*. *Frontiers in Plant Science*, 13, 994154. <https://doi.org/10.3389/fpls.2022.994154>
182. Zhao, Y., Zhang, Y., Zhang, W., Shi, Y., Jiang, C., Song, X., Tuskan, G. A., Zeng, W., Zhang, J., & Lu, M. (2022). The *PagKNAT2/6b-PagBOP1/2a* regulatory module controls leaf morphogenesis in *Populus*. *International Journal of Molecular Sciences*, 23(10), 5581. <https://doi.org/10.3390/ijms23105581>
183. Zhao, Y., Yu, X.-H., & Liu, C.-J. (2021). The inducible accumulation of cell wall-bound *p*-hydroxybenzoates is involved in the regulation of gravitropic response of poplar. *Frontiers in Plant Science*, 12, 755576. <https://doi.org/10.3389/fpls.2021.755576>
184. Zhou, F., Wu, H., Chen, Y., Wang, M., Tuskan, G. A., & Yin, T. (2023). Function and molecular mechanism of a poplar placenta limited MIXTA gene in regulating differentiation of plant epidermal cells. *International Journal of Biological Macromolecules*, 242, 124743. <https://doi.org/10.1016/j.ijbiomac.2023.124743>
185. Zhou, J., Zhang, S., Wang, J., Shen, H., Ai, B., Gao, W., Zhang, C., Fei, Q., Yuan, D., Wu, Z., Tembrock, L. R., Li, S., Gu, C., & Liao, X. (2021). Chloroplast genomes in *Populus* (Salicaceae): Comparisons from an intensively sampled genus reveal dynamic patterns of evolution. *Scientific Reports*, 11(1), 9471. <https://doi.org/10.1038/s41598-021-88160-4>
186. Zhou, R., Jenkins, J. W., Zeng, Y., Shu, S., Jang, H., Harding, S. A., Williams, M., Plott, C., Barry, K. W., Koriabine, M., Amirebrahimi, M., Talag, J., Rajasekar, S., Grimwood, J., Schmitz, R. J., Dawe, R. K., Schmutz, J., & Tsai, C. (2023). Haplotype-resolved genome assembly of *Populus tremula* × *P. alba* reveals aspen-specific megabase satellite DNA. *The Plant Journal*, 116(4), 1003–1017. <https://doi.org/10.1111/tpj.16454>

187. Zhou, R., Macaya-Sanz, D., Schmutz, J., Jenkins, J. W., Tuskan, G. A., & DiFazio, S. P. (2020). Sequencing and analysis of the sex determination region of *Populus trichocarpa*. *Genes*, 11(8), 843. <https://doi.org/10.3390/genes11080843>
188. Zhou, Y., Zhang, Y., Wang, X., Han, X., An, Y., Lin, S., Shen, C., Wen, J., Liu, C., Yin, W., & Xia, X. (2020). Root-specific NF-Y family transcription factor, *PdNF-YB21*, positively regulates root growth and drought resistance by abscisic acid-mediated indoleacetic acid transport in *Populus*. *New Phytologist*, 227(2), 407–426. <https://doi.org/10.1111/nph.16524>
189. Zhu, X., Sun, F., Sang, M., Ye, M., Bo, W., Dong, A., & Wu, R. (2022). Genetic architecture of heterophylly: Single and multi-leaf genome-wide association mapping in *Populus euphratica*. *Frontiers in Plant Science*, 13, 870876. <https://doi.org/10.3389/fpls.2022.870876>
190. Zinkgraf, M., Zhao, S., Canning, C., Gerttula, S., Lu, M., Filkov, V., & Groover, A. (2020). Evolutionary network genomics of wood formation in a phylogenetic survey of angiosperm forest trees. *New Phytologist*, 228(6), 1811–1823. (cross-listed with *Salix* and *Eucalyptus*) <https://doi.org/10.1111/nph.16819>

Plant Health, Resilience to Threats and Climate Change

191. Alvesshere, B. C., Bennett, P., Kim, M.-S., Klopfenstein, N. B., & LeBoldus, J. M. (2021). First report of *Armillaria cepistipes* causing root disease on *Populus trichocarpa* (black cottonwood) in Oregon, U.S.A. *Plant Disease*, 105(9), 2729. <https://doi.org/10.1094/PDIS-09-20-1993-PDN>
192. Andrus, R. A., Hart, S. J., Tutland, N., & Veblen, T. T. (2021). Future dominance by quaking aspen expected following short-interval, compounded disturbance interaction. *Ecosphere*, 12(1), e03345. <https://doi.org/10.1002/ecs2.3345>
193. Andrus, R. A., Hart, S. J., & Veblen, T. T. (2020). Forest recovery following synchronous outbreaks of spruce and western balsam bark beetle is slowed by ungulate browsing. *Ecology*, 101(5), e02998. <https://doi.org/10.1002/ecy.2998>
194. Baldacci-Cresp, F., Behr, M., Kohler, A., Badalato, N., Morreel, K., Goeminne, G., Mol, A., Engler, J. de A., Boerjan, W., El Jaziri, M., & Baucher, M. (2020). Molecular changes concomitant with vascular system development in mature galls induced by root-knot nematodes in the model tree host *Populus tremula* × *P. alba*. *International Journal of Molecular Sciences*, 21(2), 406. <https://doi.org/10.3390/ijms21020406>
195. Banan, D., Sher, A., Doty, S. L., & Kim, S.-H. (2024). Endophyte mediated *Populus trichocarpa* water use efficiency is dependent on time of day and plant water status. *Phytobiomes Journal*, PBIOMES-11-22-0077-R. <https://doi.org/10.1094/PBIOMES-11-22-0077-R>
196. Barua, N., Herken, A. M., Stern, K. R., Reese, S., Powers, R. L., Morrell-Falvey, J. L., Platt, T. G., & Hansen, R. R. (2021). Simultaneous discovery of positive and negative interactions among rhizosphere bacteria using microwell recovery arrays. *Frontiers in Microbiology*, 11, 601788. <https://doi.org/10.3389/fmicb.2020.601788>
197. Basso, V., Kohler, A., Miyauchi, S., Singan, V., Guinet, F., Šimura, J., Novák, O., Barry, K. W., Amirebrahimi, M., Block, J., Daguerre, Y., Na, H., Grigoriev, I. V., Martin, F., & Veneault-Fourrey, C. (2020). An ectomycorrhizal fungus alters sensitivity to jasmonate, salicylate, gibberellin, and

ethylene in host roots. *Plant, Cell & Environment*, 43(4), 1047–1068.
<https://doi.org/10.1111/pce.13702>

198. Bayliss, S. L. J., Mueller, L. O., Ware, I. M., Schweitzer, J. A., & Bailey, J. K. (2022). Stacked distribution models predict climate-driven loss of variation in leaf phenology at continental scales. *Communications Biology*, 5(1), 1213. <https://doi.org/10.1038/s42003-022-04131-z>
199. Becklin, K. M., Viele, B. M., & Coleman, H. D. (2023). Nutrient conditions mediate mycorrhizal effects on biomass production and cell wall chemistry in poplar. *Tree Physiology*, 43(9), 1571–1583. <https://doi.org/10.1093/treephys/tpad064>
200. Behr, M., Baldacci-Cresp, F., Kohler, A., Morreel, K., Goeminne, G., Van Acker, R., Veneault-Fourrey, C., Mol, A., Pilate, G., Boerjan, W., Engler, J. de A., El Jaziri, M., & Baucher, M. (2020). Alterations in the phenylpropanoid pathway affect poplar ability for ectomycorrhizal colonisation and susceptibility to root-knot nematodes. *Mycorrhiza*, 30(5), 555–566. <https://doi.org/10.1007/s00572-020-00976-6>
201. Bennett, A. E., Rubert-Nason, K. R., & Lindroth, R. L. (2020). Response of aspen genotypes to browsing damage is not influenced by soil community diversity. *Plant and Soil*, 452(1–2), 153–170. <https://doi.org/10.1007/s11104-020-04466-8>
202. Bennett, S. K., Lambert, A. M., Carey, S. P., & Braman, C. A. (2022). Evaluating pole cutting survival and growth for riparian forest restoration during invasion by polyphagous shot hole borer. *Restoration Ecology*, 30(5), e13578. (cross-listed with *Salix*)
<https://doi.org/10.1111/rec.13578>
203. Beschta, R. L., & Ripple, W. J. (2020). Large carnivore extirpation linked to loss of overstory aspen in yellowstone. *Food Webs*, 22, e00140. <https://doi.org/10.1016/j.fooweb.2019.e00140>
204. Beschta, R. L., Ripple, W. J., Kauffman, J. B., & Painter, L. E. (2020). Bison limit ecosystem recovery in northern yellowstone. *Food Webs*, 23, e00142.
<https://doi.org/10.1016/j.fooweb.2020.e00142>
205. Blumstein, M., Richardson, A., Weston, D., Zhang, J., Muchero, W., & Hopkins, R. (2020). A new perspective on ecological prediction reveals limits to climate adaptation in a temperate tree species. *Current Biology*, 30(8), 1447–1453.e4. <https://doi.org/10.1016/j.cub.2020.02.001>
206. Bothwell, H. M., Evans, L. M., Hersch-Green, E. I., Woolbright, S. A., Allan, G. J., & Whitham, T. G. (2021). Genetic data improves niche model discrimination and alters the direction and magnitude of climate change forecasts. *Ecological Applications*, 31(3), e02254.
<https://doi.org/10.1002/eap.2254>
207. Bothwell, H. M., Keith, A. R., Cooper, H. F., Hull, J. B., Andrews, L. V., Wehenkel, C., Hultine, K. R., Gehring, C. A., Cushman, S. A., Whitham, T. G., & Allan, G. J. (2023). Microevolutionary processes in a foundation tree inform macrosystem patterns of community biodiversity and structure. *Forests*, 14(5), 943. <https://doi.org/10.3390/f14050943>
208. Boyd, M. A., Berner, L. T., Foster, A. C., Goetz, S. J., Rogers, B. M., Walker, X. J., & Mack, M. C. (2021). Historic declines in growth portend trembling aspen death during a contemporary leaf miner outbreak in Alaska. *Ecosphere*, 12(6), e03569. <https://doi.org/10.1002/ecs2.3569>

209. Braziunas, K. H., Kiel, N. G., & Turner, M. G. (2023). Less fuel for the next fire? Short-interval fire delays forest recovery and interacting drivers amplify effects. *Ecology*, 104(6), e4042. <https://doi.org/10.1002/ecy.4042>
210. Brewen, C. J., Berrill, J.-P., Ritchie, M. W., Boston, K., Dagley, C. M., Jones, B., Coppoletta, M., & Burnett, C. L. (2021). 76-year decline and recovery of aspen mediated by contrasting fire regimes: Long-unburned, infrequent and frequent mixed-severity wildfire. *PLOS ONE*, 16(2), e0232995. <https://doi.org/10.1371/journal.pone.0232995>
211. Bryant, N., Muchero, W., Weber, R. A., Barros, J., Chen, J.-G., Tschaplinski, T. J., Pu, Y., & Ragauskas, A. J. (2023). Cell wall response of field grown *Populus* to *Septoria* infection. *Frontiers in Plant Science*, 14, 1089011. <https://doi.org/10.3389/fpls.2023.1089011>
212. Carper, D. L., Schadt, C. W., Burdick, L. H., Kalluri, U. C., & Pelletier, D. A. (2020). Draft genome sequence of *Tumebacillus* sp. Strain BK434, isolated from the roots of eastern cottonwood. *Microbiology Resource Announcements*, 9(22), e00351-20. <https://doi.org/10.1128/MRA.00351-20>
213. Carper, D. L., Weston, D. J., Barde, A., Timm, C. M., Lu, T.-Y., Burdick, L. H., Jawdy, S. S., Klingeman, D. M., Robeson, M. S., Veach, A. M., Cregger, M. A., Kalluri, U. C., Schadt, C. W., Podar, M., Doktycz, M. J., & Pelletier, D. A. (2021). Cultivating the bacterial Microbiota of *Populus* Roots. *mSystems*, 6(3), e01306-20. <https://doi.org/10.1128/mSystems.01306-20>
214. Carrell, A. A., Clark, M., Jawdy, S., Muchero, W., Alexandre, G., Labbé, J. L., & Rush, T. A. (2023). Interactions with microbial consortia have variable effects in organic carbon and production of exometabolites among genotypes of *Populus trichocarpa*. *Plant Direct*, 7(11), e544. <https://doi.org/10.1002/pld3.544>
215. Carrell, A. A., Hicks, B. B., Sidelinger, E., Johnston, E. R., Jawdy, S. S., Clark, M. M., Klingeman, D. M., & Cregger, M. A. (2023). Nitrogen addition alters soil fungal communities, but root fungal communities are resistant to change. *Frontiers in Microbiology*, 13, 1033631. <https://doi.org/10.3389/fmicb.2022.1033631>
216. Castillo, B. T., Franklin, R. B., Amses, K. R., Leite, M. F. A., Kuramae, E. E., Gough, C. M., James, T. Y., Faller, L., & Syring, J. (2023). Fungal community succession of *Populus grandidentata* (bigtooth aspen) during wood decomposition. *Forests*, 14(10), 2086. <https://doi.org/10.3390/f14102086>
217. Chu, C., Momayyezi, M., Stobbs, J. A., Soolanayakanahally, R. Y., McElrone, A. J., & Knipfer, T. (2023). Drought-induced fiber water release and xylem embolism susceptibility of intact balsam poplar saplings. *Physiologia Plantarum*, 175(5), e14040. <https://doi.org/10.1111/ppl.14040>
218. Coleman, H. D., Brunner, A. M., & Tsai, C.-J. (2021). Synergies and entanglement in secondary cell wall development and abiotic stress response in trees. *Frontiers in Plant Science*, 12, 639769. <https://doi.org/10.3389/fpls.2021.639769>
219. Cooper, H. F., Best, R. J., Andrews, L. V., Corbin, J. P. M., Garthwaite, I., Grady, K. C., Gehring, C. A., Hultine, K. R., Whitham, T. G., & Allan, G. J. (2022). Evidence of climate-driven selection on tree traits and trait plasticity across the climatic range of a riparian foundation species. *Molecular Ecology*, 31(19), 5024–5040. <https://doi.org/10.1111/mec.16645>

220. Cope, K. R., Prates, E. T., Miller, J. I., Demerdash, O. N. A., Shah, M., Kainer, D., Cliff, A., Sullivan, K. A., Cashman, M., Lane, M., Matthiadis, A., Labbé, J., Tschaplinski, T. J., Jacobson, D. A., & Kalluri, U. C. (2023). Exploring the role of plant lysin motif receptor-like kinases in regulating plant-microbe interactions in the bioenergy crop *Populus*. *Computational and Structural Biotechnology Journal*, 21, 1122–1139. <https://doi.org/10.1016/j.csbj.2022.12.052>
221. Cope, K. R., Irving, T. B., Chakraborty, S., & Ané, J.-M. (2021). Perception of lipo-chitooligosaccharides by the bioenergy crop *Populus*. *Plant Signaling & Behavior*, 16(6), 1903758. <https://doi.org/10.1080/15592324.2021.1903758>
222. Corenblit, D., Corbara, B., Cereghino, R., Dejean, A., Duran, P., Garófano-Gómez, V., Gilbert, F., González-Sargas, E., Julien, F., Lambs, L., Mainguin, C., Mazal, L., Otto, T., Steiger, J., Tabacchi, E., Till-Bottraud, I., & Travaillard, Y. (2023). Effects of ants on riparian poplars: An ex situ experiment of biotic interaction. *Arthropod-Plant Interactions*, 17(2), 237–252. <https://doi.org/10.1007/s11829-023-09950-3>
223. Cregger, M. A., Carper, D. L., Christel, S., Doktycz, M. J., Labbé, J., Michener, J. K., Dove, N. C., Johnston, E. R., Moore, J. A. M., Vélez, J. M., Morrell-Falvey, J., Muchero, W., Pelletier, D. A., Retterer, S., Tschaplinski, T. J., Tuskan, G. A., Weston, D. J., & Schadt, C. W. (2021). Plant-microbe interactions: From genes to ecosystems using *Populus* as a model system. *Phytobiomes Journal*, 5(1), 29–38. <https://doi.org/10.1094/PBIOMES-01-20-0009-FI>
224. Crouch, C. D., Grady, A. M., Wilhelmi, N. P., Hofstetter, R. W., DePinte, D. E., & Waring, K. M. (2021). Oystershell scale: An emerging invasive threat to aspen in the southwestern US. *Biological Invasions*, 23(9), 2893–2912. <https://doi.org/10.1007/s10530-021-02545-0>
225. Crouch, C. D., Rogers, P. C., Moore, M. M., & Waring, K. M. (2023). Building ecosystem resilience and adaptive capacity: A systematic review of aspen ecology and management in the Southwest. *Forest Science*, 69(3), 334–354. <https://doi.org/10.1093/forsci/xfad004>
226. Darsaraei, H., Khodaparast, S. A., Takamatsu, S., Abbasi, M., Asgari, B., Sajedi, S., Götz, M., Liu, S.-Y., Feng, J., Bradshaw, M., Bulgakov, T., & Braun, U. (2021). Phylogeny and taxonomy of the *Erysiphe adunca* complex (Erysiphaceae, Helotiales) on poplars and willows. *Mycological Progress*, 20(4), 517–537. (cross-listed with *Salix*) <https://doi.org/10.1007/s11557-021-01688-7>
227. DeRose, R. J., Gardner, R. S., Lindroth, R. L., & Mock, K. E. (2022). Polyploidy and growth—Defense tradeoffs in natural populations of western quaking aspen. *Journal of Chemical Ecology*, 48(4), 431–440. <https://doi.org/10.1007/s10886-022-01355-5>
228. Ding, C., Hamann, A., Yang, R.-C., & Brouard, J. S. (2020). Genetic parameters of growth and adaptive traits in aspen (*Populus tremuloides*): Implications for tree breeding in a warming world. *PLOS ONE*, 15(3), e0229225. <https://doi.org/10.1371/journal.pone.0229225>
229. Dong, F., Wang, Y., Tao, J., Xu, T., & Tang, M. (2023). Arbuscular mycorrhizal fungi affect the expression of *PxNHX* gene family, improve photosynthesis and promote *Populus simonii* × *P. nigra* growth under saline-alkali stress. *Frontiers in Plant Science*, 14, 1104095. <https://doi.org/10.3389/fpls.2023.1104095>
230. Doty, S. L., Joubert, P. M., Firrincieli, A., Sher, A. W., Tournay, R., Kill, C., Parikh, S. S., & Okubara, P. (2022). Potential biocontrol activities of *Populus* endophytes against several plant

pathogens using different inhibitory mechanisms. *Pathogens*, 12(1), 13.
<https://doi.org/10.3390/pathogens12010013>

231. Dove, N. C., Carrell, A. A., Engle, N. L., Klingeman, D. M., Rodriguez, M., Wahl, T., Tschaplinski, T. J., Muchero, W., Schadt, C. W., & Cregger, M. A. (2022). Relationships between *Sphaerulina musiva* infection and the *Populus* microbiome and metabolome. *mSystems*, 7(4), e00120-22.
<https://doi.org/10.1128/msystems.00120-22>
232. Dove, N. C., Veach, A. M., Muchero, W., Wahl, T., Stegen, J. C., Schadt, C. W., & Cregger, M. A. (2021). Assembly of the *Populus* microbiome is temporally dynamic and determined by selective and stochastic factors. *mSphere*, 6(3), e01316-20.
<https://doi.org/10.1128/mSphere.01316-20>
233. Dowell, R. V., & Scriber, J. M. (2023). Host plant mediated differences in *Papilio rutulus* Lucas larval growth (Papilionidae). *The Journal of the Lepidopterists' Society*, 77(2).
<https://doi.org/10.18473/lepi.77i2.a3>
234. Dudley, M. M., Tisserat, N. A., Jacobi, W. R., Negrón, J., & Stewart, J. E. (2020). Pathogenicity and distribution of two species of *Cytospora* on *Populus tremuloides* in portions of the rocky mountains and midwest in the United States. *Forest Ecology and Management*, 468, 118168.
<https://doi.org/10.1016/j.foreco.2020.118168>
235. Durben, R. M., Walker, F. M., Holeski, L., Keith, A. R., Kovacs, Z., Hurteau, S. R., Lindroth, R. L., Shuster, S. M., & Whitham, T. G. (2021). Beavers, bugs and chemistry: A mammalian herbivore changes chemistry composition and arthropod communities in foundation tree species. *Forests*, 12(7), 877. <https://doi.org/10.3390/f12070877>
236. Eckert, C., Wildhagen, H., Paulo, M. J., Scalabrin, S., Ballauff, J., Schnabel, S. K., Vendramin, V., Keurentjes, J. J. B., Bogeat-Triboulot, M.-B., Taylor, G., & Polle, A. (2022). Genotypic and tissue-specific variation of *Populus nigra* transcriptome profiles in response to drought. *Scientific Data*, 9(1), 297. <https://doi.org/10.1038/s41597-022-01417-z>
237. Eisenring, M., Lindroth, R. L., Flansburg, A., Giezendanner, N., Mock, K. E., & Kruger, E. L. (2023). Genotypic variation rather than ploidy level determines functional trait expression in a foundation tree species in the presence and absence of environmental stress. *Annals of Botany*, 131(1), 229–242. <https://doi.org/10.1093/aob/mcac071>
238. Eisenring, M., Best, R. J., Zierden, M. R., Cooper, H. F., Norstrom, M. A., Whitham, T. G., Grady, K., Allan, G. J., & Lindroth, R. L. (2022). Genetic divergence along a climate gradient shapes chemical plasticity of a foundation tree species to both changing climate and herbivore damage. *Global Change Biology*, 28(15), 4684–4700. <https://doi.org/10.1111/gcb.16275>
239. Eisenring, M., Unsicker, S. B., & Lindroth, R. L. (2021). Spatial, genetic and biotic factors shape within-crown leaf trait variation and herbivore performance in a foundation tree species. *Functional Ecology*, 35(1), 54–66. <https://doi.org/10.1111/1365-2435.13699>
240. Elferjani, R., Benomar, L., Momayyezi, M., Tognetti, R., Niinemets, Ü., Soolanayakanahally, R. Y., Thérout-Rancourt, G., Tosens, T., Ripullone, F., Bilodeau-Gauthier, S., Lamhamedi, M. S., Calfapietra, C., & Lamara, M. (2021). A meta-analysis of mesophyll conductance to CO₂ in relation to major abiotic stresses in poplar species. *Journal of Experimental Botany*, 72(12), 4384–4400. <https://doi.org/10.1093/jxb/erab127>

241. Feau, N., Dhillon, B. D., Sakalidis, M., Dale, A. L., Søndreli, K. L., Goodwin, S. B., LeBoldus, J. M., & Hamelin, R. C. (2023). Forest health in the Anthropocene: The emergence of a novel tree disease is associated with poplar cultivation. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 378(1873), 20220008. <https://doi.org/10.1098/rstb.2022.0008>
242. Feng, J., Liu, S.-Y., Braun, U., Takamatsu, S., Bradshaw, M., Khodaparast, S. A., Bulgakov, T. S., Guan, G.-X., Zhao, F.-Y., & Tang, S.-R. (2022). Discovery of a cryptic species, *Erysiphe salicina* sp. Nov., and reconstruction of the phylogeny of powdery mildews on *Populus* and *Salix* spp. *Mycological Progress*, 21(6), 54. (cross-listed with *Salix*) <https://doi.org/10.1007/s11557-022-01793-1>
243. Fetter, K. C., & Keller, S. R. (2023). Admixture mapping and selection scans identify genomic regions associated with stomatal patterning and disease resistance in hybrid poplars. *Ecology and Evolution*, 13(10), e10579. <https://doi.org/10.1002/ece3.10579>
244. Fetter, K. C., Nelson, D. M., & Keller, S. R. (2021). Growth-defense trade-offs masked in unadmixed populations are revealed by hybridization. *Evolution*, 75(6), 1450–1465. <https://doi.org/10.1111/evo.14227>
245. Firrincieli, A., Khorasani, M., Frank, A. C., & Doty, S. L. (2020). Influences of climate on phyllosphere endophytic bacterial communities of wild poplar. *Frontiers in Plant Science*, 11, 203. <https://doi.org/10.3389/fpls.2020.00203>
246. Fracchia, F., Mangeot-Peter, L., Jacquot, L., Martin, F., Veneault-Fourrey, C., & Deveau, A. (2021). Colonization of naive roots from *Populus tremula* × *alba* involves successive waves of fungi and bacteria with different trophic abilities. *Applied and Environmental Microbiology*, 87(6), e02541-20. <https://doi.org/10.1128/AEM.02541-20>
247. Gao, Y., Zhang, Z., Chen, J., McNulty, S., Xu, H., Chen, L., Chen, Z., & Jin, Z. (2021). Atmospheric aerosols elevated ecosystem productivity of a poplar plantation in Beijing, China. *Canadian Journal of Forest Research*, 51(10), 1440–1449. <https://doi.org/10.1139/cjfr-2020-0396>
248. Garcia, B. J., Simha, R., Garvin, M., Furches, A., Jones, P., Gazolla, J. G. F. M., Hyatt, P. D., Schadt, C. W., Pelletier, D., & Jacobson, D. (2021). A k-mer based approach for classifying viruses without taxonomy identifies viral associations in human autism and plant microbiomes. *Computational and Structural Biotechnology Journal*, 19, 5911–5919. <https://doi.org/10.1016/j.csbj.2021.10.029>
249. Garvie, L. A. J. (2022). Seasonal formation of ikaite in slime flux jelly on an infected tree (*Populus fremontii*) wound from the sonoran desert. *The Science of Nature*, 109(5), 48. <https://doi.org/10.1007/s00114-022-01818-5>
250. Goessen, R., Isabel, N., Wehenkel, C., Pavy, N., Tischenko, L., Touchette, L., Giguère, I., Gros-Louis, M., Laroche, J., Boyle, B., Soolanayakanahally, R., Mock, K., Hernández-Velasco, J., Simental-Rodriguez, S. L., Bousquet, J., & Porth, I. M. (2022). Coping with environmental constraints: Geographically divergent adaptive evolution and germination plasticity in the transcontinental *Populus tremuloides*. *PLANTS, PEOPLE, PLANET*, 4(6), 638–654. <https://doi.org/10.1002/ppp3.10297>

251. Gougherty, A. V., Keller, S. R., & Fitzpatrick, M. C. (2021). Maladaptation, migration and extirpation fuel climate change risk in a forest tree species. *Nature Climate Change*, 11(2), 166–171. <https://doi.org/10.1038/s41558-020-00968-6>
252. Han, Y., Xu, T., Chen, H., & Tang, M. (2023). Sugar metabolism and 14-3-3 protein genes expression induced by arbuscular mycorrhizal fungi and phosphorus addition to response drought stress in *Populus cathayana*. *Journal of Plant Physiology*, 288, 154075. <https://doi.org/10.1016/j.jplph.2023.154075>
253. Han, Y., Zhang, W., Xu, T., & Tang, M. (2022). Effect of arbuscular mycorrhizal fungi and phosphorus on drought-induced oxidative stress and 14-3-3 proteins gene expression of *Populus cathayana*. *Frontiers in Microbiology*, 13, 934964. <https://doi.org/10.3389/fmicb.2022.934964>
254. Heitmann, S., Bergmann, G. E., Barge, E., Ridout, M., Newcombe, G., & Busby, P. E. (2021). Culturable seed microbiota of *Populus trichocarpa*. *Pathogens*, 10(6), 653. <https://doi.org/10.3390/pathogens10060653>
255. Helm, L. T., Shi, H., Lerdau, M. T., & Yang, X. (2020). Solar-induced chlorophyll fluorescence and short-term photosynthetic response to drought. *Ecological Applications*, 30(5), e02101. <https://doi.org/10.1002/eap.2101>
256. Himes, A., Emerson, P., McClung, R., Renninger, H., Rosenstiel, T., & Stanton, B. (2021). Leaf traits indicative of drought resistance in hybrid poplar. *Agricultural Water Management*, 246, 106676. <https://doi.org/10.1016/j.agwat.2020.106676>
257. Hori, C., Yu, X., Mortimer, J. C., Sano, R., Matsumoto, T., Kikuchi, J., Demura, T., & Ohtani, M. (2020). Impact of abiotic stress on the regulation of cell wall biosynthesis in *Populus trichocarpa*. *Plant Biotechnology*, 37(3), 273–283. <https://doi.org/10.5511/plantbiotechnology.20.0326a>
258. Hu, X.-L., Zhang, J., Kaundal, R., Kataria, R., Labbé, J. L., Mitchell, J. C., Tschaplinski, T. J., Tuskan, G. A., Cheng, Z.-M. (Max), & Yang, X. (2022). Diversity and conservation of plant small secreted proteins associated with arbuscular mycorrhizal symbiosis. *Horticulture Research*, 9, uhac043. <https://doi.org/10.1093/hr/uhac043>
259. Hultine, K. R., Allan, G. J., Blasini, D., Bothwell, H. M., Cadmus, A., Cooper, H. F., Doughty, C. E., Gehring, C. A., Gitlin, A. R., Grady, K. C., Hull, J. B., Keith, A. R., Koepke, D. F., Markovchick, L., Corbin Parker, J. M., Sankey, T. T., & Whitham, T. G. (2020). Adaptive capacity in the foundation tree species *Populus fremontii*: Implications for resilience to climate change and non-native species invasion in the American Southwest. *Conservation Physiology*, 8(1), coaa061. <https://doi.org/10.1093/conphys/coaa061>
260. Inglis, N. C., & Vukomanovic, J. (2020). Climate change disproportionately affects visual quality of cultural ecosystem services in a mountain region. *Ecosystem Services*, 45, 101190. <https://doi.org/10.1016/j.ecoser.2020.101190>
261. Kang, H., Chen, X., Kemppainen, M., Pardo, A. G., Veneault-Fourrey, C., Kohler, A., & Martin, F. M. (2020). The small secreted effector protein MiSSP7.6 of *Laccaria bicolor* is required for the establishment of ectomycorrhizal symbiosis. *Environmental Microbiology*, 22(4), 1435–1446. <https://doi.org/10.1111/1462-2920.14959>

262. Kard, B. M., Page-Dumroese, D. S., Cook, S. P., Jurgensen, M. F., & Tirocke, J. M. (2022). Termite feeding on aspen and pine stakes on a high elevation sagebrush-steppe rangeland in southeastern Idaho. *Southwestern Entomologist*, 47(3). <https://doi.org/10.3958/059.047.0304>
263. Kerr, K. L., Fickle, J. C., & Anderegg, W. R. L. (2023). Decoupling of functional traits from intraspecific patterns of growth and drought stress resistance. *New Phytologist*, 239(1), 174–188. <https://doi.org/10.1111/nph.18937>
264. Kerr, K. L., Anderegg, L. D. L., Zenes, N., & Anderegg, W. R. L. (2022). Quantifying within-species trait variation in space and time reveals limits to trait-mediated drought response. *Functional Ecology*, 36(9), 2399–2411. <https://doi.org/10.1111/1365-2435.14112>
265. Kerr, K. L., Zenes, N., Trugman, A. T., & Anderegg, W. R. L. (2021). Testing the effects of species interactions and water limitation on tree seedling biomass allocation and physiology. *Tree Physiology*, 41(8), 1323–1335. <https://doi.org/10.1093/treephys/tpab005>
266. Kong, R. S., Way, D. A., Henry, H. A. L., & Smith, N. G. (2022). Stomatal conductance, not biochemistry, drives low temperature acclimation of photosynthesis in *Populus balsamifera*, regardless of nitrogen availability. *Plant Biology*, 24(5), 766–779. <https://doi.org/10.1111/plb.13428>
267. Konôpka, B., Pajtík, J., Bošeľa, M., Šebeň, V., & Shipley, L. A. (2020). Modeling forage potential for red deer (*Cervus elaphus*): A tree-level approach. *European Journal of Forest Research*, 139(3), 419–430. (cross-listed with *Salix*) <https://doi.org/10.1007/s10342-019-01250-x>
268. Konôpka, B., Šebeň, V., Pajtík, J., & Shipley, L. A. (2022). Influence of tree species and size on bark browsing by large wild herbivores. *Plants*, 11(21), 2925. (cross-listed with *Salix*) <https://doi.org/10.3390/plants11212925>
269. Kristy, B., Carrell, A. A., Johnston, E., Cumming, J. R., Klingeman, D. M., Gwinn, K., Syring, K. C., Skalla, C., Emrich, S., & Cregger, M. A. (2022). Chronic drought differentially alters the belowground microbiome of drought-tolerant and drought-susceptible genotypes of *Populus trichocarpa*. *Phytobiomes Journal*, 6(4), 317–330. <https://doi.org/10.1094/PBIOMES-12-21-0076-R>
270. Kruger, E. L., Keefover-Ring, K., Holeski, L. M., & Lindroth, R. L. (2020). To compete or defend: Linking functional trait variation with life-history tradeoffs in a foundation tree species. *Oecologia*, 192(4), 893–907. <https://doi.org/10.1007/s00442-020-04622-y>
271. Lackus, N. D., Müller, A., Kröber, T. D. U., Reichelt, M., Schmidt, A., Nakamura, Y., Paetz, C., Luck, K., Lindroth, R. L., Constabel, C. P., Unsicker, S. B., Gershenson, J., & Köllner, T. G. (2020). The occurrence of sulfated salicinoids in poplar and their formation by sulfotransferase1. *Plant Physiology*, 183(1), 137–151. <https://doi.org/10.1104/pp.19.01447>
272. Lamit, L. J., Meinhardt, K. A., Flores-Rentería, L., Kovacs, Z. I., Zinkgraf, M., Whitham, T. G., & Gehring, C. A. (2021). Ectomycorrhizal fungal communities differ among parental and hybrid *Populus* cross types within a natural riparian habitat. *Fungal Ecology*, 52, 101059. <https://doi.org/10.1016/j.funeco.2021.101059>
273. Lenz, R. R., Louie, K. B., Søndreli, K. L., Galanie, S. S., Chen, J.-G., Muchero, W., Bowen, B. P., Northen, T. R., & LeBoldus, J. M. (2021). Metabolomic patterns of Septoria canker resistant

- and susceptible *Populus trichocarpa* genotypes 24 hours postinoculation. *Phytopathology*[®], 111(11), 2052–2066. <https://doi.org/10.1094/PHYTO-02-21-0053-R>
274. LeRoy, C. J., Fischer, D., Schweitzer, J. A., & Bailey, J. K. (2020). Aphid gall interactions with forest tree genotypes influence leaf litter decomposition in streams. *Forests*, 11(2), 182. <https://doi.org/10.3390/f11020182>
 275. Li, G., Lin, R., Egekwu, C., Blakeslee, J., Lin, J., Pettengill, E., Murphy, A. S., Peer, W. A., Islam, N., Babst, B. A., Gao, F., Komarov, S., Tai, Y.-C., & Coleman, G. D. (2020). Seasonal nitrogen remobilization and the role of auxin transport in poplar trees. *Journal of Experimental Botany*, 71(15), 4512–4530. <https://doi.org/10.1093/jxb/eraa130>
 276. Li, Z., Rubert-Nason, K. F., Jamieson, M. A., Raffa, K. F., & Lindroth, R. L. (2021). Root secondary metabolites in *Populus tremuloides*: Effects of simulated climate warming, defoliation, and genotype. *Journal of Chemical Ecology*, 47(3), 313–321. <https://doi.org/10.1007/s10886-021-01259-w>
 277. Li, Z., Yang, J., Shang, B., Agathokleous, E., Rubert-Nason, K. F., Xu, Y., & Feng, Z. (2021). Nonlinear responses of foliar phenylpropanoids to increasing O₃ exposure: Ecological implications in a *Populus* model system. *Science of The Total Environment*, 767, 144358. <https://doi.org/10.1016/j.scitotenv.2020.144358>
 278. Li, Z., Yang, J., Shang, B., Xu, Y., Couture, J. J., Yuan, X., Kobayashi, K., & Feng, Z. (2020). Water stress rather than N addition mitigates impacts of elevated O₃ on foliar chemical profiles in poplar saplings. *Science of The Total Environment*, 707, 135935. <https://doi.org/10.1016/j.scitotenv.2019.135935>
 279. Liao, H.-L., Bonito, G., Hameed, K., Wu, S. H., Chen, K.-H., Labbé, J., Schadt, C. W., Tuskan, G. A., Martin, F., Kuo, A., Barry, K., Grigoriev, I. V., & Vilgalys, R. (2021). Heterospecific neighbor plants impact root microbiome diversity and molecular function of root fungi. *Frontiers in Microbiology*, 12, 680267. <https://doi.org/10.3389/fmicb.2021.680267>
 280. Lindroth, R. L., Wooley, S. C., Donaldson, J. R., Rubert-Nason, K. F., Morrow, C. J., & Mock, K. E. (2023). Phenotypic variation in phytochemical defense of trembling aspen in western north america: Genetics, development, and geography. *Journal of Chemical Ecology*, 49(5–6), 235–250. <https://doi.org/10.1007/s10886-023-01409-2>
 281. Liu, J., Li, D., Fernández, J.-E., Coleman, M., Hu, W., Di, N., Zou, S., Liu, Y., Xi, B., & Clothier, B. (2022). Variations in water-balance components and carbon stocks in poplar plantations with differing water inputs over a whole rotation: Implications for sustainable forest management under climate change. *Agricultural and Forest Meteorology*, 320, 108958. <https://doi.org/10.1016/j.agrformet.2022.108958>
 282. Looney, C. E., Previant, W. J., Bradford, J. B., & Nagel, L. M. (2021). Species mixture effects and climate influence growth, recruitment and mortality in Interior West USA *Populus tremuloides*-conifer communities. *Journal of Ecology*, 109(8), 2934–2949. <https://doi.org/10.1111/1365-2745.13709>
 283. Mangeot-Peter, L., Tschaplinski, T. J., Engle, N. L., Veneault-Fourrey, C., Martin, F., & Deveau, A. (2020). Impacts of soil microbiome variations on root colonization by fungi and bacteria and

on the metabolome of *Populus tremula* × *alba*. *Phytobiomes Journal*, 4(2), 142–155.
<https://doi.org/10.1094/PBIOMES-08-19-0042-R>

284. Markovchick, L. M., Schaefer, E. A., Deringer, T., Kovacs, Z. I., Deckert, R. J., Yazzie, J., Dixit, A., Propster, J. R., Patterson, A., Hultine, K. R., Grady, K., Allan, G. J., Whitham, T. G., & Gehring, C. A. (2023). Postrestoration colonization suggests slow regeneration, plant translocation barriers, and other host/symbiont lessons during the United Nations' decade on ecosystem restoration. *Restoration Ecology*, 31(5), e13940. <https://doi.org/10.1111/rec.13940>
285. Marqués-Gálvez, J. E., Basso, V., Kohler, A., Barry, K., Keymanesh, K., Johnson, J., Singan, V., Grigoriev, I. V., Vilgalys, R., Martin, F., & Veneault-Fourrey, C. (2022). *The establishment of Populus × Laccaria bicolor ectomycorrhiza requires the inactivation of MYC2 coordinated defense response with a key role for root terpene synthases*.
<https://doi.org/10.1101/2022.09.06.505662>
286. Mason, C. J., Rubert-Nason, K., Lindroth, R. L., Shi, J., & Hoover, K. (2021). Salicinoid phenolics reduce adult *Anoplophora glabripennis* (Cerambycidae: Lamiinae) feeding and egg production. *Arthropod-Plant Interactions*, 15(1), 127–136. <https://doi.org/10.1007/s11829-020-09802-4>
287. Mayes, M., Caylor, K. K., Singer, M. B., Stella, J. C., Roberts, D., & Nagler, P. (2020). Climate sensitivity of water use by riparian woodlands at landscape scales. *Hydrological Processes*, 34(25), 4884–4903. (cross-listed with *Salix*) <https://doi.org/10.1002/hyp.13942>
288. Moran, M. E., Aparecido, L. M. T., Koepke, D. F., Cooper, H. F., Doughty, C. E., Gehring, C. A., Throop, H. L., Whitham, T. G., Allan, G. J., & Hultine, K. R. (2023). Limits of thermal and hydrological tolerance in a foundation tree species (*Populus fremontii*) in the desert southwestern United States. *New Phytologist*, 240(6), 2298–2311.
<https://doi.org/10.1111/nph.19247>
289. Morrow, C. J., Jaeger, S. J., & Lindroth, R. L. (2022). Intraspecific variation in plant economic traits predicts trembling aspen resistance to a generalist insect herbivore. *Oecologia*, 199(1), 119–128. <https://doi.org/10.1007/s00442-022-05158-z>
290. Neary, A., Mata-González, R., & Schmalz, H. (2021). Topographic, edaphic and climate influences on aspen (*Populus tremuloides*) drought stress on an intermountain bunchgrass prairie. *Forest Ecology and Management*, 479, 118530.
<https://doi.org/10.1016/j.foreco.2020.118530>
291. Newcombe, G., Fraser, S. J., Ridout, M., & Busby, P. E. (2020). Leaf endophytes of *Populus trichocarpa* act as pathogens of neighboring plant species. *Frontiers in Microbiology*, 11, 573056. <https://doi.org/10.3389/fmicb.2020.573056>
292. O'Banion, B. S., Jones, P., Demetros, A. A., Kelley, B. R., Knoor, L. H., Wagner, A. S., Chen, J.-G., Muchero, W., Reynolds, T. B., Jacobson, D., & Lebeis, S. L. (2023). Plant myo-inositol transport influences bacterial colonization phenotypes. *Current Biology*, 33(15), 3111–3124.e5.
<https://doi.org/10.1016/j.cub.2023.06.057>
293. Ouyang, Y., Leininger, T. D., Renninger, H., Gardiner, E. S., & Samuelson, L. (2021). A model to assess eastern cottonwood water flow using adjusted vapor pressure deficit associated with a climate change impact application. *Climate*, 9(2), 22. <https://doi.org/10.3390/cli9020022>

294. Painter, L. E., Beschta, R. L., & Ripple, W. J. (2023). Bison alter the northern yellowstone ecosystem by breaking aspen saplings. *Ecology and Evolution*, 13(8), e10369. <https://doi.org/10.1002/ece3.10369>
295. Pereira, M., Cohen, D., Auer, L., Aubry, N., Bogeat-Triboulot, M., Buré, C., Engle, N. L., Jolivet, Y., Kohler, A., Novák, O., Pavlović, I., Priault, P., Tschaplinski, T. J., Hummel, I., Vaultier, M., & Veneault-Fourrey, C. (2023). Ectomycorrhizal symbiosis prepares its host locally and systemically for abiotic cue signaling. *The Plant Journal*, 116(6), 1784–1803. <https://doi.org/10.1111/tpj.16465>
296. Perez, J. M., Sener, C., Misra, S., Umana, G. E., Coplien, J., Haak, D., Li, Y., Maravelias, C. T., Karlen, S. D., Ralph, J., Donohue, T. J., & Noguera, D. R. (2022). Integrating lignin depolymerization with microbial funneling processes using agronomically relevant feedstocks. *Green Chemistry*, 24(7), 2795–2811. <https://doi.org/10.1039/D1GC03592D>
297. Persoons, A., Maupetit, A., Louet, C., Andrieux, A., Lipzen, A., Barry, K. W., Na, H., Adam, C., Grigoriev, I. V., Segura, V., Duplessis, S., Frey, P., Halkett, F., & De Mita, S. (2022). Genomic signatures of a major adaptive event in the pathogenic fungus *Melampsora larici-populina*. *Genome Biology and Evolution*, 14(1), evab279. <https://doi.org/10.1093/gbe/evab279>
298. Refsland, T. K., & Cushman, J. H. (2021). Continent-wide synthesis of the long-term population dynamics of quaking aspen in the face of accelerating human impacts. *Oecologia*, 197(1), 25–42. <https://doi.org/10.1007/s00442-021-05013-7>
299. Reikowski, E. S., Refsland, T., & Cushman, J. H. (2022). Ungulate herbivores as drivers of aspen recruitment and understory composition throughout arid montane landscapes. *Ecosphere*, 13(9), e4225. <https://doi.org/10.1002/ecs2.4225>
300. Ruess, R. W., Winton, L. M., & Adams, G. C. (2021). Widespread mortality of trembling aspen (*Populus tremuloides*) throughout interior alaskan boreal forests resulting from a novel canker disease. *PLOS ONE*, 16(4), e0250078. <https://doi.org/10.1371/journal.pone.0250078>
301. Ruytinx, J., Miyauchi, S., Hartmann-Wittulsky, S., Pereira, M. de F., Guinet, F., Churin, J.-L., Put, C., Le Tacon, F., Veneault-Fourrey, C., Martin, F., & Kohler, A. (2021). A transcriptomic atlas of the ectomycorrhizal fungus *Laccaria bicolor*. *Microorganisms*, 9(12), 2612. <https://doi.org/10.3390/microorganisms9122612>
302. Sawyer, C. R., & Labbé, J. L. (2021). Small RNAs at the interface of the plant-fungal interactions. *Fungal Genomics & Biology*, 11(1). No DOI Available
303. Schmiede, S. C., Sharkey, T. D., Walker, B., Hammer, J., & Way, D. A. (2023). Laisk measurements in the nonsteady state: Tests in plants exposed to warming and variable CO₂ concentrations. *Plant Physiology*, 193(2), 1045–1057. <https://doi.org/10.1093/plphys/kiad305>
304. Sepúlveda, S. L., Neale, D. B., Holliday, J. A., Famula, R., Fiehn, O., Stanton, B. J., & Guerra, F. P. (2023). GWAS on the attack by aspen borer *Saperda calcarata* on black cottonwood trees reveals a response mechanism involving secondary metabolism and independence of tree architecture. *Forests*, 14(6), 1129. <https://doi.org/10.3390/f14061129>
305. Sher, A. W., Aufrecht, J. A., Herrera, D., Zimmerman, A. E., Kim, Y.-M., Munoz, N. M., Trejo, J. B., Paurus, V. L., Cliff, J. B., Hu, D., Chrisler, W. B., Tournay, R. J., Gomez-Rivas, E., Orr, G.,

- Ahkami, A. H., & Doty, S. L. (2024). Dynamic nitrogen fixation in an aerobic endophyte of *Populus*. *The ISME Journal*, wrad012. <https://doi.org/10.1093/ismejo/wrad012>
306. Shrestha, H., Yao, T., Qiao, Z., Muchero, W., Hettich, R. L., Chen, J.-G., & Abraham, P. E. (2023). Lectin receptor-like kinase signaling during engineered ectomycorrhiza colonization. *Cells*, 12(7), 1082. <https://doi.org/10.3390/cells12071082>
 307. Shrestha, H. K., Appidi, M. R., Villalobos Solis, M. I., Wang, J., Carper, D. L., Burdick, L., Pelletier, D. A., Doktycz, M. J., Hettich, R. L., & Abraham, P. E. (2021). Metaproteomics reveals insights into microbial structure, interactions, and dynamic regulation in defined communities as they respond to environmental disturbance. *BMC Microbiology*, 21(1), 308. <https://doi.org/10.1186/s12866-021-02370-4>
 308. Simon, S. J., Tschaplinski, T. J., LeBoldus, J. M., Keefover-Ring, K., Azeem, M., Chen, J., Macaya-Sanz, D., MacDonald, W. L., Muchero, W., & DiFazio, S. P. (2020). Host plant genetic control of associated fungal and insect species in a *Populus* hybrid cross. *Ecology and Evolution*, 10(11), 5119–5134. <https://doi.org/10.1002/ece3.6266>
 309. Søndreli, K. L., Keriö, S., Frost, K., Muchero, W., Chen, J.-G., Haiby, K., Gantz, C., Tuskan, G., & LeBoldus, J. M. (2020). Outbreak of Septoria canker caused by *Sphaerulina musiva* on *Populus trichocarpa* in eastern oregon. *Plant Disease*, 104(12), 3266–3266. <https://doi.org/10.1094/PDIS-03-20-0494-PDN>
 310. Stanosz, G. R., Smith, D. R., Ramsfield, T. D., McMahon, B. G., & Berguson, W. E. (2021). Response of balsam poplar to inoculation with *Marssonina balsamiferae* from Minnesota, USA. *Forest Pathology*, 51(5), e12714. <https://doi.org/10.1111/efp.12714>
 311. Šuran, J., Radić, B., Trevisan-Silva, D., Cindrić, M., & Hozic, A. (2023). First proteome analysis of poplar-type propolis. *Plant Foods for Human Nutrition*. <https://doi.org/10.1007/s11130-023-01127-w>
 312. Tabima, J. F., Søndreli, K. L., Keriö, S., Feau, N., Sakalidis, M. L., Hamelin, R. C., & LeBoldus, J. M. (2020). Population genomic analyses reveal connectivity via human-mediated transport across *Populus* plantations in North America and an undescribed subpopulation of *Sphaerulina musiva*. *Molecular Plant-Microbe Interactions*®, 33(2), 189–199. <https://doi.org/10.1094/MPMI-05-19-0131-R>
 313. Ullah, C., Chen, Y.-H., Ortega, M. A., & Tsai, C.-J. (2023). The diversity of salicylic acid biosynthesis and defense signaling in plants: Knowledge gaps and future opportunities. *Current Opinion in Plant Biology*, 72, 102349. <https://doi.org/10.1016/j.pbi.2023.102349>
 314. Ullah, C., Schmidt, A., Reichelt, M., Tsai, C., & Gershenzon, J. (2022). Lack of antagonism between salicylic acid and jasmonate signalling pathways in poplar. *New Phytologist*, 235(2), 701–717. <https://doi.org/10.1111/nph.18148>
 315. Van Nuland, M. E., Daws, S. C., Bailey, J. K., Schweitzer, J. A., Busby, P. E., & Peay, K. G. (2023). Above- and belowground fungal biodiversity of *Populus* trees on a continental scale. *Nature Microbiology*, 8(12), 2406–2419. <https://doi.org/10.1038/s41564-023-01514-8>
 316. Van Nuland, M. E., Ke, P., Wan, J., & Peay, K. G. (2023). Mycorrhizal nutrient acquisition strategies shape tree competition and coexistence dynamics. *Journal of Ecology*, 111(3), 564–577. <https://doi.org/10.1111/1365-2745.14040>

317. Van Nuland, M. E., Ware, I. M., Schadt, C. W., Yang, Z., Bailey, J. K., & Schweitzer, J. A. (2021). Natural soil microbiome variation affects spring foliar phenology with consequences for plant productivity and climate-driven range shifts. *New Phytologist*, 232(2), 762–775. <https://doi.org/10.1111/nph.17599>
318. Van Nuland, M. E., Vincent, J. B., Ware, I. M., Mueller, L. O., Bayliss, S. L. J., Beals, K. K., Schweitzer, J. A., & Bailey, J. K. (2020). Intraspecific trait variation across elevation predicts a widespread tree species' climate niche and range limits. *Ecology and Evolution*, 10(9), 3856–3867. <https://doi.org/10.1002/ece3.5969>
319. Varga, T., Hixson, K. K., Ahkami, A. H., Sher, A. W., Barnes, M. E., Chu, R. K., Battu, A. K., Nicora, C. D., Winkler, T. E., Reno, L. R., Fakra, S. C., Antipova, O., Parkinson, D. Y., Hall, J. R., & Doty, S. L. (2020). Endophyte-promoted phosphorus solubilization in *Populus*. *Frontiers in Plant Science*, 11, 567918. <https://doi.org/10.3389/fpls.2020.567918>
320. Veach, A. M., Chen, H., Yang, Z. K., Labbe, A. D., Engle, N. L., Tschaplinski, T. J., Schadt, C. W., & Cregger, M. A. (2020). Plant hosts modify belowground microbial community response to extreme drought. *mSystems*, 5(3), e00092-20. <https://doi.org/10.1128/mSystems.00092-20>
321. Vélez, J. M., Morris, R. M., Vilgalys, R., Labbé, J., & Schadt, C. W. (2021). Phylogenetic diversity of 200+ isolates of the ectomycorrhizal fungus *Cenococcum geophilum* associated with *Populus trichocarpa* soils in the Pacific Northwest, USA and comparison to globally distributed representatives. *PLOS ONE*, 16(1), e0231367. <https://doi.org/10.1371/journal.pone.0231367>
322. Vigneaud, J., Kohler, A., Sow, M. D., Delaunay, A., Fauchery, L., Guinet, F., Daviaud, C., Barry, K. W., Keymanesh, K., Johnson, J., Singan, V., Grigoriev, I., Fichot, R., Conde, D., Perales, M., Tost, J., Martin, F. M., Allona, I., Strauss, S. H., ... Maury, S. (2023). DNA hypomethylation of the host tree impairs interaction with mutualistic ectomycorrhizal fungus. *New Phytologist*, 238(6), 2561–2577. <https://doi.org/10.1111/nph.18734>
323. Villalobos Solis, M. I., Poudel, S., Bonnot, C., Shrestha, H. K., Hettich, R. L., Veneault-Fourrey, C., Martin, F., & Abraham, P. E. (2020). A viable new strategy for the discovery of peptide proteolytic cleavage products in plant-microbe interactions. *Molecular Plant-Microbe Interactions*®, 33(10), 1177–1188. <https://doi.org/10.1094/MPMI-04-20-0082-TA>
324. Voothuluru, P., Hamilton, C., Kim, K., Beason, J., McCord, J., Rousseau, R., Muchero, W., Rials, T., & Labbé, N. (2023). Bark morphological and chemical features are differentially correlated with disease resistance and yield in hybrid poplar taxa. *GCB Bioenergy*, 15(9), 1140–1153. <https://doi.org/10.1111/gcbb.13085>
325. Wagner, D., Wheeler, J. M., & Burr, S. J. (2020). The leaf miner *Phyllocnistis populiella* negatively impacts water relations in aspen. *Tree Physiology*, 40(5), 580–590. <https://doi.org/10.1093/treephys/tpz109>
326. Wang, J., Carper, D. L., Burdick, L. H., Shrestha, H. K., Appidi, M. R., Abraham, P. E., Timm, C. M., Hettich, R. L., Pelletier, D. A., & Doktycz, M. J. (2021). Formation, characterization, and modeling of emergent synthetic microbial communities. *Computational and Structural Biotechnology Journal*, 19, 1917–1927. <https://doi.org/10.1016/j.csbj.2021.03.034>
327. Wang, W., Bai, X.-D., Chen, K., Gu, C.-R., Yu, Q.-B., Jiang, J., & Liu, G.-F. (2022). Role of PsnWRKY70 in regulatory network response to infection with *Alternaria alternata* (Fr.) Keissl in

Populus. *International Journal of Molecular Sciences*, 23(14), 7537.

<https://doi.org/10.3390/ijms23147537>

328. Wei, H., Movahedi, A., Liu, G., Li, Y., Liu, S., Yu, C., Chen, Y., Zhong, F., & Zhang, J. (2022a). Comprehensive analysis of carotenoid cleavage dioxygenases gene family and its expression in response to abiotic stress in poplar. *International Journal of Molecular Sciences*, 23(3), 1418. <https://doi.org/10.3390/ijms23031418>
329. Wei, H., Movahedi, A., Liu, G., Li, Y., Liu, S., Yu, C., Chen, Y., Zhong, F., & Zhang, J. (2022b). Genome-wide characterization and abiotic stresses expression analysis of annexin family genes in poplar. *International Journal of Molecular Sciences*, 23(1), 515. <https://doi.org/10.3390/ijms23010515>
330. Wei, H., Movahedi, A., Liu, G., Zhu, S., Chen, Y., Yu, C., Zhong, F., & Zhang, J. (2022a). Characteristics, expression profile, and function of non-specific lipid transfer proteins of *Populus trichocarpa*. *International Journal of Biological Macromolecules*, 202, 468–481. <https://doi.org/10.1016/j.ijbiomac.2022.01.058>
331. Wei, H., Movahedi, A., Liu, G., Zhu, S., Chen, Y., Yu, C., Zhong, F., & Zhang, J. (2022b). Poplar glycosylphosphatidylinositol-anchored lipid transfer proteins respond to osmotic stress by regulating fatty acid biosynthesis. *Industrial Crops and Products*, 179, 114683. <https://doi.org/10.1016/j.indcrop.2022.114683>
332. Wei, H., Movahedi, A., Xu, S., Zhang, Y., Liu, G., Aghaei-Dargiri, S., Ghaderi Zefrehei, M., Zhu, S., Yu, C., Chen, Y., Zhong, F., & Zhang, J. (2022). Genome-wide characterization and expression analysis of fatty acid desaturase gene family in poplar. *International Journal of Molecular Sciences*, 23(19), 11109. <https://doi.org/10.3390/ijms231911109>
333. Wei, H., Movahedi, A., Yang, J., Zhang, Y., Liu, G., Zhu, S., Yu, C., Chen, Y., Zhong, F., & Zhang, J. (2022). Characteristics and molecular identification of glyceraldehyde-3-phosphate dehydrogenases in poplar. *International Journal of Biological Macromolecules*, 219, 185–198. <https://doi.org/10.1016/j.ijbiomac.2022.08.001>
334. Wei, H., Movahedi, A., Zhang, Y., Aghaei-Dargiri, S., Liu, G., Zhu, S., Yu, C., Chen, Y., Zhong, F., & Zhang, J. (2022). Long-chain acyl-CoA synthetases promote poplar resistance to abiotic stress by regulating long-chain fatty acid biosynthesis. *International Journal of Molecular Sciences*, 23(15), 8401. <https://doi.org/10.3390/ijms23158401>
335. Williams, J., Stella, J. C., Voelker, S. L., Lambert, A. M., Pelletier, L. M., Drake, J. E., Friedman, J. M., Roberts, D. A., & Singer, M. B. (2022). Local groundwater decline exacerbates response of dryland riparian woodlands to climatic drought. *Global Change Biology*, 28(22), 6771–6788. <https://doi.org/10.1111/gcb.16376>
336. Woods, H. A., Legault, G., Kingsolver, J. G., Pincebourde, S., Shah, A. A., & Larkin, B. G. (2022). Climate-driven thermal opportunities and risks for leaf miners in aspen canopies. *Ecological Monographs*, 92(4), e1544. <https://doi.org/10.1002/ecm.1544>
337. Wooley, S. C., Smith, D. S., Lonsdorf, E. V., Brown, S. C., Whitham, T. G., Shuster, S. M., & Lindroth, R. L. (2020). Local adaptation and rapid evolution of aphids in response to genetic interactions with their cottonwood hosts. *Ecology and Evolution*, 10(19), 10532–10542. <https://doi.org/10.1002/ece3.6709>

338. Xing, J., Li, P., Zhang, Y., Li, J., Liu, Y., Lachenbruch, B., Su, X., & Zhao, J. (2020). Fungal pathogens of canker disease trigger canopy dieback in poplar saplings by inducing functional failure of the phloem and cambium and carbon starvation in the xylem. *Physiological and Molecular Plant Pathology*, 112, 101523. <https://doi.org/10.1016/j.pmpp.2020.101523>
339. Yáñez-Segovia, S., Ramírez, C. C., Lindroth, R. L., & Fuentes-Contreras, E. (2023). Resistance against *Leucoptera sinuella* (Lepidoptera: Lyonetiidae) among hybrid clones of *Populus* spp. in central Chile. *Journal of Economic Entomology*, 116(5), 1662–1670. <https://doi.org/10.1093/jee/toad129>
340. Yu, X.-Y., Zhu, Y.-J., Wang, B., Liu, D., Bai, H., Jin, L., Wang, B.-T., Ruan, H.-H., Mao, L., Jin, F.-J., & Yang, N. (2021). Effects of nitrogen addition on rhizospheric soil microbial communities of poplar plantations at different ages. *Forest Ecology and Management*, 494, 119328. <https://doi.org/10.1016/j.foreco.2021.119328>
341. Zeigenfuss, L. C., & Schoenecker, K. A. (2021). Effects of elk and bison herbivory on narrowleaf cottonwood. *Western North American Naturalist*, 81(1). <https://doi.org/10.3398/064.081.0109>
342. Zhao, Y., Zheng, X., Tabima, J. F., Zhu, S., Søndreli, K. L., Hundley, H., Bauer, D., Barry, K., Zhang, Y., Schmutz, J., Wang, Y., LeBoldus, J. M., & Xiong, Q. (2023). Secreted effector proteins of poplar leaf spot and stem canker pathogen *Sphaerulina musiva* manipulate plant immunity and contribute to virulence in diverse ways. *Molecular Plant-Microbe Interactions*®, 36(12), 779–795. <https://doi.org/10.1094/MPMI-07-23-0091-R>
343. Zhang, F., Labourel, A., Haon, M., Kemppainen, M., Da Silva Machado, E., Brouilly, N., Veneault-Fourrey, C., Kohler, A., Rosso, M., Pardo, A., Henrissat, B., Berrin, J., & Martin, F. (2022). The ectomycorrhizal basidiomycete *Laccaria bicolor* releases a GH28 polygalacturonase that plays a key role in symbiosis establishment. *New Phytologist*, 233(6), 2534–2547. <https://doi.org/10.1111/nph.17940>
344. Zheng, Y., Lv, G. B., Chen, K., Yu, Q., Niu, B., Jiang, J., & Liu, G. (2022). Impact of *PaGLK* transgenic poplar on microbial community and soil enzyme activity in rhizosphere soil. *Frontiers in Bioengineering and Biotechnology*, 10, 965209. <https://doi.org/10.3389/fbioe.2022.965209>
345. Zhu, W., Zhao, D., Di, N., Li, D., Zhou, O., Sun, Y., Jia, L., Ding, C., & Xi, B. (2023). Matching root water uptake patterns to fine root and soil water distributions. *Plant and Soil*. <https://doi.org/10.1007/s11104-023-06349-0>

Production Systems for the Bioeconomy

346. Adler, A., Kumaniaev, I., Karacic, A., Baddigam, K. R., Hanes, R. J., Subbotina, E., Bartling, A. W., Huertas-Alonso, A. J., Moreno, A., Håkansson, H., Mathew, A. P., Beckham, G. T., & Samec, J. S. M. (2022). Lignin-first biorefining of Nordic poplar to produce cellulose fibers could displace cotton production on agricultural lands. *Joule*, 6(8), 1845–1858. <https://doi.org/10.1016/j.joule.2022.06.021>
347. Aggarwal, S., Magner, J., Srinivas, R., & Sajith, G. (2022). Managing nitrate-nitrogen in the intensively drained Upper Mississippi River Basin, USA under uncertainty: A perennial path

forward. *Environmental Monitoring and Assessment*, 194(10), 704. (cross-listed with *Salix*)
<https://doi.org/10.1007/s10661-022-10401-4>

348. Aghaaminiha, M., Mehrani, R., Reza, T., & Sharma, S. (2023). Comparison of machine learning methodologies for predicting kinetics of hydrothermal carbonization of selective biomass. *Biomass Conversion and Biorefinery*, 13(11), 9855–9864. <https://doi.org/10.1007/s13399-021-01858-3>
349. Alherech, M., Omolabake, S., Holland, C. M., Klinger, G. E., Hegg, E. L., & Stahl, S. S. (2021). From lignin to valuable aromatic chemicals: Lignin depolymerization and monomer separation via centrifugal partition chromatography. *ACS Central Science*, 7(11), 1831–1837. <https://doi.org/10.1021/acscentsci.1c00729>
350. Amarasekara, A. S., & Nwankwo, V. C. (2023). A comparison of dilute aqueous isethionic acid and sulfuric acid in hydrolysis of three different untreated lignocellulosic biomass varieties. *Industrial & Engineering Chemistry Research*, 62(46), 20037–20043. <https://doi.org/10.1021/acs.iecr.3c02314>
351. Andrews, H. B., Martin, M. Z., Wymore, A. M., & Kalluri, U. C. (2024). Rapid in situ nutrient element distribution in plants and soils using laser-induced breakdown spectroscopy (LIBS). *Plant and Soil*, 495(1–2), Article 1–2. <https://doi.org/10.1007/s11104-023-05988-7>
352. Andrews, H. B., Wymore, A. M., Wetter, E. E., Herndon, E. M., Li, H., Martin, S. A., Griffiths, N. A., Yang, X., Muchero, W., Weston, D. J., & Martin, M. Z. (2023). Rapid screening of wood and leaf tissues: Investigating silicon-based phytoliths in *Populus trichocarpa* for carbon storage applications using laser-induced breakdown spectroscopy and scanning electron microscopy–energy dispersive X-ray spectroscopy. *Journal of Analytical Atomic Spectrometry*, 38(11), Article 11. <https://doi.org/10.1039/D3JA00186E>
353. Aubrey, D. P., & Teskey, R. O. (2021). Xylem transport of root-derived CO₂ caused a substantial underestimation of belowground respiration during a growing season. *Global Change Biology*, 27(12), 2991–3000. <https://doi.org/10.1111/gcb.15624>
354. Azad, T., Auad, M. L., Elder, T., & Adamczyk, A. J. (2023). Toward native hardwood lignin pyrolysis: Insights into reaction energetics from density functional theory. *Energy & Fuels*, 37(1), 401–423. <https://doi.org/10.1021/acs.energyfuels.2c03025>
355. Baer, A. B., Fickle, J. C., Medina, J., Robles, C., Pratt, R. B., & Jacobsen, A. L. (2021). Xylem biomechanics, water storage, and density within roots and shoots of an angiosperm tree species. *Journal of Experimental Botany*, 72(22), 7984–7997. <https://doi.org/10.1093/jxb/erab384>
356. Bai, Y., Liu, Y., Kueppers, L. M., Li, E., Zhang, C., Yu, K., Yang, X., & Li, X. (2022). Hydraulic sensitivity and stomatal regulation of two desert riparian species. *Journal of Geophysical Research: Biogeosciences*, 127(10), e2022JG006971. <https://doi.org/10.1029/2022JG006971>
357. Balmant, K. M., Noble, J. D., Alves, F. C., Dervinis, C., Conde, D., Schmidt, H. W., Vazquez, A. I., Barbazuk, W. B., Campos, G. D. L., Resende, M. F. R., & Kirst, M. (2020). Xylem systems genetics analysis reveals a key regulator of lignin biosynthesis in *Populus deltoides*. *Genome Research*, 30(8), 1131–1143. <https://doi.org/10.1101/gr.261438.120>

358. Bay, M. S., Karimi, K., Esfahany, M. N., & Kumar, R. (2020). Structural modification of pine and poplar wood by alkali pretreatment to improve ethanol production. *Industrial Crops and Products*, 152, 112506. <https://doi.org/10.1016/j.indcrop.2020.112506>
359. Berrill, J.-P., Dagley, C. M., Kim, Y. G., & Morgan Varner, J. (2024). Pile burning after conifer removal from aspen stands affects tree mortality, regeneration, and understory recovery. *Forest Ecology and Management*, 553, 121602. <https://doi.org/10.1016/j.foreco.2023.121602>
360. Beverly, D. P., Guadagno, C. R., & Ewers, B. E. (2020). Biophysically informed imaging acquisition of plant water status. *Frontiers in Forests and Global Change*, 3, 589493. <https://doi.org/10.3389/ffgc.2020.589493>
361. Bhagia, S., Đurković, J., Lagaña, R., Kardošová, M., Kačík, F., Cernescu, A., Schäfer, P., Yoo, C. G., & Ragauskas, A. J. (2022). Nanoscale FTIR and mechanical mapping of plant cell walls for understanding biomass deconstruction. *ACS Sustainable Chemistry & Engineering*, 10(9), 3016–3026. <https://doi.org/10.1021/acssuschemeng.1c08163>
362. Bhagia, S., & Ragauskas, A. J. (2020). Preserving aryl ether linkages and higher yields of isolated lignin through biomass fibrillation. *ACS Sustainable Chemistry & Engineering*, 8(1), 34–37. <https://doi.org/10.1021/acssuschemeng.9b06390>
363. Bhagia, S., Dunlap, J. R., Khuraishi, M. Z. A., Lowden, R. R., Muchero, W., Vaidya, U. K., Pu, Y., & Ragauskas, A. J. (2020). Fabrication of lignocellulosic biomass paper containing nanofibrillated biomass. *BioResources*, 16(1), 209–222. <https://doi.org/10.15376/biores.16.1.209-222>
364. Bhagia, S., Lowden, R. R., Erdman, D., Rodriguez, M., Haga, B. A., Solano, I. R. M., Gallego, N. C., Pu, Y., Muchero, W., Kunc, V., & Ragauskas, A. J. (2020). Tensile properties of 3D-printed wood-filled PLA materials using poplar trees. *Applied Materials Today*, 21, 100832. <https://doi.org/10.1016/j.apmt.2020.100832>
365. Bing, R. G., Straub, C. T., Sulis, D. B., Wang, J. P., Adams, M. W. W., & Kelly, R. M. (2022). Plant biomass fermentation by the extreme thermophile *Caldicellulosiruptor bescii* for co-production of green hydrogen and acetone: Technoeconomic analysis. *Bioresource Technology*, 348, 126780. <https://doi.org/10.1016/j.biortech.2022.126780>
366. Birch, J. D., Chikamoto, Y., DeRose, R. J., Manvailer, V., Hogg, E. H., Karst, J., Love, D. M., & Lutz, J. A. (2023). Frost-associated defoliation in *Populus tremuloides* causes repeated growth reductions over 185 years. *Ecosystems*, 26(4), 843–859. <https://doi.org/10.1007/s10021-022-00799-w>
367. Biswal, A. K., Hengge, N. N., Black, I. M., Atmodjo, M. A., Mohanty, S. S., Ryno, D., Himmel, M. E., Azadi, P., Bomble, Y. J., & Mohnen, D. (2022). Composition and yield of non-cellulosic and cellulosic sugars in soluble and particulate fractions during consolidated bioprocessing of poplar biomass by *Clostridium thermocellum*. *Biotechnology for Biofuels and Bioproducts*, 15(1), 23. <https://doi.org/10.1186/s13068-022-02119-9>
368. Biswas, R., Teller, P. J., Khan, M. U., & Ahring, B. K. (2020). Sugar production from hybrid poplar sawdust: Optimization of enzymatic hydrolysis and wet explosion pretreatment. *Molecules*, 25(15), 3396. <https://doi.org/10.3390/molecules25153396>
369. Blasini, D. E., Koepke, D. F., Bush, S. E., Allan, G. J., Gehring, C. A., Whitham, T. G., Day, T. A., & Hultine, K. R. (2022). Tradeoffs between leaf cooling and hydraulic safety in a dominant arid

land riparian tree species. *Plant, Cell & Environment*, 45(6), 1664–1681.

<https://doi.org/10.1111/pce.14292>

370. Blasini, D. E., Koepke, D. F., Grady, K. C., Allan, G. J., Gehring, C. A., Whitham, T. G., Cushman, S. A., & Hultine, K. R. (2021). Adaptive trait syndromes along multiple economic spectra define cold and warm adapted ecotypes in a widely distributed foundation tree species. *Journal of Ecology*, 109(3), 1298–1318. <https://doi.org/10.1111/1365-2745.13557>
371. Blonder, B. W., Brodrick, P. G., Chadwick, K. D., Carroll, E., Cruz-de Hoyos, R. M., Expósito-Alonso, M., Hateley, S., Moon, M., Ray, C. A., Tran, H., & Walton, J. A. (2023). Climate lags and genetics determine phenology in quaking aspen (*Populus tremuloides*). *New Phytologist*, 238(6), 2313–2328. <https://doi.org/10.1111/nph.18850>
372. Blonder, B., Brodrick, P. G., Walton, J. A., Chadwick, K. D., Breckheimer, I. K., Marchetti, S., Ray, C. A., & Mock, K. E. (2022). Remote sensing of cytotype and its consequences for canopy damage in quaking aspen. *Global Change Biology*, 28(7), 2491–2504. <https://doi.org/10.1111/gcb.16064>
373. Blonder, B., Ray, C. A., Walton, J. A., Castaneda, M., Chadwick, K. D., Clyne, M. O., Gaüzère, P., Iversen, L. L., Lusk, M., Strimbeck, G. R., Troy, S., & Mock, K. E. (2021). Cytotype and genotype predict mortality and recruitment in Colorado quaking aspen (*Populus tremuloides*). *Ecological Applications*, 31(8), e02438. <https://doi.org/10.1002/eap.2438>
374. Blumstein, M., Sala, A., Weston, D. J., Holbrook, N. M., & Hopkins, R. (2022). Plant carbohydrate storage: Intra- and inter-specific trade-offs reveal a major life history trait. *New Phytologist*, 235(6), 2211–2222. <https://doi.org/10.1111/nph.18213>
375. Blumstein, M., & Hopkins, R. (2021). Adaptive variation and plasticity in non-structural carbohydrate storage in a temperate tree species. *Plant, Cell & Environment*, 44(8), 2494–2505. <https://doi.org/10.1111/pce.13959>
376. Boča, A., Jacobson, A. R., & Van Miegroet, H. (2020). Aspen soils retain more dissolved organic carbon than conifer soils in a sorption experiment. *Frontiers in Forests and Global Change*, 3, 594473. <https://doi.org/10.3389/ffgc.2020.594473>
377. Borukanlu, M. R., Zadeh, O. H., Moradpour, P., & Khedive, E. (2021). Effects of growth rate of eastern poplar trees on the chemical and morphological characteristics of wood fibers. *European Journal of Wood and Wood Products*, 79(6), 1479–1494. <https://doi.org/10.1007/s00107-021-01711-4>
378. Brédoire, F., Kayler, Z. E., Dupouey, J.-L., Derrien, D., Zeller, B., Barsukov, P. A., Rusalimova, O., Nikitich, P., Bakker, M. R., & Legout, A. (2020). Limiting factors of aspen radial growth along a climatic and soil water budget gradient in south-western Siberia. *Agricultural and Forest Meteorology*, 282–283, 107870. <https://doi.org/10.1016/j.agrformet.2019.107870>
379. Bryant, N., Engle, N., Tschaplinski, T., Pu, Y., & Ragauskas, A. J. (2023). Variable lignin structure revealed in *Populus* leaves. *RSC Advances*, 13(29), 20187–20197. <https://doi.org/10.1039/D3RA03142J>
380. Bryant, N., G. Yoo, C., Pu, Y., & Ragauskas, A. J. (2020). 2D HSQC chemical shifts of impurities from biomass pretreatment. *ChemistrySelect*, 5(11), 3359–3364. <https://doi.org/10.1002/slct.202000406>

381. Buckardt Thomas, A., McNeil, D. J., Larkin, J. L., Johnson, K. E., & Roth, A. M. (2023). Evaluating golden-winged warbler use of alder and aspen communities managed with shearing in the western Great Lakes. *Ecosphere*, 14(3), e4443. <https://doi.org/10.1002/ecs2.4443>
382. Budsberg, E., Parker, N., Bandaru, V., Bura, R., & Gustafson, R. (2022). Hydrocarbon bio-jet fuel from bioconversion of poplar biomass: Life cycle assessment of site-specific impacts. *Forests*, 13(4), 549. <https://doi.org/10.3390/f13040549>
383. Budsberg, E., Morales-Vera, R., Crawford, J. T., Bura, R., & Gustafson, R. (2020). Production routes to bio-acetic acid: Life cycle assessment. *Biotechnology for Biofuels*, 13(1), 154. <https://doi.org/10.1186/s13068-020-01784-y>
384. Buell, C. R., Dardick, C., Parrott, W., Schmitz, R. J., Shih, P. M., Tsai, C.-J., & Urbanowicz, B. (2023). Engineering custom morpho- and chemotypes of *Populus* for sustainable production of biofuels, bioproducts, and biomaterials. *Frontiers in Plant Science*, 14, 1288826. <https://doi.org/10.3389/fpls.2023.1288826>
385. Bustamante-Bailon, A. P. S., Margenot, A., Cooke, R. A. C., & Christianson, L. E. (2022). Phosphorus removal in denitrifying woodchip bioreactors varies by wood type and water chemistry. *Environmental Science and Pollution Research*, 29(5), 6733–6743. <https://doi.org/10.1007/s11356-021-15835-w>
386. Bywater-Reyes, S., Diehl, R. M., Wilcox, A. C., Stella, J. C., & Kui, L. (2022). A green new balance: Interactions among riparian vegetation plant traits and morphodynamics in alluvial rivers. *Earth Surface Processes and Landforms*, 47(10), 2410–2436. <https://doi.org/10.1002/esp.5385>
387. Chen, S., Wang, J., Liu, Y., Chen, Z., Lei, Y., & Yan, L. (2022). The relationship between color and mechanical properties of heat-treated wood predicted based on support vector machines model. *Holzforschung*, 76(11–12), 994–1002. <https://doi.org/10.1515/hf-2022-0075>
388. Chen, Y., Wang, Z., Chen, G., Wang, Q., Sun, T., Zhang, M., Du, Z., Wu, M., Guo, S., Lei, T., Burra, K. G., & Gupta, A. K. (2024). Synergistic effects and products yield analyses based on co-pyrolysis of poplar tree and rape stalks with polyethylene terephthalate and polypropylene. *Journal of the Energy Institute*, 112, 101461. <https://doi.org/10.1016/j.joei.2023.101461>
389. Cheng, F., Liu, S., Karlen, S. D., Kim, H., Lu, F., Ralph, J., Ramos, L. M. V., Huber, G. W., & Dumesic, J. A. (2023). Poplar lignin structural changes during extraction in γ -valerolactone (GVL). *Green Chemistry*, 25(1), 336–347. <https://doi.org/10.1039/D2GC03446H>
390. Cho, J.-S., Kim, M.-H., Bae, E.-K., Choi, Y.-I., Jeon, H.-W., Han, K.-H., & Ko, J.-H. (2021). Field evaluation of transgenic hybrid poplars with desirable wood properties and enhanced growth for biofuel production by bicistronic expression of *PdGA20ox1* and *PtrMYB3* in wood-forming tissue. *Biotechnology for Biofuels*, 14(1), 177. <https://doi.org/10.1186/s13068-021-02029-2>
391. Chowyuk, A. N., El-Husseini, H., Gustafson, R. R., Parker, N., Bura, R., & Gough, H. L. (2021). Economics of growing poplar for the dual purpose of biorefinery feedstock and wastewater treatment. *Biomass and Bioenergy*, 153, 106213. <https://doi.org/10.1016/j.biombioe.2021.106213>

392. Christensen, C., Katz, G., Friedman, J., Redmond, M. D., & Norton, A. (2023). No evidence for cottonwood forest decline along a flow-augmented western U.S. river. *River Research and Applications*, 39(8), 1602–1615. <https://doi.org/10.1002/rra.4151>
393. Clay, C., Nave, L., Nadelhoffer, K., Vogel, C., Propson, B., Den Uyl, J., Hickey, L. J., Barry, A., & Gough, C. M. (2022). Fire after clear-cut harvesting minimally affects the recovery of ecosystem carbon pools and fluxes in a Great Lakes forest. *Forest Ecology and Management*, 519, 120301. <https://doi.org/10.1016/j.foreco.2022.120301>
394. Collins, H. P., Kimura, E., Polley, W., Fay, P. A., & Fransen, S. (2020). Intercropping switchgrass with hybrid poplar increased carbon sequestration on a sand soil. *Biomass and Bioenergy*, 138, 105558. <https://doi.org/10.1016/j.biombioe.2020.105558>
395. Curzon, M. T., Kern, C. C., Baker, S. C., Palik, B. J., & D'Amato, A. W. (2020). Retention forestry influences understory diversity and functional identity. *Ecological Applications*, 30(5), e02097. <https://doi.org/10.1002/eap.2097>
396. Cusack, C. J., Fernald, A. G., VanLeeuwen, D. M., Guldán, S. J., Baker, T. T., & Heyduck, R. F. (2022). Supporting riparian habitat with traditional irrigation systems of the northern Rio Grande Region. *The Southwestern Naturalist*, 66(4). <https://doi.org/10.1894/0038-4909-66.4.280>
397. Dagley, C. M., Berrill, J., Coppeto, S. A., & Eschtruth, A. K. (2020). Understory responses to restoration in aspen-conifer forests around the Lake Tahoe Basin: Residual stand attributes predict recovery. *Restoration Ecology*, 28(3), 603–611. <https://doi.org/10.1111/rec.13121>
398. Dahal, B., Poudel, K. P., Renninger, H. J., Granger, J. J., Leininger, T. D., Gardiner, E. S., Souter, R. A., & Rousseau, R. J. (2022). Aboveground biomass equations for black willow (*Salix nigra* Marsh.) and eastern cottonwood (*Populus deltoides* Bartr. Ex Marsh.). *Trees, Forests and People*, 7, 100195. (cross-listed with *Salix*) <https://doi.org/10.1016/j.tfp.2022.100195>
399. Danise, T., Andriuzzi, W. S., Battipaglia, G., Certini, G., Guggenberger, G., Innangi, M., Mastrolonardo, G., Niccoli, F., Pelleri, F., & Fioretto, A. (2021). Mixed-species plantation effects on soil biological and chemical quality and tree growth of a former agricultural land. *Forests*, 12(7), 842. <https://doi.org/10.3390/f12070842>
400. Davidson, K. J., Lamour, J., Rogers, A., & Serbin, S. P. (2022). Late-day measurement of excised branches results in uncertainty in the estimation of two stomatal parameters derived from response curves in *Populus deltoides* Bartr. × *Populus nigra* L. *Tree Physiology*, 42(7), 1377–1395. <https://doi.org/10.1093/treephys/tpac006>
401. De Meester, B., Oyarce, P., Vanholme, R., Van Acker, R., Tsuji, Y., Vangeel, T., Van den Bosch, S., Van Doorselaere, J., Sels, B., Ralph, J., & Boerjan, W. (2022). Engineering curcumin biosynthesis in poplar affects lignification and biomass yield. *Frontiers in Plant Science*, 13, 943349. <https://doi.org/10.3389/fpls.2022.943349>
402. De Meester, B., Madariaga Calderón, B., de Vries, L., Pollier, J., Goeminne, G., Van Doorselaere, J., Chen, M., Ralph, J., Vanholme, R., & Boerjan, W. (2020). Tailoring poplar lignin without yield penalty by combining a null and haploinsufficient *CINNAMOYL-CoA REDUCTASE2* allele. *Nature Communications*, 11(1), 5020. <https://doi.org/10.1038/s41467-020-18822-w>

403. Dewhirst, R. A., Handakumbura, P., Clendinen, C. S., Arm, E., Tate, K., Wang, W., Washton, N. M., Young, R. P., Mortimer, J. C., McDowell, N. G., & Jardine, K. J. (2021). High temperature acclimation of leaf gas exchange, photochemistry, and metabolomic profiles in *Populus trichocarpa*. *ACS Earth and Space Chemistry*, 5(8), Article 8. <https://doi.org/10.1021/acsearthspacechem.0c00299>
404. Dewhirst, R. A., Lei, J., Afseth, C. A., Castanha, C., Wistrom, C. M., Mortimer, J. C., & Jardine, K. J. (2021). Are methanol-derived foliar methyl acetate emissions a tracer of acetate-mediated drought survival in plants? *Plants*, 10(2), 411. <https://doi.org/10.3390/plants10020411>
405. Dewhirst, R. A., Afseth, C. A., Castanha, C., Mortimer, J. C., & Jardine, K. J. (2020). Cell wall O-acetyl and methyl esterification patterns of leaves reflected in atmospheric emission signatures of acetic acid and methanol. *PLOS ONE*, 15(5), e0227591. <https://doi.org/10.1371/journal.pone.0227591>
406. Dobarco, M. R., Jacobson, A. R., & Van Miegroet, H. (2021). Chemical composition of soil organic carbon from mixed aspen-conifer forests characterized with fourier transform infrared spectroscopy. *European Journal of Soil Science*, 72(3), 1410–1430. <https://doi.org/10.1111/ejss.13065>
407. Edmunds, C. W., Peralta, P., Sharma-Shivappa, R. R., Kelley, S. S., Chiang, V. L., Miller, Z. D., Giles, R. L., Sykes, R. W., Deoppke, C., Gjersing, E., Davis, M. F., & Peszlen, I. (2020). Fungal pretreatment and enzymatic hydrolysis of genetically-modified *Populus trichocarpa*. *BioResources*, 15(3), 6488–6505. <https://doi.org/10.15376/biores.15.3.6488-6505>
408. Englund, O., Dimitriou, I., Dale, V. H., Kline, K. L., Mola-Yudego, B., Murphy, F., English, B., McGrath, J., Busch, G., Negri, M. C., Brown, M., Goss, K., Jackson, S., Parish, E. S., Cacho, J., Zumpf, C., Quinn, J., & Mishra, S. K. (2020). Multifunctional perennial production systems for bioenergy: Performance and progress. *WIREs Energy and Environment*, 9(5), e375. (cross-listed with *Salix* and *Eucalyptus*) <https://doi.org/10.1002/wene.375>
409. Falk, M. A., Donaldson, J. R., Stevens, M. T., Raffa, K. F., & Lindroth, R. L. (2020). Phenological responses to prior-season defoliation and soil-nutrient availability vary among early- and late-flushing aspen (*Populus tremuloides* Michx.) genotypes. *Forest Ecology and Management*, 458, 117771. <https://doi.org/10.1016/j.foreco.2019.117771>
410. Farahi, R. H., Lereu, A. L., Charrier, A. M., Kalluri, U. C., Davison, B. H., & Passian, A. (2020). Nanomechanics and Raman spectroscopy of in situ native carbohydrate storage granules for enhancing starch quality and lignocellulosic biomass production. *ACS Omega*, 5(6), 2594–2602. <https://doi.org/10.1021/acsomega.9b02849>
411. Ferreira, G. W. D., Coleman, M. D., Coyle, D. R., & Aubrey, D. P. (2023). Whole-tree dormant season nitrogen pools for different species receiving combinations of fertilization and irrigation after one short rotation. *Forest Ecology and Management*, 529, 120730. <https://doi.org/10.1016/j.foreco.2022.120730>
412. Fu, J., Zou, S., Coleman, M., Li, X., Hu, W., Wang, A., Zhang, P., Zeng, Z., Ding, C., Xi, B., & Di, N. (2022). Is it necessary to apply chemical weed control in short-rotation poplar plantations on deep soil sites? *Industrial Crops and Products*, 184, 115025. <https://doi.org/10.1016/j.indcrop.2022.115025>

413. Gamble, J. D., Johnson, G. A., Current, D. A., Wyse, D. L., Zamora, D. S., & Sheaffer, C. C. (2020). Alley cropping affects perennial bioenergy crop root distribution, carbon, and nutrient stocks. *Agronomy Journal*, 112(5), 3718–3732. (cross-listed with *Salix*) <https://doi.org/10.1002/agj2.20350>
414. Geiselman, G. M., Kirby, J., Landera, A., Otoupal, P., Papa, G., Barcelos, C., Sundstrom, E. R., Das, L., Magurudeniya, H. D., Wehrs, M., Rodriguez, A., Simmons, B. A., Magnuson, J. K., Mukhopadhyay, A., Lee, T. S., George, A., & Gladden, J. M. (2020). Conversion of poplar biomass into high-energy density tricyclic sesquiterpene jet fuel blendstocks. *Microbial Cell Factories*, 19(1), 208. <https://doi.org/10.1186/s12934-020-01456-4>
415. Gelfand, I., Hamilton, S. K., Kravchenko, A. N., Jackson, R. D., Thelen, K. D., & Robertson, G. P. (2020). Empirical evidence for the potential climate benefits of decarbonizing light vehicle transport in the U.S. with bioenergy from purpose-grown biomass with and without BECCS. *Environmental Science & Technology*, 54(5), 2961–2974. <https://doi.org/10.1021/acs.est.9b07019>
416. Geng, Q., Ma, X., Liao, J., Wu, W., Niu, S., Luo, Y., & Xu, X. (2021). Contrasting nutrient-mediated responses between surface and deep fine root biomass to N addition in poplar plantations on the east coast of China. *Forest Ecology and Management*, 490, 119152. <https://doi.org/10.1016/j.foreco.2021.119152>
417. Geng, W., Venditti, R. A., Pawlak, J. J., Chang, H., Pal, L., & Ford, E. (2020). Carboxymethylation of hemicellulose isolated from poplar (*Populus grandidentata*) and its potential in water-soluble oxygen barrier films. *Cellulose*, 27(6), 3359–3377. <https://doi.org/10.1007/s10570-020-02993-2>
418. Ghezehei, S. B., & Saloni, D. (2023). Assessment of feedstock quality of poplars (*Populus* L.) using selected pellet-quality parameters. *BioEnergy Research*, 17(2), 993–1003. <https://doi.org/10.1007/s12155-023-10711-6>
419. Ghezehei, S. B., Ewald, A. L., Hazel, D. W., Zalesny, R. S., & Nichols, E. G. (2021). Productivity and profitability of poplars on fertile and marginal sandy soils under different density and fertilization treatments. *Forests*, 12(7), 869. <https://doi.org/10.3390/f12070869>
420. Ghezehei, S. B., Wright, J., Zalesny, R. S., Nichols, E. G., & Hazel, D. W. (2020). Matching site-suitable poplars to rotation length for optimized productivity. *Forest Ecology and Management*, 457, 117670. <https://doi.org/10.1016/j.foreco.2019.117670>
421. Ghezehei, S., Hazel, D., & Nichols, E. (2020). SRWC-PEAM: A comprehensive appraisal tool for short-rotation woody crops in the southeast. *Journal of Extension*, 58(5). <https://doi.org/10.34068/joe.58.05.07>
422. Gifford, T. S., Zobel, J. M., Hillard, S. C., & Edgar, C. B. (2023). Comprehensive review and evaluation of whole stand volume yield models for the aspen forest type in Minnesota, USA. *Forestry*, 96(4), 561–574. <https://doi.org/10.1093/forestry/cpac061>
423. Goacher, R. E., Mottiar, Y., & Mansfield, S. D. (2021). ToF-SIMS imaging reveals that *p* - hydroxybenzoate groups specifically decorate the lignin of fibres in the xylem of poplar and willow. *Holzforschung*, 75(5), 452–462. (cross-listed with *Salix*) <https://doi.org/10.1515/hf-2020-0130>

424. Gregory, S. A., McGettigan, C. P., McGuinness, E. K., Rodin, D. M., Yee, S. K., & Losego, M. D. (2020). Single-cycle atomic layer deposition on bulk wood lumber for managing moisture content, mold growth, and thermal conductivity. *Langmuir*, 36(7), 1633–1641. <https://doi.org/10.1021/acs.langmuir.9b03273>
425. Grinde, A. R., Slesak, R. A., D'Amato, A. W., & Palik, B. P. (2020). Effects of tree retention and woody biomass removal on bird and small mammal communities. *Forest Ecology and Management*, 465, 118090. <https://doi.org/10.1016/j.foreco.2020.118090>
426. Guo, J., Ren, X., Li, S., Huang, Z., Manzo, M., Cai, L., & Chang, J. (2023). The impact of blending with poplar wood on the co-pyrolysis characteristics of waste particleboards. *Biomass Conversion and Biorefinery*, 13(6), 4949–4956. <https://doi.org/10.1007/s13399-021-01520-y>
427. Guo, Y., Wang, S., Yu, K., Wang, H.-L., Xu, H., Song, C., Zhao, Y., Wen, J., Fu, C., Li, Y., Wang, S., Zhang, X., Zhang, Y., Cao, Y., Shao, F., Wang, X., Deng, X., Chen, T., Zhao, Q., ... Lin, J. (2023). Manipulating microRNA *mir408* enhances both biomass yield and saccharification efficiency in poplar. *Nature Communications*, 14(1), Article 1. <https://doi.org/10.1038/s41467-023-39930-3>
428. Guo, Y., Tian, Z., Ji, X., Lyu, G., Chen, J., Yang, G., & Lucia, L. A. (2020). Staged alkali and hydrogen peroxide treatment of poplar chemi-mechanical pulp. *BioResources*, 15(1), 1062–1073. <https://doi.org/10.15376/biores.15.1.1062-1073>
429. Haftkhani, A. R., & Hematabadi, H. (2022). Effect of layer arrangement on bending strength of cross-laminated timber (CLT) manufactured from Poplar (*Populus deltoides* L.). *Buildings*, 12(5), 608. <https://doi.org/10.3390/buildings12050608>
430. Halsted, M. C., Bible, A. N., Morrell-Falvey, J. L., & Retterer, S. T. (2022). Quantifying biofilm propagation on chemically modified surfaces. *Biofilm*, 4, 100088. <https://doi.org/10.1016/j.biofilm.2022.100088>
431. Happs, R. M., Bartling, A. W., Doepcke, C., Harman-Ware, A. E., Clark, R., Webb, E. G., Biddy, M. J., Chen, J., Tuskan, G. A., Davis, M. F., Muchero, W., & Davison, B. H. (2021). Economic impact of yield and composition variation in bioenergy crops: *Populus trichocarpa*. *Biofuels, Bioproducts and Biorefining*, 15(1), 176–188. <https://doi.org/10.1002/bbb.2148>
432. Hart, A. T., Merlin, M., Wiley, E., & Landhäusser, S. M. (2021). Splitting the difference: Heterogeneous soil moisture availability affects aboveground and belowground reserve and mass allocation in trembling aspen. *Frontiers in Plant Science*, 12, 654159. <https://doi.org/10.3389/fpls.2021.654159>
433. He, D., Zhuang, J., Jiang, Y., Xie, D., Yoo, C. G., & Yang, Q. (2021). Fractionation of poplar wood using a bifunctional aromatic acid under mild conditions. *ACS Sustainable Chemistry & Engineering*, 9(15), 5364–5376. <https://doi.org/10.1021/acssuschemeng.1c00123>
434. He, D., Wang, Y., Yoo, C. G., Chen, Q.-J., & Yang, Q. (2020). The fractionation of woody biomass under mild conditions using bifunctional phenol-4-sulfonic acid as a catalyst and lignin solvent. *Green Chemistry*, 22(16), 5414–5422. <https://doi.org/10.1039/D0GC01722A>
435. He, Y., Xi, B., Li, G., Wang, Y., Jia, L., & Zhao, D. (2021). Influence of drip irrigation, nitrogen fertigation, and precipitation on soil water and nitrogen distribution, tree seasonal growth and nitrogen uptake in young triploid poplar (*Populus tomentosa*) plantations. *Agricultural Water Management*, 243, 106460. <https://doi.org/10.1016/j.agwat.2020.106460>

436. He, Y., Xi, B., Bloomberg, M., Jia, L., & Zhao, D. (2020). Effects of drip irrigation and nitrogen fertigation on stand growth and biomass allocation in young triploid *Populus tomentosa* plantations. *Forest Ecology and Management*, 461, 117937. <https://doi.org/10.1016/j.foreco.2020.117937>
437. Hematabadi, H., Madhoushi, M., Khazaeian, A., & Ebrahimi, G. (2021). Structural performance of hybrid poplar-beech cross-laminated-timber (clt). *Journal of Building Engineering*, 44, 102959. <https://doi.org/10.1016/j.jobbe.2021.102959>
438. Hematabadi, H., Madhoushi, M., Khazaeyan, A., Ebrahimi, G., Hindman, D., & Loferski, J. (2020). Bending and shear properties of cross-laminated timber panels made of poplar (*Populus alba*). *Construction and Building Materials*, 265, 120326. <https://doi.org/10.1016/j.conbuildmat.2020.120326>
439. Henner, D. N., Hastings, A., Pogson, M., McNamara, N. P., Davies, C. A., & Smith, P. (2020). PopFor: A new model for estimating poplar yields. *Biomass and Bioenergy*, 134, 105470. <https://doi.org/10.1016/j.biombioe.2020.105470>
440. Henry, C., Tindall, G., Thies, M. C., & Nejad, M. (2023). Fractionated and purified hybrid poplar lignins as a polyol replacement in rigid polyurethane/polyisocyanurate foams. *Journal of Applied Polymer Science*, 140(45), e54648. <https://doi.org/10.1002/app.54648>
441. Hillman, E. T., Li, M., Hooker, C. A., Englaender, J. A., Wheeldon, I., & Solomon, K. V. (2021). Hydrolysis of lignocellulose by anaerobic fungi produces free sugars and organic acids for two-stage fine chemical production with *Kluyveromyces marxianus*. *Biotechnology Progress*, 37(5), e3172. <https://doi.org/10.1002/btpr.3172>
442. Himes, A., Leavengood, S., & Polinko, A. (2021). Variation in wood properties of hybrid poplar lumber by radial and vertical position in stem: A case study from Boardman, Oregon. *Wood and Fiber Science*, 53(3), 161–177. <https://doi.org/10.22382/wfs-2021-17>
443. Hogan, J. A., Baraloto, C., Ficken, C., Clark, M. D., Weston, D. J., & Warren, J. M. (2021). The physiological acclimation and growth response of *Populus trichocarpa*. *Physiologia Plantarum*, 173(3), 1008–1029. <https://doi.org/10.1111/ppl.13498>
444. Hori, C., Takata, N., Lam, P. Y., Tobimatsu, Y., Nagano, S., Mortimer, J. C., & Cullen, D. (2020). Identifying transcription factors that reduce wood recalcitrance and improve enzymatic degradation of xylem cell wall in *Populus*. *Scientific Reports*, 10(1), 22043. <https://doi.org/10.1038/s41598-020-78781-6>
445. Howe, A. A., Landhäusser, S. M., Burney, O. T., Long, J. N., & Mock, K. E. (2020). Regional differences in aspen (*Populus tremuloides* Michx.) seedling response to an established nursery protocol. *New Forests*, 51(2), 367–378. <https://doi.org/10.1007/s11056-019-09727-8>
446. Howe, A. A., Landhäusser, S. M., Burney, O. T., Long, J. N., Violett, R. D., & Mock, K. E. (2020). Exploring seedling-based aspen (*Populus tremuloides*) restoration near range limits in the Intermountain West, USA. *Forest Ecology and Management*, 476, 118470. <https://doi.org/10.1016/j.foreco.2020.118470>
447. Huang, K., Mohan, M., George, A., Simmons, B. A., Xu, Y., & Gladden, J. M. (2021). Integration of acetic acid catalysis with one-pot protic ionic liquid configuration to achieve high-efficient

biorefinery of poplar biomass. *Green Chemistry*, 23(16), 6036–6049.
<https://doi.org/10.1039/D1GC01727F>

448. Hunter, C., Sun, Z., Mansfield, S. D., Shahbaz, M., Pilon, M., & Gleason, S. M. (2023). The effects of copper deficiency on lignification, xylem vessel structure, and hydraulic traits in hybrid poplar. *Physiologia Plantarum*, 175(5), e14006. <https://doi.org/10.1111/ppl.14006>
449. Hunter, C., Stewart, J. J., Gleason, S. M., & Pilon, M. (2022). Age dependent partitioning patterns of essential nutrients induced by copper feeding status in leaves and stems of poplar. *Frontiers in Plant Science*, 13, 930344. <https://doi.org/10.3389/fpls.2022.930344>
450. Hunter, C., Ware, M. A., Gleason, S. M., Pilon-Smits, E., & Pilon, M. (2022). Recovery after deficiency: Systemic copper prioritization and partitioning in the leaves and stems of hybrid poplar. *Tree Physiology*, 42(9), 1776–1785. <https://doi.org/10.1093/treephys/tpac038>
451. Ile, O. J., McCormick, H., Skrabacz, S., Bhattacharya, S., Aguilos, M., Carvalho, H. D. R., Idassi, J., Baker, J., Heitman, J. L., & King, J. S. (2022). Integrating short rotation woody crops into conventional agricultural practices in the southeastern United States: A review. *Land*, 12(1), 10. <https://doi.org/10.3390/land12010010>
452. Ireland, K. B., Moore, M. M., Fulé, P. Z., Yocom, L. L., & Ziegler, T. J. (2020). Warm, dry conditions inhibit aspen growth, but tree growth and size predict mortality risk in the southwestern United States. *Canadian Journal of Forest Research*, 50(11), 1206–1214. <https://doi.org/10.1139/cjfr-2019-0222>
453. Isebrands, J. G., & Zalesny, R. S. (2021). Reflections on the contributions of *Populus* research at Rhinelander, Wisconsin, USA. *Canadian Journal of Forest Research*, 51(2), 139–153. <https://doi.org/10.1139/cjfr-2020-0248>
454. Jang, J. H., Morais, A. R. C., Browning, M., Brandner, D. G., Kenny, J. K., Stanley, L. M., Happs, R. M., Kovvali, A. S., Cutler, J. I., Román-Leshkov, Y., Bielenberg, J. R., & Beckham, G. T. (2023). Feedstock-agnostic reductive catalytic fractionation in alcohol and alcohol–water mixtures. *Green Chemistry*, 25(9), 3660–3670. <https://doi.org/10.1039/D2GC04464A>
455. Jardine, K. J., Lei, J., Som, S., Souza, D., Clendinen, C. S., Mehta, H., Handakumbura, P., Bill, M., & Young, R. P. (2022). Light-dependence of formate (C1) and acetate (C2) transport and oxidation in poplar trees. *Plants*, 11(16), 2080. <https://doi.org/10.3390/plants11162080>
456. Jayawardena, D. M., Robertson, G. P., Sanford, G. R., & Thelen, K. D. (2023). Comparative productivity of six bioenergy cropping systems on marginal lands in the Great Lakes Region, United States. *Agronomy Journal*, 115(5), 2451–2468. <https://doi.org/10.1002/agj2.21416>
457. Jeong, D., Tyner, W. E., Meilan, R., Brown, T. R., & Doering, O. C. (2020). Stochastic techno-economic analysis of electricity produced from poplar plantations in Indiana. *Renewable Energy*, 149, 189–197. <https://doi.org/10.1016/j.renene.2019.11.061>
458. Jiang, P., Ma, N., Sun, F., Ye, M., & Wu, R. (2021). Functional mapping of gravitropism and phototropism for a desert tree, *Populus euphratica*. *Frontiers in Bioscience-Landmark*, 26(11), 988. <https://doi.org/10.52586/5003>

459. Kanhere, S. V., Tindall, G. W., Ogale, A. A., & Thies, M. C. (2022). Carbon fibers derived from liquefied and fractionated poplar lignins: The effect of molecular weight. *iScience*, 25(12), 105449. <https://doi.org/10.1016/j.isci.2022.105449>
460. Kheam, R., Zhang, K., Elder, T., Bryant, N., Ragauskas, A., & Li, M. (2023). Totally chlorine-free peracetic acid pulping for nanocellulose isolation from hemp and poplar, TAPPI. *TAPPI Journal*, 22(8), 529–540. <https://doi.org/10.32964/TJ22.8.529>
461. Kim, K. H., Mottiar, Y., Jeong, K., Tran, P. H. N., Tran, N. T., Zhuang, J., Kim, C. S., Lee, H., Gong, G., Ko, J. K., Lee, S.-M., Kim, S. Y., Shin, J. Y., Jeong, H., Song, H. K., Yoo, C. G., Kim, N.-K., & Mansfield, S. D. (2022). One-pot conversion of engineered poplar into biochemicals and biofuels using biocompatible deep eutectic solvents. *Green Chemistry*, 24(23), 9055–9068. <https://doi.org/10.1039/D2GC02774G>
462. Kim, K., Voothuluru, P., Hamilton, C., McCord, J., Tamang, B., Cunningham, M., Eberhardt, T. L., Rials, T., & Labbé, N. (2021). Tradeoffs between yield, disease incidence and conversion efficiency for selection of hybrid poplar genotypes as bioenergy feedstocks. *Biomass and Bioenergy*, 154, 106259. <https://doi.org/10.1016/j.biombioe.2021.106259>
463. Kim, H., Li, Q., Karlen, S. D., Smith, R. A., Shi, R., Liu, J., Yang, C., Tunlaya-Anukit, S., Wang, J. P., Chang, H.-M., Sederoff, R. R., Ralph, J., & Chiang, V. L. (2020). Monolignol benzoates incorporate into the lignin of transgenic *Populus trichocarpa* depleted in C3H and C4H. *ACS Sustainable Chemistry & Engineering*, 8(9), 3644–3654. <https://doi.org/10.1021/acssuschemeng.9b06389>
464. Kreider, M. R., Mock, K. E., & Yocom, L. L. (2020). Methods for distinguishing aspen seedlings from suckers in the field. *Journal of Forestry*, 118(6), 561–568. <https://doi.org/10.1093/jofore/fvaa030>
465. Kreider, M. R., & Yocom, L. L. (2021a). Aspen seedling establishment, survival, and growth following a high-severity wildfire. *Forest Ecology and Management*, 493, 119248. <https://doi.org/10.1016/j.foreco.2021.119248>
466. Kreider, M. R., & Yocom, L. L. (2021b). Low-density aspen seedling establishment is widespread following recent wildfires in the western United States. *Ecology*, 102(10), e03436. <https://doi.org/10.1002/ecy.3436>
467. Kretchun, A. M., Scheller, R. M., Shinneman, D. J., Soderquist, B., Maguire, K., Link, T. E., & Strand, E. K. (2020). Long term persistence of aspen in snowdrift-dependent ecosystems. *Forest Ecology and Management*, 462, 118005. <https://doi.org/10.1016/j.foreco.2020.118005>
468. Kuai, B., Wang, Z., Gao, J., Tong, J., Zhan, T., Zhang, Y., Lu, J., & Cai, L. (2022). Development of densified wood with high strength and excellent dimensional stability by impregnating delignified poplar by sodium silicate. *Construction and Building Materials*, 344, 128282. <https://doi.org/10.1016/j.conbuildmat.2022.128282>
469. Kyaw, T. Y., Siegert, C. M., Dash, P., Poudel, K. P., Pitts, J. J., & Renninger, H. J. (2022). Using hyperspectral leaf reflectance to estimate photosynthetic capacity and nitrogen content across eastern cottonwood and hybrid poplar taxa. *PLOS ONE*, 17(3), e0264780. <https://doi.org/10.1371/journal.pone.0264780>

470. Landhäusser, S. M., Wiley, E. T., Solarik, K. A., Kulbaba, S. P., & Goepfel, A. E. (2023). The importance of initial seedling characteristics in controlling allocation to growth and reserves under different soil moisture conditions. *Forests*, 14(4), 796. <https://doi.org/10.3390/f14040796>
471. Li, W., Ciais, P., Stehfest, E., van Vuuren, D., Popp, A., Arneth, A., Di Fulvio, F., Doelman, J., Humpeöder, F., Harper, A. B., Park, T., Makowski, D., Havlik, P., Obersteiner, M., Wang, J., Krause, A., & Liu, W. (2020). Mapping the yields of lignocellulosic bioenergy crops from observations at the global scale. *Earth System Science Data*, 12(2), 789–804. (cross-listed with *Salix* and *Eucalyptus*) <https://doi.org/10.5194/essd-12-789-2020>
472. Li, Y., Wang, G., Guo, G., Li, Y., Via, B. K., & Pei, Z. (2022). Spectral pre-processing and multivariate calibration methods for the prediction of wood density in Chinese white poplar by visible and near infrared spectroscopy. *Forests*, 13(1), 62. <https://doi.org/10.3390/f13010062>
473. Lin, C.-Y., Geiselman, G. M., Liu, D., Magurudeniya, H. D., Rodriguez, A., Chen, Y.-C., Pidatala, V., Unda, F., Amer, B., Baidoo, E. E. K., Mansfield, S. D., Simmons, B. A., Singh, S., Scheller, H. V., Gladden, J. M., & Eudes, A. (2022). Evaluation of engineered low-lignin poplar for conversion into advanced bioproducts. *Biotechnology for Biofuels and Bioproducts*, 15(1), 145. <https://doi.org/10.1186/s13068-022-02245-4>
474. Lin, X., Kong, L., Ren, X., Zhang, D., Cai, H., & Lei, H. (2021). Catalytic co-pyrolysis of torrefied poplar wood and high-density polyethylene over hierarchical HZSM-5 for mono-aromatics production. *Renewable Energy*, 164, 87–95. <https://doi.org/10.1016/j.renene.2020.09.071>
475. Liu, J., Qi, L., Yang, G., Xue, Y., He, M., Lucia, L. A., & Chen, J. (2020). Enhancement of lignin extraction of poplar by treatment of deep eutectic solvent with low halogen content. *Polymers*, 12(7), 1599. <https://doi.org/10.3390/polym12071599>
476. Liu, W., Zhang, S., Liu, K., Yang, H., Lin, Q., Xu, T., Song, X., Du, H., Bai, L., Yao, S., & Si, C. (2023). Sustainable preparation of lignocellulosic nanofibrils and cellulose nanopaper from poplar sawdust. *Journal of Cleaner Production*, 384, 135582. <https://doi.org/10.1016/j.jclepro.2022.135582>
477. Liu, X., & Biondi, F. (2021). Inter-specific transpiration differences between aspen, spruce, and pine in a sky-island ecosystem of the North American Great Basin. *Forest Ecology and Management*, 491, 119157. <https://doi.org/10.1016/j.foreco.2021.119157>
478. Liu, Z., Wen, L., Wang, X., Zhang, Y., & Cai, L. (2020). Leachability of ACQ-D after three different preservative treatments. *Wood Research*, 65(4), 591–604. <https://doi.org/10.37763/wr.1336-4561/65.4.591604>
479. Lu, W., Zhang, Y., Yao, Y., Wu, Y., Y. H. Chen, H., Zhang, H., Yu, J., Shen, C., Liu, Q., & Ruan, H. (2020). Biochar-induced priming effects in young and old poplar plantation soils. *Phyton*, 89(1), 13–26. <https://doi.org/10.32604/phyton.2020.09125>
480. Ma, C.-Y., Gao, X., Peng, X.-P., Gao, Y.-F., Liu, J., Wen, J.-L., & Yuan, T.-Q. (2021). Microwave-assisted deep eutectic solvents (DES) pretreatment of control and transgenic poplars for boosting the lignin valorization and cellulose bioconversion. *Industrial Crops and Products*, 164, 113415. <https://doi.org/10.1016/j.indcrop.2021.113415>

481. Markel, K., Belcher, M. S., & Shih, P. M. (2020). Defining and engineering bioenergy plant feedstock ideotypes. *Current Opinion in Biotechnology*, 62, 196–201. <https://doi.org/10.1016/j.copbio.2019.11.014>
482. Mattheus, C. R., Diggins, T. P., & Santoro, J. A. (2021). Punctuated decadal morphodynamics and coupled vegetation succession along a nontidal, wave-dominated Great Lakes barrier spit, gull point, Lake Erie, U.S.A. *Journal of Great Lakes Research*, 47(6), 1538–1553. <https://doi.org/10.1016/j.jglr.2021.04.019>
483. McIlroy, S. K., & Shinneman, D. J. (2020). Post-fire aspen (*Populus tremuloides*) regeneration varies in response to winter precipitation across a regional climate gradient. *Forest Ecology and Management*, 455, 117681. <https://doi.org/10.1016/j.foreco.2019.117681>
484. Meng, F., & Wang, D. (2020). Effects of vacuum freeze drying pretreatment on biomass and biochar properties. *Renewable Energy*, 155, 1–9. <https://doi.org/10.1016/j.renene.2020.03.113>
485. Meng, F., Wang, D., & Zhang, M. (2021). Effect of ultrasonic vibration-assisted pelleting of biomass on biochar properties. *Journal of Cleaner Production*, 279, 123900. <https://doi.org/10.1016/j.jclepro.2020.123900>
486. Meyer, J. R., Li, H., Zhang, J., & Foston, M. B. (2020). Kinetics of secondary reactions affecting the organosolv lignin structure. *ChemSusChem*, 13(17), 4557–4566. <https://doi.org/10.1002/cssc.202000942>
487. Meyer, M., Tavares Wahren, F., Weber, N., Zalesny, R. S., & Weih, M. (2021). Sustainable biomass value chains based on poplar plantations in European rural areas. *BioEnergy Research*, 14(2), 355–356. <https://doi.org/10.1007/s12155-021-10275-3>
488. Meyer, P. A., Snowden-Swan, L. J., Jones, S. B., Rappé, K. G., & Hartley, D. S. (2020). The effect of feedstock composition on fast pyrolysis and upgrading to transportation fuels: Techno-economic analysis and greenhouse gas life cycle analysis. *Fuel*, 259, 116218. <https://doi.org/10.1016/j.fuel.2019.116218>
489. Miller, A. E., Stanfield, R. C., & Hacke, U. G. (2021). Seasonal patterns of callose deposition and xylem embolism in five boreal deciduous tree species. *American Journal of Botany*, 108(8), 1568–1575. <https://doi.org/10.1002/ajb2.1718>
490. Molinari, R. L., Bekker, M. F., St. Clair, B. D., Bartholomew, J., DeRose, R. J., Kitchen, S. G., & St. Clair, S. B. (2022). Facilitation differentially affects competitive responses of aspen and subalpine fir through stages of stand development. *Ecosphere*, 13(3), e3957. <https://doi.org/10.1002/ecs2.3957>
491. Monson, R. K., Winkler, B., Rosenstiel, T. N., Block, K., Merl-Pham, J., Strauss, S. H., Ault, K., Maxfield, J., Moore, D. J. P., Trahan, N. A., Neice, A. A., Shiach, I., Barron-Gafford, G. A., Ibsen, P., McCorkel, J. T., Bernhardt, J., & Schnitzler, J.-P. (2020). High productivity in hybrid-poplar plantations without isoprene emission to the atmosphere. *Proceedings of the National Academy of Sciences*, 117(3), 1596–1605. <https://doi.org/10.1073/pnas.1912327117>
492. Montagnoli, A., Dumroese, R. K., Negri, G., Scippa, G. S., Chiatante, D., & Terzaghi, M. (2022). Asymmetrical copper root pruning may improve root traits for reforestation steep and/or windy sites. *New Forests*, 53(6), 1093–1112. <https://doi.org/10.1007/s11056-022-09913-1>

493. Morales-Vera, R., Vásquez-Ibarra, L., Scott, F., Puettmann, M., & Gustafson, R. (2022). Life cycle assessment of bioethanol production: A case study from poplar biomass growth in the U.S. Pacific Northwest. *Fermentation*, 8(12), 734. <https://doi.org/10.3390/fermentation8120734>
494. Morales-Vera, R., Crawford, J., Dou, C., Bura, R., & Gustafson, R. (2020). Techno-economic analysis of producing glacial acetic acid from poplar biomass via bioconversion. *Molecules*, 25(18), 4328. <https://doi.org/10.3390/molecules25184328>
495. Mueller, L. O., Borstein, S. R., Tague, E. D., Dearth, S. P., Castro, H. F., Campagna, S. R., Bailey, J. K., & Schweitzer, J. A. (2020). Populations of *Populus angustifolia* have evolved distinct metabolic profiles that influence their surrounding soil. *Plant and Soil*, 448(1–2), 399–411. <https://doi.org/10.1007/s11104-019-04405-2>
496. Mukherjee, A., & Coleman, M. D. (2020). Converting conventional agriculture to poplar bioenergy crops: Soil chemistry. *Communications in Soil Science and Plant Analysis*, 51(3), 364–379. <https://doi.org/10.1080/00103624.2019.1709485>
497. Mustafa, R., Hamza, M., Rehman, A. U., Kamal, H., Tahir, M. N., Mansoor, S., Scheffler, B. E., Briddon, R. W., & Amin, I. (2022). Asymptomatic *Populus alba*: A tree serving as a reservoir of begomoviruses and associated satellites. *Australasian Plant Pathology*, 51(6), 577–586. <https://doi.org/10.1007/s13313-022-00886-5>
498. Nabuqi, Nuoendagula, Wu, S., Takata, N., Sakamoto, S., Yamamoto, M., Uesugi, M., Déjardin, A., Pilate, G., Taniguchi, T., Mitsuda, N., & Kajita, S. (2020). Simultaneous manipulation of lignin structure and secondary cell wall formation in transgenic poplar. *Journal of Wood Science*, 66(1), 56. <https://doi.org/10.1186/s10086-020-01902-2>
499. Nesbit, K. A., Yocom, L. L., Trudgeon, A. M., DeRose, R. J., & Rogers, P. C. (2023). Tamm review: Quaking aspen's influence on fire occurrence, behavior, and severity. *Forest Ecology and Management*, 531, 120752. <https://doi.org/10.1016/j.foreco.2022.120752>
500. Nigro, K. M., Rocca, M. E., Battaglia, M. A., Coop, J. D., & Redmond, M. D. (2022). Wildfire catalyzes upward range expansion of trembling aspen in southern Rocky Mountain beetle-killed forests. *Journal of Biogeography*, 49(1), 201–214. <https://doi.org/10.1111/jbi.14302>
501. Omer, M., Idowu, O. J., Pietrasiak, N., VanLeeuwen, D., Ulery, A. L., Dominguez, A. J., Ghimire, R., & Marsalis, M. (2023). Agricultural practices influence biological soil quality indicators in an irrigated semiarid agro-ecosystem. *Pedobiologia*, 96, 150862. <https://doi.org/10.1016/j.pedobi.2022.150862>
502. Omer, M., Idowu, O. J., Ulery, A. L., VanLeeuwen, D., Guldán, S. J., Marsalis, M. A., & Ghimire, R. (2020). Impacts of selected management practices on soil quality in an irrigated arid agroecosystem. *Journal of Soil and Water Conservation*, 75(2), 143–152. <https://doi.org/10.2489/jswc.75.2.143>
503. Ossler, F., Finney, C. E. A., Warren, J. M., Bilheux, J.-C., Zhang, Y., Mills, R. A., Santodonato, L. J., & Bilheux, H. Z. (2021). Dynamics of hydrogen loss and structural changes in pyrolyzing biomass utilizing neutron imaging. *Carbon*, 176, 511–529. <https://doi.org/10.1016/j.carbon.2020.11.060>

504. Ouyang, Y., Dev, S., Grace Iii, J. M., Amatya, D. M., & Leininger, T. D. (2022). Simulating biomass production and water use of poplars in a plantation using a STELLA-based model. *Forests*, 13(4), 547. <https://doi.org/10.3390/f13040547>
505. Palmquist, E. C., Allan, G. J., Ogle, K., Whitham, T. G., Butterfield, B. J., & Shafroth, P. B. (2021). Riverine complexity and life history inform restoration in riparian environments in the southwestern United States. *Restoration Ecology*, 29(7), e13418. <https://doi.org/10.1111/rec.13418>
506. Pascoli, D. U., Suko, A., Gustafson, R., Gough, H. L., & Bura, R. (2021). Novel ethanol production using biomass preprocessing to increase ethanol yield and reduce overall costs. *Biotechnology for Biofuels*, 14(1), 9. <https://doi.org/10.1186/s13068-020-01839-0>
507. Peterson, S. C. (2020). Coppiced biochars as partial replacement of carbon black filler in polybutadiene/natural rubber composites. *Journal of Composites Science*, 4(4), 147. <https://doi.org/10.3390/jcs4040147>
508. Peterson, S. C., & McMahan, C. M. (2023). Replacement of carbon black with coppiced biochar in guayule rubber composites improves tensile properties. *Journal of Composites Science*, 7(12), 499. <https://doi.org/10.3390/jcs7120499>
509. Pilipović, A., Headlee, W. L., Zalesny Jr., R. S., Pekeč, S., & Bauer, E. O. (2022). Water use efficiency of poplars grown for biomass production in the Midwestern United States. *GCB Bioenergy*, 14(3), 287–306. <https://doi.org/10.1111/gcbb.12887>
510. Priano, M. E., Zorita, F., & Trasar-Cepeda, C. (2023). Legacy of prior management of cropland after afforestation with *Populus × euroamericana* (Dode) Guinier: Effects on soil respiration. *Forests*, 14(5), 1048. <https://doi.org/10.3390/f14051048>
511. Rajan, K., Kim, K., Elder, T. J., Naskar, A. K., & Labbé, N. (2022). Ionic-liquid-assisted fabrication of lignocellulosic thin films with tunable Hydrophobicity. *ACS Sustainable Chemistry & Engineering*, 10(27), 8835–8845. <https://doi.org/10.1021/acssuschemeng.2c01741>
512. Rajan, K., D'Souza, D. H., Kim, K., Choi, J. M., Elder, T., Carrier, D. J., & Labbé, N. (2021). Production and characterization of high value prebiotics from biorefinery-relevant feedstocks. *Frontiers in Microbiology*, 12, 675314. <https://doi.org/10.3389/fmicb.2021.675314>
513. Renninger, H. J., Pitts, J. J., & Rousseau, R. J. (2023). Comparisons of biomass, water use efficiency and water use strategies across five genomic groups of *Populus* and its hybrids. *GCB Bioenergy*, 15(1), 99–112. <https://doi.org/10.1111/gcbb.13014>
514. Renninger, H. J., Stewart, L. F., Freeman, J. L., & Rousseau, R. J. (2022). Physiological functioning and productivity in eastern cottonwood and hybrid poplars on contrasting sites in the Southeastern US. *BioEnergy Research*, 15(2), 1057–1070. <https://doi.org/10.1007/s12155-021-10377-y>
515. Renninger, H. J., Stewart, L. F., & Rousseau, R. J. (2021). Water use, efficiency, and stomatal sensitivity in eastern cottonwood and hybrid poplar varieties on contrasting sites in the southeastern United States. *Frontiers in Forests and Global Change*, 4, 704799. <https://doi.org/10.3389/ffgc.2021.704799>

516. Rogers, E. R., Zalesny, R. S., Lin, C.-H., & Vinhal, R. A. (2023). Intrinsic and extrinsic factors influencing *Populus* water use: A literature review. *Journal of Environmental Management*, 348, 119180. <https://doi.org/10.1016/j.jenvman.2023.119180>
517. Rogers, P. C. (2022). Pando's pulse: Vital signs signal need for course correction at world-renowned aspen forest. *Conservation Science and Practice*, 4(10), e12804. <https://doi.org/10.1111/csp2.12804>
518. Rogers, P. C., Pinno, B. D., Šebesta, J., Albrechtsen, B. R., Li, G., Ivanova, N., Kusbach, A., Kuuluvainen, T., Landhäusser, S. M., Liu, H., Myking, T., Pulkkinen, P., Wen, Z., & Kulakowski, D. (2020). A global view of aspen: Conservation science for widespread keystone systems. *Global Ecology and Conservation*, 21, e00828. <https://doi.org/10.1016/j.gecco.2019.e00828>
519. Rubert-Nason, K. F., Yang, P., Morrow, C. J., & Lindroth, R. L. (2023). Environment and genotype influence quantitative and qualitative variation in condensed tannins in aspen. *Journal of Chemical Ecology*, 49(5–6), 325–339. <https://doi.org/10.1007/s10886-023-01430-5>
520. Salceda, M., Udawatta, R. P., Anderson, S. H., Mendis, S. S., & Liu, F. (2023). Agroforestry buffers on nitrogen reduction in groundwater on a grazed hillslope. *Agrosystems, Geosciences & Environment*, 6(2), e20370. <https://doi.org/10.1002/agg2.20370>
521. Schepetkin, I. A., Özek, G., Özek, T., Kirpotina, L. N., Kokorina, P. I., Khlebnikov, A. I., & Quinn, M. T. (2022). Neutrophil immunomodulatory activity of nerolidol, a major component of essential oils from *Populus balsamifera* buds and propolis. *Plants*, 11(23), 3399. <https://doi.org/10.3390/plants11233399>
522. Schook, D. M., Friedman, J. M., Stricker, C. A., Csank, A. Z., & Cooper, D. J. (2020). Short- and long-term responses of riparian cottonwoods (*Populus* spp.) to flow diversion: Analysis of tree-ring radial growth and stable carbon isotopes. *Science of The Total Environment*, 735, 139523. <https://doi.org/10.1016/j.scitotenv.2020.139523>
523. Schultz, J. A., & Coleman, H. D. (2021). Pectin and xylan biosynthesis in poplar: Implications and opportunities for biofuels production. *Frontiers in Plant Science*, 12, 712083. <https://doi.org/10.3389/fpls.2021.712083>
524. Semerci, A., Guevara, C. A., & Gonzalez-Benecke, C. A. (2021). Water availability effects on growth and phenology of 11 poplar cultivars growing in semiarid areas in Turkey. *New Forests*, 52(3), 411–430. <https://doi.org/10.1007/s11056-020-09802-5>
525. Seufitelli, G. V. S., El-Husseini, H., Pascoli, D. U., Bura, R., & Gustafson, R. (2022). Techno-economic analysis of an integrated biorefinery to convert poplar into jet fuel, xylitol, and formic acid. *Biotechnology for Biofuels and Bioproducts*, 15(1), 143. <https://doi.org/10.1186/s13068-022-02246-3>
526. Shah, R., Bhagia, S., Keum, J. K., Pingali, S. V., Ragauskas, A. J., Davison, B. H., & O'Neill, H. (2020). Structural insights into low and high recalcitrance natural poplar variants using neutron and X-ray scattering. *ACS Sustainable Chemistry & Engineering*, 8(36), 13838–13849. <https://doi.org/10.1021/acssuschemeng.0c05251>
527. Shao, Y., Li, L., Chen, Z., Wang, S., & Wang, X. (2020). Effects of thermo-hydro-mechanical treatments on various physical and mechanical properties of poplar (*Populus*) wood. *BioResources*, 15(4), 9596–9610. <https://doi.org/10.15376/biores.15.4.9596-9610>

528. Sheehan, J. D., Ebikade, E., Vlachos, D. G., & Lobo, R. F. (2022). Lignin-based water-soluble polymers exhibiting biodegradability and activity as flocculating agents. *ACS Sustainable Chemistry & Engineering*, 10(34), 11117–11129. <https://doi.org/10.1021/acssuschemeng.2c01428>
529. Shi, F., Wang, Y., Davaritouchee, M., Yao, Y., & Kang, K. (2020). Directional structure modification of poplar biomass-inspired high efficacy of enzymatic hydrolysis by sequential dilute acid–alkali treatment. *ACS Omega*, 5(38), 24780–24789. <https://doi.org/10.1021/acsomega.0c03419>
530. Shults, P., Nzokou, P., & Koc, I. (2020). Nitrogen contributions of alley cropped *Trifolium pratense* may sustain short rotation woody crop yields on marginal lands. *Nutrient Cycling in Agroecosystems*, 117(2), 261–272. <https://doi.org/10.1007/s10705-020-10068-8>
531. Sloan, J. L., Burney, O. T., & Pinto, J. R. (2020). Drought-conditioning of quaking aspen (*Populus tremuloides* Michx.) seedlings during nursery production modifies seedling anatomy and physiology. *Frontiers in Plant Science*, 11, 557894. <https://doi.org/10.3389/fpls.2020.557894>
532. Stack, S., Jones, C., Bockstette, J., Jacobs, D. F., & Landhäusser, S. M. (2020). Surface and subsurface material selections influence the early outcomes of boreal upland forest restoration. *Ecological Engineering*, 144, 105705. <https://doi.org/10.1016/j.ecoleng.2019.105705>
533. Stanton, B. J., Bourque, A., Coleman, M., Eisenbies, M., Emerson, R. M., Espinoza, J., Gantz, C., Himes, A., Rodstrom, A., Shuren, R., Stonex, R., Volk, T., & Zepa, J. (2021). The practice and economics of hybrid poplar biomass production for biofuels and bioproducts in the Pacific Northwest. *BioEnergy Research*, 14(2), 543–560. <https://doi.org/10.1007/s12155-020-10164-1>
534. Straub, C. T., Bing, R. G., Wang, J. P., Chiang, V. L., Adams, M. W. W., & Kelly, R. M. (2020). Use of the lignocellulose-degrading bacterium *Caldicellulosiruptor bescii* to assess recalcitrance and conversion of wild-type and transgenic poplar. *Biotechnology for Biofuels*, 13(1), 43. <https://doi.org/10.1186/s13068-020-01675-2>
535. Struhs, E., Sotoudehnia, F., Mirkouei, A., McDonald, A. G., & Ramirez-Corredores, M. M. (2022). Effect of feedstocks and free-fall pyrolysis on bio-oil and biochar attributes. *Journal of Analytical and Applied Pyrolysis*, 166, 105616. <https://doi.org/10.1016/j.jaap.2022.105616>
536. Sun, H., Gong, Y., Yan, Y., Fu, C., Zhan, K., Dong, C., Xie, L., & Chen, T. (2023). Leachability of fast-growing wood impregnated with low concentrations of furfuryl alcohol. *Forests*, 14(9), 1901. <https://doi.org/10.3390/f14091901>
537. Tatum, J., & Wallin, D. (2021). Using discrete-point LiDAR to classify tree species in the riparian pacific northwest, USA. *Remote Sensing*, 13(14), 2647. <https://doi.org/10.3390/rs13142647>
538. Terrell, E., & Garcia-Perez, M. (2020). Vacuum pyrolysis of hybrid poplar milled wood lignin with fourier transform-ion cyclotron resonance mass spectrometry analysis of feedstock and products for the elucidation of reaction mechanisms. *Energy & Fuels*, 34(11), 14249–14263. <https://doi.org/10.1021/acs.energyfuels.0c02928>
539. Thibault, E., Soolanayakanahally, R., & Keller, S. R. (2020). Latitudinal clines in bud flush phenology reflect genetic variation in chilling requirements in balsam poplar, *Populus*

balsamifera. *American Journal of Botany*, 107(11), 1597–1605.

<https://doi.org/10.1002/ajb2.1564>

540. Tian, B., Wang, X., Zhao, W., Xu, L., & Bai, L. (2021). Pyrolysis behaviors, kinetics and gaseous product evolutions of two typical biomass wastes. *Catalysis Today*, 374, 77–85. <https://doi.org/10.1016/j.cattod.2020.12.023>
541. Tindall, G., Lynn, B., Fitzgerald, C., Valladares, L., Pittman, Z., Bécsy-Jakab, V., Hodge, D., & Thies, M. (2021). Ultraclean hybrid poplar lignins via liquid–liquid fractionation using ethanol–water solutions. *MRS Communications*, 11(5), 692–698. <https://doi.org/10.1557/s43579-021-00090-4>
542. Valdovinos-Ayala, J., Robles, C., Fickle, J. C., Pérez-de-Lis, G., Pratt, R. B., & Jacobsen, A. L. (2022). Seasonal patterns of increases in stem girth, vessel development, and hydraulic function in deciduous tree species. *Annals of Botany*, 130(3), 355–365. <https://doi.org/10.1093/aob/mcac032>
543. Walton, C. L., Khalid, M., Bible, A. N., Kertesz, V., Retterer, S. T., Morrell-Falvey, J., & Cahill, J. F. (2022). *In situ* detection of amino acids from bacterial biofilms and plant root exudates by liquid microjunction surface-sampling probe mass spectrometry. *Journal of the American Society for Mass Spectrometry*, 33(9), 1615–1625. <https://doi.org/10.1021/jasms.2c00081>
544. Wang, B., Wang, Z., Wang, C., Wang, X., Li, J., Jia, Z., Li, P., Wu, J., Chen, M., & Liu, L. (2021). Field evidence reveals conservative water use of poplar saplings under high aerosol conditions. *Journal of Ecology*, 109(5), 2190–2202. <https://doi.org/10.1111/1365-2745.13633>
545. Wang, J., Bi, Z., Chen, Z., Yan, L., & Lei, Y. (2021). Decay resistance, dimensional stability and mechanical strength of poplar wood modified with plant-derived compounds. *Wood Research*, 66(4), 556–568. <https://doi.org/10.37763/wr.1336-4561/66.4.556568>
546. Wang, J., Chai, Y., Liu, J., & Zhu, J. Y. (2023). The viscoelastic and hygroscopicity behavior of delignified and densified poplar wood. *Forests*, 14(9), 1721. <https://doi.org/10.3390/f14091721>
547. Wang, J., Fishwild, S. J., Begel, M., & Zhu, J. Y. (2020). Properties of densified poplar wood through partial delignification with alkali and acid pretreatment. *Journal of Materials Science*, 55(29), 14664–14676. <https://doi.org/10.1007/s10853-020-05034-2>
548. Wang, J., Liu, J., Li, J., & Zhu, J. Y. (2021). Characterization of microstructure, chemical, and physical properties of delignified and densified poplar wood. *Materials*, 14(19), 5709. <https://doi.org/10.3390/ma14195709>
549. Wang, J., Rajan, K., Annamraju, A., Chmely, S. C., Pingali, S. V., Carrier, D. J., & Labbé, N. (2021). A sequential autohydrolysis-ionic liquid fractionation process for high quality lignin production. *Energy & Fuels*, 35(3), 2293–2302. <https://doi.org/10.1021/acs.energyfuels.0c03849>
550. Wang, J., Simson, R., Gleisner, R., Fishwild, S. J., Begel, M., & Zhu, J. Y. (2022). Carbonate pre-treatment of wood for transformative structural applications through densification. *Industrial Crops and Products*, 188, 115645. <https://doi.org/10.1016/j.indcrop.2022.115645>

551. Wang, J., Xu, Y., Meng, X., Pu, Y., Ragauskas, A., & Zhang, J. (2021). Production of xylo-oligosaccharides from poplar by acetic acid pretreatment and its impact on inhibitory effect of poplar lignin. *Bioresource Technology*, 323, 124593. <https://doi.org/10.1016/j.biortech.2020.124593>
552. Wang, H., Liu, Z., Zheng, X., Pan, X., Hui, L., Li, J., & Zhang, H. (2020). Assessment on temperature-pressure severally controlled explosion pretreatment of poplar. *Carbohydrate Polymers*, 230, 115622. <https://doi.org/10.1016/j.carbpol.2019.115622>
553. Wang, X., Chen, H., Feng, X., Zhang, Q., Labbé, N., Kim, K., Huang, J., Ragauskas, A. J., Wang, S., & Zhang, Y. (2020). Isolation and characterization of lignocellulosic nanofibers from four kinds of organosolv-fractionated lignocellulosic materials. *Wood Science and Technology*, 54(3), 503–517. <https://doi.org/10.1007/s00226-020-01167-4>
554. Wang, Y., Meng, X., Jeong, K., Li, S., Leem, G., Kim, K. H., Pu, Y., Ragauskas, A. J., & Yoo, C. G. (2020). Investigation of a lignin-based deep eutectic solvent using *p*-hydroxybenzoic acid for efficient woody biomass conversion. *ACS Sustainable Chemistry & Engineering*, 8(33), 12542–12553. <https://doi.org/10.1021/acssuschemeng.0c03533>
555. Wang, Y.-Y., Scheidemantle, B., Wyman, C. E., Cai, C. M., & Ragauskas, A. J. (2021). Polyurethanes based on unmodified and refined technical lignins: Correlation between molecular structure and material properties. *Biomacromolecules*, 22(5), 2129–2136. <https://doi.org/10.1021/acs.biomac.1c00223>
556. Wang, Y.-Y., Sengupta, P., Scheidemantle, B., Pu, Y., Wyman, C. E., Cai, C. M., & Ragauskas, A. J. (2020). Effects of CELF pretreatment severity on lignin structure and the lignin-based polyurethane properties. *Frontiers in Energy Research*, 8, 149. <https://doi.org/10.3389/fenrg.2020.00149>
557. Wang, Z., Yan, W., Peng, Y., Wan, M., Farooq, T. H., Fan, W., Lei, J., Yuan, C., Wang, W., Qi, Y., & Chen, X. (2023). Biomass production and carbon stocks in poplar-crop agroforestry chronosequence in subtropical Central China. *Plants*, 12(13), 2451. <https://doi.org/10.3390/plants12132451>
558. Warren, J. M., DeCarlo, K. F., Bilheux, H., Bilheux, J.-C., & Caylor, K. (2023). Integrating fine root diameter and watershed mapping to characterize rhizosphere hydrology. *Rhizosphere*, 27, 100738. <https://doi.org/10.1016/j.rhisph.2023.100738>
559. Wei, W., Tian, Z., Ji, X., Wang, Q., Chen, J., Zhang, G., & Lucia, L. A. (2020). Understanding the effect of severity factor of prehydrolysis on dissolving pulp production using prehydrolysis kraft pulping and elemental chlorine-free bleaching sequence. *BioResources*, 15(2), 4323–4336. <https://doi.org/10.15376/biores.15.2.4323-4336>
560. Wen, P., Zhang, T., Wei, L., Wang, J., Ragauskas, A. J., Zhang, Y., Xu, Y., & Zhang, J. (2021). Effect of dilute acetic acid hydrolysis on xylooligosaccharide production and the inhibitory effect of cellulolytic enzyme lignin from poplar. *ACS Sustainable Chemistry & Engineering*, 9(34), 11361–11371. <https://doi.org/10.1021/acssuschemeng.1c02937>
561. Wong, T., Sullivan, J., & Eisenstein, E. (2022). Acclimation and compensating metabolite responses to UV-B radiation in natural and transgenic *Populus* spp. defective in lignin biosynthesis. *Metabolites*, 12(8), 767. <https://doi.org/10.3390/metabo12080767>

562. Wu, R., & Baxter, L. L. (2023). Dynamic biomass char porosity during gasification: Model compared with data. *Energy*, 267, 126603. <https://doi.org/10.1016/j.energy.2022.126603>
563. Wu, R., Beutler, J., & Baxter, L. L. (2023). Biomass char gasification kinetic rates compared to data, including ash effects. *Energy*, 266, 126392. <https://doi.org/10.1016/j.energy.2022.126392>
564. Wu, R., Beutler, J., Price, C., & Baxter, L. L. (2020). Biomass char particle surface area and porosity dynamics during gasification. *Fuel*, 264, 116833. <https://doi.org/10.1016/j.fuel.2019.116833>
565. Wu, Y., Cai, Y., Yang, F., Gan, J., & Zhang, J. (2022). Chemical modification of poplar wood featuring compressible rebound 3D structure as water treatment absorbents. *Journal of Cleaner Production*, 331, 129952. <https://doi.org/10.1016/j.jclepro.2021.129952>
566. Xi, B., Clothier, B., Coleman, M., Duan, J., Hu, W., Li, D., Di, N., Liu, Y., Fu, J., Li, J., Jia, L., & Fernández, J.-E. (2021). Irrigation management in poplar (*Populus* spp.) plantations: A review. *Forest Ecology and Management*, 494, 119330. <https://doi.org/10.1016/j.foreco.2021.119330>
567. Xu, L., Ma, X., Murria, P., Talpade, A., Sheng, H., Meilan, R., Chapple, C., Agrawal, R., Delgass, W. N., Ribeiro, F. H., & Kenttämää, H. I. (2021). Fast determination of the lignin monomer compositions of genetic variants of poplar *via* fast pyrolysis/atmospheric pressure chemical ionization mass spectrometry. *Journal of the American Society for Mass Spectrometry*, 32(10), 2546–2551. <https://doi.org/10.1021/jasms.1c00186>
568. Yan, L., Ji, X., Zeng, F., Chen, Z., & Lei, Y. (2022). Quercetin-grafted modification to improve wood decay resistance. *Holzforschung*, 76(11–12), 1013–1022. <https://doi.org/10.1515/hf-2022-0110>
569. Yan, L., Zeng, F., Chen, Z., Chen, S., & Lei, Y. (2021). Improvement of wood decay resistance by salicylic acid / silica microcapsule: Effects on the salicylic leaching, microscopic structure and decay resistance. *International Biodeterioration & Biodegradation*, 156, 105134. <https://doi.org/10.1016/j.ibiod.2020.105134>
570. Yang, H., Yoo, C. G., Meng, X., Pu, Y., Muchero, W., Tuskan, G. A., Tschaplinski, T. J., Ragauskas, A. J., & Yao, L. (2020). Structural changes of lignins in natural *Populus* variants during different pretreatments. *Bioresource Technology*, 295, 122240. <https://doi.org/10.1016/j.biortech.2019.122240>
571. Yuan, Z., Bals, B. D., Hegg, E. L., & Hodge, D. B. (2022). Technoeconomic evaluation of recent process improvements in production of sugar and high-value lignin co-products via two-stage Cu-catalyzed alkaline-oxidative pretreatment. *Biotechnology for Biofuels and Bioproducts*, 15(1), 45. <https://doi.org/10.1186/s13068-022-02139-5>
572. Yuan, Z., Klinger, G. E., Nikafshar, S., Cui, Y., Fang, Z., Alherech, M., Goes, S., Anson, C., Singh, S. K., Bals, B., Hodge, D. B., Nejad, M., Stahl, S. S., & Hegg, E. L. (2021). Effective biomass fractionation through oxygen-enhanced alkaline–oxidative pretreatment. *ACS Sustainable Chemistry & Engineering*, 9(3), 1118–1127. <https://doi.org/10.1021/acssuschemeng.0c06170>
573. Yue, K., Wang, F., Lu, W., Tang, Z., Chen, Z., & Liu, W. (2021). Tensile properties of finger-jointed lumber under high-temperature and oxygen-free conditions. *Holzforschung*, 75(9), 838–846. <https://doi.org/10.1515/hf-2020-0066>

574. Yue, K., Song, X., Jiao, X., Wang, L., Jia, C., Chen, Z., & Lu, W. (2020). An experimental study of flexural behavior of glulam beams made out of thermally treated fast-growing poplar laminae. *Wood and Fiber Science*, 52(2), 152–164. <https://doi.org/10.22382/wfs-2020-014>
575. Yuldashev, F., Illukpitiya, P., Tegegne, F., & Ekanem, E. (2020). Techno-economic analysis of plantation biomass production and small-scale wood pellet processing for bioenergy market. *International Wood Products Journal*, 11(4), 173–188. (cross-listed with *Salix* and *Eucalyptus*) <https://doi.org/10.1080/20426445.2020.1816766>
576. Zhang, J., Wang, X., Wang, H.-T., Qiao, Z., Yao, T., Xie, M., Urbanowicz, B. R., Zeng, W., Jawdy, S. S., Gunter, L. E., Yang, X., Czarnecki, O., Regan, S., Seguin, A., Rottmann, W., Winkeler, K. A., Sykes, R., Lipzen, A., Daum, C., ... Chen, J.-G. (2023). Overexpression of *REDUCED WALL ACETYLATION C* increases xylan acetylation and biomass recalcitrance in *Populus*. *Plant Physiology*, 194(1), 243–257. <https://doi.org/10.1093/plphys/kiad377>
577. Zhang, J., Jiang, Y., Easterling, L. F., Anstner, A., Li, W., Alzarini, K. Z., Dong, X., Bozell, J., & Kenttämää, H. I. (2021). Compositional analysis of organosolv poplar lignin by using high-performance liquid chromatography/high-resolution multi-stage tandem mass spectrometry. *Green Chemistry*, 23(2), 983–1000. <https://doi.org/10.1039/D0GC03398G>
578. Zhang, J., Tuskan, G. A., Tschaplinski, T. J., Muchero, W., & Chen, J.-G. (2020). Transcriptional and post-transcriptional regulation of lignin biosynthesis pathway genes in *Populus*. *Frontiers in Plant Science*, 11, 652. <https://doi.org/10.3389/fpls.2020.00652>
579. Zhao, J., Yang, Y., Zhang, M., & Wang, D. (2021a). Effects of post-washing on pretreated biomass and hydrolysis of the mixture of acetic acid and sodium hydroxide pretreated biomass and their mixed filtrate. *Bioresource Technology*, 339, 125605. <https://doi.org/10.1016/j.biortech.2021.125605>
580. Zhao, J., Yang, Y., Zhang, M., & Wang, D. (2021b). Minimizing water consumption for sugar and lignin recovery via the integration of acid and alkali pretreated biomass and their mixed filtrate without post-washing. *Bioresource Technology*, 337, 125389. <https://doi.org/10.1016/j.biortech.2021.125389>
581. Zhao, R., Page-Dumroese, D. S., Liu, Y., Wang, K., & Dumroese, R. K. (2024). Biochar and manure additions increased above- and belowground wood decomposition, and soil enzyme activities in a sandy loam soil. *GCB Bioenergy*, 16(1), e13110. <https://doi.org/10.1111/gcbb.13110>
582. Zhao, W., Wen, L., Liu, Z., Liang, R., Zhang, Y., Huang, Z., & Cai, L. (2022). Decay resistance of preservative injected poplar and its process of preventing white rot fungi infection. *Wood Research*, 67(2), 198–212. <https://doi.org/10.37763/wr.1336-4561/67.2.198212>
583. Zhao, X., Bian, B., Huang, C., Lai, C., Song, J., Jin, Y., Meng, X., Ragauskas, A., & Yong, Q. (2023). Elucidating the nonproductive adsorption mechanism of cellulase with lignin fractions from hydrothermally pretreated poplar using multi-dimensional spectroscopic technologies. *Green Chemistry*, 25(16), 6383–6397. <https://doi.org/10.1039/D3GC00907F>
584. Zhao, X., Li, K., Wang, Y., Tekinalp, H., Richard, A., Webb, E., & Ozcan, S. (2020). Bio-treatment of poplar via amino acid for interface control in biocomposites. *Composites Part B: Engineering*, 199, 108276. <https://doi.org/10.1016/j.compositesb.2020.108276>

585. Zhao, Y., Yu, X., Lam, P.-Y., Zhang, K., Tobimatsu, Y., & Liu, C.-J. (2021). Monolignol acyltransferase for lignin phydroxybenzoylation in *Populus*. *Nature Plants*, 7(9), 1288–1300. <https://doi.org/10.1038/s41477-021-00975-1>
586. Zhao, Y., Xiao, S., Yue, J., Zheng, D., & Cai, L. (2020). Effect of enzymatic hydrolysis lignin on the mechanical strength and hydrophobic properties of molded fiber materials. *Holzforschung*, 74(5), 469–475. <https://doi.org/10.1515/hf-2018-0295>
587. Zhong, R., Phillips, D. R., Adams, E. R., & Ye, Z.-H. (2023). An Arabidopsis family GT106 glycosyltransferase is essential for xylan biosynthesis and secondary wall deposition. *Planta*, 257(2), 43. <https://doi.org/10.1007/s00425-023-04077-4>
588. Zhong, R., Lee, C., Haghighat, M., & Ye, Z. (2021). Xylem vessel-specific SND5 and its homologs regulate secondary wall biosynthesis through activating secondary wall NAC binding elements. *New Phytologist*, 231(4), Article 4. <https://doi.org/10.1111/nph.17425>
589. Zhou, X., Yang, S., Lu, M., Zhao, S., Cai, L., Zhang, Y., Zhao, R., & Lv, J. (2020). Structure and monomer ratio of lignin in C3H and HCT RNAi transgenic poplar saplings. *ChemistrySelect*, 5(24), Article 24. <https://doi.org/10.1002/slct.202000365>
590. Ziegler, J. P., Hoffman, C. M., Collins, B. M., Long, J. W., Dagley, C. M., & Mell, W. (2020). Simulated fire behavior and fine-scale forest structure following conifer removal in aspen-conifer forests in the Lake Tahoe Basin, USA. *Fire*, 3(3), 51. <https://doi.org/10.3390/fire3030051>
591. Zimmerman, O. R., Pearce, D. W., Woodman, S. G., Rood, S. B., & Flanagan, L. B. (2023). Increasing contribution of alluvial groundwater to riparian cottonwood forest water use through warm and dry summers. *Agricultural and Forest Meteorology*, 329, 109292. <https://doi.org/10.1016/j.agrformet.2022.109292>
592. Zou, S., Li, D., Di, N., Liu, J., Li, L., Liu, Y., Xi, B., & Coleman, M. (2022). Stand development modifies effects of soil water availability on poplar fine-root traits: Evidence from a six-year experiment. *Plant and Soil*, 480(1–2), 165–184. <https://doi.org/10.1007/s11104-022-05568-1>

Environmental and Ecosystem Services

593. Al-Lami, M. K., Oustriere, N., Gonzales, E., & Burken, J. G. (2022). Phytomanagement of Pb/Zn/Cu tailings using biosolids-biochar or -humus combinations: Enhancement of bioenergy crop production, substrate functionality, and ecosystem services. *Science of The Total Environment*, 836, 155676. (cross-listed with *Salix*) <https://doi.org/10.1016/j.scitotenv.2022.155676>
594. Bazgir, M., Omidipour, R., Heydari, M., Zainali, N., Hamidi, M., & Dey, D. C. (2020). Prioritizing woody species for the rehabilitation of arid lands in western iran based on soil properties and carbon sequestration. *Journal of Arid Land*, 12(4), 640–652. (cross-listed with *Eucalyptus*) <https://doi.org/10.1007/s40333-020-0013-x>
595. BenIsrael, M., Wanner, P., Fernandes, J., Burken, J. G., Aravena, R., Parker, B. L., Haack, E. A., Tsao, D. T., & Dunfield, K. E. (2020). Quantification of toluene phytoextraction rates and microbial biodegradation functional profiles at a fractured bedrock phytoremediation site.

Science of The Total Environment, 707, 135890.
<https://doi.org/10.1016/j.scitotenv.2019.135890>

596. Chavan, S. B., Dhillon, R. S., Sirohi, C., Keerthika, A., Kumari, S., Bharadwaj, K. K., Jinger, D., Kakade, V., Chichaghare, A. R., El-Abedin, T. K. Z., Mahmoud, E. A., Casini, R., Sharma, H., Elansary, H. O., & Yessoufou, K. (2022). Enhancing farm income through boundary plantation of poplar (*Populus deltoides*): An economic analysis. *Sustainability*, 14(14), 8663. <https://doi.org/10.3390/su14148663>
597. Cundy, A. B., LaFreniere, L., Bardos, R. P., Yan, E., Sedivy, R., & Roe, C. (2021). Integrated phytomanagement of a carbon tetrachloride-contaminated site in Murdock, Nebraska (USA). *Journal of Cleaner Production*, 290, 125190. (cross-listed with *Salix*) <https://doi.org/10.1016/j.jclepro.2020.125190>
598. El-Husseini, H., Chowyuk, A. N., Gustafson, R. R., Gough, H. L., & Bura, R. (2023). Roadside vegetated filter strips to simultaneously lower stormwater pollution loadings and improve economics of biorefinery feedstocks. *Journal of Environmental Management*, 347, 119168. <https://doi.org/10.1016/j.jenvman.2023.119168>
599. Garvin, M. C., Schijf, J., Kaufman, S. R., Konow, C., Liang, D., Nigra, A. E., Stracker, N. H., Whelan, R. J., & Wiles, G. C. (2020). A survey of trace metal burdens in increment cores from eastern cottonwood (*Populus deltoides*) across a childhood cancer cluster, Sandusky County, OH, USA. *Chemosphere*, 238, 124528. <https://doi.org/10.1016/j.chemosphere.2019.124528>
600. Giagnoni, L., Borges, L. G. dos A., Giongo, A., Silveira, A. de O., Ardisson, A. N., Triplett, E. W., Mench, M., & Renella, G. (2020). Dolomite and compost amendments enhance Cu phytostabilization and increase microbiota of the leachates from a Cu-contaminated soil. *Agronomy*, 10(5), 719. <https://doi.org/10.3390/agronomy10050719>
601. Hansen, S. M., Gunnell, J., Whaley, A., Dai, X., Harding, C., & Black, B. L. (2020). Adaptability of tree species as windbreaks for urban farms in the U.S. Intermountain West. *Horticulturae*, 6(1), 17. <https://doi.org/10.3390/horticulturae6010017>
602. Hussain, M. Z., Hamilton, S. K., & Robertson, G. P. (2023). Soil phosphorus drawdown by perennial bioenergy cropping systems in the Midwestern US. *GCB Bioenergy*, 15(2), 254–263. <https://doi.org/10.1111/gcbb.13020>
603. Hussain, M. Z., Hamilton, S. K., Robertson, G. P., & Basso, B. (2021). Phosphorus availability and leaching losses in annual and perennial cropping systems in an upper US Midwest landscape. *Scientific Reports*, 11(1), 20367. <https://doi.org/10.1038/s41598-021-99877-7>
604. Hussain, M. Z., Robertson, G. P., Basso, B., & Hamilton, S. K. (2020). Leaching losses of dissolved organic carbon and nitrogen from agricultural soils in the upper US Midwest. *Science of The Total Environment*, 734, 139379. <https://doi.org/10.1016/j.scitotenv.2020.139379>
605. Krebsbach, S., He, J., Adhikari, S., Olshansky, Y., Feyzbar, F., Davis, L. C., Oh, T.-S., & Wang, D. (2023). Mechanistic understanding of perfluorooctane sulfonate (PFOS) sorption by biochars. *Chemosphere*, 330, 138661. (cross-listed with *Eucalyptus*) <https://doi.org/10.1016/j.chemosphere.2023.138661>
606. Landmeyer, J. E., Rock, S., Freeman, J. L., Nagle, G., Samolis, M., Levine, H., Cook, A., & O'Neill, H. (2020). Phytoremediation of slightly brackish, polycyclic aromatic hydrocarbon-

contaminated groundwater from 250 ft below land surface: A pilot-scale study using salt-tolerant, endophyte-enhanced hybrid poplar trees at a superfund site in the central valley of California, April–November 2019. *Remediation Journal*, 31(1), 73–89.

<https://doi.org/10.1002/rem.21664>

607. Li, X., Tian, J., Li, X., Wang, L., Gong, H., Shi, C., Nie, S., Zhu, L., Chen, B., Pan, Y., He, J., Ni, R., & Diao, C. (2022). Developing a sub-meter phenological spectral feature for mapping poplars and willows in urban environment. *ISPRS Journal of Photogrammetry and Remote Sensing*, 193, 77–89. (cross-listed with *Salix*) <https://doi.org/10.1016/j.isprsjprs.2022.09.002>
608. Mazari, K., & Filippelli, G. M. (2020). Using deciduous trees as bioindicators of trace element deposition in a small urban watershed, Indianapolis, IN, USA. *Journal of Environmental Quality*, 49(1), 163–171. <https://doi.org/10.1002/jeq2.20009>
609. Pilipović, A., Zalesny Jr., R. S., Orlović, S., Drekić, M., Pekeč, S., Katanić, M., & Poljaković-Pajnik, L. (2020). Growth and physiological responses of three poplar clones grown on soils artificially contaminated with heavy metals, diesel fuel, and herbicides. *International Journal of Phytoremediation*, 22(4), 436–450. <https://doi.org/10.1080/15226514.2019.1670616>
610. Pilipović, A., Zalesny Jr., R. S., Rogers, E. R., McMahon, B. G., Nelson, N. D., Burken, J. G., Hallett, R. A., & Lin, C.-H. (2021). Establishment of regional phytoremediation buffer systems for ecological restoration in the Great Lakes Basin, USA. II. New clones show exceptional promise. *Forests*, 12(4), 474. <https://doi.org/10.3390/f12040474>
611. Rabideau, A. J., Swensen, K., Chen, Z., Santiago-Collazo, F. L., Bronner, C., Lowry, C., Gnat, R., & Lockwood, T. (2022). Hydrologic evaluation of a poplar phytoextraction system. *International Journal of Phytoremediation*, 24(2), 145–155. <https://doi.org/10.1080/15226514.2021.1929824>
612. Rogers, E. R., Zalesny, R. S., Benzel, T., & Wiese, A. H. (2021). Establishment of poplars in soils amended with fibercake residuals from paper and containerboard production. *Agroforestry Systems*, 95(8), 1719–1733. <https://doi.org/10.1007/s10457-021-00680-0>
613. Salehi, A., Zalesny, R. S., & Calagari, M. (2023). Effects of urban wastewater application on growth, biomass, nutrition, and heavy-metal accumulation of *Populus nigra* L. “62/154,” *P. alba* L. “20/45,” *P. euramericana* (Dode) Guinier “92/40,” and *Salix excelsa* S.G. Gmel grown in heavy-metal contaminated soil. *International Journal of Phytoremediation*, 25(10), 1371–1383. (cross-listed with *Salix*) <https://doi.org/10.1080/15226514.2022.2158783>
614. Simmer, R. A., & Schnoor, J. L. (2022). Phytoremediation, bioaugmentation, and the plant microbiome. *Environmental Science & Technology*, 56(23), 16602–16610. <https://doi.org/10.1021/acs.est.2c05970>
615. Simmer, R., Mathieu, J., Da Silva, M. L. B., Lashmit, P., Gopishetty, S., Alvarez, P. J. J., & Schnoor, J. L. (2020). Bioaugmenting the poplar rhizosphere to enhance treatment of 1,4-dioxane. *Science of The Total Environment*, 744, 140823. <https://doi.org/10.1016/j.scitotenv.2020.140823>
616. Tobin, T., Gustafson, R., Bura, R., & Gough, H. L. (2020). Integration of wastewater treatment into process design of lignocellulosic biorefineries for improved economic viability. *Biotechnology for Biofuels*, 13(1), 24. <https://doi.org/10.1186/s13068-020-1657-7>

617. Wolski, P., Blankenship, B. W., Umar, A., Cabrera, M., Simmons, B. A., Sale, K. L., & Achinivu, E. C. (2023). Factors that influence the activity of biomass-degrading enzymes in the presence of ionic liquids—A review. *Frontiers in Energy Research*, 11, 1212719. (cross-listed with *Eucalyptus* and *Salix*) <https://doi.org/10.3389/fenrg.2023.1212719>
618. Wood, J., Aryal, N., & Subedi, K. (2023). Wastewater treatment using poplar plants: Processes. *Water*, 15(21), 3812. <https://doi.org/10.3390/w15213812>
619. Yung, L., Bertheau, C., Tafforeau, F., Zappellini, C., Valot, B., Maillard, F., Selosse, M.-A., Viotti, C., Binet, P., Chiapusio, G., & Chalot, M. (2021). Partial overlap of fungal communities associated with nettle and poplar roots when co-occurring at a trace metal contaminated site. *Science of The Total Environment*, 782, 146692. <https://doi.org/10.1016/j.scitotenv.2021.146692>
620. Zalesny Jr., R. S., Zhu, J. Y., Headlee, W. L., Gleisner, R., Pilipović, A., Acker, J. V., Bauer, E. O., Birr, B. A., & Wiese, A. H. (2020). Ecosystem services, physiology, and biofuels recalcitrance of poplars grown for landfill phytoremediation. *Plants*, 9(10), 1357. <https://doi.org/10.3390/plants9101357>
621. Zalesny, R. S., Pilipović, A., Rogers, E. R., Burken, J. G., Hallett, R. A., Lin, C.-H., McMahon, B. G., Nelson, N. D., Wiese, A. H., Bauer, E. O., Buechel, L., DeBauche, B. S., Peterson, M., Seegers, R., & Vinhal, R. A. (2021). Establishment of regional phytoremediation buffer systems for ecological restoration in the Great Lakes Basin, USA. I. Genotype × Environment Interactions. *Forests*, 12(4), 430. <https://doi.org/10.3390/f12040430>

Salix

Taxonomy, Nomenclature, and Registration

622. Kuzovkina, Y. A., & Vietto, L. (2021). International registration of cultivar names for *Salix* L. (willow) in 2016–20. *HortScience*, 56(1), 106–107. <https://doi.org/10.21273/HORTSCI15356-20>
623. Marchenko, A., & Kuzovkina, Y. (2023). Notes on the Taxonomy of *Salix vitellina* (Salicaceae). *Plants*, 12(14), 2610. <https://doi.org/10.3390/plants12142610>
624. Marchenko, A. M., & Kuzovkina, Y. A. (2023). The ovule number variation provides new insights into taxa delimitation in willows (*Salix* subgen. *Salix*; salicaceae). *Plants*, 12(3), Article 3. <https://doi.org/10.3390/plants12030497>
625. Marchenko, A. M., & Kuzovkina, Y. A. (2021a). Calculation of the ovule number in the genus *Salix*: A method for taxa differentiation. *Applications in Plant Sciences*, 9(11–12), e11450. <https://doi.org/10.1002/aps3.11450>
626. Marchenko, A. M., & Kuzovkina, Y. A. (2021b). Identification of hybrid formulae of a few willows (*Salix*) using ovule numbers. *Silvae Genetica*, 70(1), 75–83. <https://doi.org/10.2478/sg-2021-0006>
627. Marchenko, A. M., & Kuzovkina, Y. A. (2022). Notes on the nomenclature and taxonomy of *Salix fragilis* (Salicaceae). *TAXON*, 71(4), 721–732. <https://doi.org/10.1002/tax.12685>

628. McGovern, P. N., Kuzovkina, Y. A., & Soolanayakanahally, R. Y. (2021). Short communication: IPC *Salix* cultivar database proof-of-concept. *Forests*, 12(5), 631. <https://doi.org/10.3390/f12050631>

Domestication, Breeding, and Selection of Genetic Resources

629. Carlson, C. H., & Smart, L. B. (2022). Heterosis for biomass-related traits in interspecific triploid hybrids of willow (*Salix* spp.). *BioEnergy Research*, 15(2), 1042–1056. <https://doi.org/10.1007/s12155-021-10305-0>
630. Carlson, C. H., Choi, Y., Chan, A. P., Town, C. D., & Smart, L. B. (2022). Nonadditive gene expression is correlated with nonadditive phenotypic expression in interspecific triploid hybrids of willow (*Salix* spp.). *G3 Genes/Genomes/Genetics*, 12(3), jkab436. <https://doi.org/10.1093/g3journal/jkab436>
631. Feng, G., Sanderson, B. J., Keefover-Ring, K., Liu, J., Ma, T., Yin, T., Smart, L. B., DiFazio, S. P., & Olson, M. S. (2020). Pathways to sex determination in plants: How many roads lead to Rome? *Current Opinion in Plant Biology*, 54, 61–68. (cross-listed with *Populus*) <https://doi.org/10.1016/j.pbi.2020.01.004>
632. Gouker, F. E., Carlson, C. H., Zou, J., Evans, L., Crowell, C. R., Smart, C. D., DiFazio, S. P., & Smart, L. B. (2021). Sexual dimorphism in the dioecious willow *Salix purpurea*. *American Journal of Botany*, 108(8), 1374–1387. <https://doi.org/10.1002/ajb2.1704>
633. Gulyaev, S., Cai, X.-J., Guo, F.-Y., Kikuchi, S., Applequist, W. L., Zhang, Z.-X., Hörandl, E., & He, L. (2022). The phylogeny of *Salix* revealed by whole genome re-sequencing suggests different sex-determination systems in major groups of the genus. *Annals of Botany*, 129(4), 485–498. <https://doi.org/10.1093/aob/mcac012>
634. Hu, N., Sanderson, B. J., Guo, M., Feng, G., Gambhir, D., Hale, H., Wang, D., Hyden, B., Liu, J., Smart, L. B., DiFazio, S. P., Ma, T., & Olson, M. S. (2023). Evolution of a ZW sex chromosome system in willows. *Nature Communications*, 14(1), 7144. <https://doi.org/10.1038/s41467-023-42880-5>
635. Hyden, B., Carper, D. L., Abraham, P. E., Yuan, G., Yao, T., Baumgart, L., Zhang, Y., Chen, C., O'Malley, R., Chen, J., Yang, X., Hettich, R. L., Tuskan, G. A., & Smart, L. B. (2023). Functional analysis of *Salix purpurea* genes support roles for *ARR17* and *GATA15* as master regulators of sex determination. *Plant Direct*, 7(11), e546. <https://doi.org/10.1002/pld3.546>
636. Hyden, B., Feng, K., Yates, T. B., Jawdy, S., Cereghino, C., Smart, L. B., & Muchero, W. (2023). De Novo Assembly and annotation of 11 diverse shrub willow (*Salix*) genomes reveals novel gene organization in sex-linked regions. *International Journal of Molecular Sciences*, 24(3), 2904. <https://doi.org/10.3390/ijms24032904>
637. Hyden, B., Zou, J., Wilkerson, D. G., Carlson, C. H., Robles, A. R., DiFazio, S. P., & Smart, L. B. (2023). Structural variation of a sex-linked region confers monoecy and implicates *GATA15* as a master regulator of sex in *Salix purpurea*. *New Phytologist*, 238(6), 2512–2523. <https://doi.org/10.1111/nph.18853>

638. Hyden, B., Yuan, G., Liu, Y., Smart, L. B., Tuskan, G. A., & Yang, X. (2022). Protoplast-based transient expression and gene editing in shrub willow (*Salix purpurea* L.). *Plants*, 11(24), 3490. <https://doi.org/10.3390/plants11243490>
639. Hyden, B., Carlson, C. H., Gouker, F. E., Schmutz, J., Barry, K., Lipzen, A., Sharma, A., Sandor, L., Tuskan, G. A., Feng, G., Olson, M. S., DiFazio, S. P., & Smart, L. B. (2021). Integrative genomics reveals paths to sex dimorphism in *Salix purpurea* L. *Horticulture Research*, 8, 170. <https://doi.org/10.1038/s41438-021-00606-y>
640. Keefover-Ring, K., Carlson, C. H., Hyden, B., Azeem, M., & Smart, L. B. (2022). Genetic mapping of sexually dimorphic volatile and non-volatile floral secondary chemistry of a dioecious willow. *Journal of Experimental Botany*, 73(18), 6352–6366. <https://doi.org/10.1093/jxb/erac260>
641. Muklada, H., Davidovich-Rikanati, R., Wilkerson, D. G., Klein, J. D., Deutch-Traubman, T., Zou, J., Awabdeh, S., Sweidan, R., Landau, S. Y., Schwartz, A., Lewinsohn, E., & Smart, L. B. (2020). Genotypic diversity in willow (*Salix* spp.) is associated with chemical and morphological polymorphism, suggesting human-assisted dissemination in the Eastern Mediterranean. *Biochemical Systematics and Ecology*, 91, 104081. <https://doi.org/10.1016/j.bse.2020.104081>
642. Sanderson, B. J., Gambhir, D., Feng, G., Hu, N., Cronk, Q. C., Percy, D. M., Frenner, F. M., Johnson, M. G., Smart, L. B., Keefover-Ring, K., Yin, T., Ma, T., DiFazio, S. P., Liu, J., & Olson, M. S. (2023). Phylogenomics reveals patterns of ancient hybridization and differential diversification that contribute to phylogenetic conflict in willows, poplars, and close relatives. *Systematic Biology*, 72(6), 1220–1232. (cross-listed with *Populus*) <https://doi.org/10.1093/sysbio/syad042>
643. Sanderson, B. J., Feng, G., Hu, N., Carlson, C. H., Smart, L. B., Keefover-Ring, K., Yin, T., Ma, T., Liu, J., DiFazio, S. P., & Olson, M. S. (2021). Sex determination through X–Y heterogamety in *Salix nigra*. *Heredity*, 126(4), 630–639. <https://doi.org/10.1038/s41437-020-00397-3>
644. Sanderson, B. J., DiFazio, S. P., Cronk, Q. C. B., Ma, T., & Olson, M. S. (2020). A targeted sequence capture array for phylogenetics and population genomics in the Salicaceae. *Applications in Plant Sciences*, 8(10), e11394. (cross-listed with *Populus*) <https://doi.org/10.1002/aps3.11394>
645. Wang, D., Li, Y., Li, M., Yang, W., Ma, X., Zhang, L., Wang, Y., Feng, Y., Zhang, Y., Zhou, R., Sanderson, B. J., Keefover-Ring, K., Yin, T., Smart, L. B., DiFazio, S. P., Liu, J., Olson, M., & Ma, T. (2022). Repeated turnovers keep sex chromosomes young in willows. *Genome Biology*, 23(1), 200. <https://doi.org/10.1186/s13059-022-02769-w>
646. Wilkerson, D. G., Crowell, C. R., Carlson, C. H., McMullen, P. W., Smart, C. D., & Smart, L. B. (2022). Comparative transcriptomics and eQTL mapping of response to *Melampsora americana* in selected *Salix purpurea* F2 progeny. *BMC Genomics*, 23(1), 71. <https://doi.org/10.1186/s12864-021-08254-1>
647. Wilkerson, D. G., Taskiran, B., Carlson, C. H., & Smart, L. B. (2022). Mapping the sex determination region in the *Salix* F1 hybrid common parent population confirms a ZW system in six diverse species. *G3 Genes/Genomes/Genetics*, 12(6), jkac071. <https://doi.org/10.1093/g3journal/jkac071>

648. Zhou, R., Macaya-Sanz, D., Carlson, C. H., Schmutz, J., Jenkins, J. W., Kudrna, D., Sharma, A., Sandor, L., Shu, S., Barry, K., Tuskan, G. A., Ma, T., Liu, J., Olson, M., Smart, L. B., & DiFazio, S. P. (2020). A willow sex chromosome reveals convergent evolution of complex palindromic repeats. *Genome Biology*, 21(1), 38. <https://doi.org/10.1186/s13059-020-1952-4>
649. Zinkgraf, M., Zhao, S., Canning, C., Gerttula, S., Lu, M., Filkov, V., & Groover, A. (2020). Evolutionary network genomics of wood formation in a phylogenetic survey of angiosperm forest trees. *New Phytologist*, 228(6), 1811–1823. (cross-listed with *Populus* and *Eucalyptus*) <https://doi.org/10.1111/nph.16819>

Plant Health, Resilience to Threats and Climate Change

650. Abbasi, M., Brar, G. S., & Aime, M. C. (2024). First report of *Melampsora ferrinii* (pucciniales) on babylon willow (*Salix babylonica*) in Iran. *Canadian Journal of Plant Pathology*, 46(1), 11–18. <https://doi.org/10.1080/07060661.2023.2266405>
651. Bennett, S. K., Lambert, A. M., Carey, S. P., & Braman, C. A. (2022). Evaluating pole cutting survival and growth for riparian forest restoration during invasion by polyphagous shot hole borer. *Restoration Ecology*, 30(5), e13578. (cross-listed with *Populus*) <https://doi.org/10.1111/rec.13578>
652. Crowell, C. R., Bekauri, M. M., Cala, A. R., McMullen, P., Smart, L. B., & Smart, C. D. (2020). Differential susceptibility of diverse *Salix* spp. To *Melampsora americana* and *Melampsora paradoxa*. *Plant Disease*, 104(11), 2949–2957. <https://doi.org/10.1094/PDIS-04-20-0718-RE>
653. Crowell, C. R., Wilkerson, D. G., Beckauri, M., Cala, A. R., McMullen, P. W., Mondo, S., Andreopoulos, W., Lipzen, A., Lail, K., Yan, M., Ng, V., Grigoriev, I. V., Smart, L. B., & Smart, C. D. (2022). The *Melampsora americana* population on *Salix purpurea* in the Great Lakes region is highly diverse with a contributory influence of clonality. *Phytopathology*®, 112(4), 907–916. <https://doi.org/10.1094/PHYTO-05-21-0201-R>
654. Crowell, C. R., Wilkerson, D. G., McMullen, P., Cala, A. R., Smart, L. B., & Smart, C. D. (2020). Population biology of *Melampsora americana* and eQTL mapping for rust resistance in *Salix purpurea* shrub willow. *Plant Health 2020*. APS Annual Meeting.
655. Crowell, C. R., Wilkerson, D. G., Smart, L. B., & Smart, C. D. (2022). Evidence of asexual overwintering of *Melampsora paradoxa* and mapping of stem rust host resistance in *Salix*. *Plants*, 11(18), 2385. <https://doi.org/10.3390/plants11182385>
656. Darsaraei, H., Khodaparast, S. A., Takamatsu, S., Abbasi, M., Asgari, B., Sajedi, S., Götz, M., Liu, S.-Y., Feng, J., Bradshaw, M., Bulgakov, T., & Braun, U. (2021). Phylogeny and taxonomy of the *Erysiphe adunca* complex (Erysiphaceae, Helotiales) on poplars and willows. *Mycological Progress*, 20(4), 517–537. (cross-listed with *Populus*) <https://doi.org/10.1007/s11557-021-01688-7>
657. Feng, J., Liu, S.-Y., Braun, U., Takamatsu, S., Bradshaw, M., Khodaparast, S. A., Bulgakov, T. S., Guan, G.-X., Zhao, F.-Y., & Tang, S.-R. (2022). Discovery of a cryptic species, *Erysiphe salicina* sp. Nov., and reconstruction of the phylogeny of powdery mildews on *Populus* and *Salix* spp. *Mycological Progress*, 21(6), 54. (cross-listed with *Populus*) <https://doi.org/10.1007/s11557-022-01793-1>

658. Houghton, D. C., Erickson, L. G., Kintz, R. C., Rowland, K. L., Gipe, N. C., & Adams, C. (2022). The distinctive plant and insect assemblages of an experimental forest in northern Lower Michigan (United States). *Environmental Entomology*, 51(4), 716–727. <https://doi.org/10.1093/ee/nvac035>
659. Keita, N., Bourgeois, B., Evette, A., Tisserant, M., González, E., Breton, V., Goulet, C., & Poulin, M. (2021). Growth response of cuttings to drought and intermittent flooding for three *Salix* species and implications for riverbank soil bioengineering. *Environmental Management*, 67(6), 1137–1144. <https://doi.org/10.1007/s00267-021-01444-3>
660. Konôpka, B., Pajtík, J., Bošeľa, M., Šebeň, V., & Shipley, L. A. (2020). Modeling forage potential for red deer (*Cervus elaphus*): A tree-level approach. *European Journal of Forest Research*, 139(3), 419–430. (cross-listed with *Populus*) <https://doi.org/10.1007/s10342-019-01250-x>
661. Konôpka, B., Šebeň, V., Pajtík, J., & Shipley, L. A. (2022). Influence of tree species and size on bark browsing by large wild herbivores. *Plants*, 11(21), 2925. (cross-listed with *Populus*) <https://doi.org/10.3390/plants11212925>
662. Mayes, M., Caylor, K. K., Singer, M. B., Stella, J. C., Roberts, D., & Nagler, P. (2020). Climate sensitivity of water use by riparian woodlands at landscape scales. *Hydrological Processes*, 34(25), 4884–4903. (cross-listed with *Populus*) <https://doi.org/10.1002/hyp.13942>
663. Wang, W., Carlson, C. H., Smart, L. B., & Carlson, J. E. (2020). Transcriptome analysis of contrasting resistance to herbivory by *Empoasca fabae* in two shrub willow species and their hybrid progeny. *PLOS ONE*, 15(7), e0236586. <https://doi.org/10.1371/journal.pone.0236586>
664. Zhu, L.-H., Xu, W., Huang, L., Ye, J.-R., & Li, D.-W. (2022). Pathogenicity and biological characteristics of *Septotinia populiperda* causing leaf blotch of willow. *Plant Disease*, 106(4), 1262–1270. <https://doi.org/10.1094/PDIS-07-21-1537-RE>

Production Systems for the Bioeconomy

665. Aggarwal, S., Magner, J., Srinivas, R., & Sajith, G. (2022). Managing nitrate-nitrogen in the intensively drained Upper Mississippi River Basin, USA under uncertainty: A perennial path forward. *Environmental Monitoring and Assessment*, 194(10), 704. (cross-listed with *Populus*) <https://doi.org/10.1007/s10661-022-10401-4>
666. Aleman, R. S., Marcia, J., Duque-Soto, C., Lozano-Sánchez, J., Montero-Fernández, I., Ruano, J. A., Hoskin, R. T., & Moncada, M. (2023). Effect of microwave and ultrasound-assisted extraction on the phytochemical and in vitro biological properties of willow (*Salix alba*) bark aqueous and ethanolic extracts. *Plants*, 12(13), 2533. <https://doi.org/10.3390/plants12132533>
667. Chahal, A., Ciolkosz, D. E., Puri, V., Liu, J., & Jacobson, M. (2023). A technoeconomic model for estimating costs of harvesting and debarking shrub willow. *Applied Engineering in Agriculture*, 39(4), 399–408. <https://doi.org/10.13031/aea.15454>
668. Chahal, A., Ciolkosz, D., Puri, V., Jacobson, M., & Liu, J. (2021). Mechanical characteristics of wood-bark interface of shrub willow. *Industrial Crops and Products*, 162, 113236. <https://doi.org/10.1016/j.indcrop.2020.113236>

669. Chahal, A., Ciolkosz, D., Puri, V., Liu, J., & Jacobson, M. (2020). Factors affecting wood-bark adhesion for debarking of shrub willow. *Biosystems Engineering*, 196, 202–209. <https://doi.org/10.1016/j.biosystemseng.2020.05.019>
670. Dahal, B., Poudel, K. P., Renninger, H. J., Granger, J. J., Leininger, T. D., Gardiner, E. S., Souter, R. A., & Rousseau, R. J. (2022). Aboveground biomass equations for black willow (*Salix nigra* Marsh.) and eastern cottonwood (*Populus deltoides* Bartr. Ex Marsh.). *Trees, Forests and People*, 7, 100195. (cross-listed with *Populus*) <https://doi.org/10.1016/j.tfp.2022.100195>
671. De Souza, D. P., Eisenbies, M. H., & Volk, T. A. (2022). Growing season harvests of shrub willow (*Salix* spp.) have higher nutrient removals and lower yields compared to dormant-season harvests. *Forests*, 13(11), 1936. <https://doi.org/10.3390/f13111936>
672. Díaz-Alba, D., Henry, A. L., García De Jalón, D., González Del Tánago, M., & Martínez-Fernández, V. (2023). *Salix* regeneration in fluvial landscapes: Empirical findings based on a systematic review. *Ecological Engineering*, 193, 107010. <https://doi.org/10.1016/j.ecoleng.2023.107010>
673. Eisenbies, M. H., Volk, T. A., DeSouza, D., Hallen, K., Stanton, B., Espinoza, J., Himes, A., Shuren, R., Stonex, R., Summers, B., & Zepa, J. (2021). An assessment of the harvesting and fuel performance of a single-pass cut-and-chip harvester in commercial-scale short-rotation poplar crops as influenced by crop and weather conditions. *Biomass and Bioenergy*, 149, 106075. <https://doi.org/10.1016/j.biombioe.2021.106075>
674. Eisenbies, M. H., Volk, T. A., De Souza, D. P. L., & Hallen, K. W. (2020). Cut-and-chip harvester material capacity and fuel performance on commercial-scale willow fields for varying ground and crop conditions. *GCB Bioenergy*, 12(6), 380–395. <https://doi.org/10.1111/gcbb.12679>
675. Elniski, A., Dongre, P., & Bujanovic, B. M. (2023). Lignin use in enhancing the properties of willow pellets. *Forests*, 14(10), 2041. <https://doi.org/10.3390/f14102041>
676. Englund, O., Dimitriou, I., Dale, V. H., Kline, K. L., Mola-Yudego, B., Murphy, F., English, B., McGrath, J., Busch, G., Negri, M. C., Brown, M., Goss, K., Jackson, S., Parish, E. S., Cacho, J., Zumpf, C., Quinn, J., & Mishra, S. K. (2020). Multifunctional perennial production systems for bioenergy: Performance and progress. *WIREs Energy and Environment*, 9(5), e375. (cross-listed with *Populus* and *Eucalyptus*) <https://doi.org/10.1002/wene.375>
677. Fabio, E. S., & Smart, L. B. (2020). Genetic and environmental influences on first rotation shrub willow (*Salix* spp.) bark and wood elemental composition. *BioEnergy Research*, 13(3), 797–809. <https://doi.org/10.1007/s12155-020-10122-x>
678. Frank, J., Therasme, O., Volk, T. A., Brown, T., Malmsheimer, R. W., Fortier, M.-O., Eisenbies, M. H., Ha, H., & Heavey, J. (2022). Integrated stochastic life cycle assessment and techno-economic analysis for shrub willow production in the northeastern United States. *Sustainability*, 14(15), 9007. <https://doi.org/10.3390/su14159007>
679. Gamble, J. D., Johnson, G. A., Current, D. A., Wyse, D. L., Zamora, D. S., & Sheaffer, C. C. (2020). Alley cropping affects perennial bioenergy crop root distribution, carbon, and nutrient stocks. *Agronomy Journal*, 112(5), 3718–3732. (cross-listed with *Populus*) <https://doi.org/10.1002/agj2.20350>

680. Goacher, R. E., Mottiar, Y., & Mansfield, S. D. (2021). ToF-SIMS imaging reveals that *p* - hydroxybenzoate groups specifically decorate the lignin of fibres in the xylem of poplar and willow. *Holzforschung*, 75(5), 452–462. (cross-listed with *Populus*) <https://doi.org/10.1515/hf-2020-0130>
681. Gouker, F. E., Fabio, E. S., Serapiglia, M. J., & Smart, L. B. (2021). Yield and biomass quality of shrub willow hybrids in differing rotation lengths and spacing designs. *Biomass and Bioenergy*, 146, 105977. <https://doi.org/10.1016/j.biombioe.2021.105977>
682. Jenderek, M. M., Ambruzs, B. D., Holman, G. E., Carstens, J. D., Ellis, D. D., & Widrlechner, M. P. (2020). *Salix* dormant bud cryotolerance varies by taxon, harvest year, and stem-segment length. *Crop Science*, 60(4), 1965–1973. <https://doi.org/10.1002/csc2.20135>
683. Karl, J. W., Yelich, J. V., Ellison, M. J., & Lauritzen, D. (2020). Estimates of willow (*Salix* spp.) canopy volume using unmanned aerial systems. *Rangeland Ecology & Management*, 73(4), 531–537. <https://doi.org/10.1016/j.rama.2020.03.001>
684. Li, W., Ciais, P., Stehfest, E., van Vuuren, D., Popp, A., Arneth, A., Di Fulvio, F., Doelman, J., Humpenöder, F., Harper, A. B., Park, T., Makowski, D., Havlik, P., Obersteiner, M., Wang, J., Krause, A., & Liu, W. (2020). Mapping the yields of lignocellulosic bioenergy crops from observations at the global scale. *Earth System Science Data*, 12(2), 789–804. (cross-listed with *Populus* and *Eucalyptus*) <https://doi.org/10.5194/essd-12-789-2020>
685. Mitu, M., Razus, D., Boldor, D., & Marculescu, C. (2023). Flammability properties of the pyrolysis gas generated from willow wood. *Processes*, 11(7), 2103. <https://doi.org/10.3390/pr11072103>
686. Montes, F., Fabio, E. S., Smart, L. B., Richard, T. L., Añó, R. M., & Kemanian, A. R. (2021). A semi-commercial case study of willow biomass production in the northeastern United States. *Agronomy Journal*, 113(2), 1287–1302. <https://doi.org/10.1002/agj2.20603>
687. Muklada, H., Fabio, E. S., & Smart, L. B. (2022). Growth, nitrogen uptake, and nutritional value of a diverse panel of shrub willow (*Salix* spp.) genotypes in response to nitrogen fertilization. *Agronomy*, 12(11), 2678. <https://doi.org/10.3390/agronomy12112678>
688. Nagardeolekar, A., Ovadias, M., Wang, K., & Bujanovic, B. (2020). Willow Lignin recovered from hot-water extraction for the production of hydrogels and thermoplastic blends. *ChemSusChem*, 13(17), 4702–4721. <https://doi.org/10.1002/cssc.202001259>
689. Ohlsson, J. A., Harman-Ware, A. E., Sandgren, M., & Schnürer, A. (2020). Biomass recalcitrance in willow under two biological conversion paradigms: Enzymatic hydrolysis and anaerobic digestion. *BioEnergy Research*, 13(1), 260–270. <https://doi.org/10.1007/s12155-019-10079-6>
690. Pečnik, J. G., Zouari, M., Schwarzkopf, M., & DeVallance, D. B. (2023). Utilization of torrefied and non-torrefied short rotation willow in wood–plastic composites. *Polymers*, 15(19), 3997. <https://doi.org/10.3390/polym15193997>
691. Sleight, N. J., Volk, T. A., & Eisenbies, M. (2023). Belowground biomass and root:shoot ratios of three willow cultivars at two sites. *Forests*, 14(3), 525. <https://doi.org/10.3390/f14030525>
692. Tamiminia, H., Salehi, B., Mahdianpari, M., Beier, C. M., Klimkowski, D. J., & Volk, T. A. (2021). Comparison of machine and deep learning methods to estimate shrub willow biomass from

UAS imagery. *Canadian Journal of Remote Sensing*, 47(2), 209–227.
<https://doi.org/10.1080/07038992.2021.1926952>

693. Tariq, Z., Volk, T. A., & Therasme, O. (2021). Life cycle assessment of a shrub willow evapotranspiration cover compared with conventional clay and geosynthetic covers in Upstate New York. *Environmental Science and Pollution Research*, 28(33), 45029–45040.
<https://doi.org/10.1007/s11356-021-13901-x>
694. Therasme, O., Volk, T. A., Eisenbies, M. H., Amidon, T. E., & Fortier, M.-O. (2021). Life cycle greenhouse gas emissions of ethanol produced via fermentation of sugars derived from shrub willow (*Salix* spp.) hot water extraction in the Northeast United States. *Biotechnology for Biofuels*, 14(1), 52. <https://doi.org/10.1186/s13068-021-01900-6>
695. Therasme, O., Volk, T. A., Eisenbies, M. H., San, H., & Usman, N. (2020). Hot water extracted and non-extracted willow biomass storage performance: Fuel quality changes and dry matter losses. *Frontiers in Energy Research*, 7, 165. <https://doi.org/10.3389/fenrg.2019.00165>
696. Wang, Y., Wang, J., Schuler, J., Hartley, D., Volk, T., & Eisenbies, M. (2020). Optimization of harvest and logistics for multiple lignocellulosic biomass feedstocks in the northeastern United States. *Energy*, 197, 117260. <https://doi.org/10.1016/j.energy.2020.117260>
697. Wood, C. D., Amidon, T. E., Volk, T. A., & Emerson, R. M. (2020). Hot water extraction: Short rotation willow, mixed hardwoods, and process considerations. *Energies*, 13(8), 2071. <https://doi.org/10.3390/en13082071>
698. Xu, J., Quackenbush, L. J., Volk, T. A., & Im, J. (2022). Estimation of shrub willow biophysical parameters across time and space from Sentinel-2 and unmanned aerial system (UAS) data. *Field Crops Research*, 287, 108655. <https://doi.org/10.1016/j.fcr.2022.108655>
699. Xu, J., Quackenbush, L. J., Volk, T. A., & Stehman, S. V. (2022). Shrub willow canopy chlorophyll content estimation from unmanned aerial systems (UAS) data: Estimation and uncertainty analysis across time, space, and scales. *International Journal of Applied Earth Observation and Geoinformation*, 108, 102737. <https://doi.org/10.1016/j.jag.2022.102737>
700. Xu, J., Volk, T. A., Quackenbush, L. J., & Stehman, S. V. (2021). Estimation of shrub willow leaf chlorophyll concentration across different growth stages using a hand-held chlorophyll meter to monitor plant health and production. *Biomass and Bioenergy*, 150, 106132. <https://doi.org/10.1016/j.biombioe.2021.106132>
701. Yuldashev, F., Illukpitiya, P., Tegegne, F., & Ekanem, E. (2020). Techno-economic analysis of plantation biomass production and small-scale wood pellet processing for bioenergy market. *International Wood Products Journal*, 11(4), 173–188. (cross-listed with *Populus* and *Eucalyptus*) <https://doi.org/10.1080/20426445.2020.1816766>
702. Zhong, L., Xu, M., Wang, C., Shao, L., Mao, J., Jiang, W., Ji, X., Yang, G., Chen, J., Lyu, G., Yoo, C. G., & Xu, F. (2020). Pretreatment of willow using the alkaline-catalyzed sulfolane/water solution for high-purity and antioxidative lignin production. *International Journal of Biological Macromolecules*, 159, 287–294. <https://doi.org/10.1016/j.ijbiomac.2020.05.074>
703. Zhong, L., Yang, L., Wang, C., Ji, X., Yang, G., Chen, J., Lyu, G., Xu, F., & Yoo, C. G. (2020). NaOH-aided sulfolane pretreatment for effective fractionation and utilization of willow (*Salix*

matsudana cv. Zhuliu). *Industrial & Engineering Chemistry Research*, 59(39), 17546–17553.
<https://doi.org/10.1021/acs.iecr.0c01208>

Environmental and Ecosystem Services

704. Al-Lami, M. K., Oustriere, N., Gonzales, E., & Burken, J. G. (2022). Phytomanagement of Pb/Zn/Cu tailings using biosolids-biochar or -humus combinations: Enhancement of bioenergy crop production, substrate functionality, and ecosystem services. *Science of The Total Environment*, 836, 155676. (cross-listed with *Populus*)
<https://doi.org/10.1016/j.scitotenv.2022.155676>
705. Amdoun, R., Bendifallah, N., Sahli, F., Moustafa, K., Hefferon, K., Makhzoum, A., & Khelifi, L. (2020). Improving zinc phytoremediation characteristics in *Salix pedicellata* with a new acclimation approach. *International Journal of Phytoremediation*, 22(7), 745–754.
<https://doi.org/10.1080/15226514.2019.1708862>
706. Canter, C. E., Zolton, K., Cacho, J. F., Negri, M. C., Zumpf, C. R., & Quinn, J. J. (2022). Impact of landscape design on the greenhouse gas emissions of shrub willow bioenergy buffers in a US midwest corn-production landscape. *Biofuels, Bioproducts and Biorefining*, 16(3), 629–639.
<https://doi.org/10.1002/bbb.2345>
707. Cao, Y., Ma, C., Chen, H., Chen, G., White, J. C., & Xing, B. (2020). Copper stress in flooded soil: Impact on enzyme activities, microbial community composition and diversity in the rhizosphere of *Salix integra*. *Science of The Total Environment*, 704, 135350.
<https://doi.org/10.1016/j.scitotenv.2019.135350>
708. Cao, Y., Ma, C., Chen, H., Zhang, J., White, J. C., Chen, G., & Xing, B. (2020). Xylem-based long-distance transport and phloem remobilization of copper in *Salix integra* Thunb. *Journal of Hazardous Materials*, 392, 122428. <https://doi.org/10.1016/j.jhazmat.2020.122428>
709. Caterino, B., Schuler, J. L., Grushecky, S. T., & Skousen, J. (2020). Early growth and survival of shrub willow on newly reclaimed mine soil. *New Forests*, 51(6), 1087–1099.
<https://doi.org/10.1007/s11056-020-09776-4>
710. Chen, Y.-H. (2022). Willows on the edge—Riparian vegetation growth and planting on anthropogenic point bars, Apalachicola River. *Southeastern Geographer*, 62(2), 85–88.
<https://doi.org/10.1353/sgo.2022.0008>
711. Cundy, A. B., LaFreniere, L., Bardos, R. P., Yan, E., Sedivy, R., & Roe, C. (2021). Integrated phytomanagement of a carbon tetrachloride-contaminated site in Murdock, Nebraska (USA). *Journal of Cleaner Production*, 290, 125190. (cross-listed with *Populus*)
<https://doi.org/10.1016/j.jclepro.2020.125190>
712. Greenplate, R., Thomas, S., Danley-Thomson, A., & Missimer, T. M. (2023). Phytoremediation potential of the coastal plain willow *Salix caroliniana* for heavy metals with emphasis on mercury. *Water*, 15(20), 3628. <https://doi.org/10.3390/w15203628>
713. Han, X., Zhang, Y., Yu, M., Zhang, J., Xu, D., Lu, Z., Qiao, G., Qiu, W., & Zhuo, R. (2020). Transporters and ascorbate–glutathione metabolism for differential cadmium accumulation

- and tolerance in two contrasting willow genotypes. *Tree Physiology*, 40(8), 1126–1142. <https://doi.org/10.1093/treephys/tpaa029>
714. Ho, J., & Chambers, L. G. (2020). Willow-shrub encroachment affects physicochemical properties differently in two subtropical freshwater marshes. *Wetlands Ecology and Management*, 28(2), 389–395. <https://doi.org/10.1007/s11273-019-09705-z>
 715. Jager, H. I., Nair, S. S., Efroymsen, R. A., DeRolph, C. R., Parish, E. S., & Wang, G. (2023). Ecosystem services from partially harvested riparian buffers can offset biomass production costs. *Science of The Total Environment*, 889, 164199. <https://doi.org/10.1016/j.scitotenv.2023.164199>
 716. Kloster, D. P., & Volk, T. A. (2023). Predicted soil loss from shrub willow production systems across the production cycle using the revised universal soil loss equation (RUSLE2). *BioEnergy Research*, 16(2), 912–923. <https://doi.org/10.1007/s12155-022-10477-3>
 717. Li, X., Tian, J., Li, X., Wang, L., Gong, H., Shi, C., Nie, S., Zhu, L., Chen, B., Pan, Y., He, J., Ni, R., & Diao, C. (2022). Developing a sub-meter phenological spectral feature for mapping poplars and willows in urban environment. *ISPRS Journal of Photogrammetry and Remote Sensing*, 193, 77–89. (cross-listed with *Populus*) <https://doi.org/10.1016/j.isprsjprs.2022.09.002>
 718. Li, X., Xiao, J., Ma, C., Salam, M. M. A., Shi, J., & Chen, G. (2021). The effect of particle size of bamboo biochar on the phytoremediation of *Salix psammophila* C. to multi-metal polluted soil. *International Journal of Phytoremediation*, 23(6), 658–668. <https://doi.org/10.1080/15226514.2020.1849012>
 719. Li, X., Xiao, J., Salam, M. M. A., Ma, C., & Chen, G. (2021). Impacts of bamboo biochar on the phytoremediation potential of *Salix psammophila* grown in multi-metals contaminated soil. *International Journal of Phytoremediation*, 23(4), 387–399. <https://doi.org/10.1080/15226514.2020.1816893>
 720. Liu, W., Xue, K., Hu, R., Zhou, J., Van Nostrand, J. D., Dimitrou, J., Giagnoni, L., & Renella, G. (2022). Long-term effects of soil remediation with willow short rotation coppice on biogeographic pattern of microbial functional genes. *Microorganisms*, 10(1), 140. <https://doi.org/10.3390/microorganisms10010140>
 721. Livingstone, D., Smyth, B. M., Lyons, G., Foley, A. M., Murray, S. T., & Johnston, C. (2022). Life cycle assessment of a short-rotation coppice willow riparian buffer strip for farm nutrient mitigation and renewable energy production. *Renewable and Sustainable Energy Reviews*, 158, 112154. <https://doi.org/10.1016/j.rser.2022.112154>
 722. Markus-Michalczyk, H., Crawford, M. L., & Baldwin, A. H. (2020). Salinity tolerance and occurrence of *Salix nigra* Marshall (black willow) in tidal wetlands of Chesapeake Bay tributaries. *Northeastern Naturalist*, 27(2), 229. <https://doi.org/10.1656/045.027.0205>
 723. Mer, K., Egiebor, N. O., Tao, W., Sajjadi, B., Wijethunga, U. K., & Leem, G. (2024). Capacitive removal of PB ions via electrosorption on novel willow biochar–manganese dioxide composites. *Environmental Technology*, 45(5), 999–1012. <https://doi.org/10.1080/09593330.2022.2135028>
 724. Salehi, A., Zalesny, R. S., & Calagari, M. (2023). Effects of urban wastewater application on growth, biomass, nutrition, and heavy-metal accumulation of *Populus nigra* L. “62/154,” *P.*

alba L. “20/45,” *P. euramericana* (Dode) Guinier “92/40,” and *Salix excelsa* S.G. Gmel grown in heavy-metal contaminated soil. *International Journal of Phytoremediation*, 25(10), 1371–1383. (cross-listed with *Populus*) <https://doi.org/10.1080/15226514.2022.2158783>

725. Vinhal, R. A., Zalesny, R. S., DeBauche, B. S., Rogers, E. R., Pilipović, A., Soolanayakanahally, R. Y., & Wiese, A. H. (2022). Establishment of willows using the novel DeValix technique: Ecological restoration mats designed for phytotechnologies. *International Journal of Phytoremediation*, 24(7), 730–743. <https://doi.org/10.1080/15226514.2021.1970102>
726. Wang, S., Volk, T. A., & Xu, J. (2021). Variability in growth and cadmium accumulation capacity among willow hybrids and their parents: Implications for yield-based selection of Cd-efficient cultivars. *Journal of Environmental Management*, 299, 113643. <https://doi.org/10.1016/j.jenvman.2021.113643>
727. Wolski, P., Blankenship, B. W., Umar, A., Cabrera, M., Simmons, B. A., Sale, K. L., & Achinivu, E. C. (2023). Factors that influence the activity of biomass-degrading enzymes in the presence of ionic liquids—A review. *Frontiers in Energy Research*, 11, 1212719. (cross-listed with *Eucalyptus* and *Populus*) <https://doi.org/10.3389/fenrg.2023.1212719>
728. Xue, K., Van Nostrand, J. D., Zhou, J., Neu, S., Müller, I., Giagnoni, L., & Renella, G. (2020). Phytomanagement reduces metal availability and microbial metal resistance in a metal contaminated soil. *Frontiers in Microbiology*, 11, 1899. <https://doi.org/10.3389/fmicb.2020.01899>
729. Yang, S., Volk, T., & Fortier, M.-O. (2020). Willow biomass crops are a carbon negative or low-carbon feedstock depending on prior land use and transportation distances to end users. *Energies*, 13(16), 4251. <https://doi.org/10.3390/en13164251>
730. Zumpf, C., Cacho, J., Grasse, N., Quinn, J., Hampton-Marcell, J., Armstrong, A., Campbell, P., Negri, M. C., & Lee, D. K. (2021). Influence of shrub willow buffers strategically integrated in an Illinois corn-soybean field on soil health and microbial community composition. *Science of The Total Environment*, 772, 145674. <https://doi.org/10.1016/j.scitotenv.2021.145674>
731. Zumpf, C., Quinn, J., Cacho, J., Grasse, N., Negri, M. C., & Lee, D. (2021). Invertebrate and plant community diversity of an Illinois corn–soybean field with integrated shrub willow bioenergy buffers. *Sustainability*, 13(21), 12280. <https://doi.org/10.3390/su132112280>

Eucalyptus

Taxonomy, Nomenclature, and Registration

None reported.

Domestication, Breeding, and Selection of Genetic Resources

732. Callister, A. N., Bermann, M., Elms, S., Bradshaw, B. P., Lourenco, D., & Brawner, J. T. (2022). Accounting for population structure in genomic predictions of *Eucalyptus globulus*. *G3 Genes/Genomes/Genetics*, 12(9), jkac180. <https://doi.org/10.1093/g3journal/jkac180>
733. Callister, A. N., Bradshaw, B. P., Elms, S., Gillies, R. A. W., Sasse, J. M., & Brawner, J. T. (2021). Single-step genomic blup enables joint analysis of disconnected breeding programs: An

example with *Eucalyptus globulus* Labill. *G3 Genes/Genomes/Genetics*, 11(10), jkab253.
<https://doi.org/10.1093/g3journal/jkab253>

734. Castro, C. A. D. O., Santos, G. A. D., Takahashi, E. K., Nunes, A. C. P., Souza, G. A. D., Resende, M. D. V. D., & Zanuncio, J. C. (2022). Top grafting to accelerate self-pollination in *Eucalyptus* breeding. *Revista Árvore*, 46, e4630. <https://doi.org/10.1590/1806-908820220000030>
735. Castro, C. A. de O., dos Santos, G. A., Takahashi, E. K., Nunes, A. C. P., Souza, G. A., & de Resende, M. D. V. (2021). Accelerating *Eucalyptus* breeding strategies through top grafting applied to young seedlings. *Industrial Crops and Products*, 171, 113906. <https://doi.org/10.1016/j.indcrop.2021.113906>
736. de Mattos, E. M., Binkley, D., Campoe, O. C., Alvares, C. A., & Stape, J. L. (2020). Variation in canopy structure, leaf area, light interception and light use efficiency among *Eucalyptus* clones. *Forest Ecology and Management*, 463, 118038. <https://doi.org/10.1016/j.foreco.2020.118038>
737. Elorriaga, E., Klocko, A. L., Ma, C., Du Plessis, M., An, X., Myburg, A. A., & Strauss, S. H. (2021). Genetic containment in vegetatively propagated forest trees: CRISPR disruption of *LEAFY* function in *Eucalyptus* gives sterile indeterminate inflorescences and normal juvenile development. *Plant Biotechnology Journal*, 19(9), 1743–1755. <https://doi.org/10.1111/pbi.13588>
738. Estopa, R. A., Paludeto, J. G. Z., Müller, B. S. F., De Oliveira, R. A., Azevedo, C. F., De Resende, M. D. V., Tambarussi, E. V., & Grattapaglia, D. (2023). Genomic prediction of growth and wood quality traits in *Eucalyptus benthamii* using different genomic models and variable SNP genotyping density. *New Forests*, 54(2), 343–362. <https://doi.org/10.1007/s11056-022-09924-y>
739. Ibarra, L., Hodge, G., & Acosta, J. J. (2023). Quantitative genetics of a hybrid population of *Eucalyptus nitens* × *Eucalyptus globulus*: Estimation of genetic parameters and implications for breeding strategies. *Forests*, 14(2), 381. <https://doi.org/10.3390/f14020381>
740. Jorge, G. L., De Paula, R. C., Mooney, B., Thelen, J. J., & Balbuena, T. S. (2023). Seasonal and water restriction-related changes in *Eucalyptus grandis* leaf proteins: Shedding light on the dark proteome. *Current Plant Biology*, 34, 100286. <https://doi.org/10.1016/j.cpb.2023.100286>
741. Lötter, A., Duong, T. A., Candotti, J., Mizrachi, E., Wegrzyn, J. L., & Myburg, A. A. (2023). Haplogenome assembly reveals structural variation in *Eucalyptus* interspecific hybrids. *GigaScience*, 12, giad064. <https://doi.org/10.1093/gigascience/giad064>
742. Mostert-O'Neill, M. M., Reynolds, S. M., Acosta, J. J., Lee, D. J., Borevitz, J. O., & Myburg, A. A. (2021). Genomic evidence of introgression and adaptation in a model subtropical tree species, *Eucalyptus grandis*. *Molecular Ecology*, 30(3), 625–638. <https://doi.org/10.1111/mec.15615>
743. Mostert-O'Neill, M. M., Tate, H., Reynolds, S. M., Mphahlele, M. M., Van Den Berg, G., Verry, S. D., Acosta, J. J., Borevitz, J. O., & Myburg, A. A. (2022). Genomic consequences of artificial selection during early domestication of a wood fibre crop. *New Phytologist*, 235(5), 1944–1956. <https://doi.org/10.1111/nph.18297>
744. Mphahlele, M. M., Isik, F., Mostert-O'Neill, M. M., Reynolds, S. M., Hodge, G. R., & Myburg, A. A. (2020). Expected benefits of genomic selection for growth and wood quality traits in

Eucalyptus grandis. *Tree Genetics & Genomes*, 16(4), 49. <https://doi.org/10.1007/s11295-020-01443-1>

745. Nagle, M. F., Nahata, S. S., Zahl, B., de Rivera, A. N., Tacker, X. V., Elorriaga, E., Ma, C., Goralogia, G. S., Klocko, A. L., Gordon, M., Joshi, S., & Strauss, S. H. (2023). Knockout of floral and meiosis genes using CRISPR/Cas9 produces male-sterility in *Eucalyptus* without impacts on vegetative growth. *Plant Direct*, 7(7), e507. <https://doi.org/10.1002/pld3.507>
746. Orr, A. J., Padovan, A., Kainer, D., Külheim, C., Bromham, L., Bustos-Segura, C., Foley, W., Haff, T., Hsieh, J.-F., Morales-Suarez, A., Cartwright, R. A., & Lanfear, R. (2020). A phylogenomic approach reveals a low somatic mutation rate in a long-lived plant. *Proceedings of the Royal Society B: Biological Sciences*, 287(1922), 20192364. <https://doi.org/10.1098/rspb.2019.2364>
747. Pereira, W. J., Pappas, M. D. C. R., Campoe, O. C., Stape, J. L., Grattapaglia, D., & Pappas Jr, G. J. (2020). Patterns of DNA methylation changes in elite *Eucalyptus* clones across contrasting environments. *Forest Ecology and Management*, 474, 118319. <https://doi.org/10.1016/j.foreco.2020.118319>
748. Pereira, W. J., Pappas, M. D. C. R., Grattapaglia, D., & Pappas, G. J. (2020). A cost-effective approach to DNA methylation detection by methyl sensitive DArT sequencing. *PLOS ONE*, 15(6), e0233800. <https://doi.org/10.1371/journal.pone.0233800>
749. Rocha, L. F., Benatti, T. R., de Siqueira, L., de Souza, I. C. G., Bianchin, I., de Souza, A. J., Fernandes, A. C. M., Oda, S., Stape, J. L., Yassue, R. M., Carvalho, H. F., Müller, N. A., Fladung, M., Acosta, J. J., Fritsche-Neto, R., & Tambarussi, E. V. (2022). Quantitative trait loci related to growth and wood quality traits in *Eucalyptus grandis* W. Hill identified through single- and multi-trait genome-wide association studies. *Tree Genetics & Genomes*, 18(6), 38. <https://doi.org/10.1007/s11295-022-01570-x>
750. Rockwood, D. L., Huber, D. A., Crawford, M. A., Rucks, P. C., Lamb, E., Fabbro, K. W., Minogue, P. J., Jump, R., & Hodge, G. R. (2022). *Eucalyptus amplifolia* and *Corymbia torelliana* in the southeastern USA: Genetic improvement and potential uses. *Forests*, 13(1), 75. <https://doi.org/10.3390/f13010075>
751. Shirazi, Z., Khakdan, F., Rafiei, F., Balalami, M. Y., & Ranjbar, M. (2023). Genome-wide identification and expression profile analysis of metal tolerance protein gene family in *Eucalyptus grandis* under metal stresses. *BMC Plant Biology*, 23(1), 240. <https://doi.org/10.1186/s12870-023-04240-9>
752. Sun, Y., Qiao, Z., Muchero, W., & Chen, J.-G. (2020). Lectin receptor-like kinases: The sensor and mediator at the plant cell surface. *Frontiers in Plant Science*, 11, 596301. (cross-listed with *Populus*) <https://doi.org/10.3389/fpls.2020.596301>
753. Visagie, C. M., Magistà, D., Ferrara, M., Balocchi, F., Duong, T. A., Eichmeier, A., Gramaje, D., Aylward, J., Baker, S. E., Barnes, I., Calhoun, S., De Angelis, M., Frisvad, J. C., Hakalova, E., Hayes, R. D., Houbraken, J., Grigoriev, I. V., LaButti, K., Leal, C., ... Wingfield, B. D. (2023). IMA genome-F18: The re-identification of *Penicillium* genomes available in NCBI and draft genomes for *Penicillium* species from dry cured meat, *Penicillium biforme*, *P. brevicompactum*, *P. solitum*, and *P. cvjetkovicii*, *Pewenomyces kutranfy*, *Pew. lalenivora*, *Pew. Tapulicola*, *Pew. kalosus*, *Teratosphaeria carnegiei*, and *Trichoderma atroviride* SC1. *IMA Fungus*, 14(1), 21. <https://doi.org/10.1186/s43008-023-00121-w>

754. Yost, J. M., Wise, S. L., Love, N. L. R., Steane, D. A., Jones, R. C., Ritter, M. K., & Potts, B. M. (2021). Origins, diversity and naturalization of *Eucalyptus globulus* (Myrtaceae) in California. *Forests*, 12(8), 1129. <https://doi.org/10.3390/f12081129>
755. Yang, X., Medford, J. I., Markel, K., Shih, P. M., De Paoli, H. C., Trinh, C. T., McCormick, A. J., Ployet, R., Hussey, S. G., Myburg, A. A., Jensen, P. E., Hassan, M. M., Zhang, J., Muchero, W., Kalluri, U. C., Yin, H., Zhuo, R., Abraham, P. E., Chen, J.-G., ... Tuskan, G. A. (2020). Plant biosystems design research roadmap 1.0. *BioDesign Research*, 2020, 8051764. (cross-listed with *Populus*) <https://doi.org/10.34133/2020/8051764>
756. Zinkgraf, M., Zhao, S., Canning, C., Gerttula, S., Lu, M., Filkov, V., & Groover, A. (2020). Evolutionary network genomics of wood formation in a phylogenetic survey of angiosperm forest trees. *New Phytologist*, 228(6), 1811–1823. (cross-listed with *Populus* and *Salix*) <https://doi.org/10.1111/nph.16819>

Plant Health, Resilience to Threats and Climate Change

757. Asao, S., Hayes, L., Aspinwall, M. J., Rymer, P. D., Blackman, C., Bryant, C. J., Cullerne, D., Egerton, J. J. G., Fan, Y., Innes, P., Millar, A. H., Tucker, J., Shah, S., Wright, I. J., Yvon-Durocher, G., Tissue, D., & Atkin, O. K. (2020). Leaf trait variation is similar among genotypes of *Eucalyptus camaldulensis* from differing climates and arises in plastic responses to the seasons rather than water availability. *New Phytologist*, 227(3), 780–793. <https://doi.org/10.1111/nph.16579>
758. Binkley, D., Campoe, O. C., Alvares, C. A., Carneiro, R. L., & Stape, J. L. (2020). Variation in whole-rotation yield among *Eucalyptus* genotypes in response to water and heat stresses: The techs project. *Forest Ecology and Management*, 462, 117953. <https://doi.org/10.1016/j.foreco.2020.117953>
759. Chen, X., Zhao, P., Ouyang, L., Zhu, L., Ni, G., & Schäfer, K. V. R. (2020). Whole-plant water hydraulic integrity to predict drought-induced *Eucalyptus urophylla* mortality under drought stress. *Forest Ecology and Management*, 468, 118179. <https://doi.org/10.1016/j.foreco.2020.118179>
760. Coleman, D., Windt, C. W., Buckley, T. N., & Merchant, A. (2023). Leaf relative water content at 50% stomatal conductance measured by noninvasive NMR is linked to climate of origin in nine species of eucalypt. *Plant, Cell & Environment*, 46(12), 3791–3805. <https://doi.org/10.1111/pce.14700>
761. du Toit, Y., Coles, D. W., Mewalal, R., Christie, N., & Naidoo, S. (2020). Ecalibrator: A comparative tool to identify key genes and pathways for *Eucalyptus* defense against biotic stressors. *Frontiers in Microbiology*, 11, 216. <https://doi.org/10.3389/fmicb.2020.00216>
762. Huang, W., Reddy, G. V. P., Shi, P., Huang, J., Hu, H., & Hu, T. (2020). Allelopathic effects of *Cinnamomum septentrionale* leaf litter on *Eucalyptus grandis* saplings. *Global Ecology and Conservation*, 21, e00872. <https://doi.org/10.1016/j.gecco.2019.e00872>
763. Mphahlele, M. M., Isik, F., Hodge, G. R., & Myburg, A. A. (2021). Genomic breeding for diameter growth and tolerance to *Leptocybe* gall wasp and *Botryosphaeria/Teratosphaeria*

fungal disease complex in *Eucalyptus grandis*. *Frontiers in Plant Science*, 12, 638969. <https://doi.org/10.3389/fpls.2021.638969>

764. Nelson, K. M., Bisbing, S., Grossenbacher, D. L., Ritter, M., & Yost, J. M. (2021). Testing an invasion mechanism for *Eucalyptus globulus*: Is there evidence of allelopathy? *American Journal of Botany*, 108(4), 607–615. <https://doi.org/10.1002/ajb2.1635>
765. Ochoa-Hueso, R., Carroll, R., Piñeiro, J., & Power, S. A. (2021). Understorey plant community assemblage of Australian *Eucalyptus* woodlands under elevated CO₂ is modulated by water and phosphorus availability. *Journal of Plant Ecology*, 14(3), 478–490. <https://doi.org/10.1093/jpe/rtab005>
766. Osburn, E. D., Moon, C., Stephenson, T., Kittipalawattanapol, K., Jones, M., Strickland, M. S., & Lynch, L. M. (2023). Disturbance of eucalypt forests alters the composition, function, and assembly of soil microbial communities. *FEMS Microbiology Ecology*, 99(9), fiad085. <https://doi.org/10.1093/femsec/fiad085>
767. Piñeiro, J., Ochoa-Hueso, R., Drake, J. E., Tjoelker, M. G., & Power, S. A. (2020). Water availability drives fine root dynamics in a *Eucalyptus* woodland under elevated atmospheric CO₂ concentration. *Functional Ecology*, 34(11), 2389–2402. <https://doi.org/10.1111/1365-2435.13660>
768. Plett, K. L., Kohler, A., Lebel, T., Singan, V. R., Bauer, D., He, G., Ng, V., Grigoriev, I. V., Martin, F., Plett, J. M., & Anderson, I. C. (2021). Intra-species genetic variability drives carbon metabolism and symbiotic host interactions in the ectomycorrhizal fungus *Pisolithus microcarpus*. *Environmental Microbiology*, 23(4), Article 4. <https://doi.org/10.1111/1462-2920.15320>
769. Ren, H., Huang, B., Fernández-García, V., Miesel, J., Yan, L., & Lv, C. (2020). Biochar and rhizobacteria amendments improve several soil properties and bacterial diversity. *Microorganisms*, 8(4), 502. <https://doi.org/10.3390/microorganisms8040502>
770. Ribeiro, M. F., Carvalho, V. R., Favoreto, A. L., De Marchi, B. R., Bello, V. H., Jordan, C., Soliman, E. P., Zanuncio, J. C., Sabattini, J. A., & Wilcken, C. F. (2023). Symbiotic bacteria in the relationship between *Anaphes nitens* (Hymenoptera: Mymaridae) and *Gonipteris platensis* (Coleoptera: Curculionidae). *Austral Ecology*, 48(1), 182–196. <https://doi.org/10.1111/aec.13259>
771. Richards, J. H., Henn, J. J., Sorenson, Q. M., Adams, M. A., Smith, D. D., McCulloh, K. A., & Givnish, T. J. (2021). Mistletoes and their eucalypt hosts differ in the response of leaf functional traits to climatic moisture supply. *Oecologia*, 195(3), 759–771. <https://doi.org/10.1007/s00442-021-04867-1>
772. Ruiz, A. M. M., Pieri, C. D., Pieroni, L. D. P., Porcena, A. S., Albuquerque, F. S., & Furtado, E. L. (2021). Diagrammatic scale for evaluating the severity of powdery mildew on eucalyptus. *Ciência Florestal*, 31(3), 1535–1546. <https://doi.org/10.5902/1980509840568>
773. Ryan, M. G., Stape, J. L., Binkley, D., & Alvares, C. A. (2020). Cross-site patterns in the response of *Eucalyptus* plantations to irrigation, climate and intra-annual weather variation. *Forest Ecology and Management*, 475, 118444. <https://doi.org/10.1016/j.foreco.2020.118444>
774. Salvi, A. M., Gosetti, S. G., Smith, D. D., Adams, M. A., Givnish, T. J., & McCulloh, K. A. (2022). Hydroscares, hydroscape plasticity and relationships to functional traits and mesophyll

- photosynthetic sensitivity to leaf water potential in *Eucalyptus* species. *Plant, Cell & Environment*, 45(9), 2573–2588. <https://doi.org/10.1111/pce.14380>
775. Salvi, A. M., Smith, D. D., Adams, M. A., McCulloh, K. A., & Givnish, T. J. (2021). Mesophyll photosynthetic sensitivity to leaf water potential in *Eucalyptus*: A new dimension of plant adaptation to native moisture supply. *New Phytologist*, 230(5), 1844–1855. <https://doi.org/10.1111/nph.17304>
 776. Sharma, A., & Raman, A. (2022). Gall-inducing Psylloidea (Insecta: Hemiptera) – plant interactions. *Journal of Plant Interactions*, 17(1), 580–594. <https://doi.org/10.1080/17429145.2022.2065371>
 777. Silva, V., Nogueira, T. A. R., Abreu-Junior, C. H., He, Z., Buzetti, S., Laclau, J.-P., Filho, M. C. M. T., Grilli, E., Murgia, I., & Capra, G. F. (2020). Influences of edaphoclimatic conditions on deep rooting and soil water availability in Brazilian *Eucalyptus* plantations. *Forest Ecology and Management*, 455, 117673. <https://doi.org/10.1016/j.foreco.2019.117673>
 778. Smith, D. D., Adams, M. A., Salvi, A. M., Krieg, C. P., Ané, C., McCulloh, K. A., & Givnish, T. J. (2023). Ecophysiological adaptations shape distributions of closely related trees along a climatic moisture gradient. *Nature Communications*, 14(1), 7173. <https://doi.org/10.1038/s41467-023-42352-w>
 779. Soares, A. A. V., Scolforo, H. F., Forrester, D. I., Carneiro, R. L., & Campoe, O. C. (2020). Exploring the relationship between stand growth, structure and growth dominance in *Eucalyptus* monoclonal plantations across a continent-wide environmental gradient in Brazil. *Forest Ecology and Management*, 474, 118340. <https://doi.org/10.1016/j.foreco.2020.118340>
 780. Teshome, D. T., Zharare, G. E., Ployet, R., & Naidoo, S. (2023). Transcriptional reprogramming during recovery from drought stress in *Eucalyptus grandis*. *Tree Physiology*, 43(6), 979–994. <https://doi.org/10.1093/treephys/tpad022>
 781. Toledo, F. H., McIntosh, T., Knothe, C., & Aubrey, D. P. (2020). *Eucalyptus* are unlikely to escape plantations and invade surrounding forests managed with prescribed fire in southeastern US. *Forests*, 11(6), 694. <https://doi.org/10.3390/f11060694>
 782. Tonet, V., Brodribb, T., & Bourbia, I. (2024). Variation in xylem vulnerability to cavitation shapes the photosynthetic legacy of drought. *Plant, Cell & Environment*, 47(4), 1160–1170. <https://doi.org/10.1111/pce.14788>
 783. Tupinambá-Simões, F., Bravo, F., Guerra-Hernández, J., & Pascual, A. (2022). Assessment of drought effects on survival and growth dynamics in eucalypt commercial forestry using remote sensing photogrammetry. A showcase in Mato Grosso, Brazil. *Forest Ecology and Management*, 505, 119930. <https://doi.org/10.1016/j.foreco.2021.119930>
 784. Wong-Bajracharya, J., Singan, V. R., Monti, R., Plett, K. L., Ng, V., Grigoriev, I. V., Martin, F. M., Anderson, I. C., & Plett, J. M. (2022). The ectomycorrhizal fungus *Pisolithus microcarpus* encodes a microRNA involved in cross-kingdom gene silencing during symbiosis. *Proceedings of the National Academy of Sciences*, 119(3), e2103527119. <https://doi.org/10.1073/pnas.2103527119>

Production Systems for the Bioeconomy

785. Acevedo, M., Rubilar, R., Dumroese, R. K., Ovalle, J. F., Sandoval, S., & Chassin-Trubert, R. (2021). Nitrogen loading of *Eucalyptus globulus* seedlings: Nutritional dynamics and influence on morphology and root growth potential. *New Forests*, 52(1), 31–46. <https://doi.org/10.1007/s11056-020-09778-2>
786. Asensio, V., Domec, J.-C., Nouvellon, Y., Laclau, J.-P., Bouillet, J.-P., Jordan-Meille, L., Lavres, J., Rojas, J. D., Guillemot, J., & Abreu-Junior, C. H. (2020). Potassium fertilization increases hydraulic redistribution and water use efficiency for stemwood production in *Eucalyptus grandis* plantations. *Environmental and Experimental Botany*, 176, 104085. <https://doi.org/10.1016/j.envexpbot.2020.104085>
787. Ayaz, I., Rizwan, M., Ullman, J. L., Haroon, H., Qayyum, A., Ahmed, N., Elesawy, B. H., Askary, A. E., Gharib, A. F., & Ismail, K. A. (2022). Lignocellulosic based biochar adsorbents for the removal of fluoride and arsenic from aqueous solution: Isotherm and kinetic modeling. *Polymers*, 14(4), 715. <https://doi.org/10.3390/polym14040715>
788. Bussoni, A., Cubbage, F., & Alvarez Giambruno, J. (2021). Silvopastoral systems and multi-criteria optimization for compatible economic and environmental outcomes. *Agricultural Systems*, 190, 103118. <https://doi.org/10.1016/j.agsy.2021.103118>
789. Campoe, O. C., & Binkley, D. (2020). Environmental and genetic influences on growth in *Eucalyptus* plantations: The TECHS special issue. *Forest Ecology and Management*, 476, 118464. <https://doi.org/10.1016/j.foreco.2020.118464>
790. Campoe, O. C., Alvares, C. A., Carneiro, R. L., Binkley, D., Ryan, M. G., Hubbard, R. M., Stahl, J., Moreira, G., Moraes, L. F., & Stape, J. L. (2020). Climate and genotype influences on carbon fluxes and partitioning in *Eucalyptus* plantations. *Forest Ecology and Management*, 475, 118445. <https://doi.org/10.1016/j.foreco.2020.118445>
791. Cerveira Junior, W. R., da Costa, Y. K. S., Carbonari, C. A., Duke, S. O., Alves, P. L. D. C. A., & Carvalho, L. B. D. (2020). Growth, morphological, metabolic and photosynthetic responses of clones of eucalyptus to glyphosate. *Forest Ecology and Management*, 470–471, 118218. <https://doi.org/10.1016/j.foreco.2020.118218>
792. Chu, S., Xian, L., Zhao, N., Lai, C., Yang, W., Wang, J., Long, M., Liao, D., Ouyang, J., Wang, Z., Jacobs, D. F., & Zeng, S. (2023). Combined addition of bagasse and zeolite stabilizes potentially toxic elements in sewage sludge compost and improves *Eucalyptus urophylla* seedling growth. *Forest Ecology and Management*, 539, 121003. <https://doi.org/10.1016/j.foreco.2023.121003>
793. Conti Junior, J. L. F., De Araujo, M. J., De Paula, R. C., Queiroz, T. B., Hakamada, R. E., & Hubbard, R. M. (2020). Quantifying turgor loss point and leaf water potential across contrasting *Eucalyptus* clones and sites within the TECHS research platform. *Forest Ecology and Management*, 475, 118454. <https://doi.org/10.1016/j.foreco.2020.118454>
794. da Silva, V. S., Silva, C. A., Mohan, M., Cardil, A., Rex, F. E., Loureiro, G. H., Almeida, D. R. A. D., Broadbent, E. N., Gorgens, E. B., Dalla Corte, A. P., Silva, E. A., Valbuena, R., & Klauber, C. (2020). Combined impact of sample size and modeling approaches for predicting stem volume in *Eucalyptus* spp. Forest plantations using field and LiDAR Data. *Remote Sensing*, 12(9), 1438. <https://doi.org/10.3390/rs12091438>

795. Davis, M. R., Kainer, D., Tuskan, G. A., Langholtz, M. H., Hellwinckel, C. M., Shedden, M., & Eaton, L. (2020). Modeled economic potential for *Eucalyptus* spp. Production for jet fuel additives in the United States. *Biomass and Bioenergy*, 143, 105807. <https://doi.org/10.1016/j.biombioe.2020.105807>
796. de Souza, H. J. P. L., Arantes, M. D. C., Vidaurre, G. B., Andrade, C. R., Carneiro, A. D. C. O., de Souza, D. P. L., & Protásio, T. D. P. (2020). Pelletization of eucalyptus wood and coffee growing wastes: Strategies for biomass valorization and sustainable bioenergy production. *Renewable Energy*, 149, 128–140. <https://doi.org/10.1016/j.renene.2019.12.015>
797. De Stefano, A., Blazier, M. A., Comer, C. E., Dean, T. J., & Wigley, T. B. (2019). Understory vegetation richness and diversity of *Eucalyptus benthamii* and *Pinus elliottii* plantations in the Midsouth US. *Forest Science*, 66(1), 66–81. <https://doi.org/10.1093/forsci/fxz051>
798. Dean, T. J. (2023). The role of height increment and marginal height cost in the production ecology of Eucalyptus plantations. *Forest Ecology and Management*, 532, 120846. <https://doi.org/10.1016/j.foreco.2023.120846>
799. Dhargay, S., Lyell, C. S., Brown, T. P., Inbar, A., Sheridan, G. J., & Lane, P. N. J. (2022). Performance of GEDI space-borne LiDAR for quantifying structural variation in the temperate forests of South-Eastern Australia. *Remote Sensing*, 14(15), 3615. <https://doi.org/10.3390/rs14153615>
800. Dix, M. J., & Aubrey, D. P. (2021). Calibration approach and range of observed sap flow influences transpiration estimates from thermal dissipation sensors. *Agricultural and Forest Meteorology*, 307, 108534. <https://doi.org/10.1016/j.agrformet.2021.108534>
801. Dong, C., Meng, X., Leu, S.-Y., Xu, L., Wu, Z., Cravotto, G., & Fang, Z. (2022). Enhancing α -etherification of lignin in Eucalyptus diol pretreatment to improve lignin monomer production. *Industrial Crops and Products*, 185, 115130. <https://doi.org/10.1016/j.indcrop.2022.115130>
802. Englund, O., Dimitriou, I., Dale, V. H., Kline, K. L., Mola-Yudego, B., Murphy, F., English, B., McGrath, J., Busch, G., Negri, M. C., Brown, M., Goss, K., Jackson, S., Parish, E. S., Cacho, J., Zumpf, C., Quinn, J., & Mishra, S. K. (2020). Multifunctional perennial production systems for bioenergy: Performance and progress. *WIREs Energy and Environment*, 9(5), e375. (cross-listed with *Populus* and *Salix*) <https://doi.org/10.1002/wene.375>
803. Fernandes, G. L., Casas, G. G., Fardin, L. P., Nogueira, G. S., Leite, R. V., Couto, L., & Leite, H. G. (2023). Effects of spacing on early growth rate and yield of hybrid *Eucalyptus* stands. *Pertanika Journal of Tropical Agricultural Science*, 46(2), 627–645. <https://doi.org/10.47836/pjtas.46.2.15>
804. Ferreira, G. W. D., Oliveira, F. C. C., Soares, E. M. B., Schnecker, J., Silva, I. R., & Grandy, A. S. (2021). Retaining eucalyptus harvest residues promotes different pathways for particulate and mineral-associated organic matter. *Ecosphere*, 12(3), e03439. <https://doi.org/10.1002/ecs2.3439>
805. Ferreto, D. O. C., Reichert, J. M., Cavalcante, R. B. L., & Srinivasan, R. (2021). Water budget fluxes in catchments under grassland and *Eucalyptus* plantations of different ages. *Canadian Journal of Forest Research*, 51(4), 513–523. <https://doi.org/10.1139/cjfr-2020-0156>

806. Ferreto, D. O. C., Reichert, J. M., Lopes Cavalcante, R. B., & Srinivasan, R. (2021). Rainfall partitioning in young clonal plantations *Eucalyptus* species in a subtropical environment, and implications for water and forest management. *International Soil and Water Conservation Research*, 9(3), 474–484. <https://doi.org/10.1016/j.iswcr.2021.01.002>
807. França, F. J. N., França, T. S. F. A., & Vidaurre, G. B. (2020). Effect of growth stress and interlocked grain on splitting of seven different hybrid clones of *Eucalyptus grandis*×*Eucalyptus urophylla* wood. *Holzforschung*, 74(10), 917–926. <https://doi.org/10.1515/hf-2019-0209>
808. Francis, R. C., Schnelle, S. T., Nicholson, D. J., Agblevor, F. A., & Kiemle, D. J. (2021). 2-Methyl-1,4,4a,9a-tetrahydroanthracene-9,10-dione (Me-THAD) as a catalyst in alkaline chemical pulping. *Journal of Wood Chemistry and Technology*, 41(6), 249–260. <https://doi.org/10.1080/02773813.2021.1976798>
809. Gimeno, T. E., Company, C. E., Drake, J. E., Barton, C. V. M., Tjoelker, M. G., Ubierna, N., & Marshall, J. D. (2021). Whole-tree mesophyll conductance reconciles isotopic and gas-exchange estimates of water-use efficiency. *New Phytologist*, 229(5), 2535–2547. <https://doi.org/10.1111/nph.17088>
810. Gonzalez-Benecke, C. A., Fernández, M. P., Albaugh, T. J., Ahumada, R., Bown, H. E., Gayoso, J., Gerding, V., Mardones, O. B., Rodríguez, A. R., & Rubilar, R. (2021). General above-stump volume and biomass functions for *Pinus radiata*, *Eucalyptus globulus* and *Eucalyptus nitens*. *Biomass and Bioenergy*, 155, 106280. <https://doi.org/10.1016/j.biombioe.2021.106280>
811. Gonzalez-Benecke, C. A., Fernández, M. P., Gayoso, J., Pincheira, M., & Wightman, M. G. (2022). Using tree height, crown area and stand-level parameters to estimate tree diameter, volume, and biomass of *Pinus radiata*, *Eucalyptus globulus* and *Eucalyptus nitens*. *Forests*, 13(12), 2043. <https://doi.org/10.3390/f13122043>
812. Grover, Z. S., Cook, R. L., Zapata, M., Byron Urrego, J., Albaugh, T. J., Zelaya, A., Ozyhar, T., Rubilar, R., Carter, D. R., & Campoe, O. C. (2021). *Eucalyptus grandis* response to calcium fertilization in Colombia. *Forest Science*, 67(6), 701–710. <https://doi.org/10.1093/forsci/fxab042>
813. Guillemot, J., Asensio, V., Bordron, B., Nouvellon, Y., Le Maire, G., Bouillet, J., Domec, J., Delgado Rojas, J. S., Abreu-Junior, C. H., Battie-Laclau, P., Cornut, I., Germon, A., De Moraes Gonçalves, J. L., Robin, A., & Laclau, J. (2021). Increased hydraulic constraints in *Eucalyptus* plantations fertilized with potassium. *Plant, Cell & Environment*, 44(9), 2938–2950. <https://doi.org/10.1111/pce.14102>
814. Hakamada, R., Binkley, D., Cegatta, I., Alvares, C., Campoe, O., & Stape, J. L. (2023). Stocking response of *Eucalyptus* growth depends on site water deficit across a 2100-km gradient in Brazil. *Forest Ecology and Management*, 546, 121325. <https://doi.org/10.1016/j.foreco.2023.121325>
815. Hakamada, R. E., Binkley, D., & Moré Mattos, E. (2023). Light use efficiency declines with water deficit and age in *Eucalyptus* plantations across Brazil. *Forest Ecology and Management*, 549, 121477. <https://doi.org/10.1016/j.foreco.2023.121477>

816. Hakamada, R. E., Hubbard, R. M., Moreira, G. G., Stape, J. L., Campoe, O., & Ferraz, S. F. D. B. (2020). Influence of stand density on growth and water use efficiency in *Eucalyptus* clones. *Forest Ecology and Management*, 466, 118125. <https://doi.org/10.1016/j.foreco.2020.118125>
817. Hakamada, R. E., Hubbard, R. M., Stape, J. L., Lima, W. D. P., Moreira, G. G., & Ferraz, S. F. D. B. (2020). Stocking effects on seasonal tree transpiration and ecosystem water balance in a fast-growing *Eucalyptus* plantation in Brazil. *Forest Ecology and Management*, 466, 118149. <https://doi.org/10.1016/j.foreco.2020.118149>
818. Hall, K. B., Stape, J., Bullock, B. P., Frederick, D., Wright, J., Scolforo, H. F., & Cook, R. (2019). A growth and yield model for *Eucalyptus benthamii* in the southeastern United States. *Forest Science*, 66(1), 25–37. <https://doi.org/10.1093/forsci/fxz061>
819. Heidari, A., Watkins, D., Mayer, A., Propato, T., Verón, S., & De Abelleira, D. (2021). Spatially variable hydrologic impact and biomass production tradeoffs associated with eucalyptus (*E. grandis*) cultivation for biofuel production in Entre Rios, Argentina. *GCB Bioenergy*, 13(5), 823–837. <https://doi.org/10.1111/gcbb.12815>
820. Huang, X., Liu, S., You, Y., Wang, J., Wen, Y., Shen, W., Tan, X., & Dahle, G. (2021). Different mechanisms underlying the divergent responses of soil respiration components to an introduction of N₂-fixer tree species into *Eucalyptus* plantations. *Agricultural and Forest Meteorology*, 308–309, 108536. <https://doi.org/10.1016/j.agrformet.2021.108536>
821. Hubbard, R. M., Carneiro, R. L., Campoe, O., Alvares, C. A., Figura, M. A., & Moreira, G. G. (2020). Contrasting water use of two *Eucalyptus* clones across a precipitation and temperature gradient in Brazil. *Forest Ecology and Management*, 475, 118407. <https://doi.org/10.1016/j.foreco.2020.118407>
822. Ile, O. J., McCormick, H., Skrabacz, S., Bhattacharya, S., Aguilos, M., Carvalho, H. D. R., Idassi, J., Baker, J., Heitman, J. L., & King, J. S. (2022). Integrating short rotation woody crops into conventional agricultural practices in the southeastern United States: A review. *Land*, 12(1), 10. (cross-listed with *Populus*) <https://doi.org/10.3390/land12010010>
823. Inkotte, J., Bomfim, B., Da Silva, S. C., Valadão, M. B. X., Da Rosa, M. G., Viana, R. B., Gatto, A., & Pereira, R. S. (2023). Litter removal impacts on soil biodiversity and eucalypt plantation development in the seasonal tropics. *Journal of Forestry Research*, 34(3), 735–748. <https://doi.org/10.1007/s11676-022-01524-y>
824. Iroumé, A., Jones, J., & Bathurst, J. C. (2021). Forest operations, tree species composition and decline in rainfall explain runoff changes in the nacimiento experimental catchments, South Central Chile. *Hydrological Processes*, 35(6), e14257. <https://doi.org/10.1002/hyp.14257>
825. Jardim, J. M., Hart, P. W., Lucia, L. A., Jameel, H., & Chang, H. (2022). The effect of the kraft pulping process, wood species, and pH on lignin recovery from black liquor. *Fibers*, 10(2), 16. <https://doi.org/10.3390/fib10020016>
826. Kruchelski, S., Trautenmüller, J. W., Deiss, L., Trevisan, R., Cabbage, F., Porfirio-da-Silva, V., & De Moraes, A. (2021). *Eucalyptus benthamii* Maiden et Cabbage growth and wood density in integrated crop-livestock systems. *Agroforestry Systems*, 95(8), 1577–1588. <https://doi.org/10.1007/s10457-021-00672-0>

827. Kulmann, M. S. D. S., Stefanello, L. O., Arruda, W. S., Sans, G. A., Parciannello, C. F., Hindersmann, J., Berghetti, Á. L. P., Araujo, M. M., Gatiboni, L. C., & Brunetto, G. (2020). Nitrogen supply methods affect the root growth dynamics in *Eucalyptus grandis*. *Forest Ecology and Management*, 473, 118320. <https://doi.org/10.1016/j.foreco.2020.118320>
828. Leite, R., Silva, C., Mohan, M., Cardil, A., Almeida, D., Carvalho, S., Jaafar, W., Guerra-Hernández, J., Weiskittel, A., Hudak, A., Broadbent, E., Prata, G., Valbuena, R., Leite, H., Taquetti, M., Soares, A., Scolforo, H., Amaral, C., Dalla Corte, A., & Klauberg, C. (2020). Individual tree attribute estimation and uniformity assessment in fast-growing *Eucalyptus* spp. Forest plantations using lidar and linear mixed-effects models. *Remote Sensing*, 12(21), 3599. <https://doi.org/10.3390/rs12213599>
829. Leite, R. V., Amaral, C. H. D., Pires, R. D. P., Silva, C. A., Soares, C. P. B., Macedo, R. P., Silva, A. A. L. D., Broadbent, E. N., Mohan, M., & Leite, H. G. (2020). Estimating stem volume in eucalyptus plantations using airborne LiDAR: A comparison of area- and individual tree-based approaches. *Remote Sensing*, 12(9), 1513. <https://doi.org/10.3390/rs12091513>
830. Li, W., Ciais, P., Stehfest, E., van Vuuren, D., Popp, A., Arneth, A., Di Fulvio, F., Doelman, J., Humpenöder, F., Harper, A. B., Park, T., Makowski, D., Havlik, P., Obersteiner, M., Wang, J., Krause, A., & Liu, W. (2020). Mapping the yields of lignocellulosic bioenergy crops from observations at the global scale. *Earth System Science Data*, 12(2), 789–804. (cross-listed with *Populus* and *Salix*) <https://doi.org/10.5194/essd-12-789-2020>
831. Lim, H., Alvares, C. A., Ryan, M. G., & Binkley, D. (2020). Assessing the cross-site and within-site response of potential production to atmospheric demand for water in *Eucalyptus* plantations. *Forest Ecology and Management*, 464, 118068. <https://doi.org/10.1016/j.foreco.2020.118068>
832. Macana, Y. A. M., Corrêa, R. S., de Toledo, F. H. S. F., Ferraz, A. de V., Ferreira, E. V. de O., Hakamada, R. E., Moreira, G. G., Arthur Junior, J. C., & Gonçalves, J. L. de M. (2023). Soil fertility, root growth, and Eucalypt productivity in response to lime and gypsum applications under soil water deficit. *New Forests*, 54(5), 833–852. <https://doi.org/10.1007/s11056-022-09943-9>
833. Masullo, L. S., Ferraz, A. D. V., Rocha, J. H. T., Valdo, G. D. C., Ávila, P. A. D., Florentino, A. L., Toledo, F. H. S. F. D., Alleoni, L. R. F., & Gonçalves, J. L. D. M. (2022). Soil Cu and Zn pools and their availability in response to forest residue management and micronutrient fertilization in a *Eucalyptus grandis* plantation. *Forest Ecology and Management*, 506, 119954. <https://doi.org/10.1016/j.foreco.2021.119954>
834. Masum, M. F. H., Dwivedi, P., & De La Torre, R. (2021). Assessing economic and environmental feasibility of wood-based electricity generation in South America: A case study from Colombia. *Forest Policy and Economics*, 124, 102381. <https://doi.org/10.1016/j.forpol.2020.102381>
835. Messick, E. J., Comer, C. E., Blazier, M. A., & Wigley, T. B. (2021). Avian breeding season diversity and community composition in camden white gum and slash pine plantations. *Forest Science*, 67(2), Article 2. <https://doi.org/10.1093/forsci/fxaa050>
836. Minogue, P. J., & Lorentz, K. A. (2021). Comparison of aminocyclopyrachlor to standard herbicides for basal stem treatment of *Eucalyptus benthamii*. *Weed Technology*, 35(2), 304–308. <https://doi.org/10.1017/wet.2020.111>

837. Neto, R. R. de O., Leite, H. G., Gleriani, J. M., & Strimbu, B. M. (2022). Estimation of Eucalyptus productivity using efficient artificial neural network. *European Journal of Forest Research*, 141(1), 129–151. <https://doi.org/10.1007/s10342-021-01431-7>
838. Oliveira, F. C. C., Ferreira, G. W. D., Dungait, J. A. J., Araújo, E. F., Soares, E. M. B., & Silva, I. R. (2021). Eucalypt harvest residue management influences microbial community structure and soil organic matter fractions in an afforested grassland. *Soil and Tillage Research*, 205, 104787. <https://doi.org/10.1016/j.still.2020.104787>
839. Oliveira, F. C. C., Ferreira, G. W. D., Santos, R. S., Aubrey, D. P., Araújo, E. F., & Silva, I. R. (2021). Soil CO₂ concentration, efflux, and partitioning in a recently afforested grassland. *New Forests*, 52(5), 737–757. <https://doi.org/10.1007/s11056-020-09824-z>
840. Oliveira, J. D. M., Tormena, C. A., Dos Santos, G. R., Zotarelli, L., Fernandes, R. B. A., & De Oliveira, T. S. (2022). Soil penetration resistance influenced by eucalypt straw management under mechanized harvesting. *Agronomy*, 12(6), 1482. <https://doi.org/10.3390/agronomy12061482>
841. Oliveira, L. A., Cardoso, A. A., Andrade, M. T., Pereira, T. S., Araújo, W. L., Santos, G. A., Damatta, F. M., & Martins, S. C. V. (2022). Exploring leaf hydraulic traits to predict drought tolerance of *Eucalyptus* clones. *Tree Physiology*, 42(9), Article 9. <https://doi.org/10.1093/treephys/tpac040>
842. Oliveira, L., Cecílio, R., Zanetti, S., Loos, R., Bressiani, D., & Srinivasan, R. (2020). Hydrological simulation of a small forested catchment under different land use and forest management. *iForest - Biogeosciences and Forestry*, 13(1), 301–308. <https://doi.org/10.3832/ifer3221-013>
843. Osuri, A. M., Gopal, A., Raman, T. R. S., DeFries, R., Cook-Patton, S. C., & Naeem, S. (2020). Greater stability of carbon capture in species-rich natural forests compared to species-poor plantations. *Environmental Research Letters*, 15(3), 034011. <https://doi.org/10.1088/1748-9326/ab5f75>
844. Ouyang, Y., Feng, G., Renninger, H., Leininger, T. D., Parajuli, P., & Grace, J. M. (2021). A STELLA-based model to simultaneously predict hydrological processes, N uptake and biomass production in a eucalyptus plantation. *Forests*, 12(5), 515. <https://doi.org/10.3390/f12050515>
845. Pascual, A., Tupinambá-Simões, F., & de Conto, T. (2022). Using multi-temporal tree inventory data in eucalypt forestry to benchmark global high-resolution canopy height models. A showcase in Mato Grosso, Brazil. *Ecological Informatics*, 70, 101748. <https://doi.org/10.1016/j.ecoinf.2022.101748>
846. Pascual, A., Tupinambá-Simões, F., Guerra-Hernández, J., & Bravo, F. (2022). High-resolution planet satellite imagery and multi-temporal surveys to predict risk of tree mortality in tropical eucalypt forestry. *Journal of Environmental Management*, 310, 114804. <https://doi.org/10.1016/j.jenvman.2022.114804>
847. Peng, J., Yang, G., Qi, L., Liu, J., Li, F., Chen, J., & Lucia, L. (2020). Enhancement of delignification by ionic liquids pretreatment and modification of hardwood kraft pulp in preparation for bleaching. *BioResources*, 15(3), 6299–6308. <https://doi.org/10.15376/biores.15.3.6299-6308>

848. Pereira, A. P. D. A., Santana, M. C., Zagatto, M. R. G., Brandani, C. B., Wang, J.-T., Verma, J. P., Singh, B. K., & Cardoso, E. J. B. N. (2021). Nitrogen-fixing trees in mixed forest systems regulate the ecology of fungal community and phosphorus cycling. *Science of The Total Environment*, 758, 143711. <https://doi.org/10.1016/j.scitotenv.2020.143711>
849. Peters, J. M. R., Gauthey, A., Lopez, R., Carins-Murphy, M. R., Brodribb, T. J., & Choat, B. (2020). Non-invasive imaging reveals convergence in root and stem vulnerability to cavitation across five tree species. *Journal of Experimental Botany*, 71(20), 6623–6637. <https://doi.org/10.1093/jxb/eraa381>
850. Piñeiro, J., Pathare, V., Ochoa-Hueso, R., Carrillo, Y., & Power, S. A. (2022). No CO₂ fertilization effect on plant growth despite enhanced rhizosphere enzyme activity in a low phosphorus soil. *Plant and Soil*, 471(1–2), 359–374. <https://doi.org/10.1007/s11104-021-05175-6>
851. Pinheiro, F. M., Nair, P. K. R., Nair, V. D., Tonucci, R. G., & Venturin, R. P. (2021). Soil carbon stock and stability under Eucalyptus-based silvopasture and other land-use systems in the Cerrado biodiversity hotspot. *Journal of Environmental Management*, 299, 113676. <https://doi.org/10.1016/j.jenvman.2021.113676>
852. Pizarro, R., Valdés-Pineda, R., Garcia-Chevesich, P. A., Ibáñez, A., Pino, J., Scott, D. F., Neary, D. G., McCray, J. E., Castillo, M., & Ubilla, P. (2022). The large-scale effect of forest cover on long-term streamflow variations in Mediterranean catchments of Central Chile. *Sustainability*, 14(8), Article 8. <https://doi.org/10.3390/su14084443>
853. Queiroz, T. B., Campoe, O. C., Montes, C. R., Alvares, C. A., Cuartas, M. Z., & Guerrini, I. A. (2020). Temperature thresholds for *Eucalyptus* genotypes growth across tropical and subtropical ranges in South America. *Forest Ecology and Management*, 472, 118248. <https://doi.org/10.1016/j.foreco.2020.118248>
854. Queiroz, T. B., Montes, C. R., & Campoe, O. C. (2023). A new method to calibrate cardinal temperatures for eucalyptus plantation †. *Forests*, 14(8), 1631. <https://doi.org/10.3390/f14081631>
855. Rachid, C. T. C. C., Balieiro, F. C., Peixoto, R. S., Fonseca, E. S., Jesus, H. E., Novotny, E. H., Chaer, G. M., Santos, F. M., Tiedje, J. M., & Rosado, A. S. (2023). Mycobiome structure does not affect field litter decomposition in *Eucalyptus* and *Acacia* plantations. *Frontiers in Microbiology*, 14, 1106422. <https://doi.org/10.3389/fmicb.2023.1106422>
856. Reichert, J. M., Morales, C. A. S., Lima, E. M., De Bastos, F., Sampietro, J. A., De Araújo, E. F., & Srinivasan, R. (2021). Best tillage practices for early-growth of clonal eucalyptus in soils with distinct granulometry, drainage and profile depth. *Soil and Tillage Research*, 212, 105038. <https://doi.org/10.1016/j.still.2021.105038>
857. Reichert, J. M., Prevedello, J., Gubiani, P. I., Vogelmann, E. S., Reinert, D. J., Consensa, C. O. B., Soares, J. C. W., & Srinivasan, R. (2021). Eucalyptus tree stockings effect on water balance and use efficiency in subtropical sandy soil. *Forest Ecology and Management*, 497, 119473. <https://doi.org/10.1016/j.foreco.2021.119473>
858. Renchon, A. A., Drake, J. E., Macdonald, C. A., Sihi, D., Hinko-Najera, N., Tjoelker, M. G., Arndt, S. K., Noh, N. J., Davidson, E., & Pendall, E. (2021). Concurrent measurements of soil and ecosystem respiration in a mature eucalypt woodland: Advantages, lessons, and questions.

859. Rivera, E. C., Assumpção, D. C., Kwon, H. J., Okonkwo, C. C., Ezeji, T. C., Filho, R. M., & Mariano, A. P. (2023). Mechanistic modeling of redox balance effects on the fermentation of eucalyptus wood-derived xylose to acetone-butanol-ethanol. *Biochemical Engineering Journal*, 190, 108738. <https://doi.org/10.1016/j.bej.2022.108738>
860. Rockwood, D. L., Tamang, B., & Ellis, M. F. (2023). Field drying for enhancing biomass quality of *Eucalyptus* logs and trees in Florida, USA. *Forests*, 14(5), 899. <https://doi.org/10.3390/f14050899>
861. Rockwood, D. L., Winandy, J. E., & Gribbins, N. R. (2022). Processing and wood factors influence medium density fiberboard production from young *Eucalyptus grandis*, *E. amplifolia*, *Corymbia torelliana*, and cottonwood grown in Florida USA. *Forests*, 13(2), 266. <https://doi.org/10.3390/f13020266>
862. Rodrigues, B. P., Oliveira, J. T. da S., Demuner, B. J., Mafia, R. G., & Vidaurre, G. B. (2022). Chemical and kraft pulping properties of young eucalypt trees affected by physiological disorders. *Forests*, 13(4), 504. <https://doi.org/10.3390/f13040504>
863. Rozas, C., Sinha, A., Montero, C., Gutierrez, P. T. V., & Vasquez, M. (2021). Prediction of end splitting in log of *Eucalyptus nitens* (H. Deane & Maiden) Maiden through regression models using longitudinal residual strain, physical and dendrometric properties. *Wood and Fiber Science*, 53(2), 79–88. <https://doi.org/10.22382/wfs-2021-10>
864. Rozas, C., Vasquez, M., Vega, P. T., Sinha, A., & Montero, C. (2021). Effect of log heat treatment on release of growth stresses in *Eucalyptus nitens*. *Wood and Fiber Science*, 53(3), 178–193. <https://doi.org/10.22382/wfs-2021-18>
865. Rubilar, R., Hubbard, R., Emhart, V., Mardones, O., Quiroga, J. J., Medina, A., Valenzuela, H., Espinoza, J., Burgos, Y., & Bozo, D. (2020). Climate and water availability impacts on early growth and growth efficiency of Eucalyptus genotypes: The importance of GxE interactions. *Forest Ecology and Management*, 458, 117763. <https://doi.org/10.1016/j.foreco.2019.117763>
866. Salem, K. S., Naithani, V., Jameel, H., Lucia, L., & Pal, L. (2021). Lignocellulosic fibers from renewable resources using green chemistry for a circular economy. *Global Challenges*, 5(2), 2000065. <https://doi.org/10.1002/gch2.202000065>
867. Santos, R. S., Oliveira, F. C. C., Ferreira, G. W. D., Ferreira, M. A., Araújo, E. F., & Silva, I. R. (2020). Carbon and nitrogen dynamics in soil organic matter fractions following eucalypt afforestation in southern Brazilian grasslands (*Pampas*). *Agriculture, Ecosystems & Environment*, 301, 106979. <https://doi.org/10.1016/j.agee.2020.106979>
868. Sarto, M. V. M., Borges, W. L. B., Bassegio, D., Nunes, M. R., Rice, C. W., & Rosolem, C. A. (2022). Deep soil water content and forage production in a tropical agroforestry system. *Agriculture*, 12(3), 359. <https://doi.org/10.3390/agriculture12030359>
869. Sarto, M. V. M., Borges, W. L. B., Sarto, J. R. W., Pires, C. A. B., Rice, C. W., & Rosolem, C. A. (2020). Soil microbial community and activity in a tropical integrated crop-livestock system. *Applied Soil Ecology*, 145, 103350. <https://doi.org/10.1016/j.apsoil.2019.08.012>

870. Sarto, M. V. M., Borges, W. L. B., Sarto, J. R. W., Rice, C. W., & Rosolem, C. A. (2020a). Deep soil carbon stock, origin, and root interaction in a tropical integrated crop–livestock system. *Agroforestry Systems*, 94(5), 1865–1877. <https://doi.org/10.1007/s10457-020-00505-6>
871. Sarto, M. V. M., Borges, W. L. B., Sarto, J. R. W., Rice, C. W., & Rosolem, C. A. (2020b). Root and shoot interactions in a tropical integrated crop–livestock–forest system. *Agricultural Systems*, 181, 102796. <https://doi.org/10.1016/j.agsy.2020.102796>
872. Schmidt, L. N., Sanquetta, M. N. I., McTague, J. P., da Silva, G. F., Fraga Filho, C. V., Sanquetta, C. R., & Soares Scolforo, J. R. (2020). On the use of the Weibull distribution in modeling and describing diameter distributions of clonal eucalypt stands. *Canadian Journal of Forest Research*, 50(10), 1050–1063. <https://doi.org/10.1139/cjfr-2020-0051>
873. Scolforo, H. F., McTague, J. P., Burkhart, H., Roise, J., Alvares, C. A., & Stape, J. L. (2020). Site index estimation for clonal eucalypt plantations in Brazil: A modeling approach refined by environmental variables. *Forest Ecology and Management*, 466, 118079. <https://doi.org/10.1016/j.foreco.2020.118079>
874. Sena, K. N., Boni, T. S., Maltoni, K. L., Cassiolato, A. M. R., & Pereira, E. I. P. (2023). Post-harvest Eucalyptus residue removal reduces soil aggregation and biological activities in Central-West Brazil. *Sustainability*, 15(11), 8790. <https://doi.org/10.3390/su15118790>
875. Shi, L., Zhang, T., Zhou, X., Yao, L., Yang, L., Yue, F., Lan, W., & Lu, F. (2022). Isolation, characterization, and depolymerization of l -cysteine substituted *Eucalyptus* lignin. *Global Challenges*, 6(4), 2100130. <https://doi.org/10.1002/gch2.202100130>
876. Silva, L. E., dos Santos, A. D. A., Torres, L., McCaffrey, Z., Klamczynski, A., Glenn, G., Neto, A. R. de S., Wood, D., Williams, T., Orts, W., Damásio, R. A. P., & Tonoli, G. H. D. (2021). Redispersion and structural change evaluation of dried microfibrillated cellulose. *Carbohydrate Polymers*, 252, 117165. <https://doi.org/10.1016/j.carbpol.2020.117165>
877. Silva, L. E., Simson, R., Torres, L., Hart-Cooper, W., Cao, T., Klamczynski, A., Glenn, G., Neto, A. R. de S., Williams, T., Wood, D., Orts, W., & Tonoli, G. H. D. (2023). Sodium chloride and sodium dodecyl sulfate as additives to enhance dispersibility in microfibrillated cellulose. *Cellulose*, 30(17), 10923–10934. <https://doi.org/10.1007/s10570-023-05555-4>
878. Silva, V., Nogueira, T. A. R., Abreu-Junior, C. H., Dilipkumar Jani, A., Buzetti, S., Filho, M. C. M. T., He, Z., Ganga, A., Penido, T. M. A., da Silva, A. R., & Capra, G. F. (2021). Variability in soil physical–chemical properties along the root-explored profile in deep Oxisols of commercial eucalypt plantations. *Forest Ecology and Management*, 494, 119334. <https://doi.org/10.1016/j.foreco.2021.119334>
879. Souza, I. F., Barros, N. F., Novais, R. F., Vergütz, L., & Silva, I. R. (2020). Potential contribution of eucalypt harvest residues to soil organic carbon in Brazil. *New Forests*, 51(6), 911–932. <https://doi.org/10.1007/s11056-019-09765-2>
880. Squizani, W., de Paula, B. V., Stefanello, L., Rozane, D. E., de Araujo, E. F., Natale, W., & Brunetto, G. (2023). Assessing the nutritional status of Southern Brazilian *Eucalyptus* plantations by the CND method. *Canadian Journal of Forest Research*, 53(6), 407–415. <https://doi.org/10.1139/cjfr-2022-0210>

881. Stape, J. L., Silva, C. R., & Binkley, D. (2022). Spacing and geometric layout effects on the productivity of clonal *Eucalyptus* plantations. *Trees, Forests and People*, 8, 100235. <https://doi.org/10.1016/j.tfp.2022.100235>
882. Tariq, K. A., Haroon, M., Mughal, A. H., & Ali, S. (2022). Physical and mechanical characterization of structural wood used in Pakistan. *Wood Research*, 67(1), 134–146. <https://doi.org/10.37763/wr.1336-4561/67.1.134146>
883. Taskhiri, M. S., Hafezi, M. H., Harle, R., Williams, D., Kundu, T., & Turner, P. (2020). Ultrasonic and thermal testing to non-destructively identify internal defects in plantation eucalypts. *Computers and Electronics in Agriculture*, 173, 105396. <https://doi.org/10.1016/j.compag.2020.105396>
884. Tuncer, F. D., Kartal, S. N., Soytürk, E. E., Arango, R. A., Ohno, K. M., Önses, M. S., Çelik, N., & Ibanez, C. M. (2024). Changes in chemical properties and microstructure of *Pinus taeda* and *Eucalyptus bosistoana* woods modified by contact charring. *European Journal of Wood and Wood Products*, 82(1), Article 1. <https://doi.org/10.1007/s00107-023-02011-9>
885. Vallejos, M. E., Olmos, G. V., Taleb, M. C., Felissia, F. E., Ehman, N. V., Peresin, M. S., Area, M. C., & Maximino, M. G. (2022). Dissolving pulp from eucalyptus sawdust for regenerated cellulose products. *Cellulose*, 29(8), 4645–4659. <https://doi.org/10.1007/s10570-022-04581-y>
886. Van Duong, D., & Schimleck, L. (2022). Prediction of static bending properties of *Eucalyptus* clones using stress wave measurements on standing trees, logs and small clear specimens. *Forests*, 13(10), 1728. <https://doi.org/10.3390/f13101728>
887. Waldy, J., Kershaw, J. A., Weiskittel, A., & Ducey, M. J. (2021). Comparison of time-based versus state–space stand growth models for tropical hybrid *Eucalyptus* clonal plantations in Sumatera, Indonesia. *Canadian Journal of Forest Research*, 51(8), 1178–1187. <https://doi.org/10.1139/cjfr-2020-0499>
888. Waldy, J., Kershaw Jr, J. A., Weiskittel, A., & Ducey, M. J. (2022). Diameter distribution model development of tropical hybrid *Eucalyptus* clonal plantations in Sumatera, Indonesia: A comparison of estimation methods. *New Zealand Journal of Forestry Science*, 52. <https://doi.org/10.33494/nzifs522022x151x>
889. Wang, Y., Lin, Y., Zhang, L., Liu, S., Wang, J., Tian, Y., Campbell, D. E., Lin, R., Ren, H., & Lu, H. (2024). Long-term effects of intercropping on multi-trophic structure and bio-thermodynamic health of mixed *Eucalyptus*-native tree plantations. *Journal of Applied Ecology*, 61(1), 103–119. <https://doi.org/10.1111/1365-2664.14558>
890. Wille, V. K. D., Gentil, M., Nunes, G. R. S., da Rosa, R. C., Jardim, J. M., Berger, C., Costa, H. W. D., Gatto, D. A., & Pedrazzi, C. (2023). Totora fibers as a new source for papermaking. *Biomass Conversion and Biorefinery*, 13(6), 5235–5241. <https://doi.org/10.1007/s13399-021-01547-1>
891. Wongchai, W., Onsree, T., Sukkam, N., Promwungkwa, A., & Tippayawong, N. (2022). Machine learning models for estimating above ground biomass of fast growing trees. *Expert Systems with Applications*, 199, 117186. <https://doi.org/10.1016/j.eswa.2022.117186>
892. Wood, K. C., Kjellow, A. W., Konkler, M. J., Presley, G., & Morrell, J. J. (2023). Preservative treatment of Tasmanian plantation *Eucalyptus nitens* using supercritical fluids. *Wood and Fiber Science*, 55(1), 83–93. <https://doi.org/10.22382/wfs-2023-08>

893. Ye, Q., Wang, Y.-H., Zhang, Z.-T., Huang, W.-L., Li, L.-P., Li, J., Liu, J., Zheng, Y., Mo, J.-M., Zhang, W., & Wang, J.-J. (2020). Dissolved organic matter characteristics in soils of tropical legume and non-legume tree plantations. *Soil Biology and Biochemistry*, 148, 107880. <https://doi.org/10.1016/j.soilbio.2020.107880>
894. Younger, S. E., Jackson, C. R., Dix, M. J., Caldwell, P. V., & Aubrey, D. P. (2023). Evapotranspiration partitioning of *Eucalyptus benthamii* and *Pinus taeda* during early stand development. *BioEnergy Research*, 16(4), 2204–2218. <https://doi.org/10.1007/s12155-023-10591-w>
895. Yuldashev, F., Illukpitiya, P., Tegegne, F., & Ekanem, E. (2020). Techno-economic analysis of plantation biomass production and small-scale wood pellet processing for bioenergy market. *International Wood Products Journal*, 11(4), 173–188. (cross-listed with *Populus* and *Salix*) <https://doi.org/10.1080/20426445.2020.1816766>
896. Zhang, C., Xiao, X., Zhao, L., Qin, Y., Doughty, R., Wang, X., Dong, J., & Yang, X. (2023). Mapping Eucalyptus plantation in Guangxi, China by using knowledge-based algorithms and PALSAR-2, Sentinel-2, and Landsat images in 2020. *International Journal of Applied Earth Observation and Geoinformation*, 120, 103348. <https://doi.org/10.1016/j.jag.2023.103348>
897. Zhuang, J., Liu, C., Xia, Y., Li, Y., Hou, Y., & Yoo, C. G. (2022). Synergistic effects of peracetic acid and oxalic acid on structural transformation of eucalyptus and its conversion. *Industrial Crops and Products*, 188, 115530. <https://doi.org/10.1016/j.indcrop.2022.115530>
898. Zolin, C. A., Matos, E. D. S., Magalhães, C. A. D. S., Paulino, J., Lal, R., Spera, S. T., & Behling, M. (2021). Short-term effect of a crop-livestock-forestry system on soil, water and nutrient loss in the Cerrado-Amazon ecotone. *Acta Amazonica*, 51(2), 102–112. <https://doi.org/10.1590/1809-4392202000391>

Environmental and Ecosystem Services

899. Abreu-Junior, C. H., Oliveira, M. G. D., Cardoso, P. H. S., Mandu, T. D. S., Florentino, A. L., Oliveira, F. C., Reis, J. V. D., Alvares, C. A., Stape, J. L., Nogueira, T. A. R., Capra, G. F., & He, Z. (2020). Sewage sludge application in *Eucalyptus urograndis* plantation: Availability of phosphorus in soil and wood production. *Frontiers in Environmental Science*, 8, 116. <https://doi.org/10.3389/fenvs.2020.00116>
900. Bazgir, M., Omidipour, R., Heydari, M., Zainali, N., Hamidi, M., & Dey, D. C. (2020). Prioritizing woody species for the rehabilitation of arid lands in western iran based on soil properties and carbon sequestration. *Journal of Arid Land*, 12(4), 640–652. (cross-listed with *Populus*) <https://doi.org/10.1007/s40333-020-0013-x>
901. Cardoso, P. H. S., Neto, A. D., Oliveira, F. C., Tomazello-Filho, M., Guedes, F. T. P., Alvares, C. A., Stape, J. L., Nogueira, T. A. R., Capra, G. F., Jani, A. D., He, Z., & Abreu-Junior, C. H. (2022). Fertilization management with sewage sludge sustains *Eucalyptus* productivity in Cerrado infertile soil. *Forest Ecology and Management*, 519, 120338. <https://doi.org/10.1016/j.foreco.2022.120338>
902. Chu, S., Yang, W., Xia, D., Liang, Z., Su, S., Zhao, N., Wang, J., Long, M., Ouyang, J., Liao, D., Jacobs, D. F., & Zeng, S. (2021). Changes in potentially toxic element concentration and

potential ecological risk in topsoil caused by sewage sludge application on forestland: A 3-year field trial. *Forest Ecology and Management*, 500, 119657.

<https://doi.org/10.1016/j.foreco.2021.119657>

903. Domingues, R. R., Sánchez-Monedero, M. A., Spokas, K. A., Melo, L. C. A., Trugilho, P. F., Valenciano, M. N., & Silva, C. A. (2020). Enhancing cation exchange capacity of weathered soils using biochar: Feedstock, pyrolysis conditions and addition rate. *Agronomy*, 10(6), 824. <https://doi.org/10.3390/agronomy10060824>
904. Feng, J., Yang, Y., Ruan, K., Wu, D., Xu, Y., Jacobs, D. F., & Zeng, S. (2023). Combined application of sewage sludge, bagasse, and molybdenum tailings ameliorates rare earth mining wasteland soil. *Journal of Soils and Sediments*, 23(4), 1775–1788. <https://doi.org/10.1007/s11368-023-03437-7>
905. Guerrini, I. A., Sampaio, T. F., Bogiani, J. C., Backes, C., Harrison, R. B., Oliveira, F. C., Gava, J. L., Traballi, R. C., Garuba De Menezes Mota, R., Roder, L. R., Grilli, E., Ganga, A., James, J. N., & Capra, G. F. (2021). Sewage sludge as a pedotechnomaterial for the recovery of soils compacted by heavy machinery on *Eucalyptus* commercial plantation. *Journal of Cleaner Production*, 325, 129320. <https://doi.org/10.1016/j.jclepro.2021.129320>
906. Hussain, M. S., Naeem, M. S., Tanvir, M. A., Nawaz, M. F., & Abd-Elrahman, A. (2021). Eco-physiological evaluation of multipurpose tree species to ameliorate saline soils. *International Journal of Phytoremediation*, 23(9), 969–981. <https://doi.org/10.1080/15226514.2020.1871321>
907. Krebsbach, S., He, J., Adhikari, S., Olshansky, Y., Feyzbar, F., Davis, L. C., Oh, T.-S., & Wang, D. (2023). Mechanistic understanding of perfluorooctane sulfonate (PFOS) sorption by biochars. *Chemosphere*, 330, 138661. (cross-listed with *Populus*) <https://doi.org/10.1016/j.chemosphere.2023.138661>
908. Olmedo, G. F., Guevara, M., Gilabert, H., Montes, C. R., Arellano, E. C., Barría-Knopf, B., Gárate, F., Mena-Quijada, P., Acuña, E., Bown, H. E., & Ryan, M. G. (2020). Baseline of carbon stocks in *Pinus radiata* and *Eucalyptus* spp. Plantations of Chile. *Forests*, 11(10), Article 10. <https://doi.org/10.3390/f11101063>
909. Rockwood, D. L., Ellis, M. F., & Fabbro, K. W. (2022). Economic potential for carbon sequestration by short rotation eucalypts using biochar in Florida, USA. *Trees, Forests and People*, 7, 100187. <https://doi.org/10.1016/j.tfp.2021.100187>
910. Sanders, A. R., Nielsen, E. A., & Abrams, J. (2021). Does information and deliberation affect ecosystem service judgments? Evidence from tree plantation expansion in Argentina. *Society & Natural Resources*, 34(11), 1433–1448. <https://doi.org/10.1080/08941920.2021.1961957>
911. Steenhuis, T. S., Alemie, T. C., Muche, H., Tilahun, S. A., Zimale, F. A., & Mhiret, D. A. (2023). Effect of eucalyptus on blue and green water availability and discharge in the tropical highlands: An interpretation of available literature. *Journal of Hydrology and Hydromechanics*, 71(3), 221–230. <https://doi.org/10.2478/johh-2023-0020>
912. Visscher, A. M., Vanek, S., Meza, K., De Goede, R. G. M., Valverde, A. A., Ccanto, R., Olivera, E., Scurrah, M., & Fonte, S. J. (2020). Eucalyptus and alder field margins differ in their impact on

ecosystem services and biodiversity within cropping fields of the Peruvian Andes. *Agriculture, Ecosystems & Environment*, 303, 107107. <https://doi.org/10.1016/j.agee.2020.107107>

913. Visscher, A. M., Vanek, S., Meza, K., Wellstein, C., Zerbe, S., Ccanto, R., Olivera, E., Huaraca, J., Scurrah, M., & Fonte, S. J. (2023). Tree-based land uses enhance the provision of ecosystem services in agricultural landscapes of the Peruvian highlands. *Agriculture, Ecosystems & Environment*, 342, 108213. <https://doi.org/10.1016/j.agee.2022.108213>
914. Wolski, P., Blankenship, B. W., Umar, A., Cabrera, M., Simmons, B. A., Sale, K. L., & Achinivu, E. C. (2023). Factors that influence the activity of biomass-degrading enzymes in the presence of ionic liquids—A review. *Frontiers in Energy Research*, 11, 1212719. (cross-listed with *Populus* and *Salix*) <https://doi.org/10.3389/fenrg.2023.1212719>
915. Yao, X., Hui, D., Hou, E., Xiong, J., Xing, S., & Deng, Q. (2023). Differential responses and mechanistic controls of soil phosphorus transformation in *Eucalyptus* plantations with N fertilization and introduced N₂-fixing tree species. *New Phytologist*, 237(6), 2039–2053. <https://doi.org/10.1111/nph.18673>
916. Zhang, J., Li, J., Ma, L., He, X., Liu, Z., Wang, F., Chu, G., & Tang, X. (2022). Accumulation of glomalin-related soil protein benefits soil carbon sequestration: Tropical coastal forest restoration experiences. *Land Degradation & Development*, 33(10), 1541–1551. <https://doi.org/10.1002/ldr.4192>

Short Rotation Woody Crops

Taxonomy, Nomenclature, and Registration

None reported.

Domestication, Breeding, and Selection of Genetic Resources

917. Devireddy, A. R., Tschaplinski, T. J., Tuskan, G. A., Muchero, W., & Chen, J.-G. (2021). Role of reactive oxygen species and hormones in plant responses to temperature changes. *International Journal of Molecular Sciences*, 22(16), 8843. <https://doi.org/10.3390/ijms22168843>
918. Nelson, C. D., Crocker, E. V., DeWald, L., Raley, E. M., Adams, J. P., Crane, B. S., Peter, G. F., Casola, C., Davis, J. M., & Merkle, S. A. (2020). Report on the Thirty-Fifth Southern Forest Tree Improvement Conference (SFTIC 2019). *Tree Genetics & Genomes*, 16(2), 31. <https://doi.org/10.1007/s11295-020-1423-9>

Plant Health, Resilience to Threats and Climate Change

919. Calvin, K., Cowie, A., Berndes, G., Arneth, A., Cherubini, F., Portugal-Pereira, J., Grassi, G., House, J., Johnson, F. X., Popp, A., Rounsevell, M., Slade, R., & Smith, P. (2021). Bioenergy for climate change mitigation: Scale and sustainability. *GCB Bioenergy*, 13(9), 1346–1371. <https://doi.org/10.1111/gcbb.12863>

Production Systems for the Bioeconomy

920. Ayoub, N., Costello, C., & Jose, S. (2021). Systematic application of a quantitative definition of marginal lands in estimating biomass energy potential in the Missouri/Mississippi River Corridor. *Biofuels*, 12(7), 839–852. <https://doi.org/10.1080/17597269.2018.1554945>
921. Bhatia, S. K., Jagtap, S. S., Bedekar, A. A., Bhatia, R. K., Patel, A. K., Pant, D., Rajesh Banu, J., Rao, C. V., Kim, Y.-G., & Yang, Y.-H. (2020). Recent developments in pretreatment technologies on lignocellulosic biomass: Effect of key parameters, technological improvements, and challenges. *Bioresource Technology*, 300, 122724. <https://doi.org/10.1016/j.biortech.2019.122724>
922. Carpita, N. C., & McCann, M. C. (2020). Redesigning plant cell walls for the biomass-based bioeconomy. *Journal of Biological Chemistry*, 295(44), 15144–15157. <https://doi.org/10.1074/jbc.REV120.014561>
923. Carter, D. R., Albaugh, T. J., Campoe, O. C., Grossman, J. J., Rubilar, R. A., Sumnall, M., Maier, C. A., Cook, R. L., & Fox, T. R. (2020). Complementarity increases production in genetic mixture of loblolly pine (*Pinus taeda* L.) throughout planted range. *Ecosphere*, 11(11), e03279. <https://doi.org/10.1002/ecs2.3279>
924. Carvalho, H. D. R., Aguilos, M. M., Ile, O. J., Howard, A. M., King, J. S., & Heitman, J. L. (2023). Water use of short-rotation coppice American sycamore (*Platanus occidentalis* L.) for bioenergy during establishment on marginal land in the North Carolina Piedmont. *Agricultural Water Management*, 276, 108071. <https://doi.org/10.1016/j.agwat.2022.108071>
925. Consalter, R., Motta, A. C. V., Barbosa, J. Z., Vezzani, F. M., Rubilar, R. A., Prior, S. A., Nisgoski, S., & Bassaco, M. V. M. (2021). Fertilization of *Pinus taeda* L. On an acidic oxisol in southern Brazil: Growth, litter accumulation, and root exploration. *European Journal of Forest Research*, 140(5), 1095–1112. <https://doi.org/10.1007/s10342-021-01390-z>
926. Ferreira, G. W. D., Rau, B. M., & Aubrey, D. P. (2020). Herbicide, fertilization, and planting density effects on intensively managed loblolly pine early stand development. *Forest Ecology and Management*, 472, 118206. <https://doi.org/10.1016/j.foreco.2020.118206>
927. Ferreira, G. W. D., Rau, B. M., & Aubrey, D. P. (2021). Temporal nitrogen dynamics in intensively managed loblolly pine early stand development. *Forest Ecology and Management*, 483, 118890. <https://doi.org/10.1016/j.foreco.2020.118890>
928. Hossain, T., Jones, D. S., Hartley, D. S., Thompson, D. N., Langholtz, M., & Davis, M. (2022). Nth-plant scenario for forest resources and short rotation woody crops: Biorefineries and depots in the contiguous US. *Applied Energy*, 325, 119881. <https://doi.org/10.1016/j.apenergy.2022.119881>
929. Ile, O. J., Aguilos, M., Morkoc, S., Heitman, J., & King, J. S. (2021). Root biomass distribution and soil physical properties of short-rotation coppice American sycamore (*Platanus occidentalis* L.) grown at different planting densities. *Forests*, 12(12), 1806. <https://doi.org/10.3390/f12121806>
930. Ile, O. J., Aguilos, M., Morkoc, S., Minick, K., Domec, J.-C., & King, J. S. (2021). Productivity of low-input short-rotation coppice American sycamore (*Platanus occidentalis* L.) grown at

different planting densities as a bioenergy feedstock over two rotation cycles. *Biomass and Bioenergy*, 146, 105983. <https://doi.org/10.1016/j.biombioe.2021.105983>

931. Ile, O., Typhina, E., Brannum, K., Parajuli, R., Bardon, R., & King, J. (2023). Toward the adoption of new farming systems among farmers: A case study of short rotation woody crops in North Carolina. *Journal of Extension*, 61(1). <https://doi.org/10.34068/joe.61.01.09>
932. Loy, X., Gruenewald, D., Gottlieb, I. G. W., Dobbs, E. K., Miljanic, A. S., Botsch, J., Lawley, B., Ober, H. K., Smith, L., Fletcher, R. J., & Brosi, B. J. (2020). The impacts of bioenergy pine plantation management practices on bee communities. *Journal of Applied Ecology*, 57(5), 952–962. <https://doi.org/10.1111/1365-2664.13586>
933. Morkoc, S., Aguilos, M., Noormets, A., Minick, K. J., Ile, O., Dickey, D. A., Hardesty, D., Kerrigan, M., Heitman, J., & King, J. (2022). Environmental and plant-derived controls on the seasonality and partitioning of soil respiration in an american sycamore (*Platanus occidentalis*) bioenergy plantation grown at different planting densities. *Forests*, 13(8), 1286. <https://doi.org/10.3390/f13081286>
934. Steele, J. K., Aust, W. M., & Seiler, J. R. (2021). Effects of mechanical site preparation, planting stock, and planting aids on the survival and growth of American sycamore in a marginal old field riparian restoration. *Forests*, 12(10), 1295. <https://doi.org/10.3390/f12101295>
935. Trlica, A., Cook, R. L., Albaugh, T. J., Parajuli, R., Carter, D. R., & Rubilar, R. A. (2021). Financial returns for biomass on short-rotation loblolly pine plantations in the southeastern United States. *Forest Science*, 67(6), 670–681. <https://doi.org/10.1093/forsci/fxab033>
936. Vache, K., Meles, M. B., Griffiths, N. A., & Jackson, C. R. (2021). Ensemble modeling of watershed-scale hydrologic effects of short-rotation woody crop production. *Biofuels, Bioproducts and Biorefining*, 15(5), 1345–1359. <https://doi.org/10.1002/bbb.2247>

Environmental and Ecosystem Services

937. Ferreira, G. W. D., & Aubrey, D. P. (2023). A functional trait framework for integrating nitrogen-fixing cover crops into short-rotation woody crop systems. *GCB Bioenergy*, 15(5), 663–679. <https://doi.org/10.1111/gcbb.13045>
938. Hou, D., O'Connor, D., Igalavithana, A. D., Alessi, D. S., Luo, J., Tsang, D. C. W., Sparks, D. L., Yamauchi, Y., Rinklebe, J., & Ok, Y. S. (2020). Metal contamination and bioremediation of agricultural soils for food safety and sustainability. *Nature Reviews Earth & Environment*, 1(7), 366–381. <https://doi.org/10.1038/s43017-020-0061-y>
939. Pisarello, K. L., Sun, G., Evans, J. M., & Fletcher, R. J. (2022). Potential long term water yield impacts from pine plantation management strategies in the southeastern United States. *Forest Ecology and Management*, 522, 120454. <https://doi.org/10.1016/j.foreco.2022.120454>
940. White, J. C., & Smith, W. K. (2020). Water source utilization under differing surface flow regimes in the riparian species *Liquidambar styraciflua*, in the southern Appalachian foothills, USA. *Plant Ecology*, 221(11), 1069–1082. <https://doi.org/10.1007/s11258-020-01062-9>
941. Zalesny, R. S., & Pilipović, A. (2022). Growth and development of short-rotation woody crops for rural and urban applications. *Forests*, 13(6), 867. <https://doi.org/10.3390/f13060867>

IV. SUMMARY STATISTICS, 2020-2023

Questionnaire on Poplars and Other Fast-Growing Trees Sustaining People and the Environment

Contact Information	
Country	USA
Contact Person	Ronald S. Zalesny Jr., USDA Forest Service
Position of Contact Person	Supervisory Research Plant Geneticist
Email	ronald.zalesny@usda.gov
Telephone	+1 715-362-1132

Terms and Definitions

The main FAO categories of land with a tree component are classified according to the terms and definitions of the [Global Forest Resources Assessment \(2020\)](#).

Naturally regenerating forest: Forest predominantly composed of trees established through natural regeneration. *Explanatory notes:* 1) Includes forests for which it is not possible to distinguish whether planted or naturally regenerated. 2) Includes forests with a mix of naturally regenerated tree species and planted/seeded trees, and where the naturally regenerated trees are expected to constitute the major part of the growing stock at stand maturity. 3) Includes coppice from trees originally established through natural regeneration. 4) Includes naturally regenerated trees of introduced species.

Planted forest: Forest predominantly composed of trees established through planting and/or deliberate seeding.

Explanatory notes: 1) In this context, predominantly means that the planted/seeded trees are expected to constitute more than 50 percent of the growing stock at maturity. 2) Includes coppice from trees that were originally planted or seeded.

Plantation forest (subcategory of planted forest): Planted Forest that is intensively managed and meet ALL the following criteria at planting and stand maturity: one or two species, even age class, and regular spacing.

Explanatory notes: 1) Specifically includes: short rotation plantation for wood, fibre and energy. 2) Specifically excludes: forest planted for protection or ecosystem restoration. 3) Specifically excludes: forest established through planting or seeding which at stand maturity resembles or will resemble naturally regenerating forest.

Agroforestry: “Other land with tree cover” with temporary agricultural crops and/or pastures/animals.

Explanatory notes: 1) Includes areas with bamboo and palms provided that land use, height and canopy cover criteria are met. 2) Includes agrisilvicultural, silvopastoral and agrosilvopastoral systems.

Trees in urban settings: “Other land with tree cover” such as: urban parks, alleys, and gardens.

Forest: Land spanning more than 0.5 hectares with trees higher than 5 meters and a canopy cover of more than 10 percent, or trees able to reach these thresholds *in situ*. It does not include land that is predominantly under agricultural or urban land use. *Explanatory notes:* 1) Forest is determined both by the presence of trees and the absence of other predominant land uses. The trees should be able to reach a minimum height of 5 meters *in situ*. 2) Includes areas with young trees that have not yet reached but which are expected to reach a canopy cover of 10 percent and tree height of 5 meters. It also includes areas that are temporarily unstocked due to clear-cutting as part of a forest management practice or natural disasters, and which are expected to be regenerated within 5 years. Local conditions may, in exceptional cases, justify that a longer time frame is used. 3) Includes forest roads, firebreaks and other small open areas; forest in national parks, nature reserves and other protected areas such as those of specific environmental, scientific, historical, cultural or spiritual interest. 4) Includes windbreaks, shelterbelts and corridors of trees with an area of more than 0.5 hectares and width of more than 20 meters. 5) Includes abandoned shifting cultivation land with a regeneration of trees that have, or are expected to reach, a canopy cover of 10 percent and tree height of 5 meters. 6) Includes areas with mangroves in tidal zones, regardless whether this area is classified as land area or not. 7) Includes rubber-wood, cork oak and Christmas tree plantations. 8) Includes areas with bamboo and palms provided that land use, height and canopy cover criteria are met. 9) Includes areas outside the legally designated forest land which meet the definition of “forest”. 10) Excludes tree stands in agricultural production systems, such as fruit tree plantations, oil palm plantations, olive orchards and agroforestry systems when crops are grown under tree cover. Note: Some agroforestry systems such as the “Taungya” system where crops are grown only during the first years of the forest rotation should be classified as forest.

Other land with tree cover: Land classified as “remaining land area”, spanning more than 0.5 hectares with a canopy cover of more than 10 percent of trees able to reach a height of 5 meters at maturity. *Explanatory notes:* 1) **Land use is the key criteria for distinguishing between forest and other land with tree cover.** 2) Specifically includes: palms (oil, coconut, dates, etc), tree orchards (fruit, nuts, olive, etc), agroforestry and trees in urban settings. 3) Includes groups of trees and scattered trees (e.g., trees outside forest) in agricultural landscapes, parks, gardens and around buildings, provided that area, height, and canopy cover criteria are met. 4) Includes tree stands in agricultural production systems, such as fruit tree plantations/orchards. In these cases, the height threshold can be lower than 5 meters. 5) Includes agroforestry systems when crops are grown under tree cover and tree plantations established mainly for other purposes than wood, such as oil palm plantations. 6) The different sub-categories of “other land with tree cover” are exclusive and area reported under one subcategory should not be reported for any other sub-categories. 7) Excludes scattered trees with a canopy cover less than 10 percent, small groups of trees covering less than 0.5 hectares and tree lines less than 20 meters wide.

Table 1. Total area (ha) of poplars, willows, and eucalypts in 2023 in the US (see Country Notes), as well as area planted (ha) from 2020-2023.

Land use category / genus	Total area (2023; ha)	Total area by forest function (%)				Area planted from 2020 to 2023 (ha)
		Production		Protection (%)	Other (%)	
		Industrial roundwood (%)	Fuelwood biomass (%)			
Naturally Regenerating Forest						
Poplars	9,296,159			7.6	92.4	N/A
Willows	440,191			6.4	93.6	N/A
Mix of P&W	218,905			4.8	95.2	N/A
Eucalypts	Unavailable			Unavailable		N/A
Planted Forest						
Poplars	142,394			2.2	97.8	
Willows	5,658			32.0	68.0	
Mix of P&W	16,748			0.0	100.0	
Eucalypts	Unavailable			Unavailable		
Plantations						
Poplars						
Willows						
Mix of P&W						
Eucalypts						
Other Planted Forest						
Poplars						
Willows						
Mix of P&W						
Eucalypts						
Other Land with Tree Cover (Agroforestry)						
Poplars						
Willows						
Mix of P&W						
Eucalypts						
Other Land with Tree Cover (Trees in Urban Settings)						
Poplars						
Willows						
Mix of P&W						
Eucalypts						
Grand Total	10,120,055					

Country Notes: "Other" is forest land available for timber production but not necessarily intended for that purpose. In other words, it is not reserved nor protected, but the owner of the land may not plan to use the trees for industrial purposes. Citation: USDA Forest Service, Forest Inventory and Analysis Program, Wednesday, 29 May 2024 12:37:57 GMT. Forest Inventory EVALIDator web-application Version 2.1.2. St. Paul, MN: U.S. Department of Agriculture, Forest Service, Northern Research Station. [Available only on internet: <https://apps.fs.usda.gov/fiadb-api/evalidator>]. The state of Alaska, which has a considerable poplar and willow resource, is excluded from these statistics due to lack of data.

Table 2. Wood removals (m³) of poplars, willows, and eucalypts in 2023 in the US (see Country Notes).

Land use category / genus	Species	Total Removals	Industrial roundwood			Fuelwood / wood chips
			Veneer / Plywood	Pulpwood	Sawnwood	
Naturally Regenerating Forest						
Poplars ¹						
SPCD 741 – Balsam Poplar	<i>P. balsamifera</i> ²	339,127.03				
SPCD 742 – Eastern Cottonwood	<i>P. deltoides</i>	523,811.15				
SPCD 743 – Bigtooth Aspen	<i>P. grandidentata</i>	2,166,649.83				
SPCD 744 – Swamp Cottonwood	<i>P. heterophylla</i>	72,708.44				
SPCD 745 – Plains Cottonwood	<i>P. deltoides</i>	9,305.70				
SPCD 746 – Quaking Aspen	<i>P. tremuloides</i>	7,517,427.18				
SPCD 747 – Black Cottonwood	<i>P. balsamifera</i>	61,867.66				
SPCD 748 – Fremont Cottonwood	<i>P. fremontii</i>	8,517.09				
Willows ³						
SPCD 920 – Willow spp.	<i>S. spp.</i>	2,189.78				
SPCD 921 – Peachleaf Willow	<i>S. amygdaloides</i>	1,415.46				
SPCD 922 – Black Willow	<i>S. nigra</i>	78,879.04				
SPCD 927 – White Willow	<i>S. alba</i>	491.28				
Mix of P&W		Unavailable				
Eucalypts		Unavailable				
Planted Forest						
Poplars						
SPCD 741 – Balsam Poplar	<i>P. balsamifera</i>	8,521.79				
SPCD 742 – Eastern Cottonwood	<i>P. deltoides</i>	170,492.47				
SPCD 743 – Bigtooth Aspen	<i>P. grandidentata</i>	44,850.66				
SPCD 746 – Quaking Aspen	<i>P. tremuloides</i>	308,061.40				
SPCD 747 – Black Cottonwood	<i>P. balsamifera</i>	91,197.95				
Willows						
SPCD 922 – Black Willow	<i>S. nigra</i>	20,781.45				
SPCD 925 – Coastal Plain Willow	<i>S. caroliniana</i>	18,859.13				
Mix of P&W		Unavailable				
Eucalypts		Unavailable				
Other Land with Tree Cover (Agroforestry)						
Poplars						
Willows						
Mix of P&W						
Eucalypts						
Grand Total		11,445,154.49				

¹ *Populus balsamifera* L.; *Populus deltoides* Bart. ex Marsh.; *Populus fremontii* S. Watson; *Populus grandidentata* Michx.; *Populus heterophylla* L.; *Populus tremuloides* Michx. | ² The US Forest Inventory and Analysis categorizes *P. trichocarpa* as *P. balsamifera* ssp. *trichocarpa*, thus numbers for *P. balsamifera* are inclusive of *P. trichocarpa*. | ³ *Salix alba* L.; *Salix amygdaloides* Andersson; *S. caroliniana* Michx.; *Salix nigra* Marsh. | Country Notes: Data were obtained from the US Forest Inventory and Analysis (FIA) program and represent the most current inventory information from each state in the United States. Some states may be missing information (particularly Alaska) and removals should be viewed as approximate. Removals data include harvest removals only, not removals due to land conversion. Species with fewer than 10 observations were omitted because of high standard error values. Removals values are **AVERAGE ANNUAL REMOVALS**.

Table 3. Forest products in roundwood equivalents (1,000 m³ r) in 2023 in the US (see Country Notes).

Land use category / genus	Wood fuel	Wood chips / particles	Sawnwood	Veneer sheets	Plywood	Particle board	Fibreboard	Wood pulp
Naturally Regenerating Forest								
	----- Roundwood equivalents (1,000 m ³ r) -----							
Poplars								
Willows								
Mix of P&W								
Eucalypts								
Planted Forest								
	----- Roundwood equivalents (1,000 m ³ r) -----							
Poplars								
Willows								
Mix of P&W								
Eucalypts								
Other Land with Tree Cover (Agroforestry)								
	----- Roundwood equivalents (1,000 m ³ r) -----							
Poplars								
Willows								
Mix of P&W								
Eucalypts								
Grand Total								

Country Note: These data are not available for the current reporting period.

Table 4. Prevailing trends until 2030 in the development of poplars, willows, and eucalypts in the United States.

	Increase	Decrease	Remain as is	No comment
1a. The conversion of naturally regenerating forests of poplar to other land uses will...	X			
1b. The conversion of naturally regenerating forests of willow to other land uses will...	X			
1c. The conversion of naturally regenerating forests of eucalypts to other land uses will...				X
2a. The conversion of planted forests of poplar to other land uses will...			X	
2b. The conversion of planted forests of willow to other land uses will...			X	
2c. The conversion of planted forests of eucalypts to other land uses will...			X	
3a. The conversion of planted forests of poplar to other species will...			X	
3b. The conversion of planted forests of willow to other species will...			X	
3C. The conversion of planted forests of eucalypts to other species will...				X
4a. The area of poplars for bioenergy plantations will ...			X	
4b. The area of willows for bioenergy plantations will ...	X			
4c. The area of eucalypts for bioenergy plantations will ...				X
5a. Government investments in poplars will ...				X
5b. Government investments in willows will ...				X
5c. Government investments in eucalypts will ...				X
6a. Private sector investments in poplars will ...				X
6b. Private sector investments in willows will ...				X
6c. Private sector investments in eucalypts will ...				x
7a. The significance of poplars for productive purposes will ...	X			
7b. The significance of willows for productive purposes will ...	X			
7c. The significance of eucalypts for productive purposes will ...			X	
8a. The significance of poplars for environmental protection purposes will ...	X			
8b. The significance of willows for environmental protection purposes will ...	X			

Table 4. Prevailing trends until 2030 in the development of poplars, willows, and eucalypts in the United States.

	Increase	Decrease	Remain as is	No comment
8c. The significance of eucalypts for environmental protection purposes will ...	X			
9a. The rejection by environmental groups of poplars will...				X
9b. The rejection by environmental groups of willows will...				X
9c. The rejection by environmental groups of eucalypts will...				X
10a. The acceptance by the general public of poplars as important natural resources will.....				X
10b. The acceptance by the general public of willows as important natural resources will.....				X
10c. The acceptance by the general public of eucalypts as important natural resources will.....				X
11a. The introduction of poplars in agroforestry systems will.....	X			
11b. The introduction of willows in agroforestry systems will.....	X			
11c. The introduction of eucalypts in agroforestry systems will.....				X

Country Note: The opinions expressed in this table are those of the authors and should not be construed to represent any official USDA or U.S. Government determination or policy.